

Supplementary Materials for  
**Structure and dynamics of enterovirus genotype networks**

Nathânia Dábilla and Patrick T. Dolan

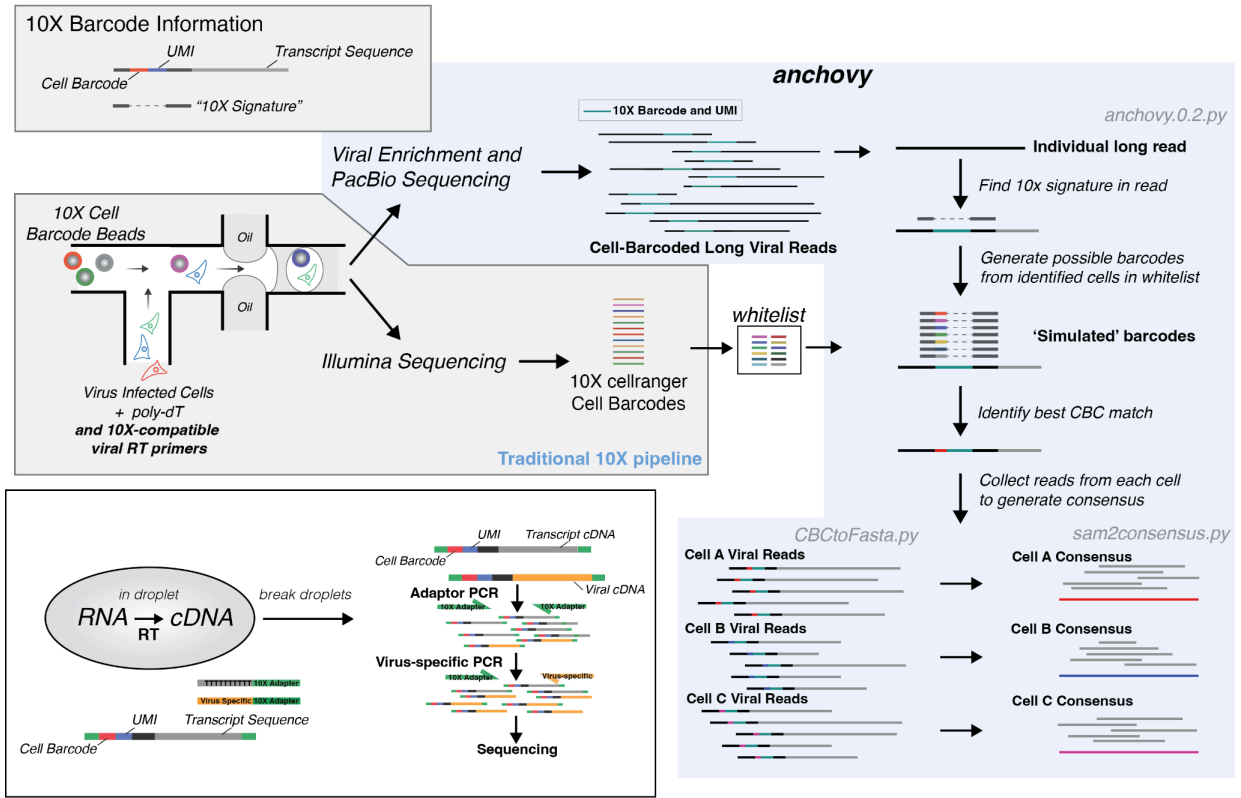
Corresponding author: Patrick T. Dolan, [patrick.dolan@nih.gov](mailto:patrick.dolan@nih.gov)

