

## Supplementary Material

# Enhancing the Potential of Phenomic and Genomic Prediction in Winter Wheat Breeding Using High-Throughput Phenotyping and Deep Learning

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## Supplementary Tables and Figures

**Supplementary Table 1:** List of vegetation indices and equations extracted from multispectral images used in this study.

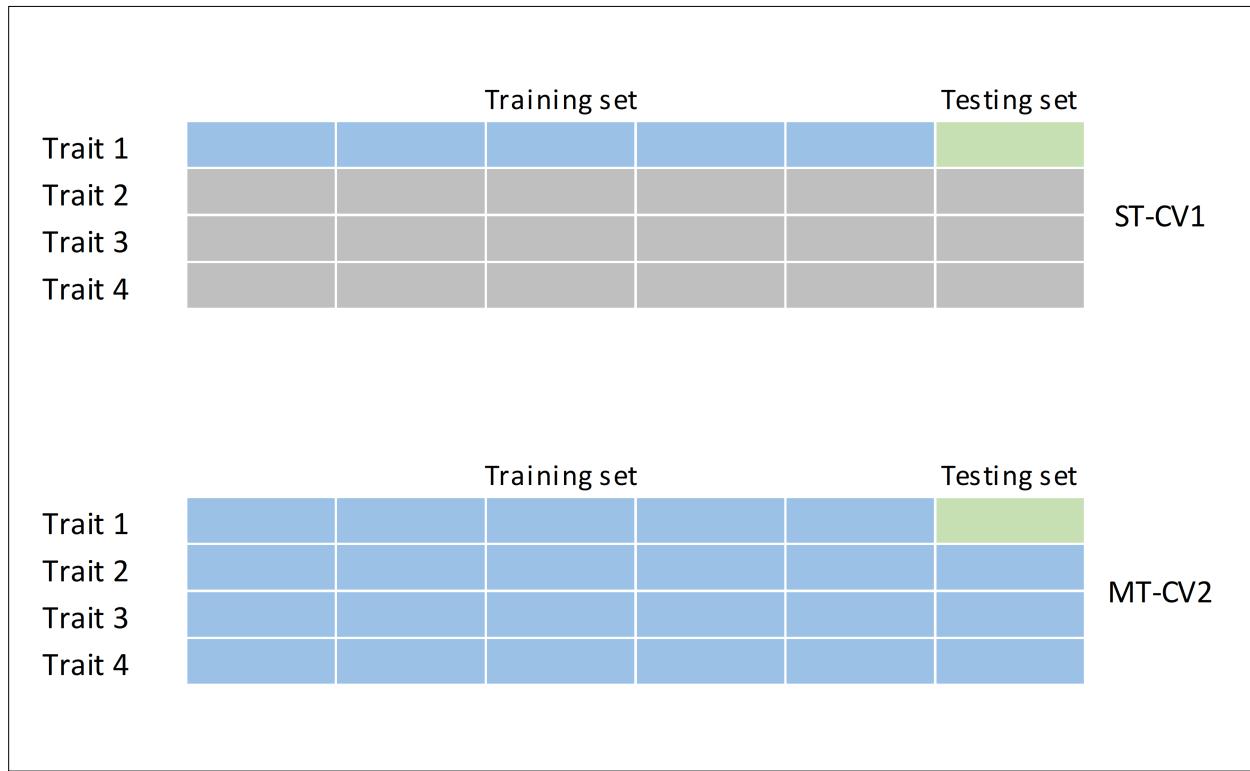
Spectral feature name	Formulation	Ref.
5 original reflectance bands: Blue (B), Green (G), Red (R), Red edge (RE), Near Infrared (NIR)	The reflectance value of each band	-
Normalized difference vegetation index	$NDVI = (NIR - R) / (NIR + R)$	(Rouse Jr et al., 1974)
Green Normalized Difference Vegetation Index (GNDVI)	$GNDVI = (NIR - G) / (NIR + G)$	(Gitelson & Merzlyak, 1997)
Ratio Vegetation Index 1	$RVI\_1 = NIR / R$	(Richardson & Wiegand, 1977)
Green Chlorophyll Index	$GCI = (NIR / G) - 1.0$	(Gitelson et al.,
Rice Growth Vegetation Index	$RGVI = R / G$	(Nuarsa et al.,
Difference Vegetation Index	$DVI = NIR - R$	(Tucker, 1979)
Soil-Adjusted Vegetation Modified Soil Adjusted Vegetation Index	$SAVI = ((NIR - R) / (NIR + R + L)) * (1.0 + L)$ , $L = np.sqrt(np.square(2.0 * NIR) + 1.0) - np.sqrt(np.square(2.0 * NIR + 1.0))$	(Huete, 1988) (Qi et al., 1994)
Optimized Soil Adjusted Vegetation Index	$OSAVI = (NIR - R) / (NIR + R + 0.16)$	(Rondeaux et al., 1996)
Renormalized Difference Vegetation Index	$RDVI = np.sqrt((np.square(NIR - R)) / (NIR + R))$	(Roujean & Breon, 1995)
Triangular Vegetation Index	$TVI = 60.0 * (NIR - G) - 100.0 * (R - G)$	(Deering, 1975)
Transformed Soil Adjusted Vegetation Index	$TSAVI = (a * (NIR - a * R - b)) / (a * NIR + R - a * b)$ , $a, b = 0.96916, 0.084726$	(Baret & Guyot, 1991)
Perpendicular Vegetation Index	$PVI = (NIR - a * R - b) / np.sqrt(1 + np.square(a))$ , $a, b =$	(Richardson & Wiegand, 1977)
Soil-Adjusted Vegetation Index 2	$SAVI\_2 = NIR / (R - (b/a))$ , $a, b = 0.96916,$ $0.084726$	(Huete, 1988)
Adjusted Transformed Soil Adjusted Vegetation Index	$ATSAVI = (a * (-a * R - b)) / (a * NIR + R - a * b + X * (1 + np.square(a)))$ , $a, b, X =$ $0.96916, 0.084726, 0.08$	(Baret & Guyot, 1991)
Normalized Difference Water Index	$NDWI = (G - NIR) / (G + NIR)$	(McFeeters, 1996)
Kawashima index	$IKAW = (R - B) / (R + B)$	(Kawashima & Nakatani, 1998)
Simple Ratio Pigment Index	$SRPI = B / R$	(Peñuelas et al., 1995)

