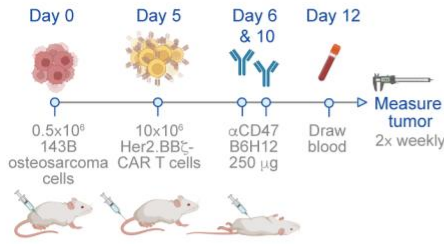

Supplementary information

Engineered CD47 protects T cells for enhanced antitumour immunity

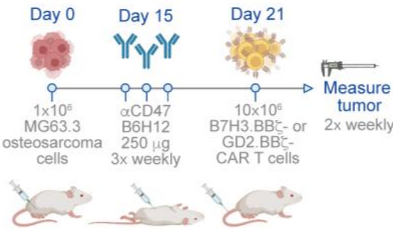
In the format provided by the authors and unedited

Supplementary Fig. 1 | Method schematics for *in vivo* studies

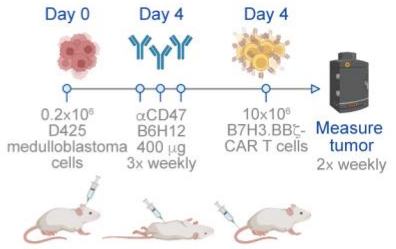
a Her2-CAR + α CD47 vs 143B Osteosarcoma
Fig. 1a | Extended Data Fig. 1a,h



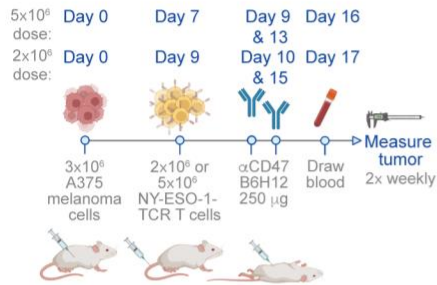
b B7H3- or GD2-CAR + α CD47 vs MG63.3 Osteosarcoma
Fig. 1b | Extended Data Fig. 1b



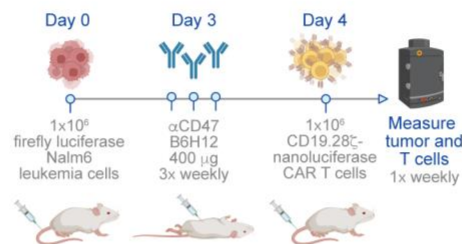
c B7H3-CAR + α CD47 vs Medulloblastoma
Extended Data Fig. 1c,d



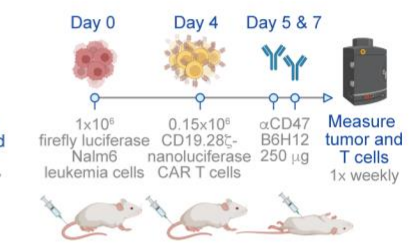
d NY-ESO-1-TCR T + α CD47 vs Melanoma
Fig. 1d,e | Extended Data Fig. 1i-k



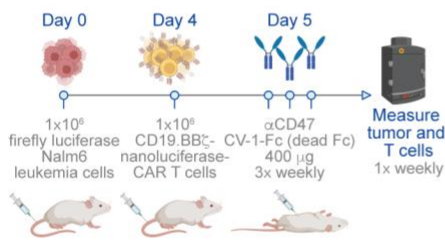
e High dose CD19-CAR + α CD47 vs Leukemia
Extended Data Fig. 1l,n,p,s



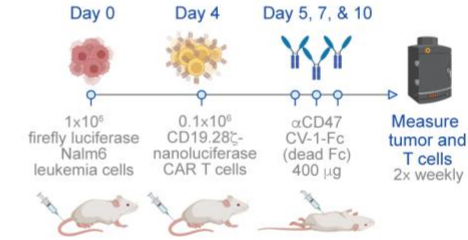
f Low dose CD19-CAR + α CD47 vs Leukemia
Extended Data Fig. 1m-o,q,r



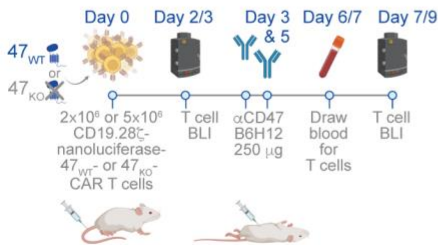
g CD19-CAR + α CD47 CV-1 vs Leukemia
Fig. 1f,g | Extended Data Fig. 2a,b,e



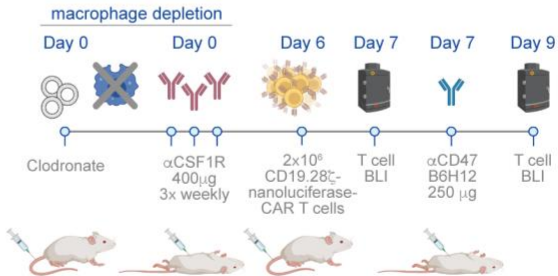
h Low dose CD19-CAR + α CD47 CV-1 vs Leukemia
Extended Data Fig. 2c-f



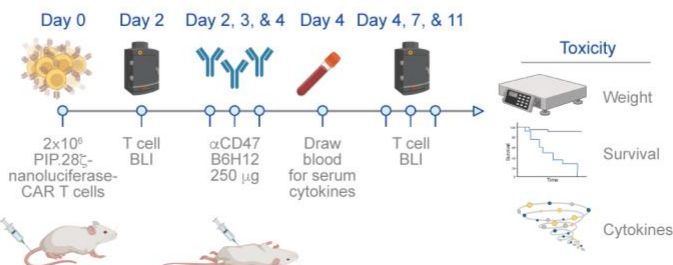
i 47_{KO}-CD19-CAR vs α CD47
Extended Data Fig. 2j-m