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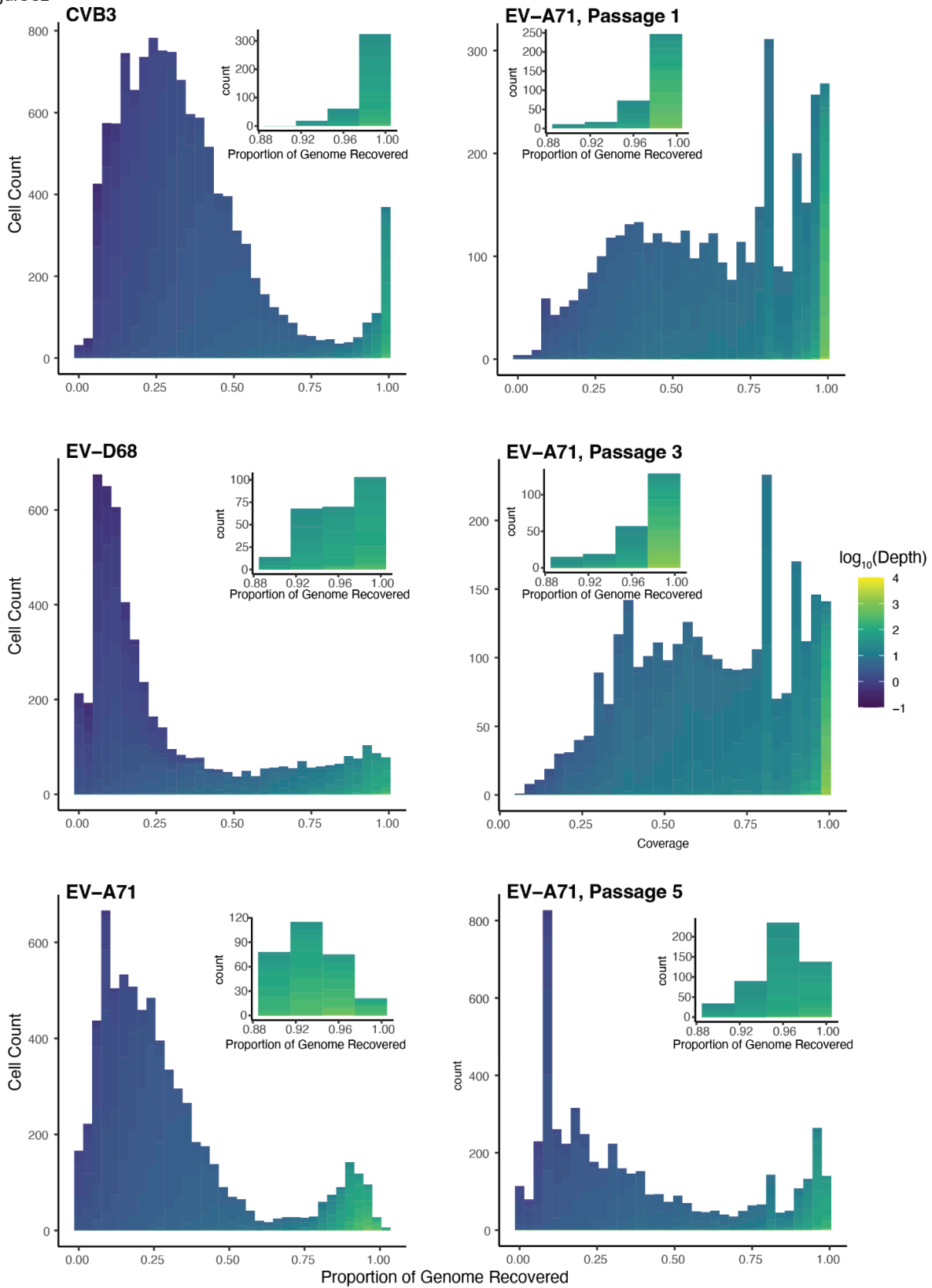
Figure S1



**Fig. S1.**

Anchovy pipeline. cDNA generated by 10x Genomics is sequenced by conventional illumina sequencing and analyzed by the CellRanger pipeline to yield the associated matrix, barcode, and gene files. The cDNA is also amplified by fragment-specific primers to yield 10x-barcoded amplicon pools that covers the full genome. The 10x barcode is identified in each read, and the cell barcode is matched to one of the barcodes identified in the CellRanger pipeline. This assures that the droplets carrying viral genome information are also 'real' cells. The viral reads associated with each cell barcode are used to generate a consensus sequence for each cell.

