OMTM, Volume 32

Supplemental information

Design and validation of a GMP stem cell

manufacturing protocol for MPSII hematopoietic

stem cell gene therapy

Stuart Ellison, Karen Buckland, Yuko Learmonth, Victoria Day, Spandan Kalra, Lauren Howe, Francisco José Roman-Rodriguez, Jose Bonafont, Laura Booth, Rebecca Holley, Jon Smythe, Simon Jones, Adrian Thrasher, Claire Booth, and Brian W. Bigger **Table S1.** Current specification for batch release testing of a cryopreserved medicinal product for

 proposed clinical trial

Test	Method	Acceptance Criteria	
Mycoplasma	PCR	Not Detected	
Total Viable Cell Count/ml	Trypan Blue	0.5 to 17.5 x 10 ⁶ TNC/ml	
Total Viable Cell Count	Trypan Blue	Record	
CD34+ Cell Purity and Identity	FACS	≥ 35% CD34+ cells	
CD34+ Cell Dose	Calculated	Record	
Cell Viability	Trypan Blue	≥ 70% viable cells	
Sterility	BacTec	No Growth	
Bacterial Endotoxins	Kinetic Turbidimetric	≤ 5 EU/kg/hr	
Vector Copy Number	ddPCR	≥ 0.5 copy/cell	

Table S2: Small scale hCD34+ stem cell transduction optimisation performed at GOSH. Low vector dose range.

					Batch Testing												
Methodology Batch MOI		OI (x 10 ⁸ IG/ml)	lG/ml)	lg/ml)	lg/ml)	0 (x 10 ⁸ IG/ml)	Cells	Test Attribute	Viability	CD34%	Clono- genicity	VCN (liquid culture)	Pass/ Fail				
			IOM	ō	ō		(x 10 [§]	(x 10 ⁸	(x 10 ⁸	(x 10 [°]	(× 10 ⁸	(x 10 ⁸	(x 10 ⁸	01 (x 10 ⁸	(x 10 ⁸ onor (Method	Trypan Blue
Metho	Ba	Vector Conc.		Healthy Donor	Acceptance Criteria	≥ 70%	For information only	≥4 CFUs/ 1000 cells plated	≥ 0.5								
		25	0.5	А		94.01	99.2	366	0.93	Pass							
lent	018	12.5	0.25	А		92.87	99.0	346	0.85	Pass							
rin	SII-	25	0.5	В		93.01	99.1	406	0.839	Pass							
xpe	МΡ	12.5	0.25	В		92.94	99.2	374	0.827	Pass							
Ш Б	- - -	N/A	N/A	А		92.87	99.0	336	0.01	N/A							
GMP Experiment	21-SU-MPSII-01a	N/A	N/A	В		99.6	99.6	398	0	N/A							

Table S3: Small scale hCD34+ stem cell transduction optimisation performed at GOSH. High vector dose range.

Batch Testing													
) ** (x 10 ⁸ IG/ml)*		lG/ml)*	lG/ml)*	IG/ml)*	Cells	Test Attribute	Viability	CD34%	VCN (liquid culture)	VCN (pooled CFUs)	Pass/ Fail
dolog	Batch MOI** nc. (x 10 ⁸		(x 10 ⁸		Method	Trypan Blue	Flow Cytometry	ddPCR	ddPCR				
Methodology Batch		M	Vector Conc.	Healthy Donor	Acceptance Criteria	≥ 70%	For information only	≥ 0.5	≥ 0.5				
L.	C	25	0.5	А		91.6	99.3	0.92	1.1	Pass			
Experiment	01k	50	1.0	А		89.13	99.5	1.19	1.08	Pass			
rin	SII-	100	2.0	А		92.46	99.0	1.63	1.56	Pass			
kpe	ЧΡ	25	0.5	С		91.07	99.2	0.99	1.5	Pass			
Ш Ц	۲-J	50	1.0	С		95.03	99.5	1.08	1.27	Pass			
GMP	21-SU-MPSII-01b	100	2.0	С		92.53	99.5	1.92	2.09	Pass			

*A minimum of 3.0×10^6 CD34+ cells/kg after transduction is required for infusion into the patient

*Cells from the same health donor A were used in both assays (table S2 and S3).**MOI = Multiplicity of infection

 Table S4: UoM/Barnsley vs GOSH equivalent vector concentration comparison.

UoM/Barnsley MOI	GOSH MOI
25	18
50	35
100	71

Table S5. Individual VCN counts in single picked colonies from CFU assay, determined by QPCR. Left panel) Large scale transduction performed at Manchester research lab with LV IDS.ApoEII at MOI25 + TE with IL3 in the media, cultured in rectronection-coatedT175 flask. Right panel) 'At scale' transduction performed at NHSBT Barnsley research and development lab with LV IDS.ApoEII at MOI25 + TE with IL3 in the media, cultured in rectronection-coatedT175 flask.

BFU-E	CFU-GM		BFU-E	CFU-GM
1.52	1.51		1.41	28.00**
5.96	6.42		1.89	1.10
1.90	1.88		1.34	6.28
4.92	4.49		2.26	10.24
3.86	1.35		1.25	8.43
3.56	5.73		2.69	3.51
1.51	4.55		4.27	2.41
6.42	2.76		10.20	8.07
1.88	1.88 1.31		0.15*	5.53
4.49	.49 1.29		7.26	4.62
1.35	7.88		5.63	2.80
5.73	1.62		2.21	5.26
* VCN below 0.3 classe	ed as 0 (colony with no		3.58	4.07
copies).			2.49	3.32
**VCN assay is accurate	te up to approximately		4.06	0.82
	s above this cannot be		3.21	3.30
accurately determined	with this method.		1.28	-

Table S6. Overview of sampling timepoints and volumes taken for sterility testing performed for
the NHSBT GMP validation runs.

Reagent / Sample	No. of Replicates/micro- organism	Inoculum volume per broth	Comments
Final product – CD11b- IDS.ApoEII LV Gene Modified Autologous CD34+ Cells in final container and condition as presented to patient	3	600µl	Cryopreserved in pilot vial 1% of final product volume
Fresh HPC Apheresis as collected from donor	1	2-4ml	1% of starting material volume depending on received volume
CD34- cell fraction post selection	1	5ml	1% of total selected cell product volumes (+ and – fractions)
Day 3 transduction media supernatent	1	5ml	>1% of total supernatant volume
Day 4 transduction media supernatent	1	5ml	>1% of total supernatant volume
Day 4 wash buffer supernatent	1	5ml	>1% of total supernatant volume