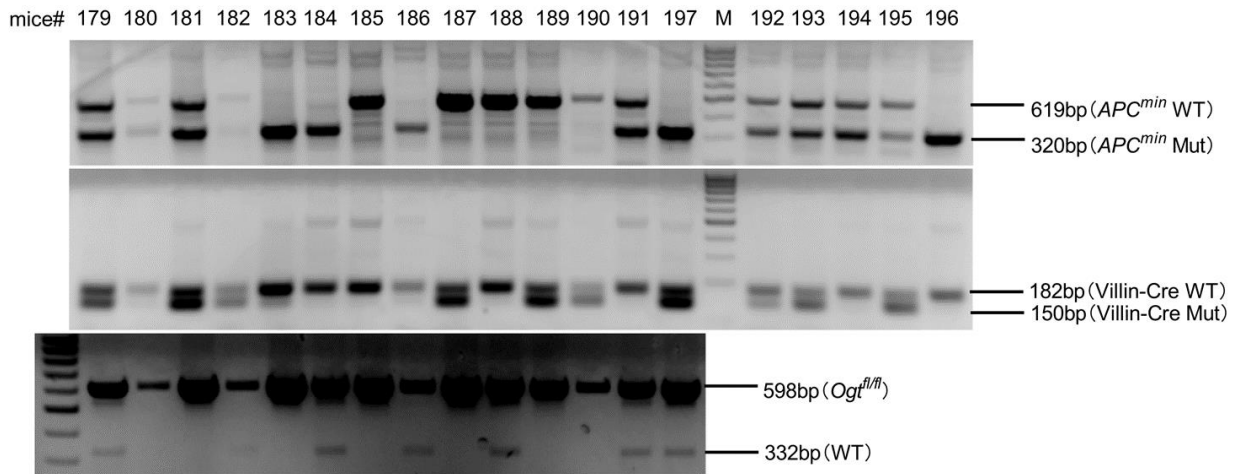


**Supplemental Fig. 1 The expression pattern of OGT in TCGA and CTPAC databases.**

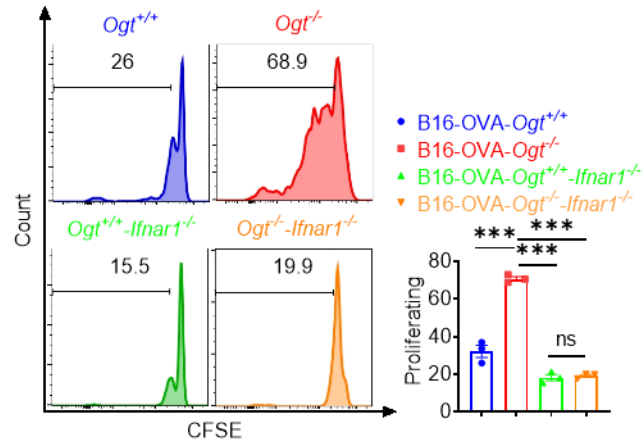
**A)** Boxplot showing mRNA expression level of *Ogt* in multiple types of cancers. The plot was generated using the GEPIA2 online server. \* $p < 0.05$ . **B-D)** Boxplot showing mRNA expression level of *Ogt* in LUAD, Normal and tumor samples (**B**), Individual stages (**C**), Nodal metastasis

status (**D**). The plot was generated using the UALCAN online server. **E-F**) Boxplot showing protein expression level of OGT in LUAD, Normal and tumor samples (**E**), Individual stages (**F**). The plot was generated using the UALCAN online server (<https://ualcan.path.uab.edu/analysis.html>). Statistical significance was determined by Pearson test, unpaired Student's t-test, \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , ns, no significant difference. Data represent the mean of  $\pm$  SD.



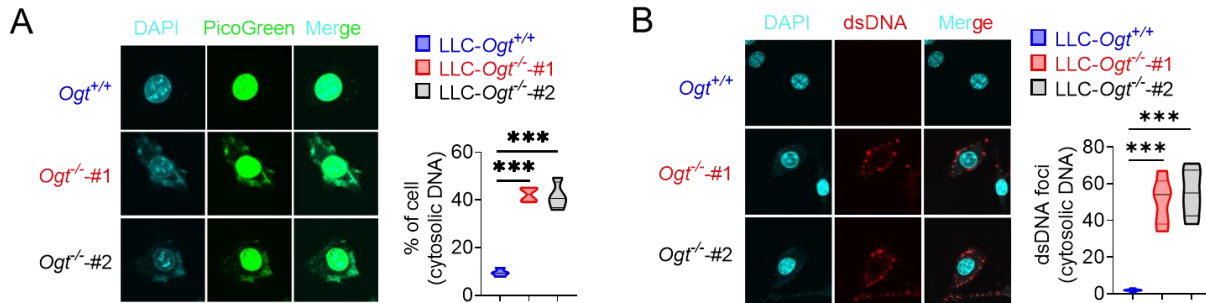
**Supplemental Fig. 2. The genotype of *APC<sup>min</sup>*, *Villin-Cre* and *Ogt<sup>fl/fl</sup>* mice.**

Genomic DNA was extracted from the tails of *APC<sup>min</sup>*, *Villin-Cre*, and *Ogt<sup>fl/fl</sup>* mice and used for PCR with various primers. The resulting products were separated by agarose gel electrophoresis to determine the genotype.