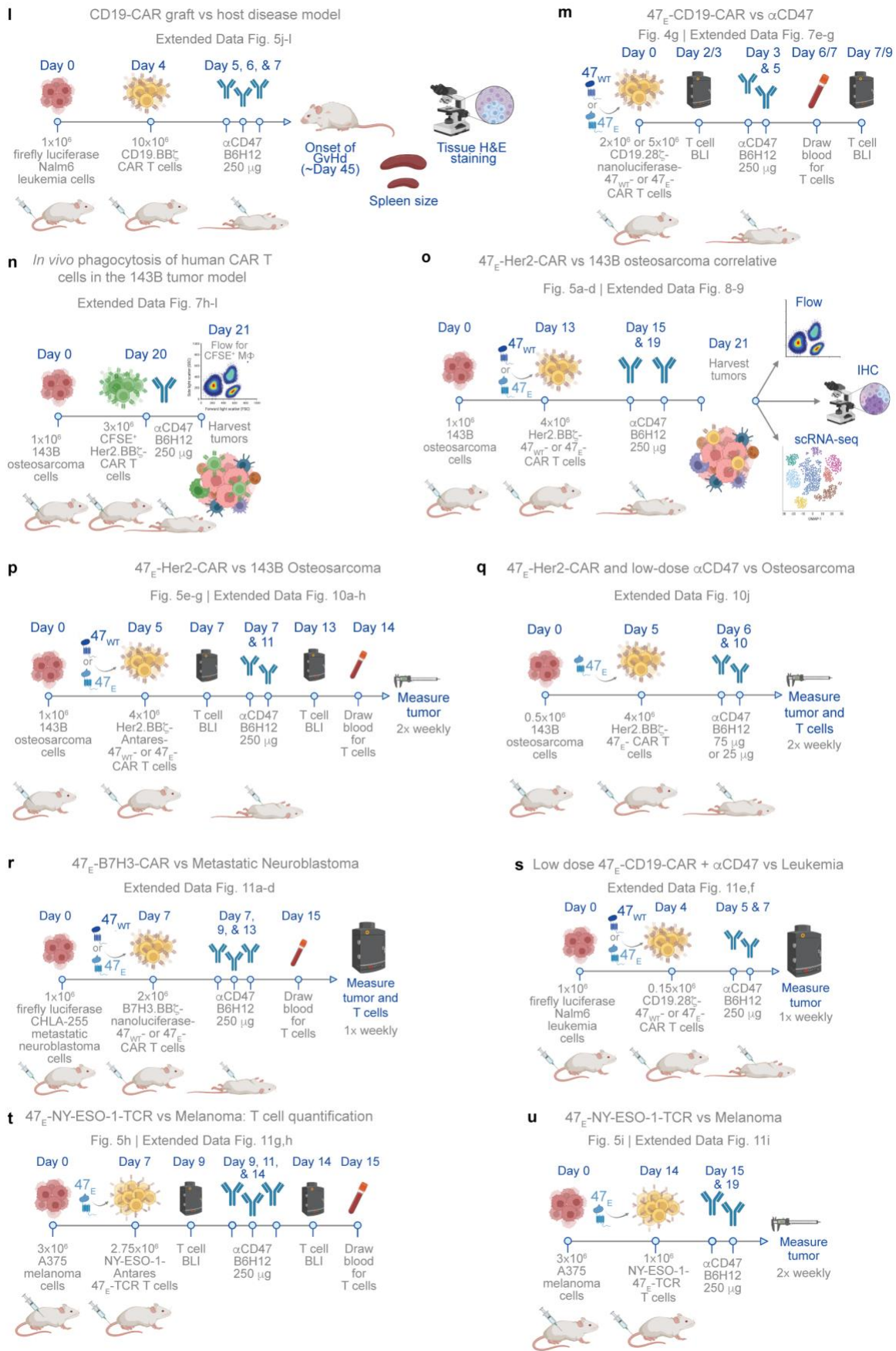


j CD19-CAR vs macrophage depletion + α CD47
Fig. 1l,m | Extended Data Fig. 3p-r



k PIP-CAR toxicity model vs α CD47
Fig. 3c-f





Supplementary Fig. 1 | Method schematics for *in vivo* studies.

(a) 143B model treatment scheme. Mice engrafted orthotopically in the tibia periosteum with 0.5×10^6 143B were treated intravenously (IV) with 10×10^6 Her2.BB ζ -CAR-T cells on day 5, and then intraperitoneally (IP) \pm B6H12 twice on days 6 and 10 (250 μ g/dose). Blood was drawn on day 12 to assess CAR-T cell expansion.

(b) MG63.3 model treatment scheme. Mice engrafted orthotopically in the tibia periosteum with 1×10^6 MG63.3 were treated IP \pm B6H12 three times per week (400 μ g/dose) starting on day 15, then IV with 10×10^6 B7H3.BB ζ - or GD2.BB ζ -CAR-T cells on day 21.

(c) D425 treatment scheme. Mice engrafted with 0.2×10^6 D425 cells in the cerebellum were treated \pm B6H12 IP three times per week (400 μ g/dose) starting on day 4. Mice were also treated IV with 10×10^6 CD19.BB ζ - (non-tumor targeting control) or B7H3.BB ζ - (tumor targeting) CAR-T cells on day 4.

(d) A375 model treatment scheme. Mice engrafted subcutaneously (SQ) with 3×10^6 A375 were treated IV with two different doses of mock or NY-ESO-1-TCR-T cells. The 2×10^6 dose was administered on day 9 \pm two doses of B6H12 (250 μ g/dose; IP) on days 10 and 15. Blood was drawn on day 17 to assess T cell expansion. The 5×10^6 dose was administered on day 7 \pm two doses of B6H12 (250 μ g/dose; IP) on days 9 and 13. Blood was drawn on day 16 to assess T cell expansion.

(e) High-dose CAR-T – Nalm6 model treatment scheme. Mice engrafted IV with 1×10^6 Nalm6-fLuc cells were treated \pm B6H12 (400 μ g/dose; IP) three times per week starting on day 3. Mice were then treated IV with 1×10^6 mock or CD19.28 ζ -nLuc-CAR-T cells on day 4. Mice were serially imaged by BLI for both tumor growth (fLuc signal) and T cell expansion (nLuc signal).

