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**Supplemental information**

**Environmental purines decrease**

***Pseudomonas aeruginosa* biofilm formation**

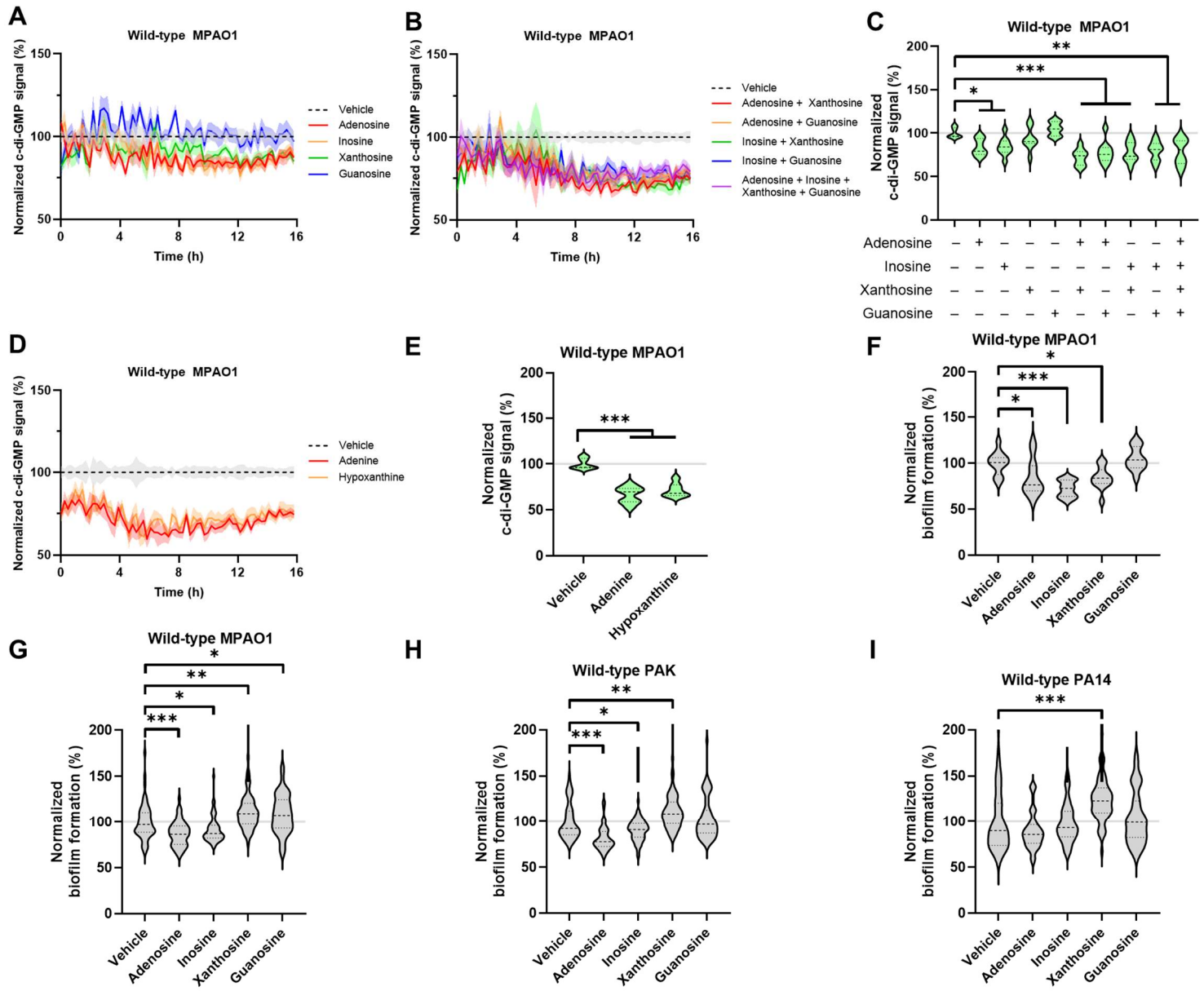
**by disrupting c-di-GMP metabolism**

**Corey Kennelly, Peter Tran, and Arthur Prindle**

**Supplementary Table 1: Oligonucleotides**

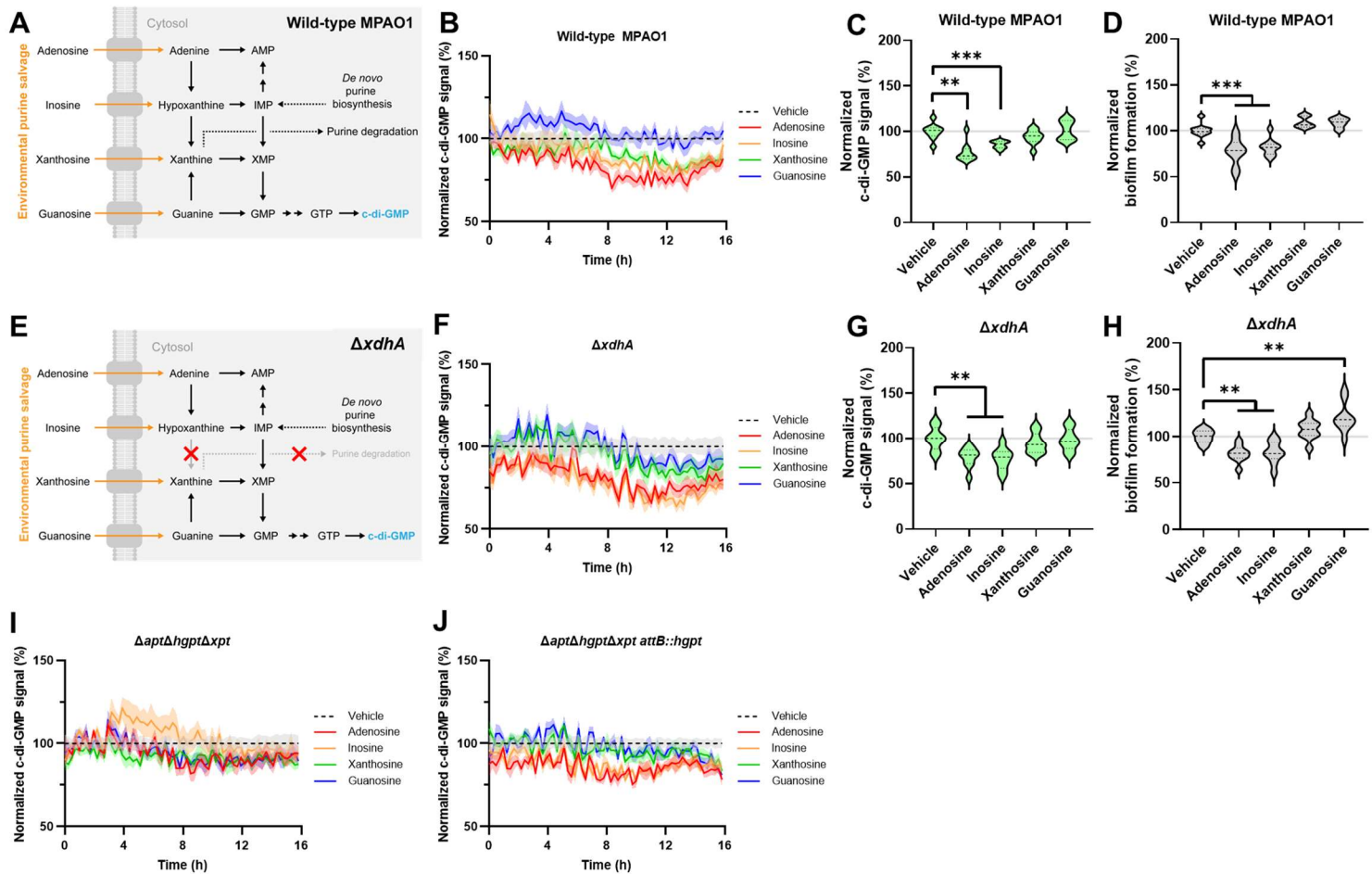
Primer name	Primer sequence (5' to 3')
PA1543-5	CCTGTCGCTGACCGTGGGA
PA1543-3	GGTGAAGTAGCCGCCGTAGAC
PA1543-5-1	GCATAAATGTAAAGCAGGTTGATCCTTTTCGGCTCGGTC
PA1543-5-2	AACTCGAGCCGCAAGCATGCTGAAAGACTTGAGGGTGAACCTCGTC
PA1543-3-1	TTCAGCATGCTTGCGGCTCGAGTTGCTCTCGACGAGCGTTGATACG
PA1543-3-2	CTAGAGTCGACCTGCAGAGCATCTGGATGGTCAGGTCTTCG
PA4645-5	GCGGATTCCTTGCTGCCG
PA4645-3	GATCTGGATCAGTTGCTGCTGCTC
PA4645-5-1	GCATAAATGTAAAGCAGCTTCCAGGGTCTCTTCGTTGC
PA4645-5-2	AACTCGAGCCGCAAGCATGCTGAAAGAGATCGACGGACATGGTGCTTC