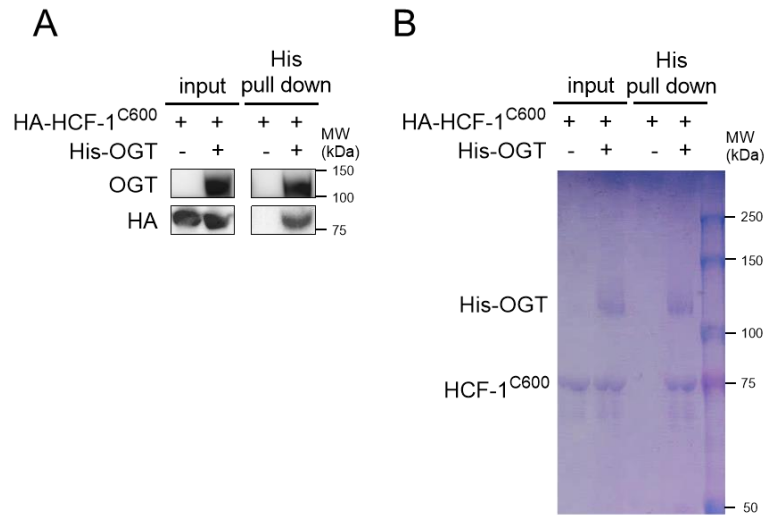
### Supplemental Fig. 3 *In vitro* Cross-Priming of T Cells by *Ifnar*<sup>-/-</sup> BMDCs.

*Ifnar1*<sup>-/-</sup> BMDCs was pre-treated with B16-OVA-*Ogt*<sup>+/+</sup> or B16-OVA-*Ogt*<sup>-/-</sup> supernatant, and co-cultured with OT-1 T cell, then T cell proliferation was evaluated by flow cytometry. Representative fluorescence-activated cell sorting histograms and statistical data are shown. Data are representative of three independent experiments. Statistical significance was determined by one-way ANOVA, \**p* < 0.05, \*\**p* < 0.01, \*\*\**p* < 0.001, ns, no significant difference. Data represent the mean of ± SD.



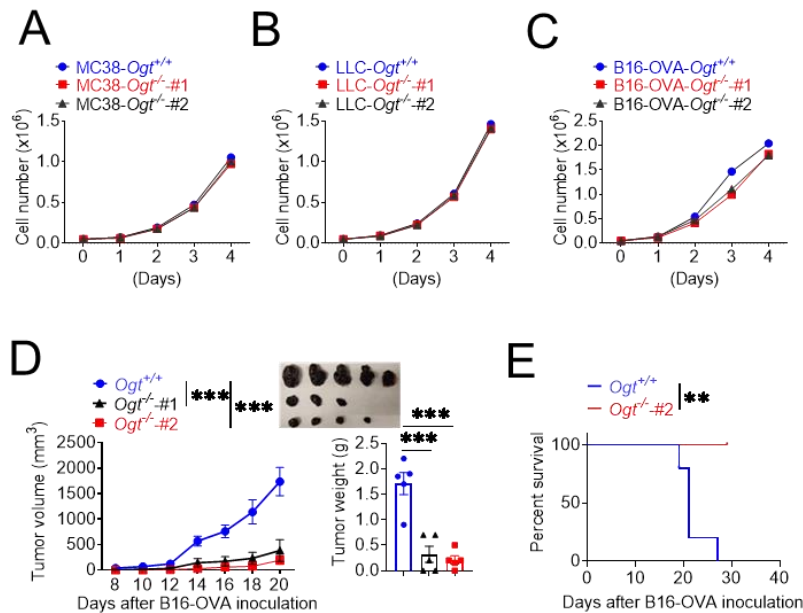
**Supplemental Fig. 4 OGT deficiency causes DNA damage and accumulates cytosolic DNA.**

**A)** The extranuclear dsDNA in different *Ogt*<sup>-/-</sup> LLC clones were determined by PicoGreen staining assay and was quantified by image J. **B)** The extranuclear dsDNA in different *Ogt*<sup>-/-</sup> LLC clones were determined by anti-dsDNA fluorescence staining assay and was quantified by image J. Data are representative of three independent experiments. Statistical significance was determined by unpaired Student's t-test, two-way ANOVA, \**p* < 0.05, \*\**p* < 0.01, \*\*\**p* < 0.001, ns, no significant difference. Data represent the mean of ± SD.