(f) Low-dose CAR-T – Nalm6 model treatment scheme. Mice engrafted IV with 1×10^6 Nalm6-fLuc cells were then treated IV with 0.15×10^6 mock or CD19.28 ζ -nLuc-CAR-T cells on day 4. Mice were then treated \pm B6H12 (250 μ g/dose; IP) on days 5 and 7. Mice were serially imaged by BLI for both tumor growth (fLuc signal) and T cell expansion (nLuc signal).

(g) Nalm6 with high-dose CAR-T and CV-1 treatment scheme. Mice engrafted IV with 1×10^6 Nalm6-fLuc were treated IV with 1×10^6 CD19.BB ζ -nLuc-CAR-T cells on day 4, \pm CV-1 (dead Fc; 400 μ g/dose; IP) three times per week starting on day 5.

(h) Nalm6 with low-dose CAR-T and CV-1 treatment scheme. Mice engrafted IV with 1×10^6 Nalm6-fLuc were treated IV with 0.1×10^6 CD19.28 ζ -nLuc-CAR-T cells on day 4, \pm CV-1 (dead Fc; 400 μ g/dose; IP) three times on days 5, 7 and 10.

(i) 47_{KO}- vs 47_{WT}-CAR-T cell depletion scheme. Non-tumor bearing mice were treated IV with two different doses of 47_{KO}-CD19.28 ζ -nLuc-CAR-T cells, with (47_{WT}) or without (47_{KO}) CD47 exogenous expression and treated twice \pm B6H12 (250 μ g/dose; IP) on days 3 and 5. Mice dosed with 2×10^6 CAR-T cells were imaged by BLI on day 2 before and on day 9 after α CD47 treatment, and had blood drawn on day 7. Mice dosed with 5×10^6 CAR-T cells were imaged by BLI on day 3 before and on day 7 after α CD47 treatment, and had blood drawn on day 6.

(j) Macrophage depletion + CAR-T treatment scheme. Non-tumor bearing mice were treated with clodronate (200 μ L; IV) on day 0 and α CSF1R (400 μ g/dose; IP) three times per week starting on day 0 to deplete macrophages. Mice were then treated IV with 2×10^6 CD19.28 ζ -nLuc-CAR-T cells on day 6, followed by a single 250 μ g IP dose of B6H12 on day 7. Mice were imaged by BLI before (day 7) and after (day 9) α CD47 treatment.

(k) PIP-CAR toxicity model safety-switch scheme. Mice were treated IV with 2×10^6 CD19.28 ζ -nLuc- or PIP.28 ζ -nLuc-CAR-T cells \pm B6H12 treatment (250 μ g daily; IP) three times, two days later. Mice were

imaged by BLI before (day 2) and after (days 4, 7, and 11) α CD47 treatment. Blood was collected on day 4 for cytokine analysis.

(l) Scheme of CAR-T GvHD model. Mice engrafted IV with 1×10^6 Nalm6-fLuc were treated IV with a high dose of 10×10^6 CD19.BB ζ -CAR T cells on day 4 \pm three doses of B6H12 (250 μ g/dose; IP) on days 5, 6, and 7. Mice were monitored for onset of GvHD around day 45.

(m) 47_E⁻ vs 47_{WT}-CAR-T cell depletion scheme. Non-tumor bearing mice were treated IV with 5×10^6 47_{WT}- or 47_E-CD19.28 ζ -nLuc-CAR-T cells (with endogenous 47_{KO}) and treated twice \pm B6H12 (250 μ g/dose; IP) on days 3 and 5. Mice dosed with 2×10^6 CAR-T cells were imaged by BLI on day 2 before and on day 9 after α CD47 treatment, and had blood drawn on day 7. Mice dosed with 5×10^6 CAR-T cells were imaged by BLI on day 3 before and on day 7 after α CD47 treatment, and had blood drawn on day 6.

(n) Scheme of *in vivo* CAR-T phagocytosis model. Mice were engrafted orthotopically in the tibia periosteum with 1×10^6 143B cells and treated intratumorally (IT) with 3×10^6 Her2.BB ζ -CAR-T cells on day 20. Mice were co-treated \pm B6H12 (250 μ g/dose; IP) on day 20. Tumors were excised on day 21 and then analyzed via flow cytometry.

(o) Scheme of correlative mechanistic study in 143B osteosarcoma. Mice were engrafted orthotopically in the tibia periosteum with 1×10^6 143B-CD19 cells and treated IV with no T cells, or 4×10^6 mock, 47_{WT}- or 47_E-Her2.BB ζ -CAR-T cells (with endogenous 47_{KO}) on day 13. Mice were then treated \pm two doses of B6H12 (250 μ g/dose; IP) on days 15 and 19. Tumors were excised on day 21 and then analyzed via flow cytometry, IHC, and scRNA-seq.

(p) 47_E-CAR-T – 143B treatment scheme. Mice were engrafted orthotopically in the tibia periosteum with 1×10^6 143B-CD19 cells and treated IV with 4×10^6 mock-Antares, or 47_{WT}- or 47_E-Her2.BB ζ -Antares-CAR-T cells (with endogenous 47_{KO}) on day 5. Mice were then treated \pm two doses of B6H12 (250 μ g/dose; IP) on days 7 and 11. Mice were imaged by BLI before (day 7) and after (day 13) α CD47 treatment, and had blood drawn on day 14.

(q) Low dose α CD47 + 47_E-CAR-T – 143B treatment scheme. Mice were engrafted orthotopically in the tibia periosteum with 0.5×10^6 143B-CD19 cells and treated IV with 4×10^6 mock or 47_E-Her2.BB ζ -CAR-T cells (with endogenous 47_{KO}) on day 5. Mice were then treated \pm two doses of B6H12 (75 μ g [\sim 3 mg/kg] or 25 μ g [\sim 1 mg/kg] per dose; IP) on days 6 and 10.

(r) 47_E-CAR-T – CHLA-255 treatment scheme. Mice were engrafted IV with 1×10^6 CHLA-255-fLuc cells and treated IV with 2×10^6 mock-nLuc, 47_{WT}- or 47_E-B7H3.BB ζ -nLuc-CAR-T cells (with endogenous 47_{KO}) on day 7. Mice were then treated \pm three doses of B6H12 (250 μ g/dose; IP) on days 7, 9 and 13. T cells were imaged by BLI on day 14 and blood was collected on day 15.