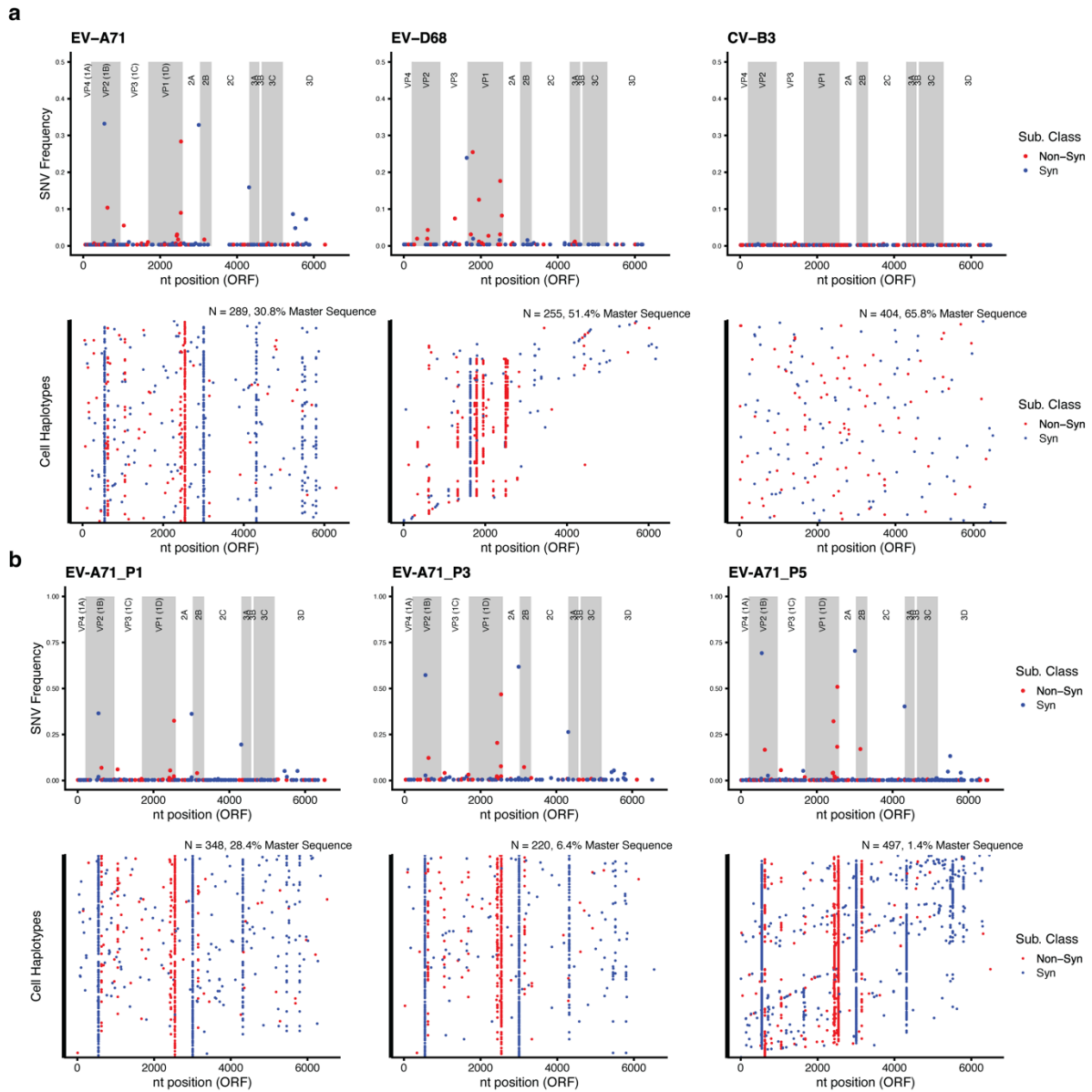
Figure S2



**Fig. S2.**

Histogram of the proportion of viral genome captured in each cell during SEARCHLIGHT sequencing experiments. Fill color represents the average depth of coverage across the viral genome in each cell. The inset shows the cells from which consensus haplotypes were recovered for subsequent analysis.