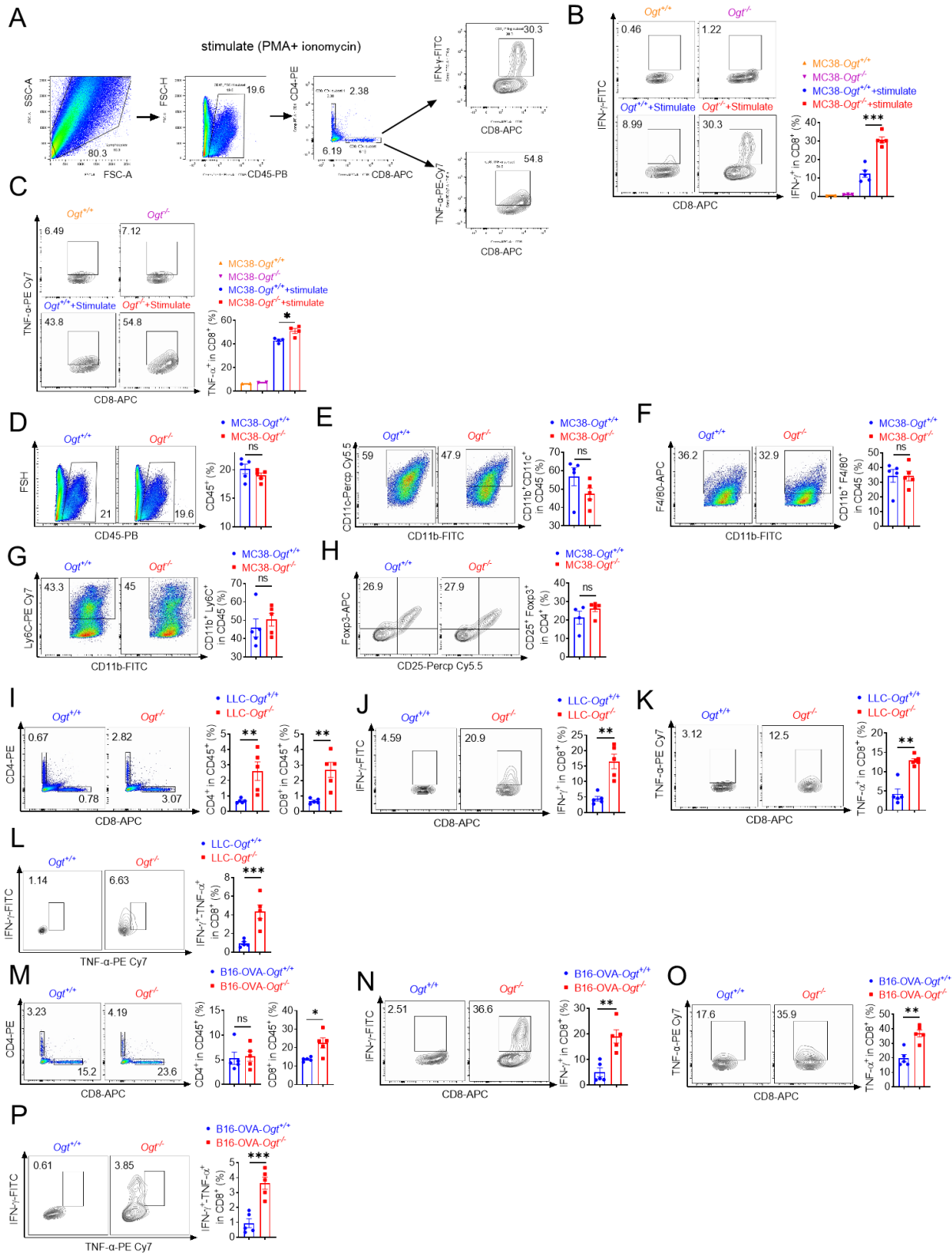
**Supplemental Fig. 5 *In vitro* pull-down assay analysis of the interaction of HCF-1<sup>C600</sup> and OGT.**

**A)** His pull-down assays were used to analyze the interaction between HCF-1C600 and OGT. **B)** His-tagged OGT and HCF1 were expressed and purified, followed by SDS-PAGE separation and staining with Coomassie blue.



**Supplemental Fig. 6 The cell proliferation of different tumor model *in vitro* and B16-OVA tumor growth analysis *in vivo*.**