(s) 47_E-CAR-T – Nalm6 treatment scheme. Mice engrafted IV with 1×10^6 Nalm6-fLuc cells were then treated IV with 0.15×10^6 mock or 47_{WT}- or 47_E-CD19.28 ζ -CAR-T cells on day 4. Mice were then treated \pm B6H12 (250 μ g/dose; IP) on days 5 and 7. Mice were serially imaged by BLI for tumor growth.

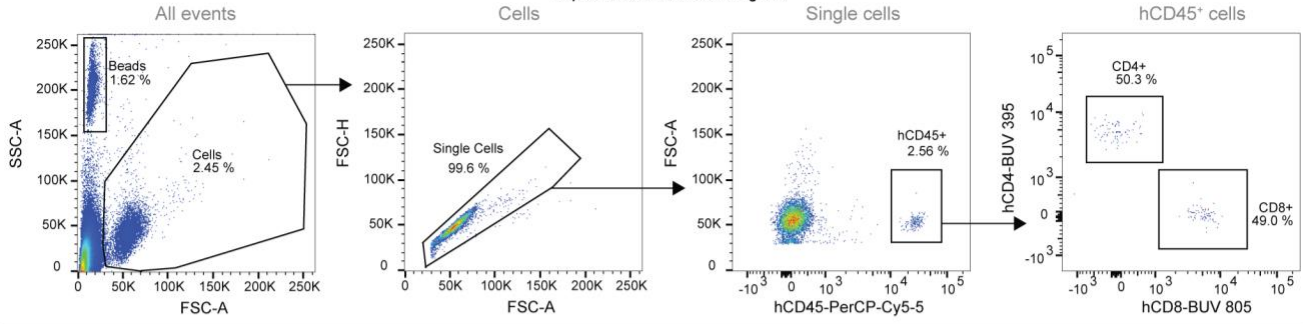
(t) Schematic of quantification of T cells in the A375 – NY-ESO-1 model. Mice engrafted SQ with 3×10^6 A375 were treated IV with 2.75×10^6 mock-Antares or 47_E-NY-ESO-1-Antares-TCR-T cells (with endogenous 47_{KO}) on day 7 \pm three doses of B6H12 (250 μ g/dose; IP) on days 9, 11, and 14. Mice were imaged by BLI before (day 9) and after (day 14) α CD47 treatment. Blood was collected on day 15.

(u) 47_E-TCR-T – A375 treatment scheme. Mice were engrafted SQ with 3×10^6 A375 cells and treated IV with 1×10^6 mock-Antares or 47_E-NY-ESO-1-Antares-TCR T cells (with endogenous 47_{KO}) on day 14. Mice were then treated \pm two doses of B6H12 (250 μ g/dose; IP) on days 15 and 19.

Supplementary Fig. 2 | Representative Flow Cytometry Gating Strategies

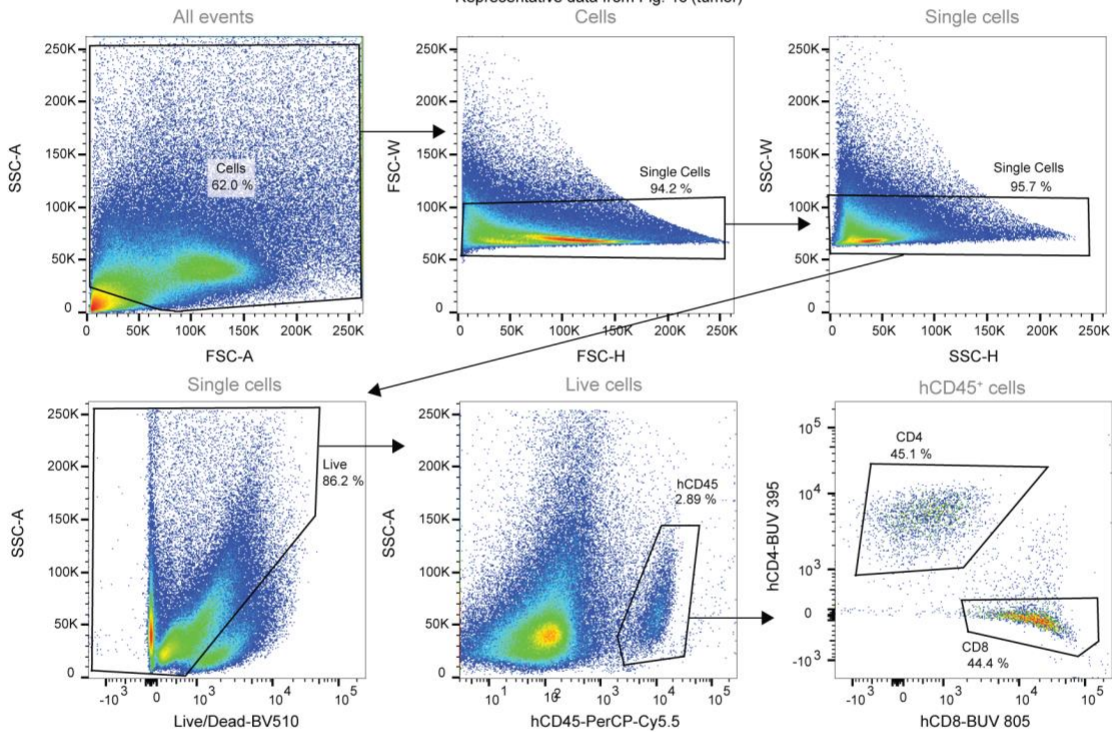
Representative gating strategy to identify human T cells from blood

Used for: Fig. 1c, 1e, 1g, 1k, 4h, 5f, and 5h | Extended Data Fig. 1e, 1f, 1g, 1h, 1k, 2j, 2m, 7g, 10a, 10b, 10c, 10i, 11a, and 11h
 Representative data from Fig. 1e