Ratio Vegetation Index 2	$RVI\_2 = NIR/G$	(Pearson & Miller, 1972)
Modified Chlorophyll Absorption in Reflective	$MCARI = (RE-R-0.2*(RE-G))*(RE/R)$	(Daughtry et al., 2000)
Modified Chlorophyll Absorption in Reflective	$MCARI\_1 = 1.2*(2.5*(NIR-R)-1.3*(NIR-G))$	(Haboudane et al., 2004)
Modified Chlorophyll Absorption in Reflective	$MCARI\_2 = 1.5*(2.5*(NIR-R)-1.3*(NIR-G))*(np.square(2.0*NIR+1))-(6.0*NIR-$	(Haboudane et al., 2004)
Modified Triangular Vegetation Index	$MTVI\_1 = 1.2*(1.2*(NIR-G)-2.5*(R-G))$	(Haboudane et al., 2004)
Modified Triangular Vegetation Index 2	$MTVI\_2 = 1.5*(1.2*(NIR-G)-2.5*(R-G))*(np.square(2*NIR+1))-(6.0*NIR-$	(Haboudane et al., 2004)
Modified chlorophyll absorption ratio index/Second modified	$R\_MCARI\_MTVI2 = ((RE-R-0.2*(RE-G))*(RE/R))/(1.5*(1.2*(NIR-G)-2.5*(R-G))*(np.square(2*NIR+1))-(6.0*NIR-$	(Eitel et al., 2007)
Enhanced Vegetation Index	$EVI = (NIR-R)/(NIR+6.0*R-7.5*B+1.0)$	(Huete et al., 1997)
Datt Index	$DATT = (NIR-RE)/(NIR-R)$	(Datt, 1999)
Double-peak Canopy Nitrogen Index	$DNCI = (RE-G)/(RE+G)$	(Chen et al., 2010)
Plant Senescence Reflectance Index	$PSRI = (R-G)/RE$	(Merzlyak et al., 1999)
Structure Intensive Pigment Vegetation Index	$SICI = (NIR-B)/(NIR+R)$	(Peñuelas et al., 1995)
Spectral Polygon Vegetation Index	$SPVI = 0.4*3.7*(NIR-R)-1.2*np.abs(G-R)$	(Vincini et al., 2006)
Transformed Chlorophyll Absorption Reflectance Index	$TCARI = 3.0*((RE-R)-0.2*(RE-G)*(RE/R))$	(Haboudane et al., 2002)
Transformed Chlorophyll Absorption in the Reflectance Index/Optimized Soil-Adjusted Vegetation Index	$R\_TCARI\_OSAVI = (3.0*((RE-R)-0.2*(RE-G)*(RE/R)))/((NIR-R)/(NIR+R+0.16))$	(Haboudane et al., 2002)
Red edge relative index	$RERI = (RE-R)/NIR$	(Haboudane et al., 2002)
Normalized difference red edge index	$NDRE = (NIR-RE)/(NIR+RE)$	(Fitzgerald et al., 2010)
MERIS Terrestrial Chlorophyll Index	$MTCI = (NIR-RE)/(RE-R)$	(Dash & Curran, 2004)
Enhanced Vegetation Index 2	$EVI\_2 = 2.5*((NIR-R)/(NIR+2.4*R+1.0))$	(Huete et al., 1997)
Red-Edge Chlorophyll Index	$RECI = (NIR/RE)-1$	(Haboudane et al., 2002)

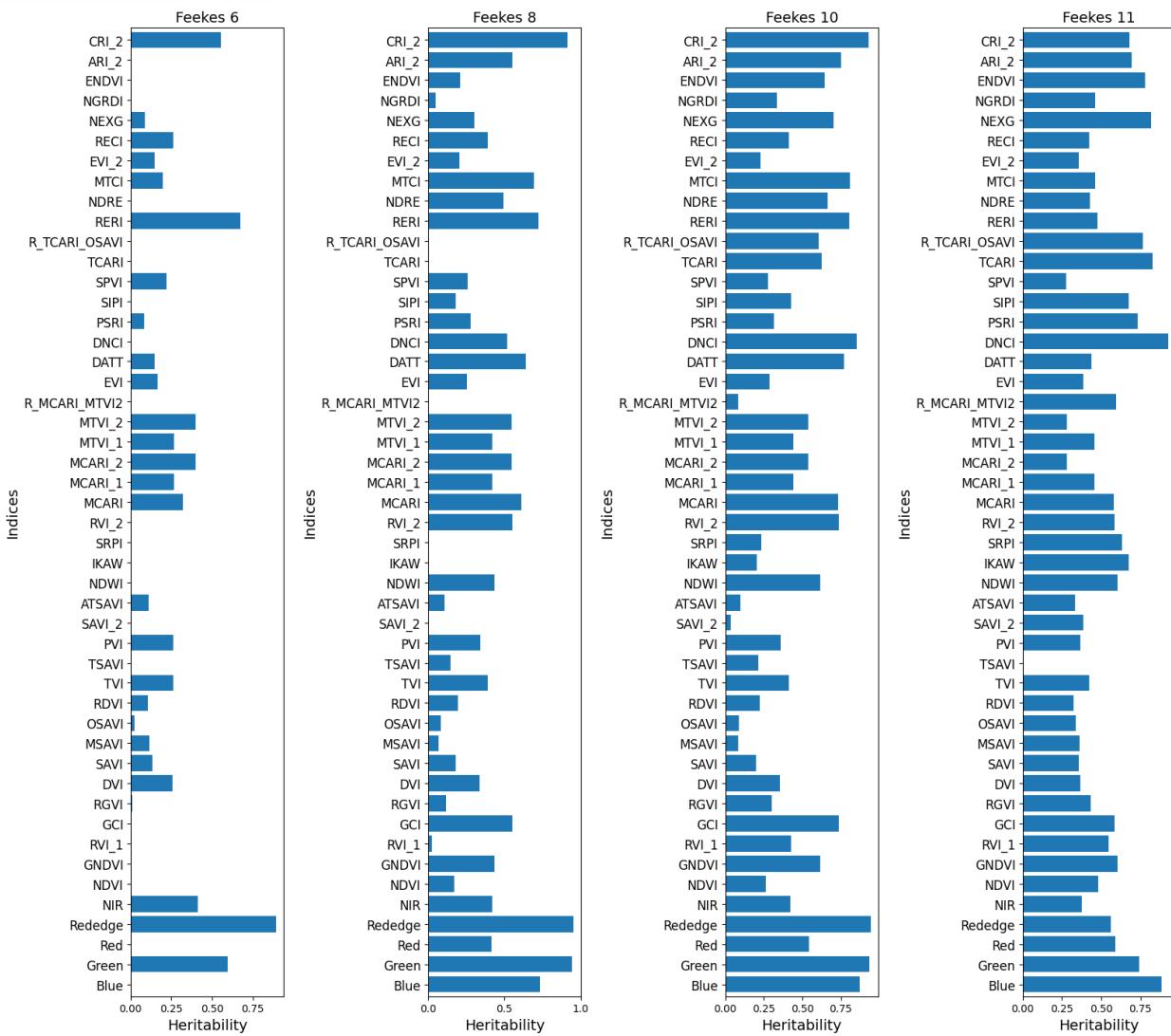
Normalized excess green vegetation index	$NEXG = (2*G-R-B)/(G+R+B)$	(Woebbecke et al., 1995)
Normalized green-red	$NGRDI = (G-R)/(G+R)$	(Tucker, 1979)
Enhanced Normalized Difference Vegetation Index	$ENDVI = (NIR+G-2.0*B)/(NIR+G+2.0*B)$	(Anchal et al., 2022)
Anthocyanin Reflectance Index 2	$ARI\_2 = NIR*(1.0/G)-(1.0/RE)$	(Gitelson et al., 2001)
Carotenoid Reflectance Index 2	$CRI\_2 = (1.0/G)-(1.0/RE)$	(Gitelson et al., 2003)