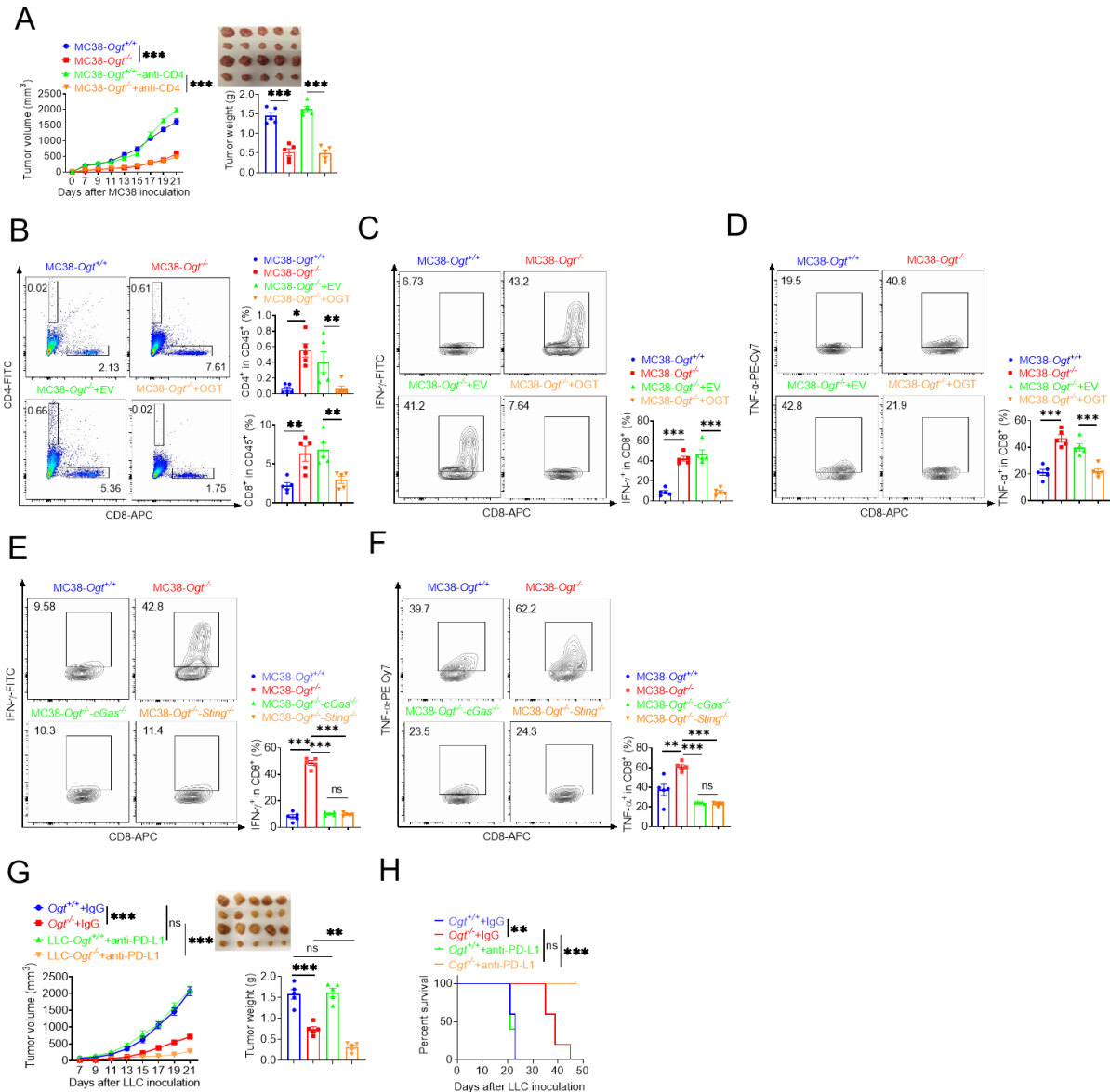
**A-C)**, The cell proliferation of different  $Ogt^{-/-}$  tumor model *in vitro*. **A)** MC38, **B)** LLC, **C)** B16-OVA  $Ogt^{-/-}$  cells proliferation *in vitro*. **D-E)** Tumor volume, weight of  $Ogt^{+/+}$  or  $Ogt^{-/-}$  B16-OVA tumors in C57BL/6J mice, and mice survival, n=5 respectively. Data are representative of two or three independent experiments. Statistical significance was determined by unpaired Student's t-test, two-way ANOVA, \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , ns, no significant difference. Data represent the mean of  $\pm$  SD.





**Supplemental Fig. 7 *Ogt* deficiency inhibits tumor progression through enhancing infiltration of CD8<sup>+</sup> T cells.**