PA4645-3-1	TTCAGCATGCTTGCGGCTCGAGTTGCTACTGGCGCAACGCCCGC
PA4645-3-2	CTAGAGTCGACCTGCAGAGCCAGACTGACGAACAGAAGCAGGGAG
PA5298-5	CTGATCCATCCGCAGCACCTG
PA5298-3	CAACTTCGTCGCCCAGGCTC
PA5298-5-1	GCATAAATGTAAAGCAGGCTGTTGCGTCCACATCCAC
PA5298-5-2	AACTCGAGCCGCAAGCATGCTGAAAGTCTTTGAGAATGTCCACGGGTGG
PA5298-3-1	TTCAGCATGCTTGCGGCTCGAGTTCTGCTGGAAAGCGAAGGCTACC
PA5298-3-2	CTAGAGTCGACCTGCAGAACTTGCCTGACATGGTGGTGAC
PA0934-5	GGTGACTGGCAACTGACTCTGG
PA0934-3	CTCGTCACCCTCTGCCATCG
PA0934-5-1	GCATAAATGTAAAGCAGCGGGTCTCTGACCTGTTT
PA0934-5-2	AACTCGAGCCGCAAGCATGCTGAAAGCTCTCACCTGTACCATCTTGC
PA0934-3-1	TTCAGCATGCTTGCGGCTCGAGTTCTGCCAACATCATCGAGGCG
PA0934-3-2	CTAGAGTCGACCTGCAGAGACAGGAACGTATCCAGCAGC
PA5338-5	CTTCATCCTCCCGCCGAGC
PA5338-3	CAGCTTGCCGTCTTGCTC
PA5338-5-1	GCATAAATGTAAAGCACCGTCAGCGAAATGAGCCACTAC
PA5338-5-2	AACTCGAGCCGCAAGCATGCTGAAAGAGTCTGTCCGCGAAGGGC
