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Supplemental information

Etiological involvement of KCND1 variants

in an X-linked neurodevelopmental

disorder with variable expressivity

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Figure S1. Structure of the ternary Kv4/KChIP/DPP channel complex

The structure of the Kv4.2/KChIP1/DPP6 complex reported by Kise and co-workers (PDB ID 7E8H)¹ was used to give an overview of the ternary channel configuration (a-subunits: grey, KChIPs: orange, DPPs: blue) and the location of *KCND1* variant-associated amino acid substitutions. Amino acid exchanges at sites homologous to Kv4.1 were simulated with the most likely backbone-dependent rotamer orientation using PyMOL (Schrödinger). Protein backbones depicted as ribbon diagrams. Substituted amino acid side chains depicted as red (group 1 variants) or purple (group 2 variants) CPK models. In each case one of the four corresponding substitutions is indicated and labeled (unlabeled structure also available as Movie S1). Note that p.Thr516Ser, p.Arg536Gly and p.Asn578Ile are absent because the structure lacks the C-terminal ends of the α -subunits. Horizontal dotted lines indicate the location of the plasma membrane separating extracellular from cytoplasmic space.

1

3 4

 \bullet **2**

Structural homology modeling: Structural homology modeling:

Figure S2. Structural side chain modification caused by *KCND1* **variant-associated amino acid substitutions**

Structural homology modeling using UniProt (https://www.uniprot.org/), SWISS MODEL (Center for Molecular Life Sciences, University of Basel, Switzerland; https://swissmodel.expasy.org/) and PyMOL (Schrödinger), was performed based on the amino acid sequences of human Kv4.1 (UniProt identifier: Q9NSA2), KChIP2 (UniProt identifier: Q9NS61) and DPP6 (UniProt identifer: P42658), and the ternary Kv4.2/KChIP1/DPP6 structure reported by Kise and co-workers (PDB ID 7E8H).¹ The structure provided by homology modeling was incomplete showing a 4:2:2 stoichiometry due to the lack of two opposite KChIP and DPP molecules (see cartoons). Computer-simulated amino acid substitutions using PyMOL (Schrödinger) were done with consideration of the most likely backbone-dependent rotamer orientation, and the putative steric consequences of the amino acid substitution within a distance of 8 Å were taken into account. Protein backbones depicted as ribbon diagrams. Native (wild-type) and novel variant-associated side chains depicted in stick mode, native residues in green, mutated residues in red (group 1 variants) or purple (group 2 variants).

Arg 146

Arg146Cys

Figure S3. Effects of auxiliary b**-subunit co-expression on Kv4.1 channel-mediated peak current amplitudes**

Mean peak current amplitudes (data from Tables S4 – S6, error bars are SD) obtained in the absence (empty bars) and presence of either KChIP or DPP, as indicated by the filled bar extensions.

(A) Effects of KChIP co-expression; p.Lys450* data plotted on two different y-scales;

(B) Effects of DPP co-expression; p.His308Tyr and p.Lys450* data plotted on two different y-scales;

Statistics are based on unpaired Student's *t*-tests, applied to the data from Tables S4 – S6; significant effects of auxiliary b-subunit coexpression (compared to Kv4.1 alone) are indicated with * (p<0.05) or ** (p<0.0001) and by color (significant KChIP effect: orange; significant DPP effect: blue; absence of or non-significant effects: black). Note that the typical incease in peak current amplitude caused by both KChIP and DPP co-expression,^{2,3} as seen for Kv4.1 WT and the majority of variants, is absent in the case of p.Arg92Cys, p.Asp115Asn and p.Arg146Cys for KChIP co-expression, and in the case of p.His308Tyr for DPP co-expression.

Figure S4. Effects of auxiliary b**-subunit co-expression on Kv4.1 channel macroscopic inactivation and recovery kinetics** Mean values of inactivation and recovery time constants (data from Tables S4 – S6, error bars are SD and in many cases smaller than symbols) obtained in the absence (empty symbols) and presence of either KChIP or DPP (filled symbols).