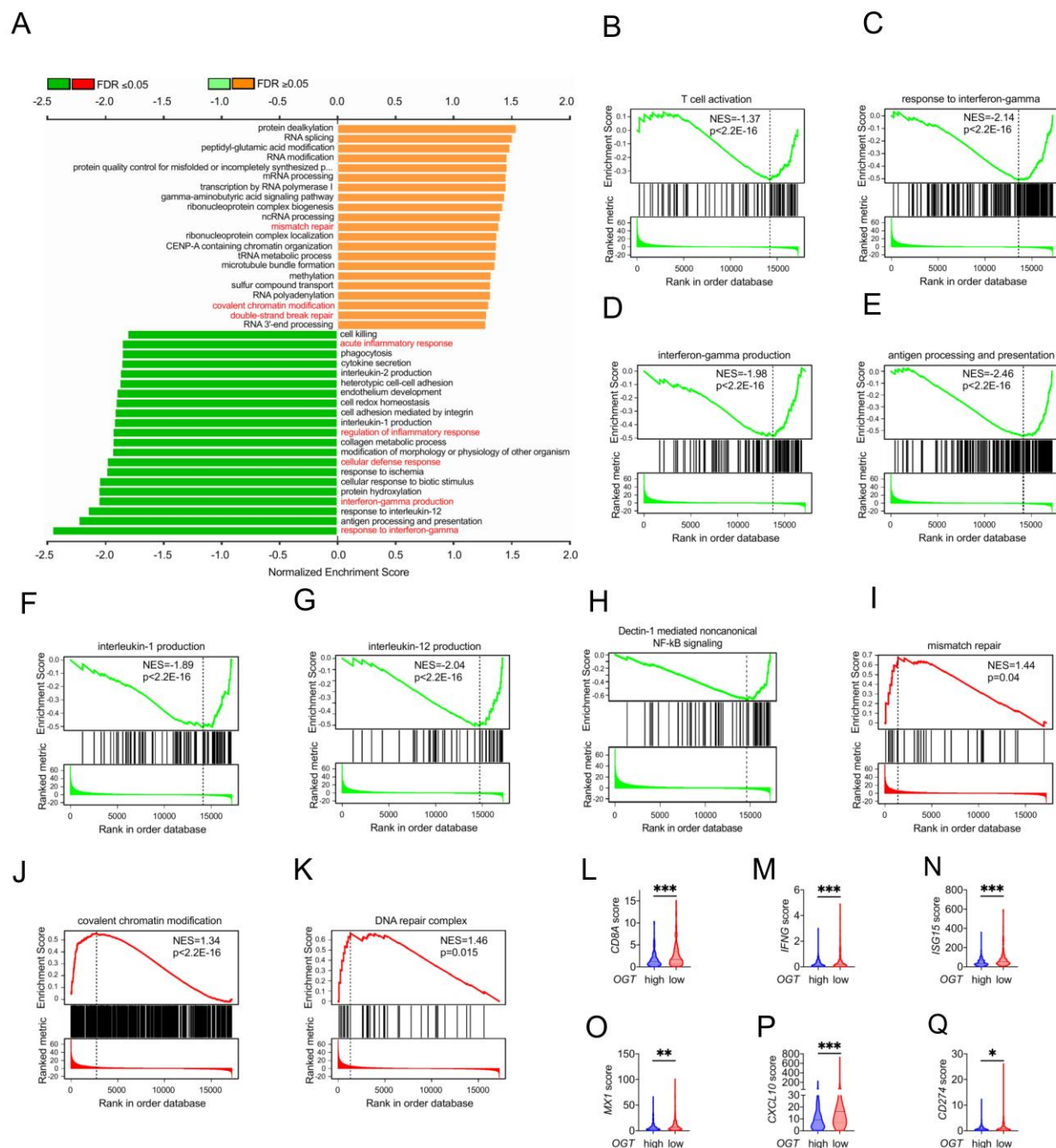
**A)** A total schematic of flow cytometry analysis. **B-C)** Flow cytometry analysis showing percentage of IFN- $\gamma$  and TNF- $\alpha$ -expressing intratumoral CD8<sup>+</sup> T cells in MC38 tumors with or without PMA and ionomycin stimulation. **D-H)** Flow cytometry analysis showing percentage of CD45<sup>+</sup> (**D**), CD11b<sup>+</sup> CD11c<sup>+</sup> (**E**), CD11b<sup>+</sup> F4/80<sup>+</sup> (**F**), CD11b<sup>+</sup> Ly6C<sup>+</sup> (**G**) and Treg cells (**H**) in MC38 tumor model, n=5 respectively. **I-P)** Flow cytometry analysis showing percentage of CD4<sup>+</sup> and CD8<sup>+</sup> T cells in LLC (**I**), B16-OVA cells (**M**). IFN- $\gamma$ <sup>+</sup>, TNF- $\alpha$ <sup>+</sup> and IFN- $\gamma$ <sup>+</sup> TNF- $\alpha$ <sup>+</sup> double positive expressing intratumoral CD8<sup>+</sup> T cells in LLC (**J-L**), or B16-OVA (**N-P**) tumors isolated at day 18 post-tumor inoculation, n=5 respectively. Data are representative of two or three independent experiments. Statistical significance was determined by unpaired Student's t-test, one-way ANOVA, \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , ns, no significant difference. Data represent the mean of  $\pm$  SD.



**Supplemental Fig. 8. *Ogt* deficiency inhibits tumor progression through enhancing infiltration of CD8<sup>+</sup> T cells.**

**A)** Tumor volume and weight of *Ogt*<sup>+/+</sup> or *Ogt*<sup>-/-</sup> MC38 tumors injected with control IgG or anti-CD4 antibody at day 0, 7 and 14 post tumor inoculation in C57BL/6J mice, n=5 respectively. **B-D)** The percentage of CD4<sup>+</sup> and CD8<sup>+</sup> T cells (**B**), CD8<sup>+</sup> IFN- $\gamma$ <sup>+</sup> (**C**) and CD8<sup>+</sup> TNF- $\alpha$ <sup>+</sup> (**D**) in *Ogt*<sup>-/-</sup> rescued MC38 tumors isolated at day 18 post-tumor inoculation. **E-F)** Flow cytometry analysis