**Supplementary Table 2.** Summary statistics and broad-sense heritability ( $H^2$ ) of recorded agronomic traits.

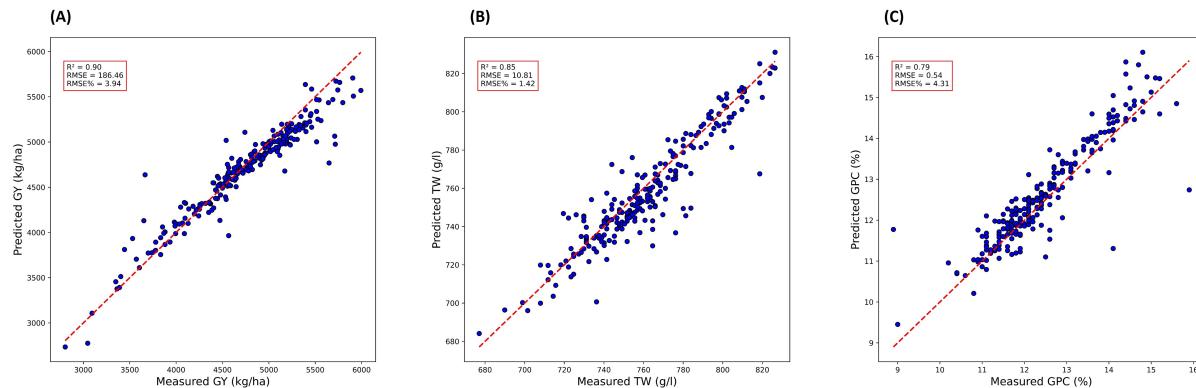
Trait	Mean	$H^2$
Grain Yield (GY)(kg/ha)	70.30	0.36
Grain Protein (GP)(%)	12.48	0.26
Test Weight (TW)(g/l)	58.88	0.87



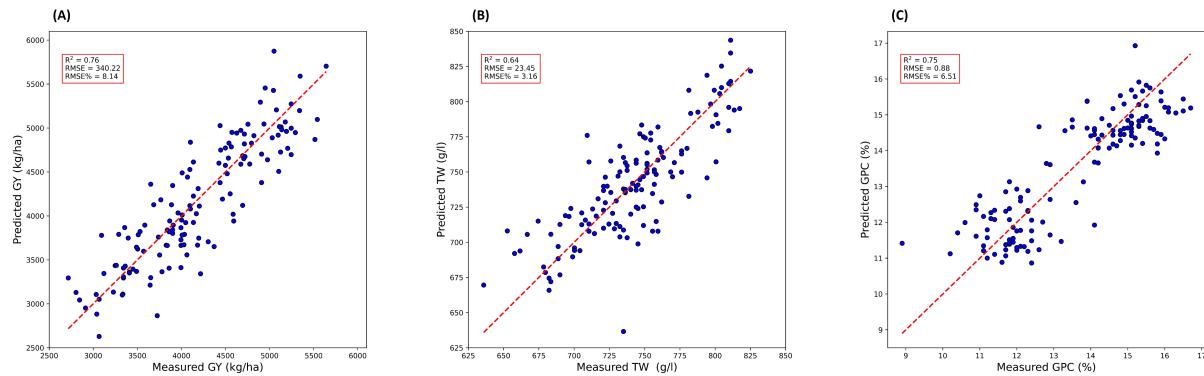
**Supplementary Figure 1.** Prediction accuracies were estimated for the single-trait (ST) model with cross-validation scheme 1 (ST-CV1) and multi-trait (MT) model with cross-validation scheme 2 (MT-CV2).



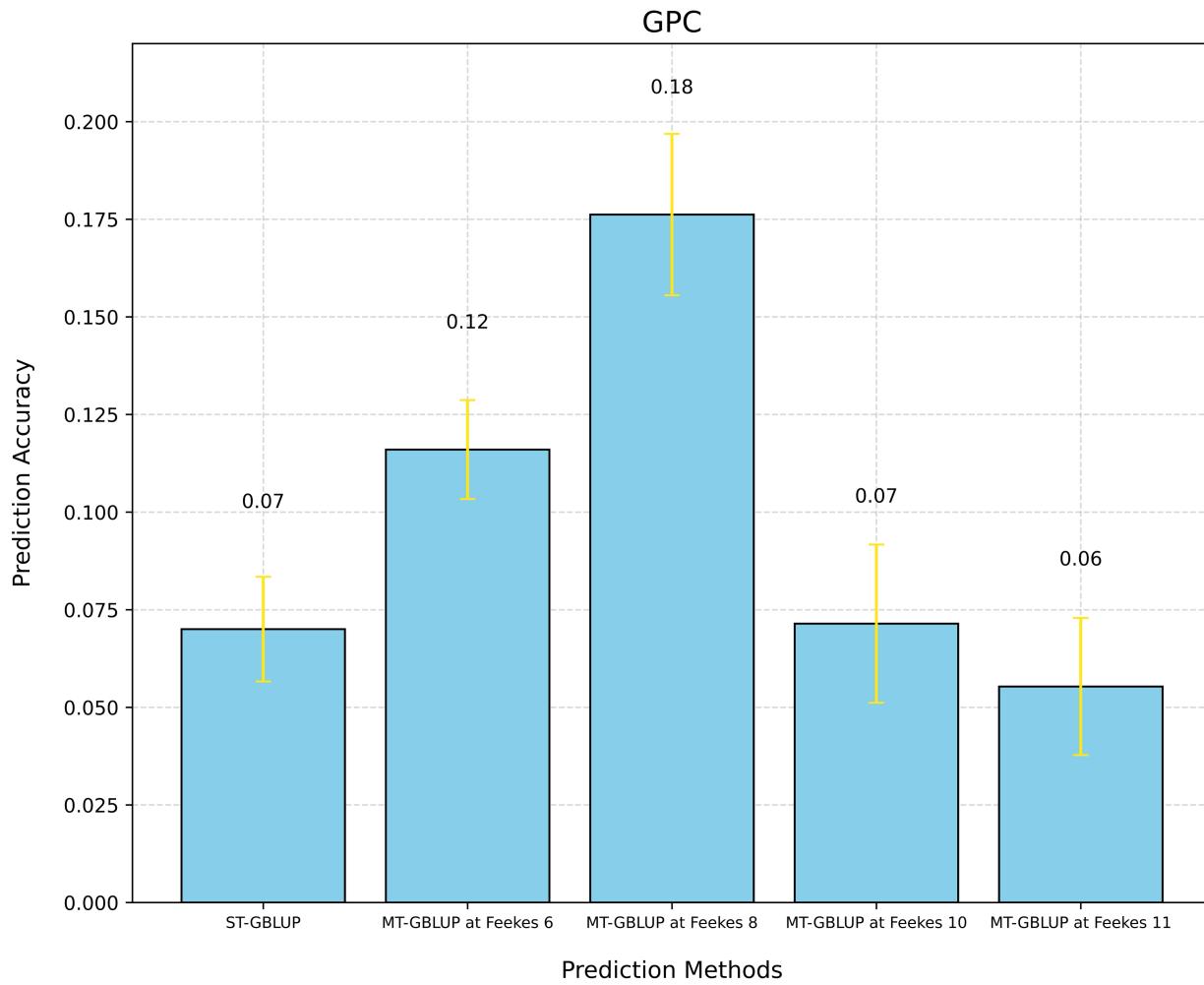
**Supplementary Figure 2.** Broad-sense heritability ( $H^2$ ) of High-Throughput Phenotyping (HTP) based traits across various growth stages (Feeekes 6: jointing, Feeekes 8: Flag leaf, Feeekes 10: Booting, Feeekes 11: Milk ripe).