PA5338-3-1	TTCAGCATGCTTGCGGCTCGAGTTCTCAAGGGAGTGATCCGAATCACC
PA5338-3-2	CTAGAGTCGACCTGCAGAGGCTGCGGCGAGGAAAATC
PA3770-5	GAGCAGCTCACCGATGTCTG
PA3770-3	GTGGAATTGCACGCCGTAGTAGG
PA3770-5-1	GCATAAATGTAAAGCAGACGGTAATGTCGTCGTGGCTG
PA3770-5-2	AACTCGAGCCGCAAGCATGCTGAAAGAGGGCTTCTTGACTGATTCCGAG
PA3770-3-1	TTCAGCATGCTTGCGGCTCGAGTTGCCGAGTCCCATGTCCACG
PA3770-3-2	CTAGAGTCGACCTGCAGACTTGTCCGCTGGCTCATCC
PA1524-5	CGAGCTGATCTGGTGGTACCG
PA1524-3	CGATGACGATGCGGTTGAAGGAAAC
PA1524-5-1	GCATAAATGTAAAGCACCTTACCCTGCTGGAGATCG
PA1524-5-2	AACTCGAGCCGCAAGCATGCTGAAAGATCTCCCGTTGAGCAGGAAC
PA1524-3-1	TTCAGCATGCTTGCGGCTCGAGTTCCCGCCGTGAAACCCG
PA1524-3-2	CTAGAGTCGACCTGCAGACCGAGGAAATCTGCGTCTCCAGGTAG
PA4645-CompF	GGCTGCAGGAATTCGATATCAGGCGACTGTCTCCAACGGG
PA4645-CompR	TCGAGGTCGACGGTATCGATATCAGAGCCCTTGACCGC
attB-5	GCTGACTTCACGCTGTTCCG
attB-3	GCTACCTGGACTGGGAGTTTCG