(A and C) Effects of KChIP co-expression;

(B and D) Effects of DPP co-expression;

Statistics are based on unpaired Student's *t*-tests, applied to the data from Tables S4 – S6; significant effects of auxiliary bsubunit co-expression (compared to Kv4.1 alone) are indicated with * (p<0.05) or ** (p<0.0001) and by color (significant KChIP effect: orange; significant DPP effect: blue; absence of or non-significant effects: black). Typically, KChIP causes a slowing of the initial (fast) and an acceleration of the second (slower) current decay component (τ_1 and τ_2 depicted as circles and triangles, respectively), whereas DPP causes an acceleration of the initial (fast) current decay component (τ_1) , while both KChIP and DPP speed up recovery from inactivation.^{2,3} These typical modifications were seen for Kv4.1 WT and, at least partially, for the majority of variants. Still, for p.Lys450* KChIP failed to accelerate the recovery kinetics, and for p.His308Tyr none of the typical DPP effects was seen. Notably, in the case of p.Arg146Cys, p.Arg536Gly and p.Asn578Ile KChIP caused an accceleration of the initial current decay component (filled green symbols).

Figure S5. Effects of auxiliary ß-subunit co-expression on the voltage dependences of Kv4.1 channel activation and steady-state inactivation

Mean values obtained for the voltages of halfmaximal activation and inactivation (data from Tables S4 – S6, error bars are SD and in many cases smaller than symbols) obtained in the absence (empty symbols) and in the presence of either KChIP or DPP (filled symbols).

(A) Effects of KChIP co-expression;

(B) Effects of DPP co-expression;

Statistics are based on unpaired Student's *t*-tests, applied to the data from Tables S4 – S6; significant effects of auxiliary B-subunit co-expression (compared to Kv4.1 alone) are indicated with * (p<0.05) or ** (p<0.0001) and by color (significant KChIP effect: orange; significant DPP effect: blue; absence of or non-significant effects: black). Typically KChIP causes a positive shift of the inactivation curve, and DPP a negative shift of both activation and inactivation curve,^{2,3} as seen for Kv4.1 WT and, at least partially, for the majority of variants. However, in the case of p.Lys450* KChIP failed to significantly shift the inactivation curve in the positive direction, and in the case of p.Asp115Asn DPP failed to significantly shift both curves in the negative direction.

Table S1. Key clinical features of subjects with hemizygous *KCND1* **variants** (group 1 and group 2)

Z-scores for birth and growth parameters were calculated using the Ped(z) Pediatric Calculator (https://www.pedz.de/en/welcome.html). Abbreviations: abt, about; ASD, autism spectrum disorder; DD, developmental delay; exam, examination; DWI, diffusion-weighted magnetic resonance imaging; FSIQ, full scale intelligence quotient; ID, intellectual disability; IUGR, intrauterine growth retardation; m, months; MI, maternally inherited; ND, no data; OFC, occipitofrontal head circumference; PAPP-A, pregnancyassociated plasma protein A; RN, reported normal; SD, standard deviation; SGA, small for gestational age; TIQ, total intelligence quotient; WISC, Wechsler intelligence scale for children; WNV, Wechsler nonverbal scale of ability; wks, weeks; WPPSI, Wechsler preschool and primary scale of intelligence; y, years; z, z-score. ^aNo clear evidence of ataxia, no broad-based gait, no coordination problems, no muscle weakness or hypertonia, but rather a somewhat unstable gait pattern possibly reflecting hypermobility or a general gross motor delay, ^bDied at eight months due to respiratory insufficiency, probably linked to progressive hypo-myelinating encephalopathy, ^cBrothers of different age, ^dAt ten years of age, decline probably due to suspected childhood disintegrative disorder, ^eIncreasingly frequent use of a wheel chair since the onset of suspected childhood disintegrative disorder

Table S2. Polar contacts involving side chains in a modeled Kv4.1/KChIP2/DPP6 structure

A computer search for polar interactions involving side chains was performed on the Kv4.1/KChIP2/DPP6 structure provided by homology modeling (see cartoons: 4 α -subunits, grey; 2 KChIPs, orange; 2 DPPs, blue; see also Figure S2). Amino acid substitutions were modeled and searches for polar side chain interactions including distance measurements were performed for wild-type and variant α -subunits. The results obtained for the residues of interest (wildtype: green; group 1 mutants: red; group 2 mutants: purple) in α -subunits 1 and 4 are illustrated. Numbers separated by dashes represent (in this order): subunit of the residue under study $(1 \text{ or } 4)$ – subunit of the interaction partner – smallest distance between interacting atoms. All three possible combinations of side chain and backbone involvement are considered. Note that in the majority of cases the amino acid substitution leads to a reduction in the number of contacts.