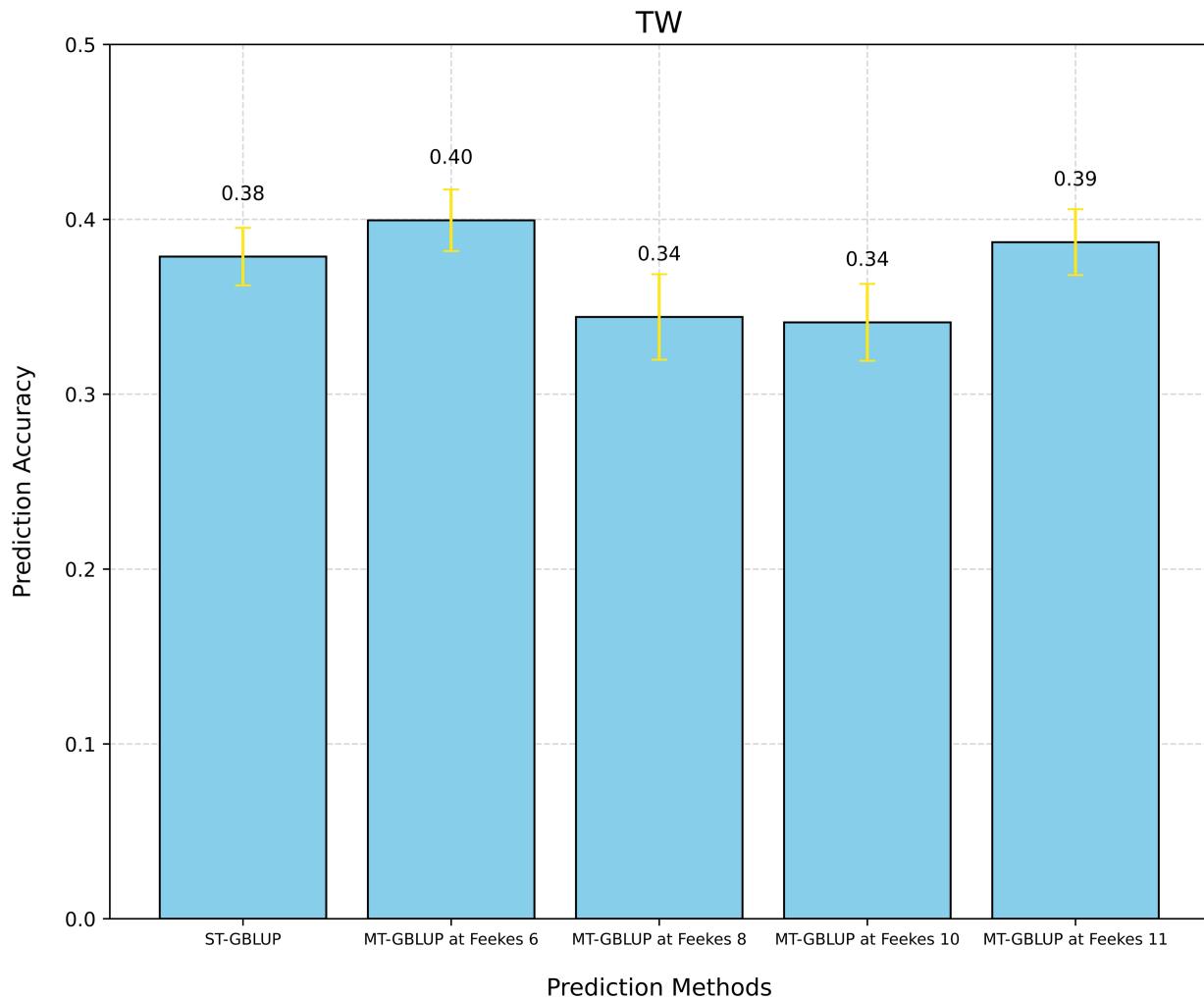
**Supplementary Figure 3.** Phenomic predictions for grain yield (GY), B) test weight (TW), and C) grain protein (GPC), using DNN with combined High-Throughput Phenotyping (HTP) based traits on the training set (70%) for ELITE and AYT at Brookings.