For PA1543 (apt), PA4645 (hgpt), PA5298 (xpt), PA0934 (relA), PA5338 (spoT), PA3770 (guaB), PA1524 (xdhA), and attB, -5 and -3 indicate primers used to amplify these loci for PCR confirmation and sequencing; -5-1 and -5-2 were used to amplify the region upstream of each gene with appropriate Gibson assembly overhangs while -3-1 and -3-2 were used to amplify the region downstream of each gene with appropriate Gibson assembly overhangs. Within each knockout primer, green font indicates homology with pEXG2; red font with italics indicates arbitrary homology created between -5-2 and -3-1 primers to facilitate Gibson assembly of pEXG2 constructs; black font indicates homology with PAO1 genome; yellow-highlighted font indicates homology with gene of interest. For CompF/R primers, green font indicates homology with pminiCTX; black font indicates homology with PAO1 genome. Grey highlight signifies linker for Gibson assembly.

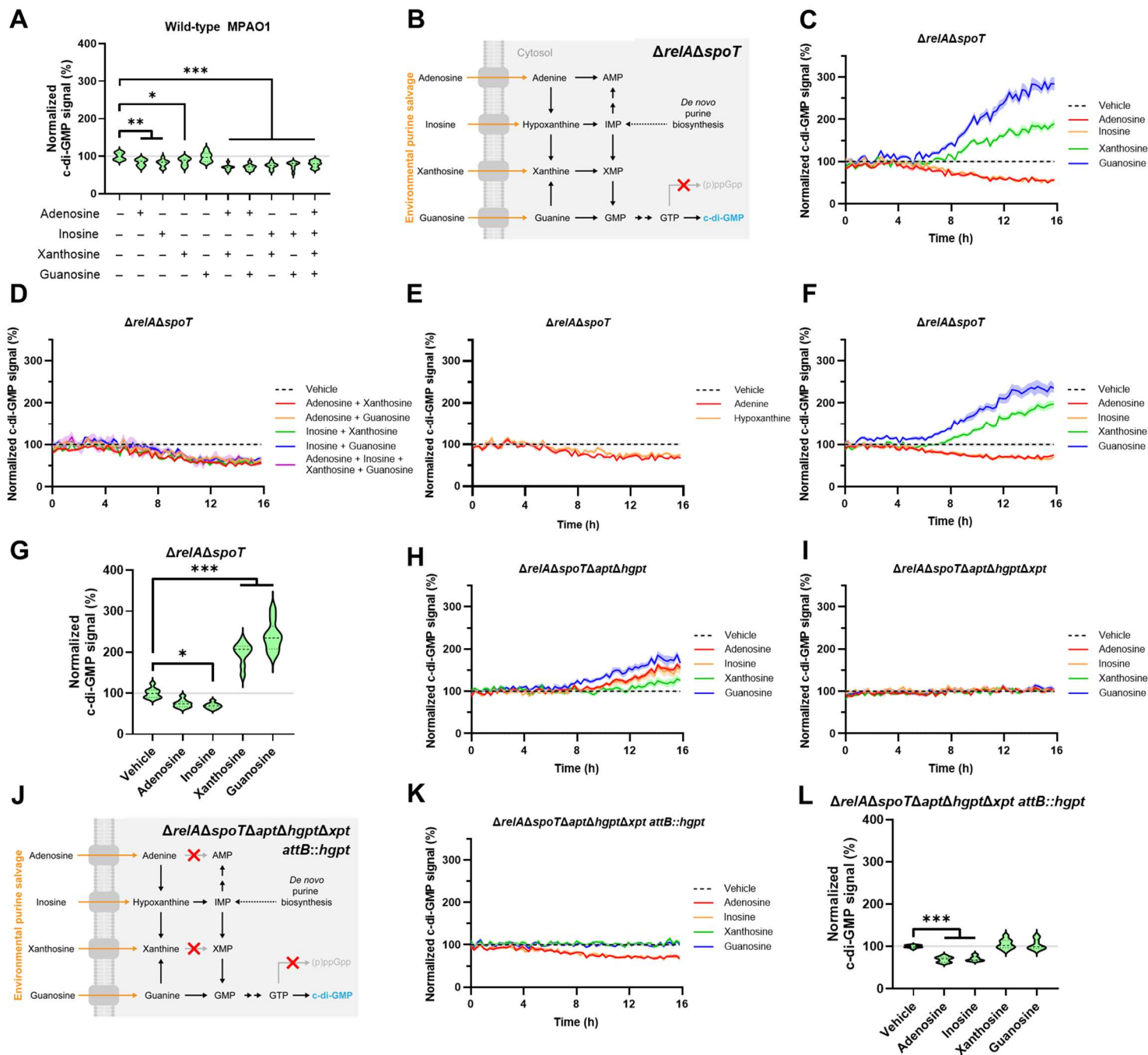


**Supplementary Figure 1: Adenosine and inosine decrease c-di-GMP and biofilm formation.** (A) c-di-GMP signal time course plots for MPAO1 exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. Data are used in Figure 1d, Supplementary Figure 1c, and Supplementary Figure 3a. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Vehicle, Xanthosine). Data represent mean  $\pm$  SE. (B) c-di-GMP signal time course plots for MPAO1 exposed to vehicle or 100  $\mu$ M combinations of compounds as indicated. Data are used in Figure 1d, Supplementary Figure 1c, and Supplementary Figure 3a. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Vehicle, Adenosine-Inosine-Xanthosine-Guanosine). Data represent mean  $\pm$  SE. (C) Violin plot of c-di-GMP signal for MPAO1 exposed to vehicle or 100  $\mu$ M indicated compounds after 8 hours of exposure at 37°C, shaking, to quantify changes in Supplementary Figure 1a,b. Data are used in Figure 1d. (D) c-di-GMP signal time course plots for MPAO1 exposed to vehicle or 100  $\mu$ M adenine or hypoxanthine. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Vehicle); n=7 (Hypoxanthine). Data represent mean  $\pm$  SE. (E) Violin plot of c-di-GMP signal for MPAO1 exposed to vehicle or 100  $\mu$ M adenine or hypoxanthine after 8 hours of exposure at 37°C, shaking, to quantify changes in Supplementary Figure 1d. All c-di-GMP data in Figure 1 and Supplementary Figure 1 were collected simultaneously. (F) Violin plots of biofilm formation from safranin-stained biomass at OD<sub>530</sub> normalized to OD<sub>600</sub> growth for MPAO1 exposed to vehicle or 100  $\mu$ M indicated compounds after 8 hours of exposure. In these experiments, biofilms were allowed to grow in 96-well plates at 37°C for 18 hours prior to compound exposure. 3 wells per condition per experiment from 4 independent experiments were included (n=12). (G) Violin plots of biofilm formation from safranin-stained biomass at

OD<sub>530</sub> normalized to OD<sub>600</sub> growth for *P. aeruginosa* exposed to vehicle or 100 μM indicated compounds after 8 hours of exposure. 9 wells per condition per experiment from 7 independent experiments were included (n=63) unless otherwise noted. Strain MPAO1 was used and n=61 (Vehicle, Adenosine, Inosine, Guanosine) and n=60 (Xanthosine) wells were included after removal of outliers. Biofilm data in Supplementary Figure 1g, 1h, and 1i were collected simultaneously. **(H)** Violin plots of biofilm formation from safranin-stained biomass at OD<sub>530</sub> normalized to OD<sub>600</sub> growth for *P. aeruginosa* exposed to vehicle or 100 μM indicated compounds after 8 hours of exposure. 9 wells per condition per experiment from 7 independent experiments were included (n=63) unless otherwise noted. Strain PAK was used and n=62 (Adenosine, Xanthosine, Guanosine) and n=61 (Inosine) wells were included after removal of outliers. Biofilm data in Supplementary Figure 1g, 1h, and 1i were collected simultaneously. **(I)** Violin plots of biofilm formation from safranin-stained biomass at OD<sub>530</sub> normalized to OD<sub>600</sub> growth for *P. aeruginosa* exposed to vehicle or 100 μM indicated compounds after 8 hours of exposure. 9 wells per condition per experiment from 7 independent experiments were included (n=63) unless otherwise noted. Strain PA14 was used and n=62 (Xanthosine) wells were included after removal of outliers. Biofilm data in Supplementary Figure 1g, 1h, and 1i were collected simultaneously. Throughout this figure, repeated measures one-way ANOVA with Dunnett's multiple comparison test comparing to vehicle was used to determine statistical significance. \*p≤0.05; \*\*p≤0.01; \*\*\*p≤0.001.



