KAT8-mediated H4K16ac is Essential for Sustaining Trophoblast Selfrenewal and Proliferation via Regulating CDX2

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Description of Supplementary Information

Supplementary Figure 1-7.

Supplementary Table S1: The clinical characteristics of the pregnant women enrolled in this study

Supplementary Table S2: Primer list of constructing overexpression vector

Supplementary Table S3. Primer list of quantitative RT-PCR



Supplementary Figure 1. Genotype analysis of the generated mouse model. (a) Genotype analysis of $Kat8^{f/+}$, $Kat8^{f/f}$, WT mouse. (b) Genotype analysis of the offspring from female $Kat8^{f/f}$ mice crossed with male $Kat8^{f/+}$; *Elf5-Cre* mice. (c) Genotype analysis of embryos obtained from the implantation site dissection of female $Kat8^{f/f}$ mice crossed with male $Kat8^{f/+}$; *Elf5-Cre* mice.



Supplementary Figure 2. Impaired trophoblast proliferation upon *Kat8* deletion. (a) Quantitative real-time PCR analysis of *Kat8* mRNA levels in *Kat8*^{f/f} and *Kat8*^{d/d} mTSCs. Data are representative of three independent experiments (n=3) and the values are normalized to ACTB. Two-tailed unpaired Student's t test. Error bars, mean \pm SEM. P < 0.0001. Source data are provided as a Source Data file. (b) The analysis of cell proliferation between *Kat8*^{f/f} and *Kat8*^{d/d} mTSCs. Data are representative of three independent experiments (n=3). Two-tailed unpaired Student's t test. Error bars, mean \pm SEM. P = 0.0042 (for D3), P = 0.0001 (for D4), P < 0.0001 (for D5). Source data are provided as a Source Data file. (c) IF staining images of pH3 in $Kat8^{f/f}$ and $Kat8^{d/d}$ mTSCs. (d) The quantitative results of C. Data are representative of three independent experiments (n=3). Two-tailed unpaired Student's t test. Error bars, mean \pm SEM. P = 0.0053. (e) Gene Ontology analysis for top 150 downregulated genes. (f) IF staining images of γ H2AX and H4K16ac in $Kat8^{f/f}$, $Kat8^{d/d}$ embryos and mTSCs. (g) Gene Ontology analysis for top 150 upregulated genes. (h) X gal staining in $Kat8^{f/f}$ and $Kat8^{d/d}$ mTSCs. (i) Heatmap of differentiation of mTSCs associated genes differentially regulated between $Kat8^{f/f}$ and $Kat8^{d/d}$ mTSCs.



Supplementary Figure 3. Impaired trophoblast proliferation upon *Kat8* deletion. (a) Genomic distribution of KAT8 and H4K16ac peaks in mTSCs. (b) Coverage profiles for KAT8 and H4K16ac. (c) Genome browser view of CUT&Tag-seq signals and RNA-seq tracks for KAT8 and H4K16ac target genes in mTSCs





Brightfield

		E8.5	6		2		
F/+	F/+	F/+	F/+	F/+	F/+	F/F	F/F
17	17-	17.	11.	111	111-		
			-	100000	and the second second	(accessed)	(COLOR)

cre





Kat8^{d/d}

Kat8^{tr}

е

E8.5

g







b

d

Kat8^{d/d}+EX527

Supplementary Figure 4. Partial restoration of abnormal placental development caused by KAT8 knockout through EX527 intervention. (a) H&E staining, immunofluorescent staining of CDX2 and H4K16ac in WT mice treated with 20mg/kg EX527 at E7.5. (b) Genotype analysis of embryos from female $Kat \delta^{f/f}$ mice crossed with male $Kat8^{f/+}$; *Elf5-Cre* mice after treatment with EX527 at E6.5. (c) Genotype analysis of embryos from female Kat8^{f/f} mice crossed with male Kat8^{f/+};Elf5-Cre mice after treatment with EX527 at E7.5. (d) Genotype analysis of embryos from female $Kat8^{f/f}$ mice crossed with male $Kat8^{f/+}$; Elf5-Cre mice after treatment with EX527 at E8.5. (e) Brightfield images of embryos from female $Kat 8^{f/f}$ mice crossed with male Kat8^{f/+};Elf5-Cre mice after restoration treatment with EX527. (f) The number of KAT8 knockout embryos and the number of rescued embryos in female Kat8^{f/f} mice crossed with male Kat8^{f/+};Elf5-Cre mice after restoration treatment with EX527. Rescued embryos were defined as $Kat8^{d/d}$ embryos exhibiting CDX2+ trophoblast at E6.5 and E7.5, and AP2 γ + trophoblast at E8.5. (g) Implantation sites in female Kat8^{f/f} mice crossed with male $Kat8^{f/+}$ Elf5-Cre mice at E12.5 with EX527 or without EX527. (h) Genotype analysis of offspring from female Kat8^{f/f} mice crossed with male Kat8^{f/+};Elf5-Cre mice after treatment with EX527.