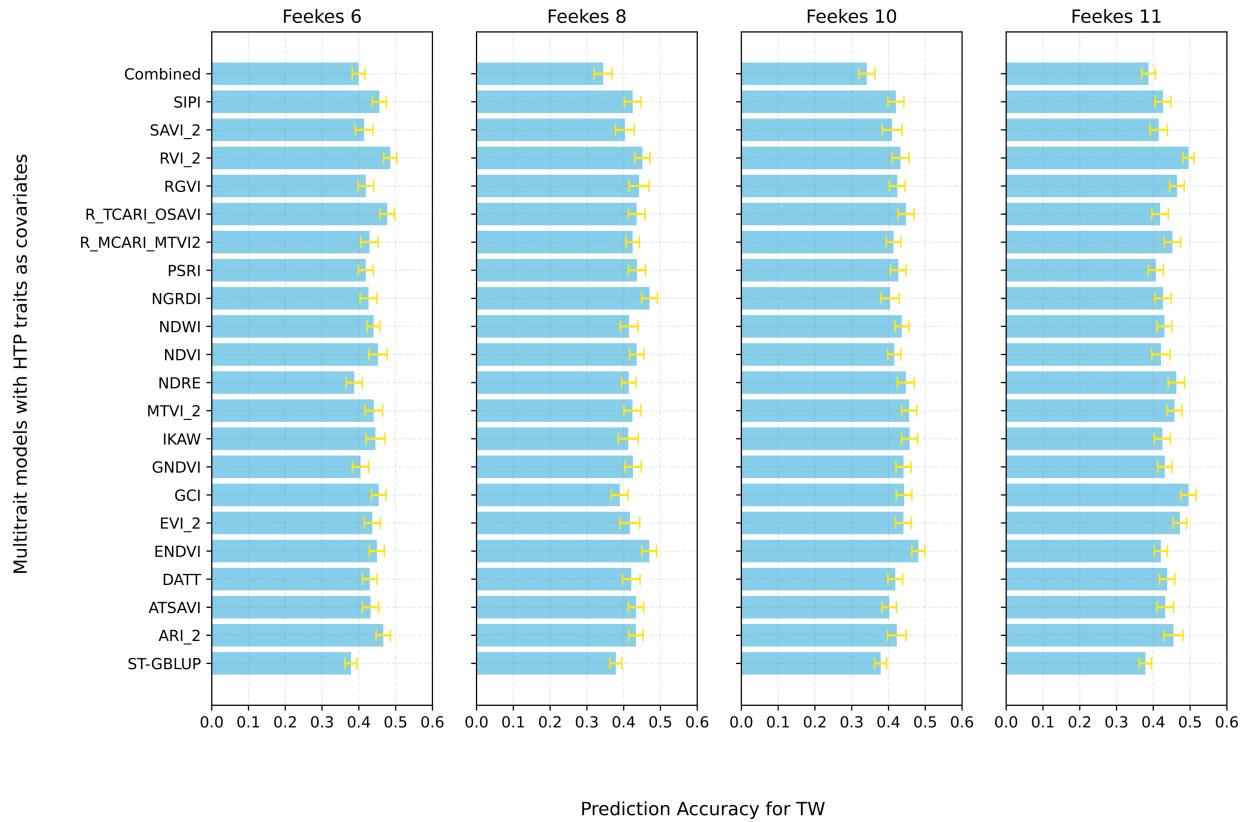
**Supplementary Figure 4.** A) Phenomic predictions for grain yield (GY), B) Test weight (TW), and C) grain protein content (GPC). Predictions were made on the testing set (30%) using DNN trained on ELITE and AYT at Brookings and Dakota Lakes (multi-locations).



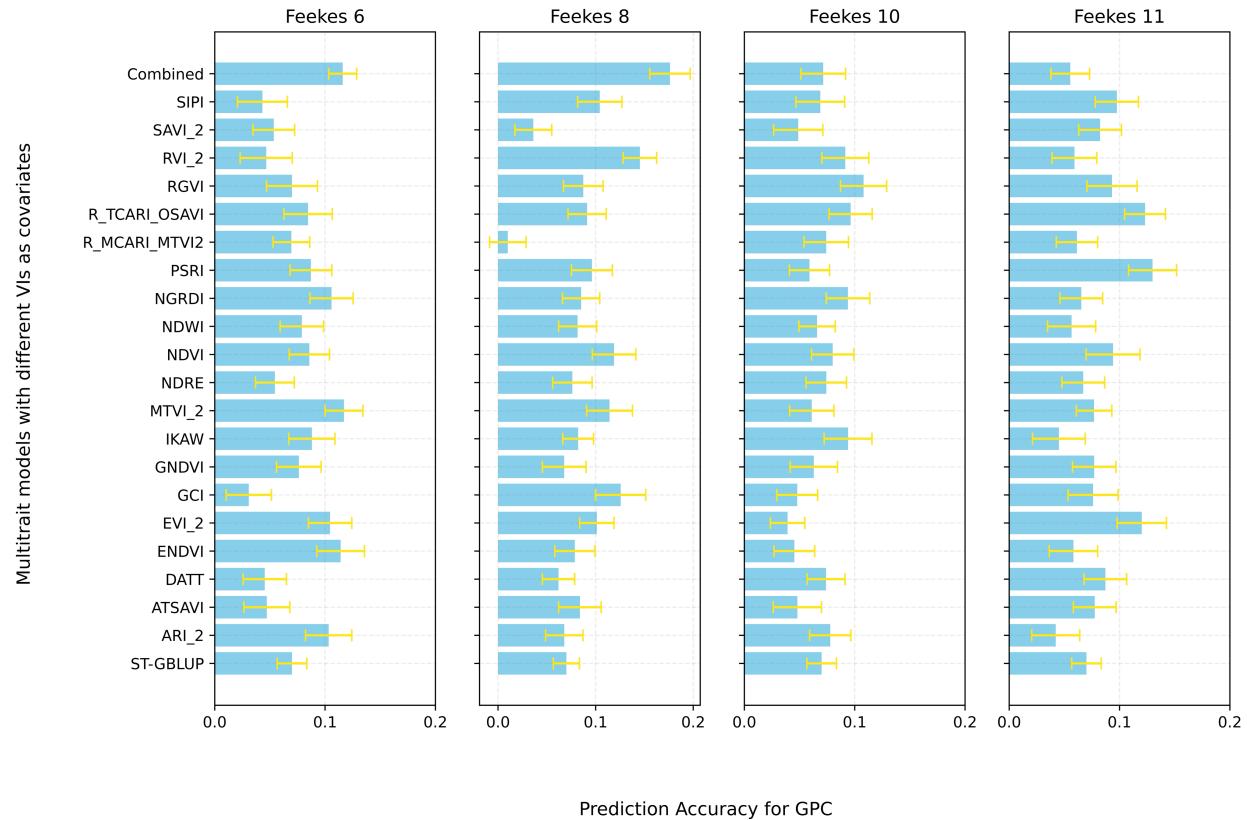
**Supplementary Figure 5.** Comparison of Prediction accuracy for Grain Protein (GPC) using Single-Trait GBLUP (ST-GBLUP) with cross-validation scheme ST-CV1 and Multi-Trait GBLUP (MT-GBLUP) incorporating High-Throughput Phenotyping (HTP) based traits with cross-validation scheme MT-CV2 across different growth stages (Feekes 6: jointing, Feekes 8: Flag leaf, Feekes 10: Booting, Feekes 11: Milk ripe).