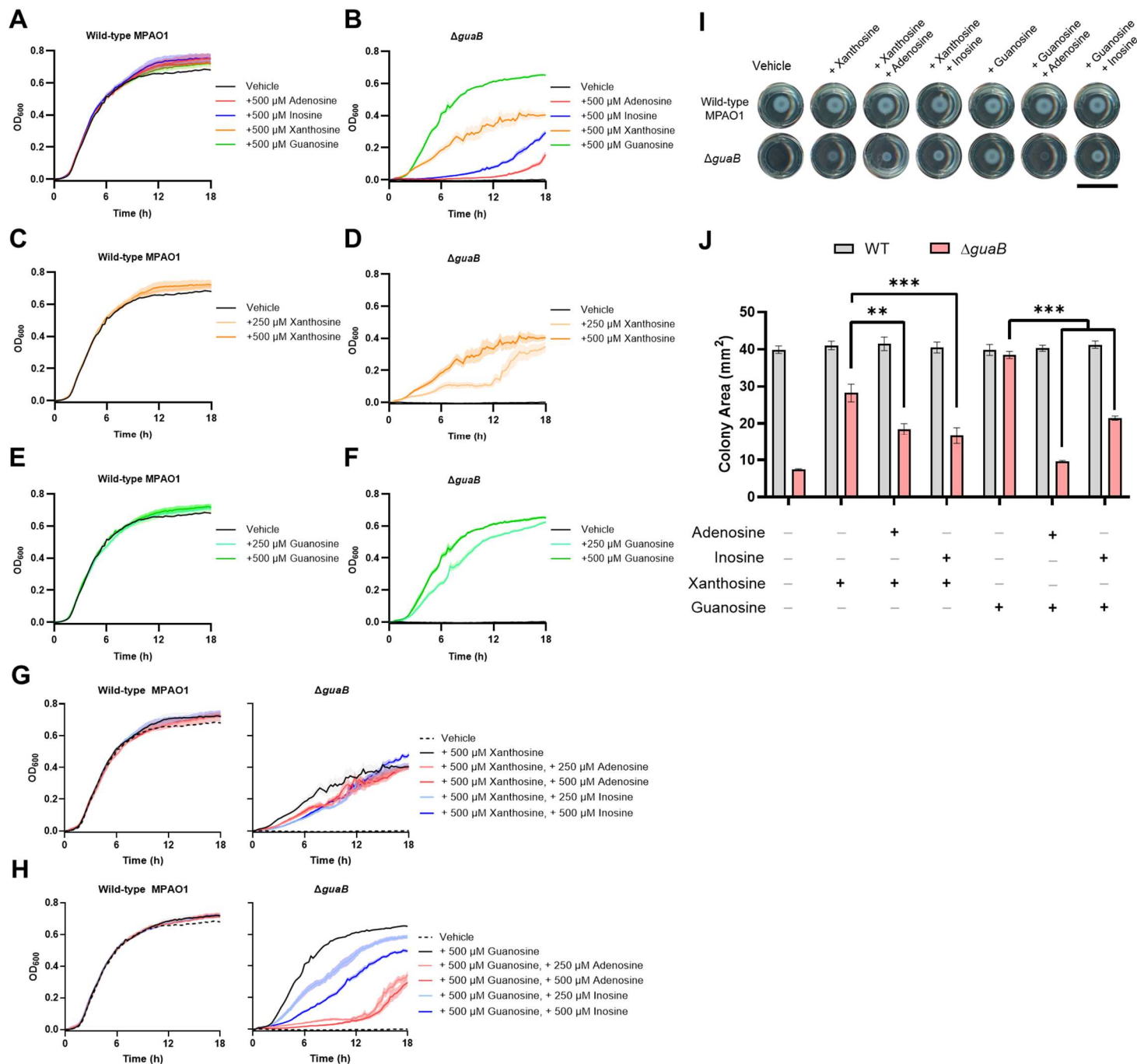
**Supplementary Figure 2: Adenosine/inosine-dependent effects require purine salvage.** (A) MPAO1 can import environmental purines and use them in both salvage and degradation pathways. (B) c-di-GMP signal time course plots for MPAO1 exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring an outlier removed due to aberrant growth: n=8 (Inosine). Data represent mean  $\pm$  SE. (C) Violin plot of c-di-GMP signal for MPAO1 exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, guanosine after 8 hours of exposure at 37°C, shaking, to quantify changes in Supplementary Figure 2b. (D) Violin plot of biofilm formation from safranin-stained biomass for MPAO1 exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine after 8 hours of exposure. 3 wells per condition per experiment from 3 independent experiments were included (n=9). (E)  $\Delta xdhA$  can import environmental purines and use them in the salvage pathway but not the degradation pathway. (F) c-di-GMP signal time course plots for  $\Delta xdhA$  exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Vehicle, Inosine). Data represent mean  $\pm$  SE. (G) Violin plot of c-di-GMP signal for  $\Delta xdhA$  exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, guanosine after 8 hours of exposure at 37°C, shaking, to quantify changes in Supplementary Figure 2f. (H) Violin plot of biofilm formation from safranin-stained biomass for  $\Delta xdhA$  exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine after 8 hours of exposure. 3 wells per condition per experiment from 3 independent experiments were included (n=9). (I) c-di-GMP signal time course plots for  $\Delta apt\Delta hgpt\Delta xpt$  exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Adenosine, Inosine, Guanosine); n=7 (Xanthosine). Data are used in Figure 2b. Data represent mean  $\pm$  SE. (J) c-di-GMP signal time course plots for  $\Delta apt\Delta hgpt\Delta xpt attB::hgpt$  exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=7 (Adenosine). Data are used in Figure 2e. Data represent mean  $\pm$  SE. All c-di-GMP data in Figure 2 and Supplementary Figure 2 were collected simultaneously. All biofilm data in Figure 2 and Supplementary Figure 2 were collected simultaneously. Throughout this figure, repeated measures one-way ANOVA with Dunnett's multiple comparison test comparing to vehicle was used to determine statistical significance. \*p $\leq$ 0.05; \*\*p $\leq$ 0.01; \*\*\*p $\leq$ 0.001.



