

1 Supplemental Figures and Legends

2 *Supplemental Figure 1. Binding to TAFV, BOMV and RESTV GP and RESTV pseudovirus neutralization*
3 *by rabbit mAb panel*

4 **A) Binding of broadly reactive rabbit mAb panel to full-length transmembrane GPs from *Ebolavirus***
5 **species that have not caused multiple outbreaks of EVD in humans.** MDCK SIAT-1 cells expressing
6 TAFV GP (dark blue), RESTV GP (dark red), BOMV GP (turquoise) or parental cells not expressing GP
7 (No GP, grey) were incubated with 50 µg/mL mAb, and mAb binding was detected using an Alexa Fluor
8 647 anti-IgG conjugate secondary antibody. R5.034 (non-anti-GP mAb) and 66-3-9C (broadly reactive
9 anti-GP mAb) were included as negative and positive controls, respectively. Background (mean of 8
10 wells per assay plate incubated with relevant secondary only) was subtracted from data. Mean and
11 SEM of duplicates shown for each mAb.

12 **B) *In vitro* neutralization of RESTV GP pseudotyped S-FLU virus.** S-FLU viruses coated with RESTV GP
13 were incubated with mAb, and MDCK SIAT-1 cells then infected with the mAb-virus mixture. Percent
14 neutralization of S-FLU virus by titrated mAb was calculated using maximal (no antibody added) and
15 minimal fluorescence (no virus added) signals within each assay. Duplicates within assay for each mAb
16 concentration and calculated non-linear regression curves are shown.

17

18 *Supplemental Figure 2. Development of chimeric GP for assessment of fusion loop binding.*

19 **(A) Alignment of Internal Fusion Loop (IFL) sequences from *Ebolaviruses* and Marburgvirus (MARV)**
20 **GP sequences obtained from NCBI Reference database.** EBOV (Ebola virus/H.sapiens-
21 wt/GIN/2014/Makona-C15; ATY51135), SUDV (Sudan virus/H.sapiens-tc/UGA/2000/Gulu-808892;
22 YP_138523.1), BDBV (Bundibugyo virus/H.sapiens-tc/UGA/2007/Butalya-811250; YP_003815435.1),
23 TAFV (Tai Forest virus/H.sapiens-tc/CIV/1994/Pauleoula-CI; YP_003815426.1), MARV (Marburg
24 virus/H.sapiens-tc/KEN/1980/Mt.Elgon-Musoke; YP_001531156.1). Numbering associated with EBOV
25 sequence. Region in box indicates sequences that are swapped to produce chimeric EBOV GP with

26 MARV IFL ("E/M chimeric GP"). Numbered according to EBOV sequence. Differences between MARV
27 and EBOV sequences are highlighted in blue in the MARV sequence.

28 **(B) Validation of chimeric GP as a tool to assess IFL binding.** (i) Confluent monolayers of MDCK-SIAT
29 1 cells expressing the GPs on their surface were treated with thermolysin to reveal the RBR, then
30 incubated with 10 µg/mL biotinylated NPC1, MR78 (pan-filovirus RBR-binding mAb control) or R5.014
31 (irrelevant malaria mAb control) in triplicate. NPC1 was detected using a streptavidin Alexa Fluor 647
32 conjugate; MR78 and R5.014 were detected using anti-human IgG Alexa Fluor 647 conjugate. Cells
33 were then stained with wheat germ agglutinin Alexa Fluor 488 conjugate to normalize for number of
34 cells remaining in each well after thermolysin treatment. The mean and SEM of the ratio between
35 fluorescence intensities of the 647 and 488 conjugates of triplicates in each assay are shown. (ii)
36 Confluent cell monolayers were incubated with reference mAbs at 10 µg/mL which were then
37 detected using anti-human IgG AlexaFluor 488 conjugate. The mean and SEM of duplicates in each
38 assay are shown. Representative data are shown for mAbs 040, 6662, 6541 and FVM02 which were
39 repeated in N=3 experiments.

40 **(C). Broadly reactive rabbit mAbs do not lose binding to chimeric GP with MARV IFL.** Confluent
41 monolayers of MDCK-SIAT 1 cells expressing the GPs on their surface were incubated with test mAbs
42 at 10 µg/mL, and mAb binding was detected using anti-rabbit IgG Alexa Fluor 488 conjugate. The mean
43 and SEM of duplicates within each assay are shown. Representative data shown from N=2 repeats of
44 each experiment. Serum from the rabbit immunized with *Ebolavirus* GPs for initial antibody discovery
45 was included as a positive control.

46

47 *Supplemental Figure 3. Binding of mAb panel to cleaved and uncleaved Ebolavirus GPs*

48 **(A) Antibody binding to thermolysin (THL) cleaved cell surface-expressed SUDV, BDBV and TAFV GP**
49 **was assessed in an immunofluorescence assay.** For each test mAb, binding to GP was tested under
50 two conditions: GP-expressing cells were pre-incubated with THL to produce GP_{CL}, then incubated with
51 test mAbs (dark grey); and mAbs were incubated with GP-expressing cells without THL (mid-grey).