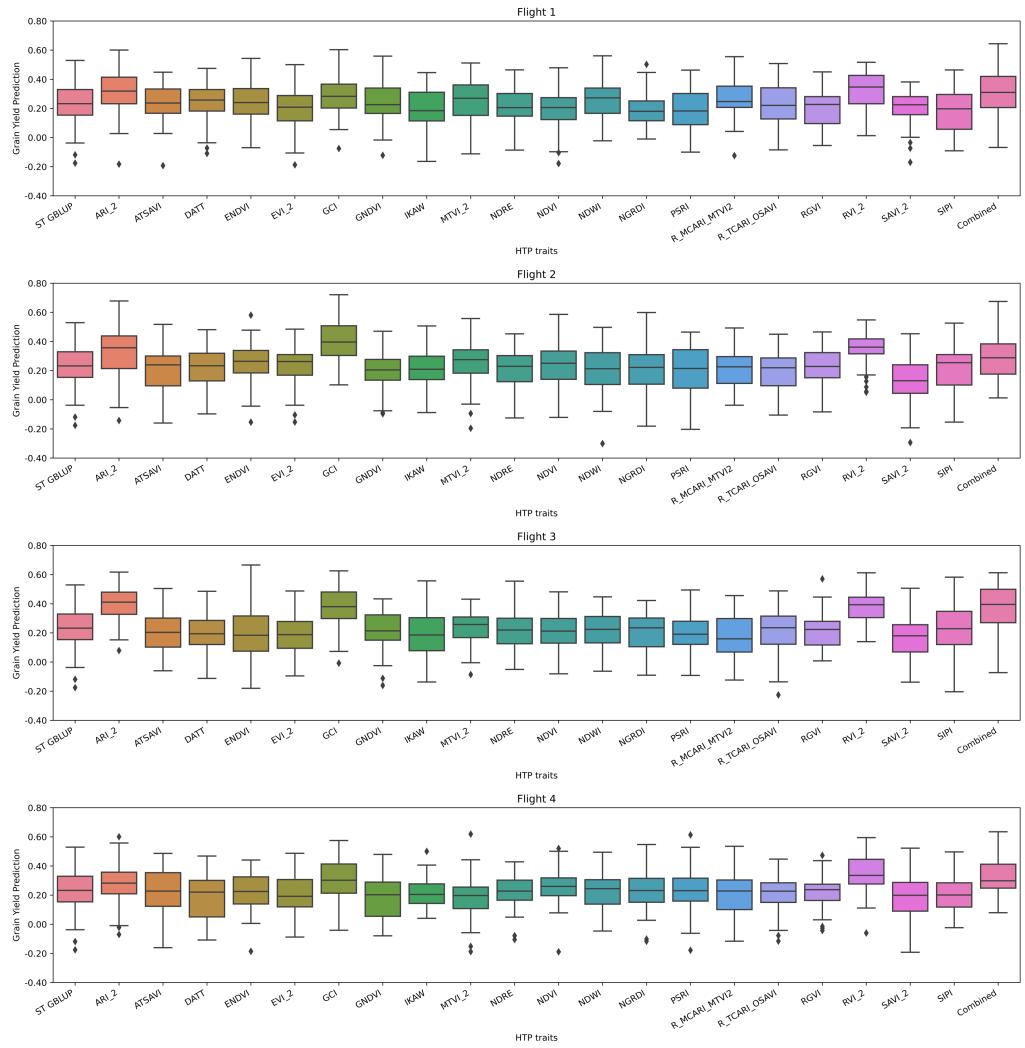
**Supplementary Figure 6.** Comparison of Prediction accuracy for Test Weight (TW) using Single-Trait GBLUP (ST-GBLUP) with cross-validation scheme ST-CV1 and Multi-Trait GBLUP (MT-GBLUP) High-Throughput Phenotyping (HTP) based traits with cross-validation scheme MT-CV2 across different growth stages (Feekes 6: jointing, Feekes 8: Flag leaf, Feekes 10: Booting, Feekes 11: Milk ripe).



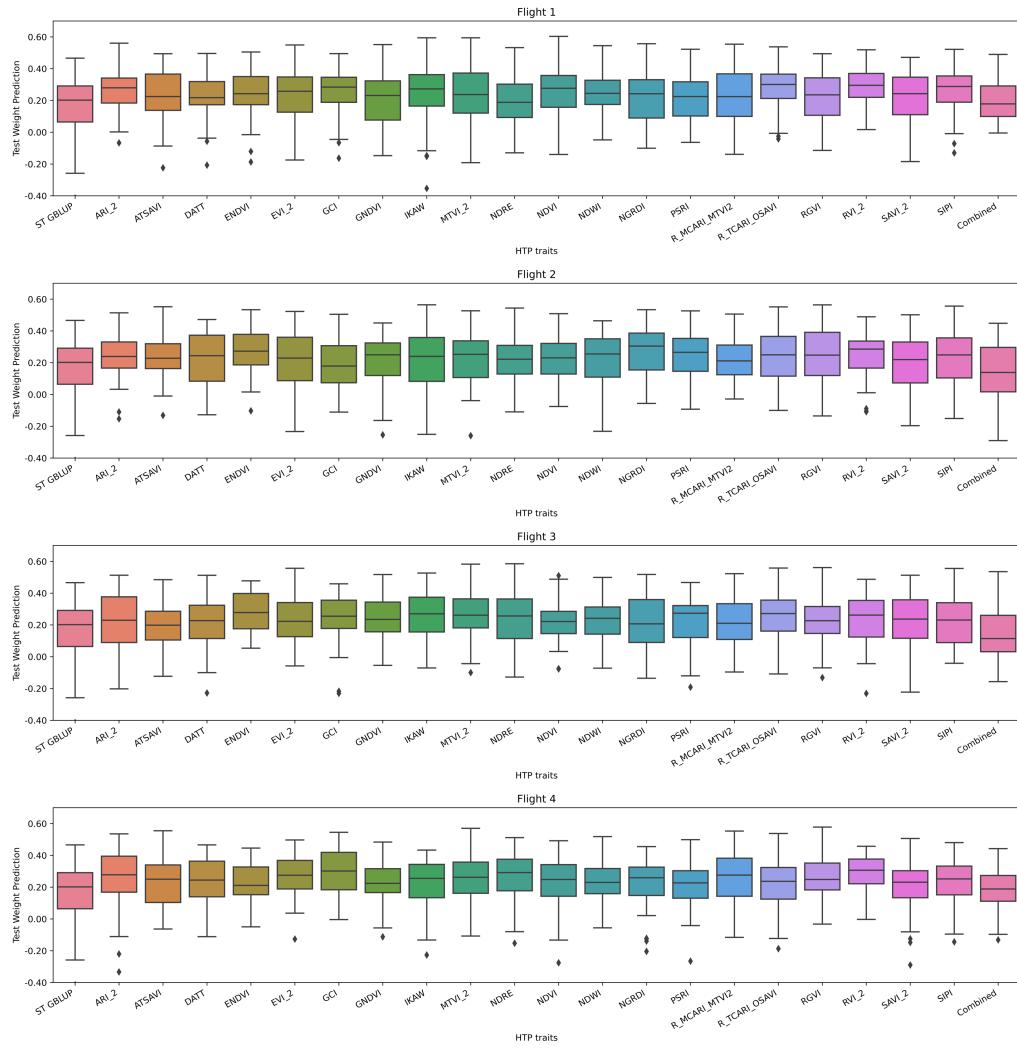
**Supplementary Figure 7.** Prediction accuracy for Test Weight (TW) using Single-Trait GBLUP (ST-GBLUP) with cross-validation scheme ST-CV1 and Multi-Trait GBLUP (MT-GBLUP) incorporating High-Throughput Phenotyping (HTP) based traits with cross-validation scheme MT-CV2 across different growth stages different growth stages (Feekes 6: jointing, Feekes 8: Flag leaf, Feekes 10: Booting, Feekes 11: Milk ripe).