**Supplementary Figure 3: (p)ppGpp prevents xanthosine/guanosine-dependent effects and their salvage is also blocked by adenosine and inosine.**

**(A)** Violin plot of c-di-GMP signal for MPAO1 exposed to vehicle or 100  $\mu$ M indicated compounds after 12 hours of exposure at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Vehicle, Xanthosine). Data are from Supplementary Figure 1a,b. **(B)**  $\Delta relA\Delta spoT$  is capable of salvaging all purines; however, extracellular purines may affect this strain differently compared to MPAO1 due to the absence of (p)ppGpp, a negative regulator of GTP homeostasis in some bacteria. **(C)** c-di-GMP signal time course plots for  $\Delta relA\Delta spoT$  exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Vehicle). Data was collected simultaneously with Supplementary Figure 1a,b,d. Data are used in Figure 3b. Data represent mean  $\pm$  SE. **(D)** c-di-GMP signal time course plots for  $\Delta relA\Delta spoT$  exposed to vehicle or 100  $\mu$ M combinations of compounds as indicated. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Vehicle, Xanthosine+Adenosine, Xanthosine+Inosine); n=7 (Guanosine+Adenosine); n=5 (Adenosine+Inosine+Xanthosine+Guanosine). Data was collected simultaneously with Supplementary Figure 1a,b,d. Data are used in Figure 3b. Data represent mean  $\pm$  SE. **(E)** c-di-GMP signal time course plots for  $\Delta relA\Delta spoT$  exposed to