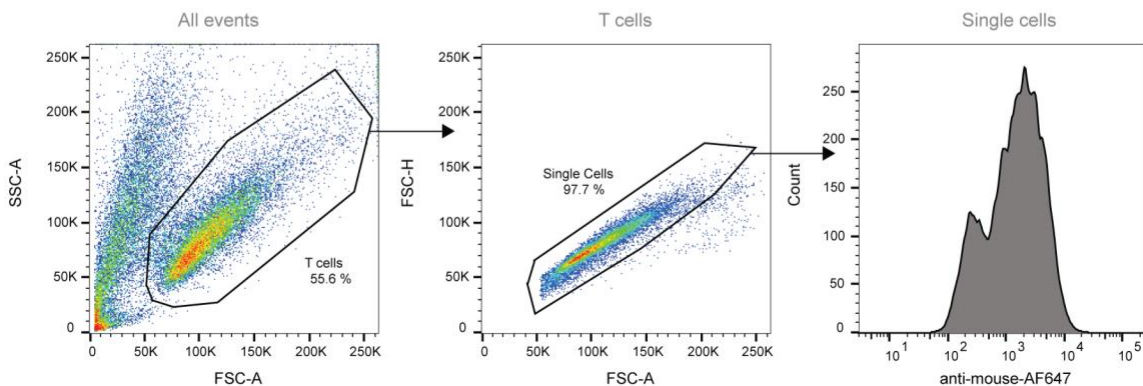
Representative gating strategy to identify human T cells from dissociated tumors and spleens

Used for: Fig. 1c | Extended Data Fig. 1e, 1s
 Representative data from Fig. 1c (tumor)



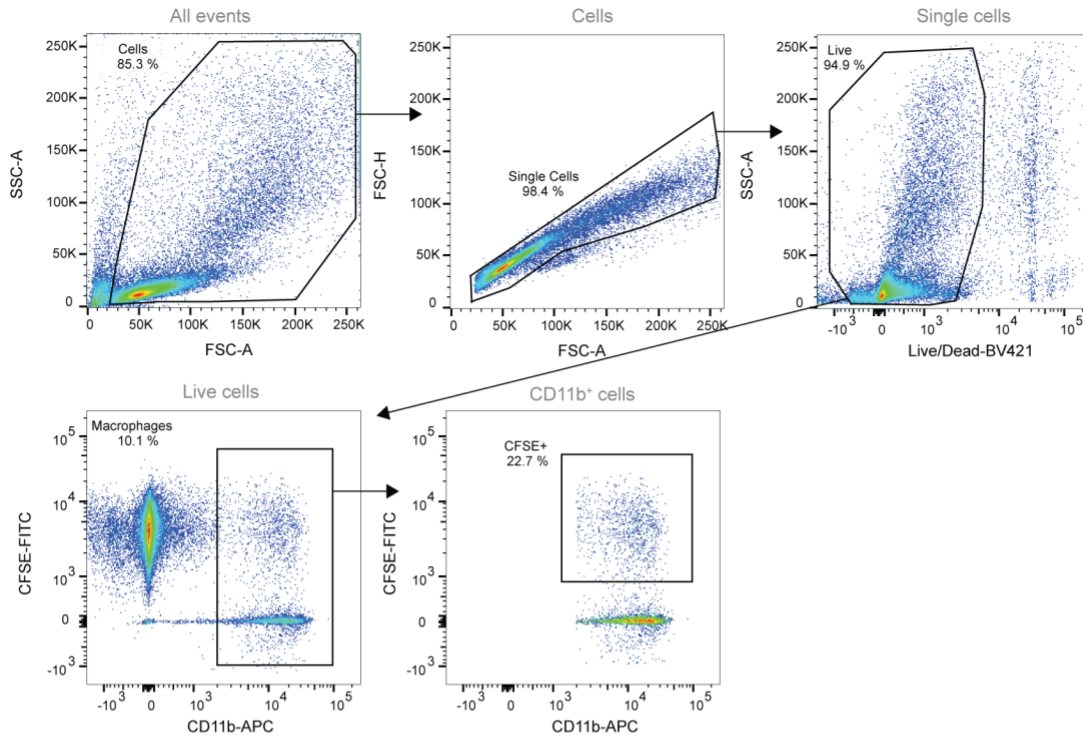
Representative gating strategy to quantify protein expression on T cells and Jurkats

Used for: Fig. 2b, 4e | Extended Data Fig. 3g-l, 3m, 4g, 4h, 5b, 7a, 7b
 Representative data from Fig. 4e



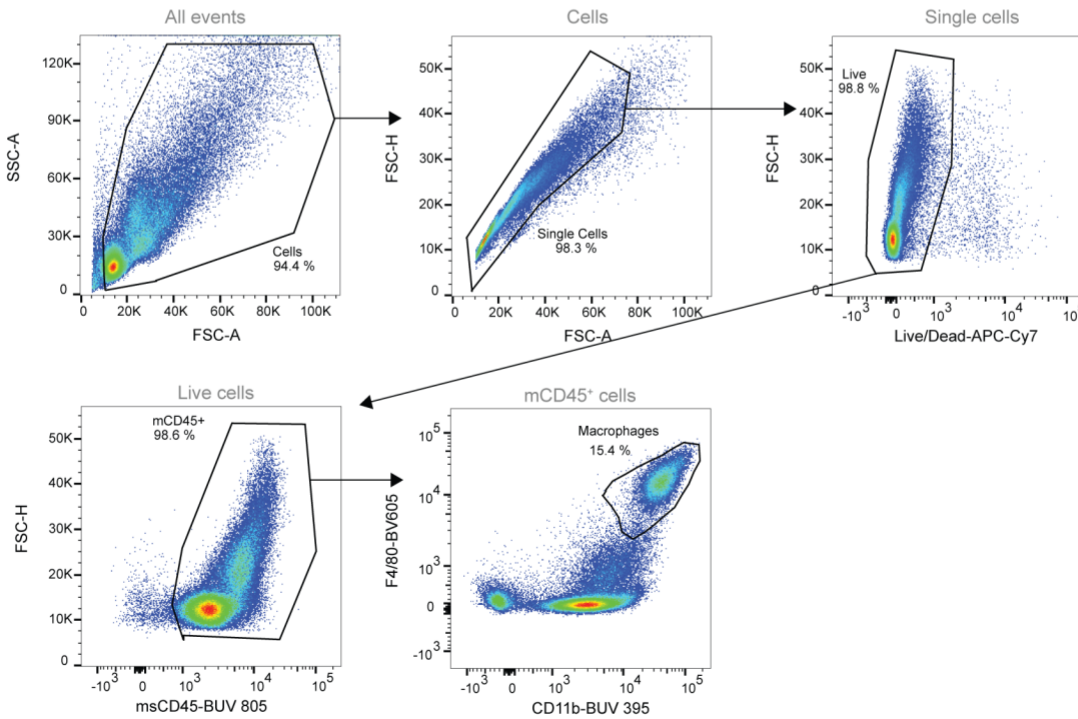
Representative gating strategy for phagocytosis assays