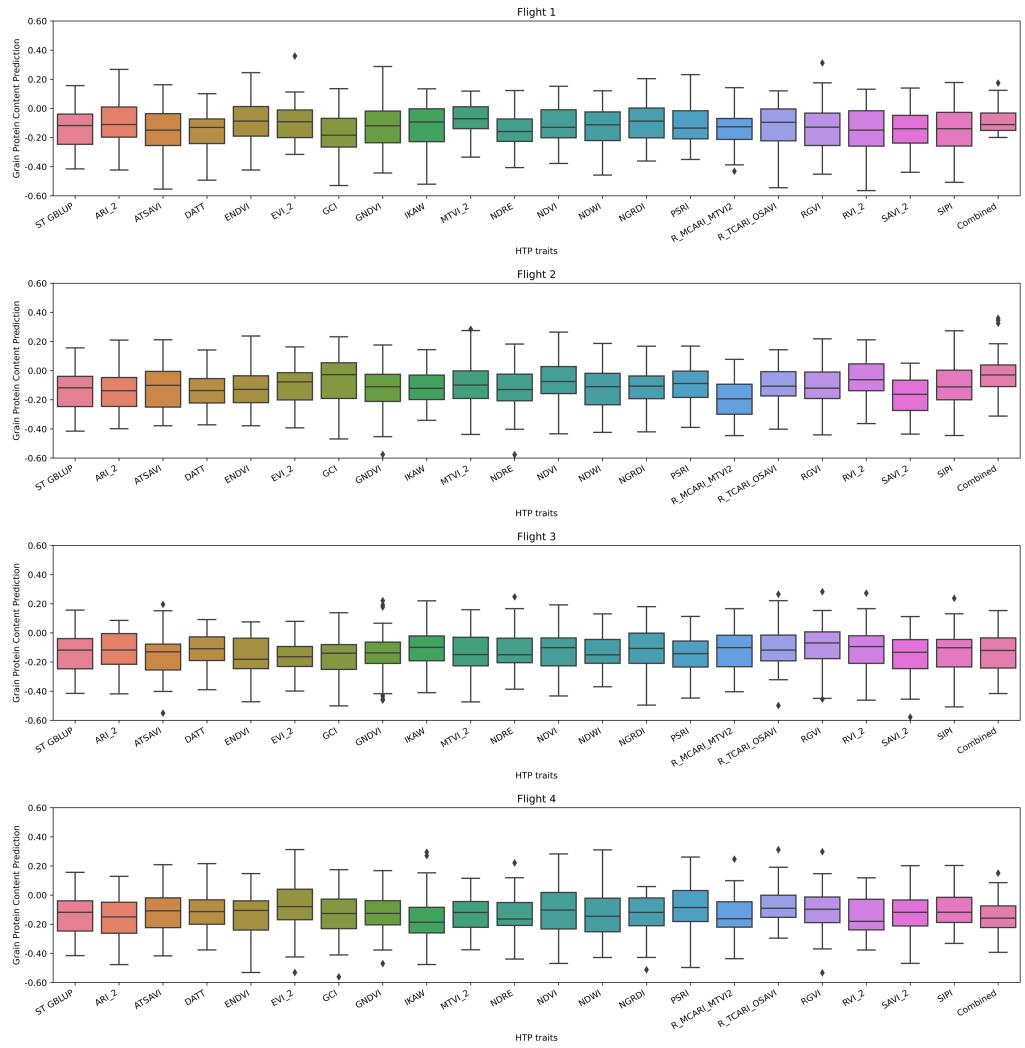
**Supplementary Figure 8.** Prediction accuracy for Grain Protein (GPC) using Single-Trait GBLUP (ST-GBLUP) with cross-validation scheme ST-CV1 and Multi-Trait GBLUP (MT-GBLUP) incorporating High-Throughput Phenotyping (HTP) based traits with cross-validation scheme MT-CV2 across different growth stages different growth stages (Feekes 6: jointing, Feekes 8: Flag leaf, Feekes 10: Booting, Feekes 11: Milk ripe).



**Supplementary Figure 9.** Distribution of prediction accuracy estimates for Grain Yield (GY) across multiple repetitions using Single-Trait GBLUP (ST-GBLUP) with cross-validation scheme ST-CV1 and Multi-Trait GBLUP (MT-GBLUP) incorporating High-Throughput Phenotyping (HTP) based traits with cross-validation scheme MT-CV2 across flights. Each boxplot captures the variation in prediction accuracy across replicates for different prediction models.



**Supplementary Figure 10.** Distribution of prediction accuracy estimates for Test Weight (TW) across multiple repetitions using Single-Trait GBLUP (ST-GBLUP) with cross-validation scheme ST-CV1 and Multi-Trait GBLUP (MT-GBLUP) incorporating High-Throughput Phenotyping (HTP) based traits with cross-validation scheme MT-CV2 across flights. Each boxplot captures the variation in prediction accuracy across replicates for different prediction models.