Figure S3

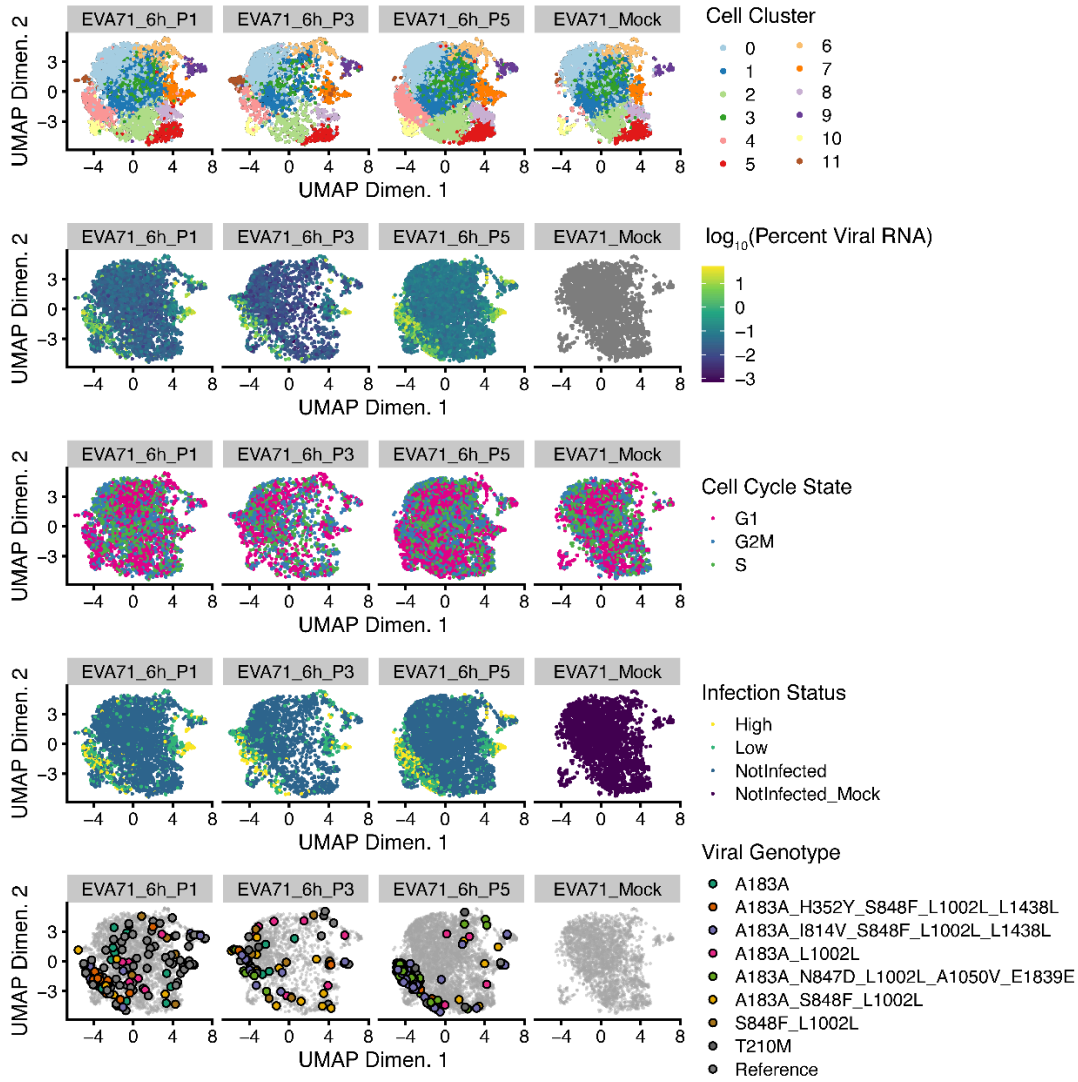


**Fig. S3.**

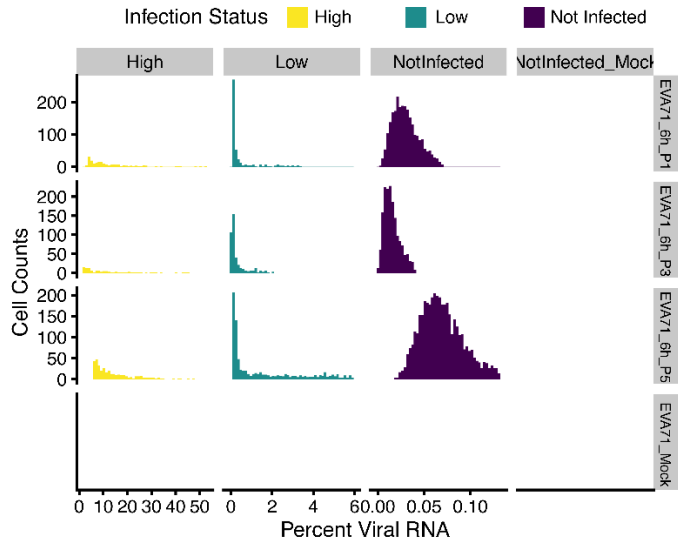
The 'bulk' view of population diversity (top) and the 'phased' haplotype alignment (bottom). In the haplotype alignment, each haplotype derived from an individual cell is shown as a single row, with differences from the modal genotype shown as points. Color indicates non-synonymous or synonymous mutations. Viral proteins are labeled and denoted by alternate shading. **(A)** First row refers to the strains Enterovirus-A71 (strain Tainan), Enterovirus-D68 (strain MO/14-18947), and Coxsackievirus B3 (strain Nancy) belonging to three distinct enterovirus species (A, B, and D). **(B)** Second row refers to Enterovirus-A71 passage 1, passage 3 and passage 5.