52 mAb binding was then detected using Alexa Fluor 647 conjugate and cells stained with wheat germ
53 agglutinin Alexa Fluor 488 conjugate. Mean and SEM for replicate wells within the assay (6 wells for
54 GP with THL, 3 wells for GP without THL treatment) are shown. 040 (GC binding mAb) and 6541 (base
55 binding mAb) are comparator mAbs showing THL sensitive and insensitive binding profiles. MR78 can
56 only bind to EBOV GP when the GL and MLD are removed exposing the RBR ¹, and is included as a
57 control to confirm THL digestion of GP in the assay.

58 **(B) Immunoprecipitation of thermolysin cleaved EBOV GP by mAbs 6541, 11889 and 11897.**

59 GP-expressing cells were treated to biotinylate surface proteins, then incubated with increasing
60 concentrations of THL. Cells were washed then lysed. GP was immunoprecipitated from cell lysates
61 using Protein A Sepharose and anti-GP mAb of interest. Samples were run on reducing SDS-PAGE, and
62 bands were revealed using streptavidin Alexa Fluor 647 conjugate. Band at ~150 kDa is full-length GP;
63 band at ~25 kDa is GP_{CL} (GP1 core and GP2); other bands are products of sequential cleavage of MLD
64 and GC. 6541 is a known base-binding mAb that can immunoprecipitate the GP1 core/GP2.

65

66 *Supplemental Figure 4. Comparison of 11883 and 11886 footprints with previously described*
67 *antibodies.*

68 **(A) Comparison of 11883 footprint with mAb114.** GP Δ muc trimer ectodomain with 11883 contacts
69 on the GP (pink) overlaid with mAb114 (teal) (PDB: 5FHC) contacts. Residues in common between the
70 two footprints are shown in yellow.

71 **(B) Comparison of 11886 footprint with other 3₁₀ pocket binding mAbs.** GP Δ muc trimer ectodomain
72 with 11886 contacts on the GP (green) overlaid with ADI-14956 contacts (light blue) (PDB: 6MAM) or
73 EBOV-520 (purple) (PDB: 6OZ9) contacts. Residues in common between the two footprints are shown
74 in yellow. For monomer with footprints shown, GP1 is shaded in light grey and GP2 in dark grey. Dotted
75 line indicates domains cleaved after cathepsin or THL treatment to generate GP_{CL}. In PDB: 6OZ9, the
76 GP does not contain the GC, therefore any contacts between EBOV-520 and the GC are not modelled.
77 However, in an additional structural study, PDB: 6PCI, a complex of EBOV GP Δ muc trimer with EBOV-

78 520 and a partner mAb that remodels the glycan cap, suggests that EBOV-520 does contact the GP1
79 in that structure, with contacts reported as Pro250, Gln251 and Gln255 with the α 2 helix displaced
80 upwards compared to the unliganded GP^{2,3}.

81 **(C) VPΔ30 EBOV with V505I or T402I mutations in the GP permits viral escape from neutralization**
82 **by mAb 11886.** PFU; Plaque-forming units. WT; wild-type. Viruses incubated with 10 μ g/mL 11886
83 mAb.

84 **(D) Alignment of *Ebolavirus* GP sequences showing conservation of 11886 and 11883 contact**
85 **residues.** mAb contact residues on EBOV GP (and conserved residues in other GPs) are highlighted.
86 11886 (green) and 11883 (pink). Location of residue Val505 highlighted by black arrow.

87

88 *Supplemental Figure 5. Additive interactions between 11886 and other broadly reactive partner mAbs*
89 *across *Ebolavirus* species in a pseudovirus neutralization assay.*

90 **(A) 11886 with partner RBR mAbs 11883, mAb114 and 6662.**

91 **(B) 11886 with partner fusion loop mAb CA45.**

92 For all data, a representative experiment of at least two independent repeats has been shown. Dotted
93 line represents the mean inhibition given by a held concentration of 11886 with 95 % confidence limits
94 shown in shaded grey. Solid lines represent mean of triplicate wells within an experiment, with shaded
95 areas indicating the standard error. Red lines indicate the inhibition given by a titration series of the
96 partner mAb alone. Blue lines indicate the inhibition given by a titration series of the partner mAb plus
97 the held concentration of mAb 11886. Grey line represents the calculated Bliss Additivity value which
98 assumes a purely independent and additive interaction between the partner mAb and 11886. Held
99 concentration of 11886 ranged from 0.2-0.6 μ g/mL between experiments to achieve target 20-45 %
100 inhibition in a given assay.

101

102 *Supplemental Figure 6. Neutralization by component antibodies of cocktail mixes.*

103 Antibodies were tested for neutralization of EBOV (purple), SUDV (orange) and BDBV (teal) GP coated
104 S-FLU pseudoviruses. Open symbols represent results where 50 % neutralization of virus was not
105 achieved at the highest concentration of the test antibody assayed in that experiment. Horizontal
106 dotted lines denote 12.5, 25 and 50 µg/mL of total antibody respectively.

107 **(A)** Median and 95 % CI of IC50 values of individual mAbs that were included in cocktail mixes (**Figure**
108 **7**) from N=2 to N=5 experiments are shown.

109 **(B)** Median and 95% CI of the IC80 values (concentration of antibody required to achieve 80 % virus
110 neutralization) for antibody mixes and 11886 alone were calculated from N=3 experiments. For
111 comparison, median and 95 % CI of IC80 values of individual antibodies 11883 and mAb114 calculated
112 from N=2 to N=5 experiments run separately are also shown.

113

114 Supplementary References

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