vehicle or 100  $\mu$ M adenine or hypoxanthine. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=8 (Vehicle, Hypoxanthine). Data was collected simultaneously with Supplementary Figure 1a,b,d. Data represent mean  $\pm$  SE. **(F)** c-di-GMP signal time course plots for  $\Delta$ relA $\Delta$ spoT exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9). Data was collected simultaneously with Supplementary Figure 3h,i,k. Data represent mean  $\pm$  SE. **(G)** Violin plot of c-di-GMP signal for  $\Delta$ relA $\Delta$ spoT exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, guanosine after 16 hours of exposure at 37°C, shaking, to quantify changes in Supplementary Figure 3f. **(H)** c-di-GMP signal time course plots for  $\Delta$ relA $\Delta$ spoT $\Delta$ apt $\Delta$ hgpt exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring outliers removed due to aberrant growth: n=7 (Xanthosine). Data was collected simultaneously with Supplementary Figure 3f, i, k. Data are used in Figure 3d. Data represent mean  $\pm$  SE. **(I)** c-di-GMP signal time course plots for  $\Delta$ relA $\Delta$ spoT $\Delta$ apt $\Delta$ hgpt $\Delta$ xpt exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9). Data was collected simultaneously with Supplementary Figure 3f, h, k. Data are used in Figure 3f. Data represent mean  $\pm$  SE. **(J)** (p)ppGpp-mediated GTP homeostasis remains absent but intracellular nucleotide pools are no longer insulated from environmental purines in  $\Delta$ relA $\Delta$ spoT $\Delta$ apt $\Delta$ hgpt $\Delta$ xpt *attB::hgpt* background due to expression of *hgpt* from native promoter at the *attB* neutral site. **(K)** c-di-GMP signal time course plots for  $\Delta$ relA $\Delta$ spoT $\Delta$ apt $\Delta$ hgpt $\Delta$ xpt *attB::hgpt* exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, or guanosine. 3 wells per condition per experiment from 3 independent experiments were included (n=9). Data was collected simultaneously with Supplementary Figure 3f, h, i. Data represent mean  $\pm$  SE. **(L)** Violin plot of c-di-GMP signal for  $\Delta$ relA $\Delta$ spoT $\Delta$ apt $\Delta$ hgpt $\Delta$ xpt *attB::hgpt* exposed to vehicle or 100  $\mu$ M adenosine, inosine, xanthosine, guanosine after 16 hours of exposure at 37°C, shaking, to quantify changes in Supplementary Figure 3k. Throughout this figure, repeated measures one-way ANOVA with Dunnett's multiple comparison test comparing to vehicle was used to determine statistical significance. \*p $\leq$ 0.05; \*\*p $\leq$ 0.01; \*\*\*p $\leq$ 0.001.



**Supplementary Figure 4: Guanine auxotroph reveals adenosine and inosine block guanosine-dependent effects even at expense of growth.**

(A) Growth of MPAO1 in M9 supplemented with vehicle or 500 μM of adenosine, inosine, xanthosine, or guanosine, respectively, at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9). (B) Growth of  $\Delta$ *guaB* in M9 supplemented with vehicle or 500 μM of adenosine, inosine, xanthosine, or guanosine, respectively, at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9). (C) Growth of MPAO1 in M9 supplemented with vehicle, 250 μM, or 500 μM xanthosine, respectively, at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9). (D) Growth of  $\Delta$ *guaB* in M9 supplemented with vehicle, 250 μM, or 500 μM xanthosine, respectively, at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring an outlier removed due to aberrant growth: n=8 ( $\Delta$ *guaB* 500 μM Guanosine, 500 μM Inosine). (E) Growth of MPAO1 in M9 supplemented with vehicle, 250 μM, or 500 μM guanosine, respectively, at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9). (F) Growth of  $\Delta$ *guaB* in M9 supplemented with vehicle, 250 μM, or 500 μM guanosine, respectively, at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9). (G) Growth of MPAO1 and  $\Delta$ *guaB* in M9 supplemented with vehicle or indicated mixtures of compounds



including xanthosine at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9). **(H)** Growth of MPAO1 and  $\Delta$ *guaB* in M9 supplemented with vehicle or indicated mixtures of compounds including guanosine at 37°C, shaking. 3 wells per condition per experiment from 3 independent experiments were included (n=9), barring an outlier removed due to aberrant growth: n=8 ( $\Delta$ *guaB* 500  $\mu$ M guanosine, 500  $\mu$ M inosine). **(I)** Image of MPAO1 and  $\Delta$ *guaB* growth on M9 agar pads supplemented with vehicle or 300  $\mu$ M of indicated compounds at 37°C after 24 hours. 2 wells per condition per experiment from 4 independent experiments were included for vehicle, guanosine, adenosine-guanosine, inosine-guanosine, and xanthosine conditions (n=8). 1 well per condition per experiment from 4 independent experiments were included for adenosine-xanthosine and inosine-xanthosine experiments (n=4). Scale indicates 15 mm. A subset of this data is displayed in Figure 4c. **(J)** Bar plot of MPAO1 and  $\Delta$ *guaB* colony size from Supplementary Figure 4i quantified using ImageJ. All data represent mean  $\pm$  SE. Repeated measures one-way ANOVA with Dunnett's multiple comparison test comparing to supplemented xanthosine or supplemented guanosine condition was used to determine statistical significance. \*\*p $\leq$ 0.01; \*\*\*p $\leq$ 0.001.