**Supplementary Figure 11.** Distribution of prediction accuracy estimates for Grain Protein Content (GPC) across multiple repetitions using Single-Trait GBLUP (ST-GBLUP) with cross-validation scheme ST-CV1 and Multi-Trait GBLUP (MT-GBLUP) incorporating High-Throughput Phenotyping (HTP) based traits with cross-validation scheme MT-CV2 across flights. Each boxplot captures the variation in prediction accuracy across replicates for different prediction models.

## REFERENCES

- Anchal, S., Bahuguna, S., Priti, Pal, P. K., Kumar, D., Murthy, P. S., & Kumar, A. (2022). Non-destructive method of biomass and nitrogen (N) level estimation in Stevia rebaudiana using various multispectral indices. *Geocarto International*, 37(22), 6409-6421.
- Baret, F., & Guyot, G. (1991). Potentials and limits of vegetation indices for LAI and APAR assessment. *Remote sensing of environment*, 35(2-3), 161-173.
- Chen, P., Haboudane, D., Tremblay, N., Wang, J., Vigneault, P., & Li, B. (2010). New spectral indicator assessing the efficiency of crop nitrogen treatment in corn and wheat. *Remote sensing of environment*, 114(9), 1987-1997.
- Dash, J., & Curran, P. (2004). The MERIS terrestrial chlorophyll index.
- Datt, B. (1999). A new reflectance index for remote sensing of chlorophyll content in higher plants: tests using Eucalyptus leaves. *Journal of plant physiology*, 154(1), 30-36.
- Daughtry, C. S., Walther, C., Kim, M., De Colstoun, E. B., & McMurtrey III, J. (2000). Estimating corn leaf chlorophyll concentration from leaf and canopy reflectance. *Remote sensing of environment*, 74(2), 229-239.
- Deering, D. (1975). Measuring "forage production" of grazing units from Landsat MSS data. Proceedings of the Tenth International Symposium of Remote Sensing of the Environment,
- Eitel, J., Long, D., Gessler, P., & Smith, A. (2007). Using in-situ measurements to evaluate the new RapidEye™ satellite series for prediction of wheat nitrogen status. *International Journal of Remote Sensing*, 28(18), 4183-4190.
- Fitzgerald, G., Rodriguez, D., & O'Leary, G. (2010). Measuring and predicting canopy nitrogen nutrition in wheat using a spectral index—The canopy chlorophyll content index (CCCI). *Field crops research*, 116(3), 318-324.
- Gitelson, A. A., Gritz, Y., & Merzlyak, M. N. (2003). Relationships between leaf chlorophyll content and spectral reflectance and algorithms for non-destructive chlorophyll assessment in higher plant leaves. *Journal of plant physiology*, 160(3), 271-282.
- Gitelson, A. A., & Merzlyak, M. N. (1997). Remote estimation of chlorophyll content in higher plant leaves. *International Journal of Remote Sensing*, 18(12), 2691-2697.
- Gitelson, A. A., Merzlyak, M. N., & Chivkunova, O. B. (2001). Optical properties and nondestructive estimation of anthocyanin content in plant leaves¶. *Photochemistry and photobiology*, 74(1), 38-45.
- Haboudane, D., Miller, J. R., Pattey, E., Zarco-Tejada, P. J., & Strachan, I. B. (2004). Hyperspectral vegetation indices and novel algorithms for predicting green LAI of crop canopies: Modeling and validation in the context of precision agriculture. *Remote sensing of environment*, 90(3), 337-352.
- Haboudane, D., Miller, J. R., Tremblay, N., Zarco-Tejada, P. J., & Dextraze, L. (2002). Integrated narrow-band vegetation indices for prediction of crop chlorophyll content for application to precision agriculture. *Remote sensing of environment*, 81(2-3), 416-426.
- Huete, A., Liu, H., Batchily, K., & Van Leeuwen, W. (1997). A comparison of vegetation indices over a global set of TM images for EOS-MODIS. *Remote sensing of environment*, 59(3), 440-451.
- Huete, A. R. (1988). A soil-adjusted vegetation index (SAVI). *Remote sensing of environment*, 25(3), 295-309.
- Kawashima, S., & Nakatani, M. (1998). An algorithm for estimating chlorophyll content in leaves using a video camera. *Annals of Botany*, 81(1), 49-54.

- McFeeters, S. K. (1996). The use of the Normalized Difference Water Index (NDWI) in the delineation of open water features. *International Journal of Remote Sensing*, 17(7), 1425-1432.
- Merzlyak, M. N., Gitelson, A. A., Chivkunova, O. B., & Rakitin, V. Y. (1999). Non-destructive optical detection of pigment changes during leaf senescence and fruit ripening. *Physiologia plantarum*, 106(1), 135-141.
- Nuarsa, I. W., Nishio, F., & Hongo, C. (2011). Relationship between rice spectral and rice yield using MODIS data. *Journal of Agricultural Science*, 3(2), 80.
- Pearson, R. L., & Miller, L. D. (1972). Remote mapping of standing crop biomass for estimation of the productivity of the shortgrass prairie. *Remote sensing of environment*, VIII, 1355.
- Peñuelas, J., Filella, I., & Gamon, J. A. (1995). Assessment of photosynthetic radiation-use efficiency with spectral reflectance. *New Phytologist*, 131(3), 291-296.
- Qi, J., Chehbouni, A., Huete, A. R., Kerr, Y. H., & Sorooshian, S. (1994). A modified soil adjusted vegetation index. *Remote sensing of environment*, 48(2), 119-126.
- Richardson, A. J., & Wiegand, C. (1977). Distinguishing vegetation from soil background information. *Photogrammetric engineering and remote sensing*, 43(12), 1541-1552.
- Rondeaux, G., Steven, M., & Baret, F. (1996). Optimization of soil-adjusted vegetation indices. *Remote sensing of environment*, 55(2), 95-107.
- Roujean, J.-L., & Breon, F.-M. (1995). Estimating PAR absorbed by vegetation from bidirectional reflectance measurements. *Remote sensing of environment*, 51(3), 375-384.
- Rouse Jr, J., Haas, R. H., Deering, D., Schell, J., & Harlan, J. C. (1974). *Monitoring the vernal advancement and retrogradation (green wave effect) of natural vegetation*.
- Tucker, C. J. (1979). Red and photographic infrared linear combinations for monitoring vegetation. *Remote sensing of environment*, 8(2), 127-150.
- Vincini, M., Frazzi, E., & D'Alessio, P. (2006). Angular dependence of maize and sugar beet VIs from directional CHRIS/Proba data. Proc. 4th ESA CHRIS PROBA Workshop,
- Woebbecke, D. M., Meyer, G. E., Von Bargen, K., & Mortensen, D. A. (1995). Color indices for weed identification under various soil, residue, and lighting conditions. *Transactions of the ASAE*, 38(1), 259-269.