Used for: Fig. 2a and 4f | Extended Data Fig. 4a and 7c
 Representative data from Extended Data Fig. 7c



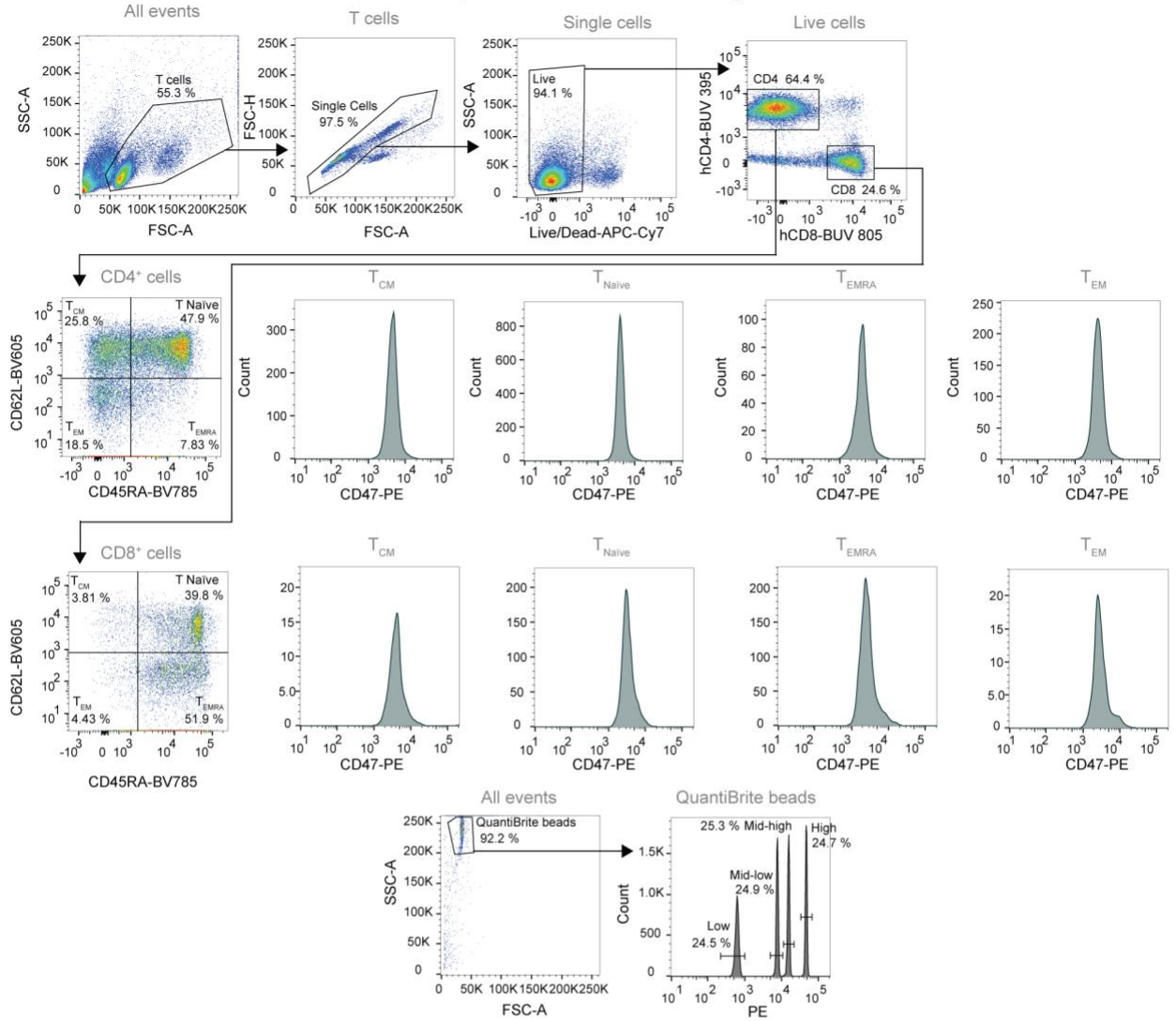
Representative gating strategy for assessing macrophage depletion after peritoneal lavage

Used for: Extended Data Fig. 3p and 3q
 Representative data from Extended Data Fig. 3q



Representative gating strategy for quantifying CD47 expression using QuantiBrite

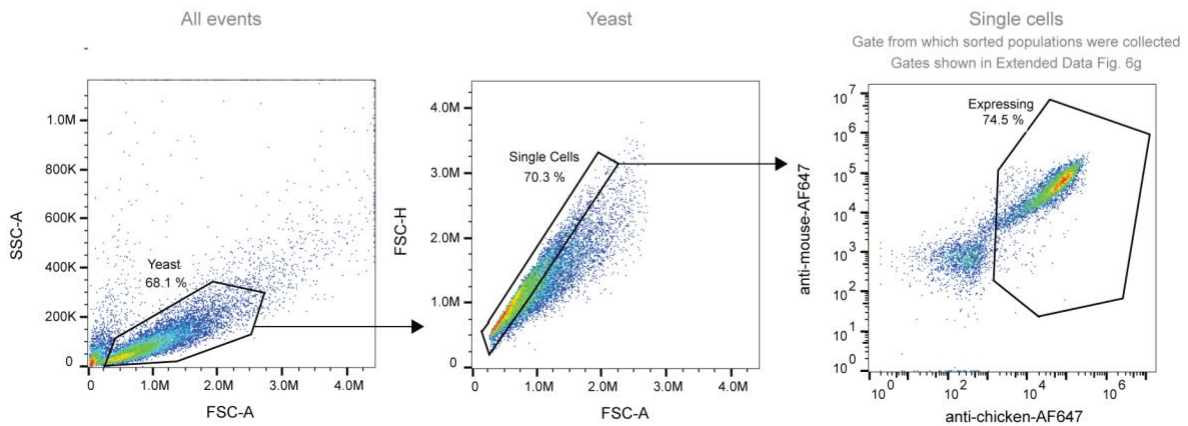
Used for: Extended Data Fig. 4c, 4d, 4e and 4f
 Representative data from Extended Data Fig. 4f



