Single cell regulatory architecture of human pancreatic islets suggests sex differences in β cell function and the pathogenesis of type 2 diabetes.

Mirza Muhammad Fahd Qadir^{1,2,3}, Ruth M. Elgamal^{4,5}, Keijing Song⁶, Parul Kudtarkar⁵, Siva S.V.P Sakamuri⁷, Prasad V. Katakam⁷, Samir El-Dahr⁸, Jay Kolls⁶, Kyle J. Gaulton⁵, Franck Mauvais-Jarvis^{1,2,3,10,}

¹Section of Endocrinology and Metabolism, John W. Deming Department of Medicine, Tulane University School of Medicine, New Orleans, LA, USA

²Southeast Louisiana Veterans Health Care System, New Orleans, LA, USA

³Tulane Center of Excellence in Sex-Based Biology & Medicine, New Orleans, LA, USA

⁴Biomedical Sciences Graduate Program, University of California, San Diego, La Jolla, CA, USA

⁵Department of Pediatrics, University of California, San Diego, La Jolla, CA, USA

⁶Center for Translational Research in Infection and Inflammation, John W. Deming Department of Medicine, Tulane University School of Medicine, New Orleans, LA, USA

⁷Department of Pharmacology, Tulane University School of Medicine, New Orleans, LA, USA

⁸Department of Pediatrics, Tulane University, School of Medicine, New Orleans, LA, USA

¹⁰Lead contact: fmauvais@tulane.edu



Extended Data Fig.1 Donor metadata, scRNAseq quality control metrics and network analysis of all nondiabetic cell cells.

a, Sunburst plot showing donor metadata of scRNAseq datasets used in this study. n=52 donors from Tulane University and the Human Pancreas Analysis Program (HPAP). Metadata is organized into source, disease status, sex, and race across all donor profiles. b, Percentage distribution across all 17 cell clusters for each of the n=52 donors used in this study. Cells are colored based on a key (legend) and metadata is organized based on information shown in (a). c, Violin plots showing genes expressed per cell, unique molecular identifiers (UMIs) per cell, percentage mitochondrial genes and library complexity [log10(number of genes)/log10(number of UMIs)] across each of the donor sources. d, Quality control metrics for cell number, mean RNA reads/cell, median genes/cell, total reads, valid barcodes, sequencing saturation, percentage of reads mapped confidently to the reference genome, reads mapped confidently to intronic regions, reads mapped confidently to exonic regions, fraction of reads/cell, total genes detected/cell, median UMI counts/cell, basal metabolic index and age, across all donors split on the basis of source. e, Pearson's correlation between normalized read coverage within a merged set of RNAseq expression profiles across 52 donors. Annotation bars highlight the sex, race, cell type and diabetes status of the donors. f, Heatmap showing normalized aggregated RNAseq expression profiles for select Y and X linked chromosome genes across race, sex, source, celltype and disease. g, UMAP plots showing RNAseq expression profiles for select cognate genes across each cell type specific cell cluster. h, Gene ontology network analysis showing regulated pathways across all non-diabetic islet cells. Size of circle equivalent to gene contribution. (d) p < 0.05 is significant (h) FDR adjusted p < 0.2 is significant. n = 52 donors



Extended Data Fig.2 Differentially expressed genes and pathways in human islet cells across sex and race.

a, Venn diagrams showing differentially expressed genes (DEGs) across sex in case of non-diabetic: beta, alpha, delta, gamma, beta+alpha, beta+delta, cycling endo, acinar, ductal, activated stellate, quiescent stellate, endothelial, lymphocyte, macrophage, mast and schwann cells. **b**, Gene ontology (GO) analysis of celltypes other than α/β shown in (a). c, Venn diagrams showing differentially expressed genes (DEGs) across ancestry for cells highlighted in (a). d, Gene ontology (GO) analysis of all celltypes other than α/β shown in (c). DGE: FDR adjusted pval < 0.1 is significant, GO: FDR adjusted pval < 0.2 is significant, n = 36 (non-diabetic).



Extended Data Fig.3 snATACseq quality control metrics and network analysis of all non-diabetic cells.

a, Density scatter showing transcriptional start site (TSS) enrichment on y-axis and number of peak counts on the x-axis. **b**, QC plots showing TSS, fragment reads in accessible peaks (FRAP) and fragment reads in promoters (FRIP) across all donors. Dashed red line signifies cutoff. **c**, QC plots showing metrics for detected cells, fraction fragments overlapping peaks, fraction cells mapped confidently to genome, annotated cells, number of fragments, total usable fragments, median fragments/cell at 50K raw reads/cell (RRPC) and TSS enrichment score across all donors. **d**, Post-filtering QC metrics showing number of peaks, TSS enrichment, blacklist ratio, nucleosome signal, percentage reads in peaks and FRIP across all donors. **e**, Pearson's correlation between aggregated read coverage from snATACseq experiments. **f**, Pearson's' correlation between aggregated predicted RNA expression in snATACseq experiments based on mapped expression models from scRNAseq experiments. **h**, Gene activity based on accessibility in 100kb windows flanking select cognate genes across cell clusters (top row). Predicted gene expression based on expression data mapping from scRNAseq atlas (bottom row). **i**, Differential analysis using chromVAR and the JASPAR 2020 motif database showing enriched motifs for beta, delta, alpha, gamma, ductal, acinar, quiescent stellate, activated stellate, endothelial and macrophage cells. DGE: FDR adjusted pval < 0.05 is significantly enriched. n = 15 non-diabetics.