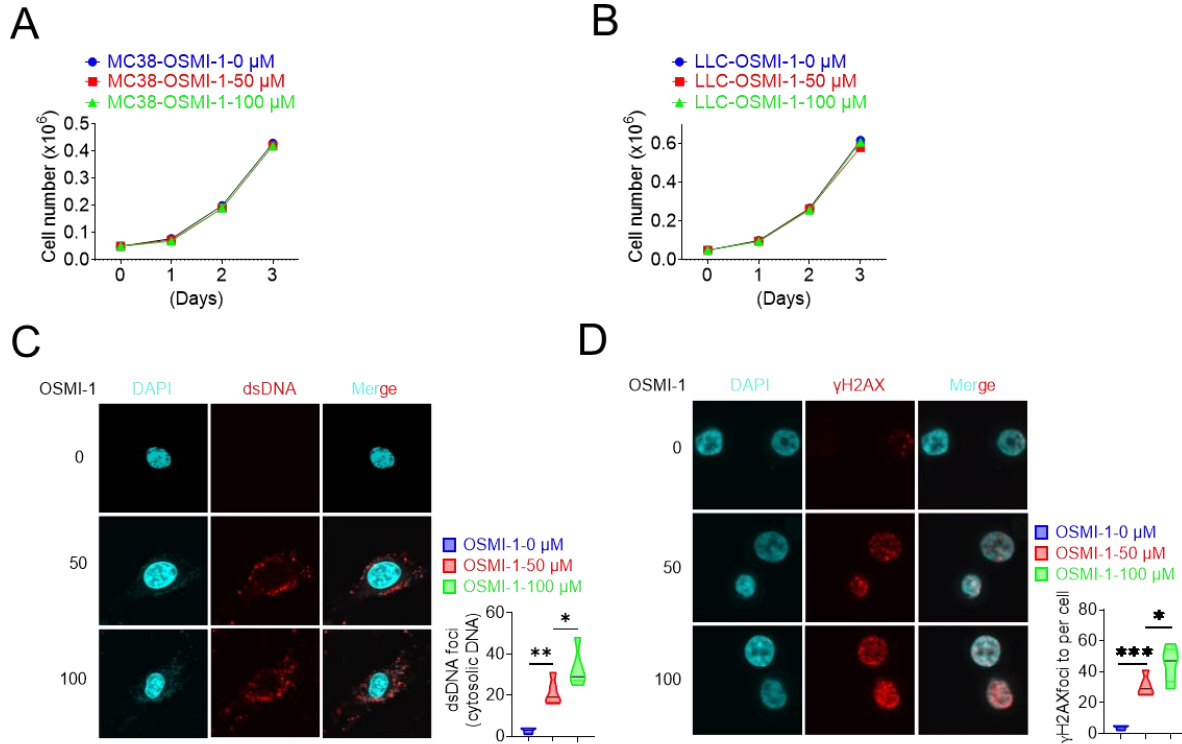
showing percentage of CD8<sup>+</sup> IFN- $\gamma$  (**E**), CD8<sup>+</sup> TNF- $\alpha$ <sup>+</sup> (**F**) in *Ogt*<sup>-/-</sup>*cGAS*<sup>-/-</sup> or *Ogt*<sup>-/-</sup>*Sting*<sup>-/-</sup> double knockout MC38 tumors in C57BL/6J mice, subcutaneous tumor isolated at day 18 post-tumor inoculation. **G-H**) Tumor volume, weight of *Ogt*<sup>+/+</sup> or *Ogt*<sup>-/-</sup> LLC tumors injected with control IgG or anti-PD-L1 antibody at day 7, 10 and 13 post tumor inoculation in C57BL/6J mice, and mice survival, n=5 respectively. Data are representative of two or three experiments. Statistical significance was determined by unpaired Student's t-test, one-way ANOVA, two-way ANOVA, \**p* < 0.05, \*\**p* < 0.01, \*\*\**p* < 0.001, ns, no significant difference. Data represent the mean of  $\pm$  SD.



**Supplemental Fig. 9** *OGT* expression is negatively related to  $CD8^+$  T cells infiltration in human colorectal cancer.