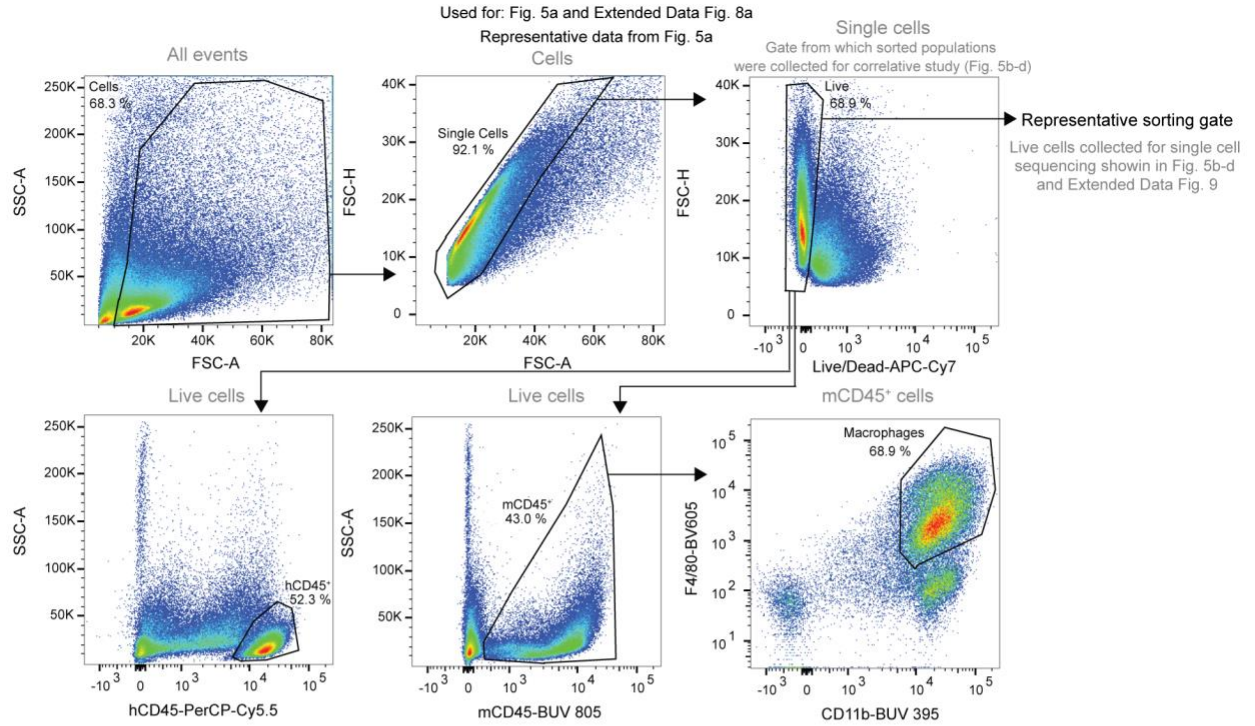
Representative gating strategy for yeast surface display binding and sorting

Used for: Fig. 4b and 4d | Extended Data Fig. 6c, 6d, 6e, 6f, 6g, 6h, 6i, 6k and 6n
 Representative data from Extended Data Fig. 6c

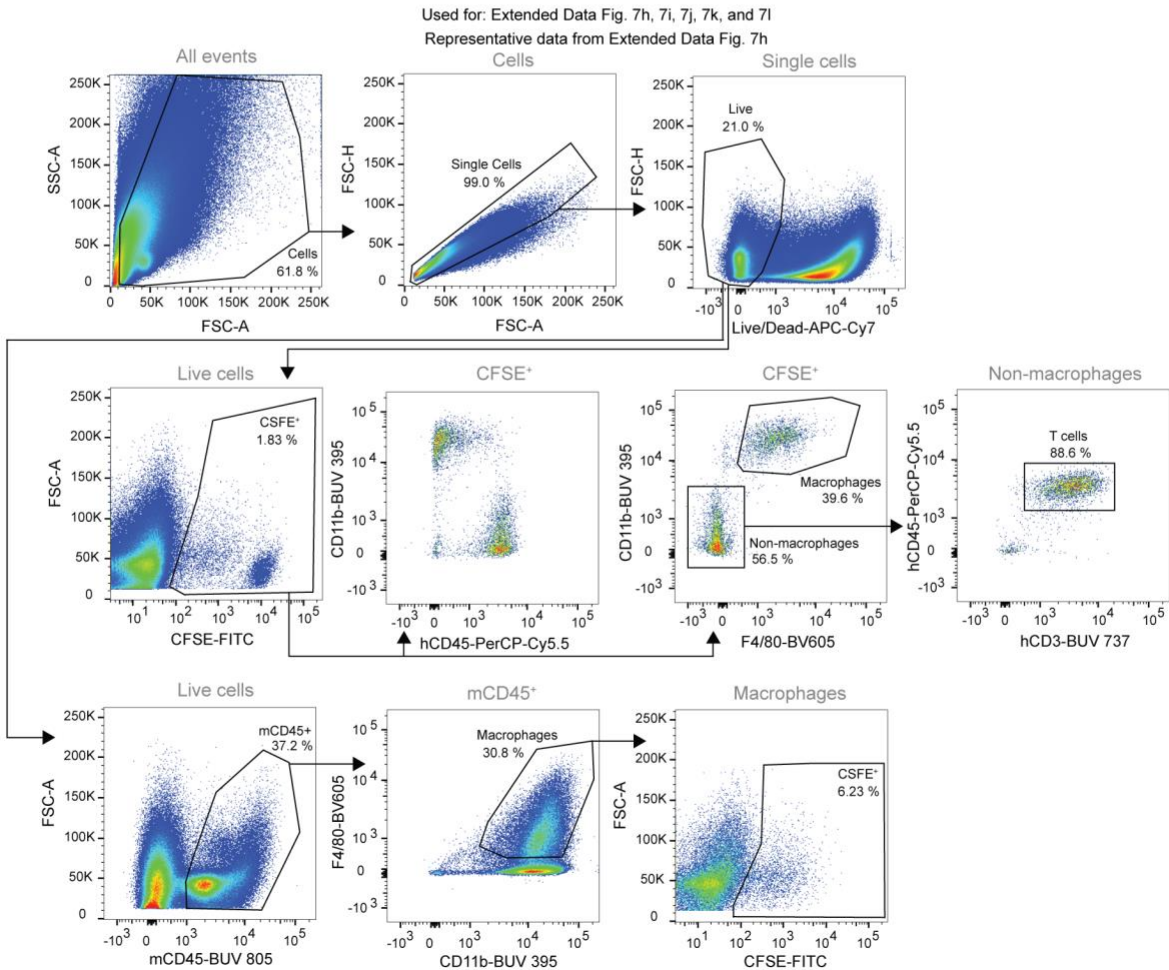
Specific sorting gates for CD47 library on single cell populations shown in Extended Data Fig. 6g