Figure S4

**A**



**B**





**Fig. S4.**

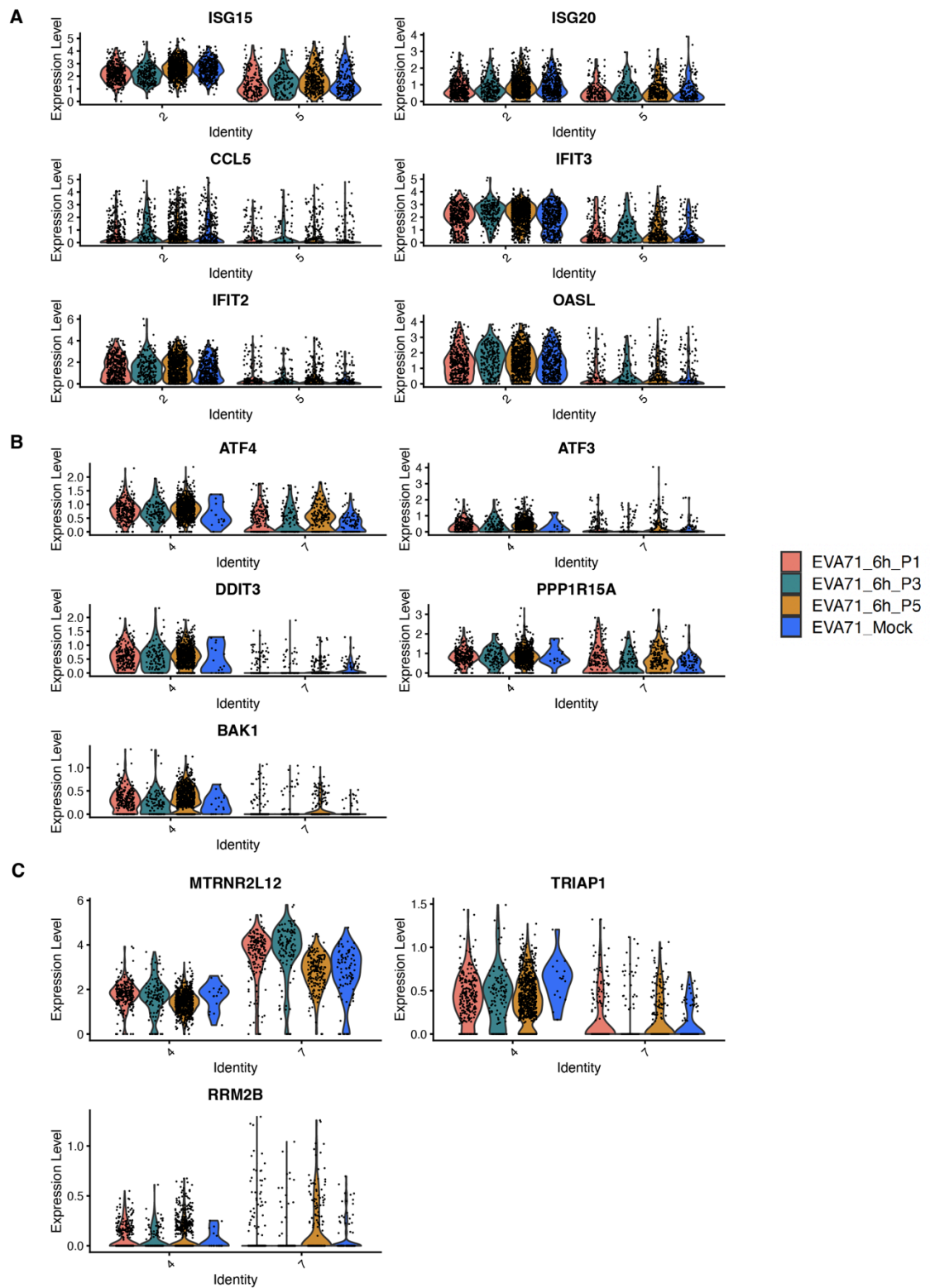
Connecting genotype with infected cell phenotype. **(A)** Uniform Manifold-Assisted Projection (UMAP) of the cell transcriptional phenotypes recovered from the passaged cell populations. (i) Clustering of transcriptional phenotypes derived from captured cells. (ii) Percent of viral RNA in each cell. (iii) Cell Cycle State of each cell. (iv) Identification of infected and uninfected cells. Cells are also classified by high- and low-viral RNA content. (v) Major genotypes captured by SEARCHLIGHT placed onto the phenotypic map of the host cells. **(B)** Histogram of the percent of viral RNA captured in each cell according to each Infection Status group, yellow for High, green for Low and dark purple for Not infected.