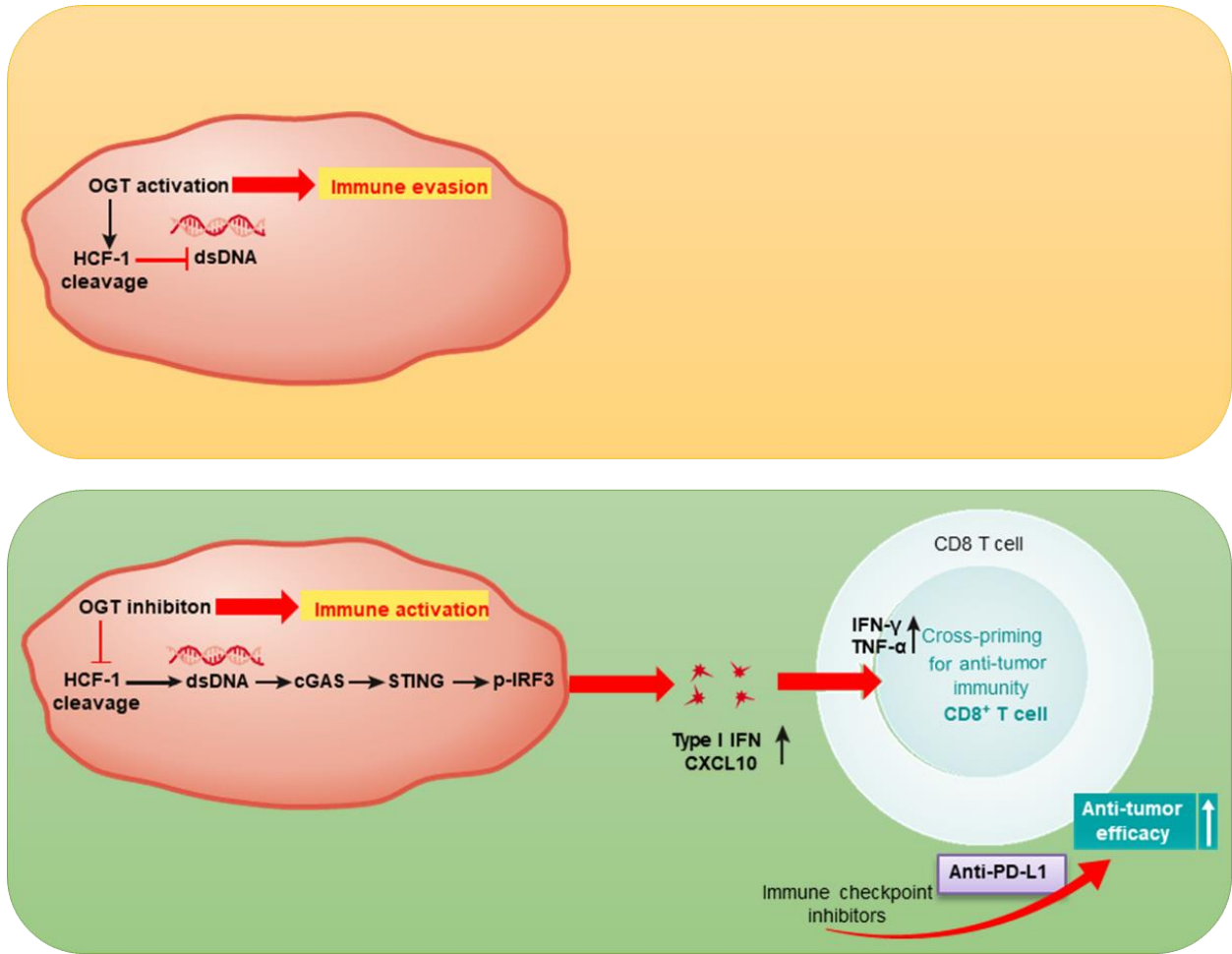
**A)** Gene Ontology (GO) enrichment and pathway analysis in *OGT* high and *OGT* low patients. **B-K)** GSEA analysis in *OGT* high and *OGT* low patients. T cells activation (**B**), response to interferon-gamma (**C**), interferon-gamma production (**D**), antigen processing and presentation (**E**),

interleukin-1 production (**F**), interleukin-12 production (**G**), dectin-1 mediated noncanonical NF- $\kappa$ B signaling (**H**), mismatch repair (**I**), covalent chromatin modification (**J**) and DNA repair complex (**K**) in *OGT* high and *OGT* low patients. **L-Q** RNAseq analysis of mRNA expression pattern in *OGT* high and *OGT* low patients, CD8A (**L**), *IFNG* (**M**), *ISG15* (**N**), *MX1* (**O**), *CD274* (**P**) and *CXCL10* (**Q**) mRNA expression patterns in *OGT* high and *OGT* low patients. Statistical significance was determined by Pearson test, unpaired Student's t-test, \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , ns, no significant difference. Data represent the mean of  $\pm$  SD.



**Supplemental Fig. 10 OSMI-1 could significantly induce a high percentage of DNA damage.**

**A-B)** The cell proliferation in different treatments in *vitro*. **A)** MC38 cell proliferation, **B)** LLC cell proliferation. **C)** The extranuclear dsDNA was measured by anti-dsDNA fluorescence staining treated with 50  $\mu\text{M}$  and 100  $\mu\text{M}$  in LLC cells, respectively. **D)** The  $\gamma\text{H2AX}$  expression was measured anti- $\gamma\text{H2AX}$  fluorescence staining treated with 50  $\mu\text{M}$  and 100  $\mu\text{M}$  in LLC cells, respectively. Data are representative of three experiments. Statistical significance was determined by one-way ANOVA, \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , ns, no significant difference. Data represent the mean of  $\pm$  SD.



**Supplementary Fig. 11 A schematic of critical role of OGT-mediated antitumor immunity.**

Inhibition of O-GlcNAc transferase promotes the activation of cGAS-STING pathway and the production of type I interferon, which enhances CD8 T cells dependent antitumor immunity.