Supplementary Figure 5. Generating human trophoblast organoid. (a) Schematic representation illustrating the workflow for generating human placenta-derived organoids. The graphic elements were created by figdraw. (b) Brightfield images, H&E staining, and IF staining of AP2 α , CK7, TP63, and CGB in hTOs. (c) IF staining of KAT8, H4K16ac and CDX2 in hTOs.



Supplementary Figure 6. Assessing the impact of EX527. (a) Immunofluorescent staining of CDX2 in villi from 6 and 7 weeks. (b) Immunoblot analysis of KAT8 and H4K16ac in hTOs treated with or without EX527. (c) Brightfield images, and immunofluorescent staining of KAT8 and H4K16ac in hTOs with or without EX527. (d) The quantification results of organoid diameter for hTOs in indicated groups. Each dot in the column represents one TO (n=105, DMSO; n=93, EX527). Source data are provided as a Source Data file. (e) The quantification results of organoid forming efficiency for hTOs in indicated groups. Data are representative of five independent experiments (n=5). Source data are provided as a Source Data file.



Fig. 5b



Fig. 6b



α-Tubuli





Fig. 6g





Supplementary Figure 7. Unprocessed images of immunoblotting.

PVDF membranes were cut into several small pieces to incubate with different antibodies for immunoblotting. Black boxes indicate images showed in relevant figures.

	Con	RPL	P value
Total sample	28	24	
Maternal age(year)	31.2±0.7	33.0±1.1	ns
Gestational age(day)	53±1.3	53±1.9	ns
Number of spontaneous			
abortions	n/a	2.4±0.18	n/a

Supplementary Table S1. The clinical characteristics of the pregnant women enrolled in this study

Two-tailed unpaired Student's t test. Data are mean \pm SEM. D&C, dilatation and curettage; ns, not statistically significant;

Supplementary Table S2. Primer list of constructing overexpression vector

Gene	Forward Primer	
mCdx2	CCAGTAACGTGGCGCGCCTTAtcactgggtgacagtg	
hCDX2	ACTAGAGGATCTATTTCCGGTGAATTCgccaccatgtacgtgagctacctc	
KAT8	ACTGCTTACTGGCTTATCGGTCGACatggcggcacagggagctacag	
Gene	Reverse Primer	
mCdx2	CGATGACAAGGCTAGCatgtacgtgagctaccttc	
hCDX2	GCGGCTTCGGCCAGTAACGTGGATCCtcactgggtgacggtggggtttagc	
KAT8	CCGTCATGGTCTTTGTAGTCGCTAGCAGAGGAGCCcttcttggaaagcttgactt	

Supplementary Table S3. Primer list of quantitative RT-PCR

quantitative RT-PCR primer for mTSCs					
Gene	Forward Primer	Reverse Primer			
Cdx2	TCCTGCTGACTGCTTTCTGA	CCCTTCCTGATTTGTGGAGA			
Eomes	CCTGGTGGTGTTTTGTTGTG	TTTAATAGCACCGGGGCACTC			
Pl1	TGGAGCCTACATTGTGGTGG	TGGCAGTTGGTTTGGAGGA			
Pl2	CCAACGTGTGATTGTGGTGT	TGCCACCATGTGTTTCAGAG			
Actb	TGTTACCAACTGGGACGACA	GGGGTGTTGAAGGTCTCAAA			
Kat8	ATGGGTGGACAAGAACCGAC	GTTTCGAGTGATCTTGCGCTC			
Esrrb	GCAGTCCTTCGTGCTGTCTCA	TACATGGGACTGGATGGGAGAT			
Elf5	ACAAGCTTGATGCCAACTGC	CCTTGCGAGCGAATGTTCTG			
Ctsq	CAGGCAAACTGATCCCACTGA	CAACGACAGCCTCTATTGCCTT			
Plf	CTCAGAGACAAAAGCCCCATGA	TGAGCCTGGCTTGTTCCTTG			
Ly6a	AGGAGGCAGCAGTTATTGTGG	CGTTGACCTTAGTACCCAGGA			
quantitative RT-PCR primer for hTOs					
Gene	Forward Primer	Reverse Primer			
CDX2	GACGTGAGCATGTACCCTAGC	GCGTAGCCATTCCAGTCCT			
KAT8	GTCACGGTGGAGATCGGAGA	CCCTCCTGGTCGTTCACTC			

ACTB CTACCTCATGAAGATCCTCACCGA TTCTCCTTAATGTCACGCACGATT