Table S3. Biophysical parameters for Kv4.1 wild-type and variant ternary channels

Kv4.1 channels (grey, variant channels with red or purple dots) were studied in a ternary configuration; i.e. in the presence of both KChIP (orange) and DPP (blue). Mean values \pm SD and number of observations (n) are given for the peak current amplitude at + 40 mV (I_{+40}), the time constants of macroscopic inactivation (τ_1 and τ_2) including the fractional amplitude of τ_1 (%), the time constant of recovery from inactivation (τ_{rec}) and the voltages of halfmaximal inactivation (V_{1/2,inact}) and activation (V_{1/2,act}) with corresponding slope factors (k_{inact} and k_{act} , respectively). Statistics are based on one-way analysis of variance (ANOVA) with Dunnett's post hoc testing. For significant variant effects (i.e., deviations from Kv4.1 WT ternary; key variants: red, maternally inherited missense variants: purple) significance levels are indicated with * (p<0.05) or ** (p<0.0001); shaded fields: no significant differences found compared to Kv4.1 WT ternary.

Table S4. Biophysical parameters for Kv4.1 wild-type and variant homotetrameric channels

Kv4.1 channels (grey, variant channels with red or purple dots) were expressed in the absence of auxiliary β -subunits. Mean values \pm SD and number of observations (n) are given for the peak current amplitude at + 40 mV (I_{+40}), the time constants of macroscopic inactivation (τ_1 and τ_2) including the fractional amplitude of τ_1 (%), the time constant of recovery from inactivation (τ_{rec}) and the voltages of halfmaximal inactivation (V_{1/2,inact}) and activation (V_{1/2,act}) with corresponding slope factors (k_{inact} and k_{act} respectively); Statistics are based on one-way analysis of variance (ANOVA) with Dunnett's post hoc testing. For significant variant effects (i.e., deviations from Kv4.1 WT; key variants: red, maternally inherited missense variants: purple) significance levels are indicated with * (p<0.05) or ** (p<0.0001); shaded fields: no significant differences found compared to Kv4.1 WT.

Table S5. Biophysical parameters for Kv4.1 wild-type and variant binary channels with KChIP

Kv4.1 channels (grey, variant channels with red or purple dots) were studied in the presence of KChIP (orange). Mean values ± SD and number of observations (n) are given for the peak current amplitude at +40 mV (I_{+40}), the time constants of macroscopic inactivation (τ_1 and τ_2) including the fractional amplitude of τ_1 (%), the time constant of recovery from inactivation (τ_{rec}) and the voltages of halfmaximal inactivation (V_{1/2,inact}) and activation (V_{1/2,act}) with corresponding slope factors (k_{inact} and k_{act}, respectively); Statistics are based on one-way analysis of variance (ANOVA) with Dunnett's post hoc testing. For significant variant effects (i.e., deviations from Kv4.1 WT + KChIP; key variants: red, maternally inherited missense variants: purple) significance levels are indicated with * (p<0.05) or ** (p<0.0001); shaded fields: no significant differences found compared to Kv4.1 WT + KChIP.

Table S6. Biophysical parameters for Kv4.1 wild-type and variant binary channels with DPP

Kv4.1 channels (grey, variant channels with red or purple dots) were studied in the presence of DPP (blue). Mean values ± SD and number of observations (n) are given for the peak current amplitude at +40 mV (I_{+40}), the time constants of macroscopic inactivation (τ_1 and τ_2) including the fractional amplitude of τ_1 (%), the time constant of recovery from inactivation (τ_{rec}) and the voltages of halfmaximal inactivation ($V_{1/2,\text{inact}}$) and activation ($V_{1/2,\text{act}}$) with corresponding slope factors (*k_{inact}* and *k_{act}*, respectively); Statistics are based on one-way analysis of variance (ANOVA) with Dunnett's post hoc testing. For significant variant effects (i.e., deviations from Kv4.1 WT + DPP; key variants: red, maternally inherited missense variants: purple) significance levels are indicated with * (p<0.05) or ** (p<0.0001).

Table S7. PS3 scoring for functionally expressed *KCND1* **variants**

Functional *KCND1* variants were assessed based on significant alterations relative to Kv4.1 WT. For each biophysical parameter and in all channel configurations (t: ternary, a: alone, K: only with KChIP, D: only with DPP) the following questions were asked: 1) Is there a significant difference relative to Kv4.1 WT? Yes (Y) or No (N); 2) Is the ß-subunit effect on this biophysical parameter absent (i.e.; no KChIP effect, = ± K or no DPP effect, = ± D)? Yes (Y) or No (N). Assessment based on the data from Tables S3 - S6; see also Figures S3 - S5.

Supplemental References

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