**Table S1. Primer sequences for genotype**

Genes	Sequence	Size
<i>Ogt<sup>fl/fl</sup></i>	Forward CATCTCTCCAGCCCCACAAACTG	WT 332 bp,
	Reverse GACGAAGCAGGAGGGGAGAGCAC	Mutant 487 bp
Villin-Cre ( $\Delta IEC$ )	WT Forward TATAGGGCAGAGCTGGAGGA	WT 182 bp,
	Mut Forward AGGCAAATTTTGGTGTACGG	Mutant 150 bp
	Common Rev GCCTTCTCCTCTAGGCTCGT	
<i>Apc<sup>min</sup></i>	WT Forward GCCATCCCTTCACGTTAG	WT 619 bp,
	Mut Forward TTCTGAGAAAGACAGAAGTTA	Mutant 320 bp
	Common Rev TTCCACTTTGGCATAAAGGC	

**Table S2. Related to Experimental Procedures. Primer sequences for RT-PCR**

Genes	Forward	Reverse
Mouse <i>Ifna4</i>	CCTGTGTGATGCAGGAACC	TCACCTCCCAGGCACAGA
Mouse <i>Ifnb1</i>	ATGAGTGGTGGTTGCAGGC	TGACCTTTCAAATGCAGTAGAGTCA
Mouse <i>Ifng</i>	TCAAGTGGCATAGATGTGGAAGAA	TGGCTCTGCAGGATTTTCATG
Mouse <i>Il1a</i>	GCACCTTACACCTACCAGAGT	AAACTTCTGCCTGACGAGCTT
Mouse <i>Il1b</i>	CTCATTGTGGCTGTGGAGAAG	ACCAGCAGGTTATCATCATCAT
Mouse <i>Il6</i>	AGCTGGAGTCACAGAAGGAG	AGGCATAACGCACTAGGTTT
Mouse <i>Il10</i>	CCCTTTGCTATGGTGTCTT	TGGTTTCTCTTCCCAAGACC
Mouse <i>Il12a</i>	GAGGACTTGAAGATGTACCAG	TCCTATCTGTGTGAGGAGGGC
Mouse <i>Tnfa</i>	GTCAGGTTGCCTCTGTCTCA	TCAGGGAAGAGTCTGGAAAG
Mouse <i>Cxcl10</i>	CCTGCCACGTGTTGAGAT	TGATGGTCTTAGATTCCGGATTC
Mouse <i>Isg15</i>	TGGAAAGGGTAAGACCGTCCT	GGTGTCCGTGACTAACTCCAT
Mouse <i>Mx1</i>	GGGGAGGAAATAGAGAAAATGAT	GTTTACAAAGGGCTTGCTTGCT
Mouse <i>Actb</i>	AGGGCTATGCTCTCCCTCAC	CTCTCAGCTGTGGTGGTGAA
Human <i>IFNB1</i>	CATTACCTGAAGGCCAAGGA	CAATTGTCCAGTCCCAGAGG
Human <i>ISG15</i>	CTGAGAGGCAGCGAACTCAT	AGCATCTTACCGTCAGGTC
Human <i>MX1</i>	AGAGAAGGTGAGAAGCTGATCC	TTCTTCCAGCTCCTTCTCCTG
Human <i>CXCL10</i>	CTCCAGTCTCAGCACCATGA	GCTCCCCTCTGGTTTTAAGG
Human <i>GADPH</i>	ATGACATCAAGAAGGTGGTG	CATACCAGGAAATGAGCTTG



**Table S3. Related to CRISPR/Cas9. Primer sequences for molecular cloning**

Genes	Forward	Reverse
<i>Ogt</i> gRNA#1	CACCGTGCCACGGAAGACGCCATC	AAACGATGGCGTCTTCCGTGGGCAC
<i>Ogt</i> gRNA#2	CACCGGCTCCAGATGGCGTCTTCCG	AAACCGGAAGACGCCATCTGGAGCC
<i>mMavs</i> gRNA#1	ACCGGCCGTCGCGAGGATGTCTGG	AACCCAGACATCCTCGCGACGGCC
<i>mMavs</i> gRNA#2	CACCGGATACCCTCTCCTAACCAGC	AACGCTGGTTAGGAGAGGGTATCC
<i>mCgas</i> gRNA	CACCGATATGGAAGATCCGCGTAGA	AAACTCTACGCGGATCTTCCATATC
<i>mSting</i> gRNA	CACCGGCTGGATGCAGGTTGGAGTA	AAACTACTCCAACCTGCATCCAGCC
<i>hcGAS</i> gRNA	CACCGAAGTGCGACTCCGCGTTCAG	AAACCTGAACGCGGAGTCGCACTT
<i>hSTING</i> gRNA	CACCGGGATGTTTCAGTGCCTGCGAG	AAACCTCGCAGGCACTGAACATCC

**Table S4. Mass spectrometry assay of OGT interactome**