Extended Data Fig.4 Dynamic insulin and glucagon secretion coupled with islet bioenergetics.

a, Dynamic insulin secretion assay, showing response to 16.7mM glucose, IBMX + 16.7mM Glucose, epinephrine + 1.7mM Glucose and potassium chloride + 5.6mM glucose. Each curve represents secretion normalized to total insulin content across sex and race. **b**, Area under the curve (AUC) measurements for incretin driven insulin secretion measurements outlined in (a). **c**, Dynamic insulin secretion assay, showing response to 16.7mM glucose, IBMX + 16.7mM Glucose, epinephrine + 1.7mM Glucose and potassium chloride + 5.6mM

glucose. Each curve represents secretion normalized to total insulin content across sex. d, Area under the curve (AUC) measurements for incretin driven insulin secretion measurements outlined in (b). e, Dynamic insulin secretion assay using an ascending glucose ramp. Stimulation modes show response to 7mM glucose, 11mM Glucose, 20mM Glucose, potassium chloride + 2mM glucose and, potassium chloride + 11mM glucose. Each curve represents secretion normalized to total insulin content across sex and race. f, Area under the curve (AUC) measurements for incretin driven insulin secretion measurements outlined in (e). g, Dynamic insulin secretion assay using an ascending glucose ramp. Stimulation modes show response to 7mM glucose, 11mM Glucose, 20mM Glucose, potassium chloride + 2mM glucose and, potassium chloride + 11mM glucose. Each curve represents secretion normalized to total insulin content across sex and race. h, Area under the curve (AUC) measurements for incretin driven insulin secretion measurements outlined in (g). i, Dynamic glucagon secretion assay, showing response to 16.7mM glucose, IBMX + 16.7mM Glucose, epinephrine + 1.7mM Glucose and potassium chloride + 5.6mM glucose. Each curve represents secretion normalized to total glucagon content across sex and race. j, Area under the curve (AUC) measurements for incretin driven insulin secretion measurements outlined in (i). k, Dynamic glucagon secretion assay, showing response to 16.7mM glucose, IBMX + 16.7mM Glucose, epinephrine + 1.7mM Glucose and potassium chloride + 5.6mM glucose. Each curve represents secretion normalized to total glucagon content across sex. I, Area under the curve (AUC) measurements for incretin driven insulin secretion measurements outlined in (k). m, Oxygen consumption ratio for islets across sex and race. n, Basal respiration, glucose mediated respiration, maximal (max) respiration, ATP mediated respiration, non-electron transport chain (ETC) respiration and coupling efficiency, across sex and race. o, Oxygen consumption ratio for islets across sex. I, Basal respiration, glucose mediated respiration, maximal (max) respiration, ATP mediated respiration, non-electron transport chain (ETC) respiration and coupling efficiency, of human islets across sex. q, Extracellular acidification rate for islets across sex and race. r, Baseline acidification and total acidification of human islets across sex and race. s, Extracellular acidification rate for islets across sex. t, Baseline acidification and total acidification of human islets across sex. *pval < 0.05, **pval < 0.01 is significant. n = 15 (non-diabetic).



Extended Data Fig.5 Dynamic hormone secretion profiles of HPAP type 2 diabetic donors.

a, Dynamic insulin secretion assay of human islets subject to Amino acid cocktail (AA) 4mM, AA 4mM + 3mM Glucose, AA 4mM +16.7mM glucose, AA 4mM + 16.7mM Glucose + IBMX 0.1mM and KCI 30mM. Each curve represents mean of each donor type across sex and disease. **b**, Area under the curve for insulin secretion at baseline, AA 4mM, AA 4mM + 3mM Glucose, AA 4mM +16.7mM glucose, AA 4mM + 16.7mM Glucose + IBMX 0.1mM and KCI 30mM. **c**, Dynamic glucagon secretion assay of human islets subject to secretagogues as in (**a**). **d**, Area under the curve for glucagon secretion like (**a**). Data analyzed from HPAP dataset. n= 21 non-diabetic and n=16 T2D diabetic donors. **pvalue < 0.01



Extended Data Fig.6 Gene ontology networks in human islet cells from type 2 diabetes donors.

a-h, Gene ontology plots highlighting upregulated and downregulated pathways in T2D across sex. Pathways compare type two diabetic males to non-diabetic males and similarly for females. Data is shown for: **a**, Ductal cells; **b**, Acinar cells; **c**, Quiescent stellate cells. **d**, Activated stellate cells; **e**, Endothelial cells; **f**, Macrophage cells; **g**, Lymphocyte cells; **h**, Mast cells. DGE: FDR adjusted pval < 0.1 is significant, GO: FDR adjusted pval < 0.2 is significant, n = 36 (non-diabetic) and n = 16 T2D.