**Fig. S5.**

Heatmap of top20 marker genes of each cluster. The genes label are colored according to the clusters colors.

Figure S6



**Fig. S6.**

Violin plot of Expression Level of markers genes per cluster per condition. **(A)** Expression Levels of interferon-stimulated genes (ISGs) in cluster 2 and cluster 5. **(B)** Expression Levels of Integrated Stress Response (ISR) and pro-apoptosis markers. **(C)** Expression Levels of anti-apoptosis markers.

Primers with 10x genomics adaptors

nameseq

CVB3\_4\_R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACGCCCTGCTAACGTAACCCATT-3'  
CVB3\_9\_R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACTTCACTCCCTGTTCCATTGCA-3'  
CVB3\_14\_R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACAAGCATTCTTGGTGGGTGTG-3'

EVD68\_MO\_15\_R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACTGTGACTTCAATGTCCACGTCA-3'  
EVD68\_MO\_12\_R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACAGTGAACCTCTCCAATTAGCATATCT-3'  
EVD68\_MO\_6\_R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACACCAATGAGTGAATATCTAGATATGTTTCC-3'

EVA71\_NS\_4\_R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACTGACGATCTGTTGTTTGTATCCGT-3'  
EVA71\_NS\_9\_R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACAACCTCGTCGGGTTTCATGAAGC-3'  
EVA71\_Cap\_9R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACTCCAATAATCCTGCTCTGCTG-3'

EVA71\_Cap\_1R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACGCCAATAGTTAATTGTGCCACCC-3'  
EVA71\_Cap\_3R\_10X 5'-AAGCAGTGGTATCAACGCAGAGTACGGTGTGCAATTCGAAGCCATC-3'

Primers without 10x genomics for virus-specific PCR enrichment

cDNA Primers - part number 2000089

From Chromium Next GEM Single Cell 5' Reagent Kit v2 (Dual index), CG000331, Rev E, August 2022

cDNA Forward Primer:

5'-CTACACGACGCTCTCCGATCT-3'

CVB3\_4\_R 5'-GCCCTGCTAACGTAACCCATT-3'  
CVB3\_9\_R 5'-CTTCACTCCCTGTTCCATTGCA-3'  
CVB3\_14\_R 5'-AAGCATTCTTGGTGGGTGTG-3'  
CVB3\_18\_R 5'-CGAATGCGGAGAATTTACCCCT-3'

EVD68\_MO\_15\_R 5'-TGTGACTTCAATGTCCACGTCA-3'  
EVD68\_MO\_12\_R 5'-AGTGAACCTCTCCAATTAGCATATCT-3'  
EVD68\_MO\_6\_R 5'-ACCAATGAGTGAATATCTAGATATGTTTCC-3'

EVA71\_NS\_4\_R 5'-TGACGATCTGTTGTTTGTATCCGT-3'  
EVA71\_NS\_9\_R 5'-AACTCGTCGGGTTTCATGAAGC-3'  
EVA71\_Cap\_9R 5'-TCCAATAATCCTGCTCTGCTG-3'  
EVA71\_Cap\_1R 5'-GCCAATAGTTAATTGTGCCACCC-3'  
EVA71\_Cap\_3R 5'-GGTGTGCAATTCGAAGCCATC-3'

**Table S1.**

Primers for SEARCHLIGHT and for virus-specific PCR enrichment of CV-B3, EV-D68\_MO and EV-A71.





**Table S2.**  
Cluster markers enrichment analysis by DAVID.