Representative gating strategy for quantifying T cells and macrophages in 143B tumor dissociations

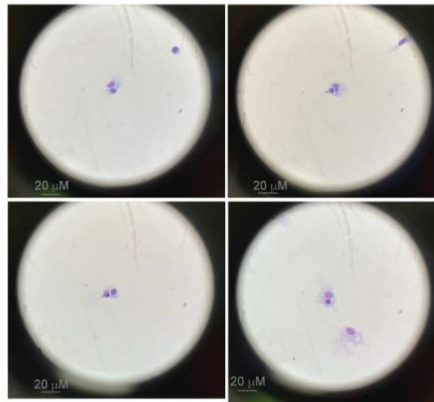


Representative gating strategy for T cells and macrophage identification in the *in vivo* phagocytosis assay



Supplementary Fig. 3 | Raw images of patient CSF sample shown in Fig. 2c

Patient CSF: raw images of histiocytes engulfing lymphocytes after CD19.28 ζ CAR T treatment



Supplementary Fig. 3 | Microscope images of histiocytes engulfing lymphocytes collected from the CSF of a CD19.28 ζ -CAR T treated large B cell lymphoma (LBCL) patient and stained with hematoxylin and eosin (1000x magnification). Fig. 2c depicts enlargements of these images, kept in the same orientation of images.

Supplementary Notes

Data duplication between figures:

- Fig. 1h: Mock and 47_{WT}-CD19.28 ζ are shared conditions duplicated in Extended Data Fig. 1m (mock) and Extended Data Fig. 11e (mock and 47_{WT}-CD19.28 ζ).
- Fig. 1j: Mock and 47_{OE}-mock are shared conditions duplicated in Extended Data Fig. 3o.
- Fig. 4g: 47_{WT} is a shared condition duplicated in T cell depletion in Extended Data Fig. 2m.
- Fig. 5a: IHC data is also represented in Extended Data Fig. 8d.
- Extended Data Fig. 1m: Mock and mock + B6H12 are shared conditions duplicated in Fig. 1h (mock) and Extended Data Fig. 11e (mock and mock + B6H12).
- Extended Data Fig. 1n: Mock treatment is a shared condition, duplicated in Extended Data Fig. 2h.
- Extended Data Fig. 1o: Mock and mock + B6H12 are shared conditions duplicated in Extended Data Fig. 2i (mock) and Extended Data Fig. 11f (mock and mock + B6H12).
- Extended Data Fig. 2h: Mock treatment is a shared condition, with the image from day 17 duplicated in Extended Data Fig. 1n.
- Extended Data Fig. 2i: Mock and 47_{WT}-CD19.28 ζ are shared conditions duplicated in Extended Data Fig. 1o (mock) and Extended Data Fig. 11f (mock and 47_{WT}-CD19.28 ζ).
- Extended Data Fig. 2k: 47_{WT} is a shared condition duplicated in T cell depletion in Extended Data Fig. 7e.
- Extended Data Fig. 2l: 47_{WT} is a shared condition duplicated in T cell depletion in Extended Data Fig. 7f.
- Extended Data Fig. 2m: 47_{WT} is a shared condition duplicated in T cell depletion in Fig. 4g and Extended Data Figure 7g.
- Extended Data Fig. 3o: Mock and 47_{OE}-mock are a shared conditions duplicated in Fig. 1j.
- Extended Data Fig. 7e: 47_{WT} is a shared condition duplicated in T cell depletion in Extended Data Fig. 2k.
- Extended Data Fig. 7f: 47_{WT} is a shared condition duplicated in T cell depletion in Extended Data Fig. 2l.
- Extended Data Fig. 7g: 47_{WT} is a shared condition duplicated in T cell depletion in Extended Data Fig. 2m.
- Extended Data Fig. 8b: Data is also represented in Extended Data Fig. 8d.
- Extended Data Fig. 8d: hCD3 is also represented in Extended Data Fig. 8b. F4/80 is also represented in Fig. 5a.
- Extended Data Fig. 11e: Mock, mock + B6H12, and 47_{WT}-CD19.28 ζ are shared conditions duplicated in Fig. 1h (mock and 47_{WT}-CD19.28 ζ) and Extended Data Fig. 1m (mock and mock + B6H12).
- Extended Data Fig. 11f: Mock, mock + B6H12, and 47_{WT}-CD19.28 ζ are shared conditions duplicated in Extended Data Fig. 1o (mock and mock + B6H12) and Extended Data Fig. 2i (mock and 47_{WT}-CD19.28 ζ).