

Supplementary Information

UPF1 Plays Critical Roles in Early B Cell Development

Supplementary Figure 1 Analysis and generation of B cell specific *Upf1* deficient mice

Supplementary Figure 2 Enrichment score plot of down-regulated genes in *Upf1*-deficient early LPre-B cells

Supplementary Figure 3 Loss of IFN α R1 signaling does not impact B cell development.

Supplementary Figure 4 Genetic pre-arrangement of *Igh* locus rescued early to late LPre B cell differentiation in *Upf1*-cKO mice

Supplementary Figure 5 Genetic pre-arrangement of *Igh* locus did not rescue the splenic B cell population in *Upf1*-cKO mice

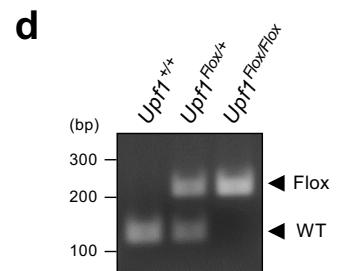
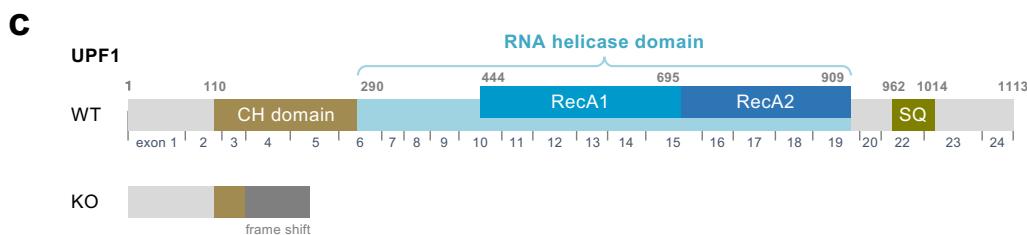
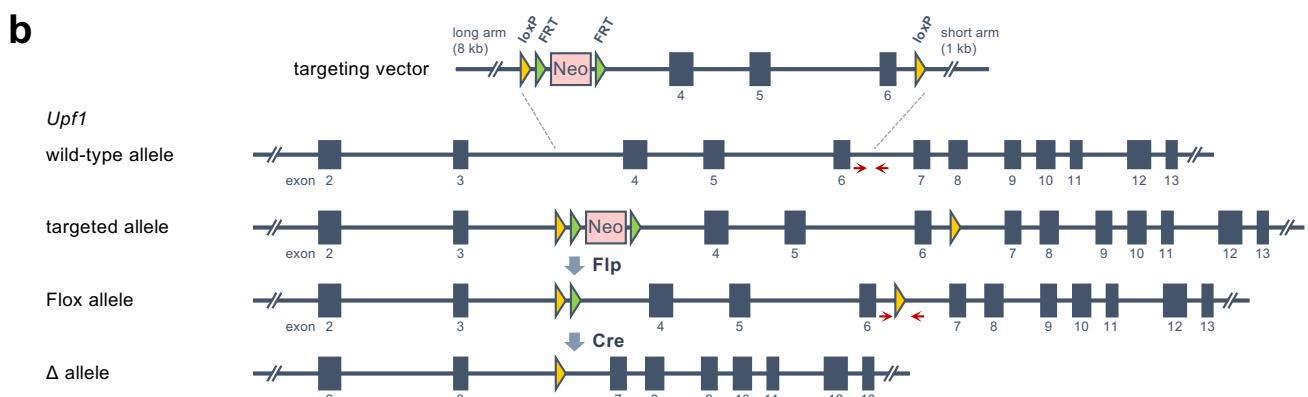
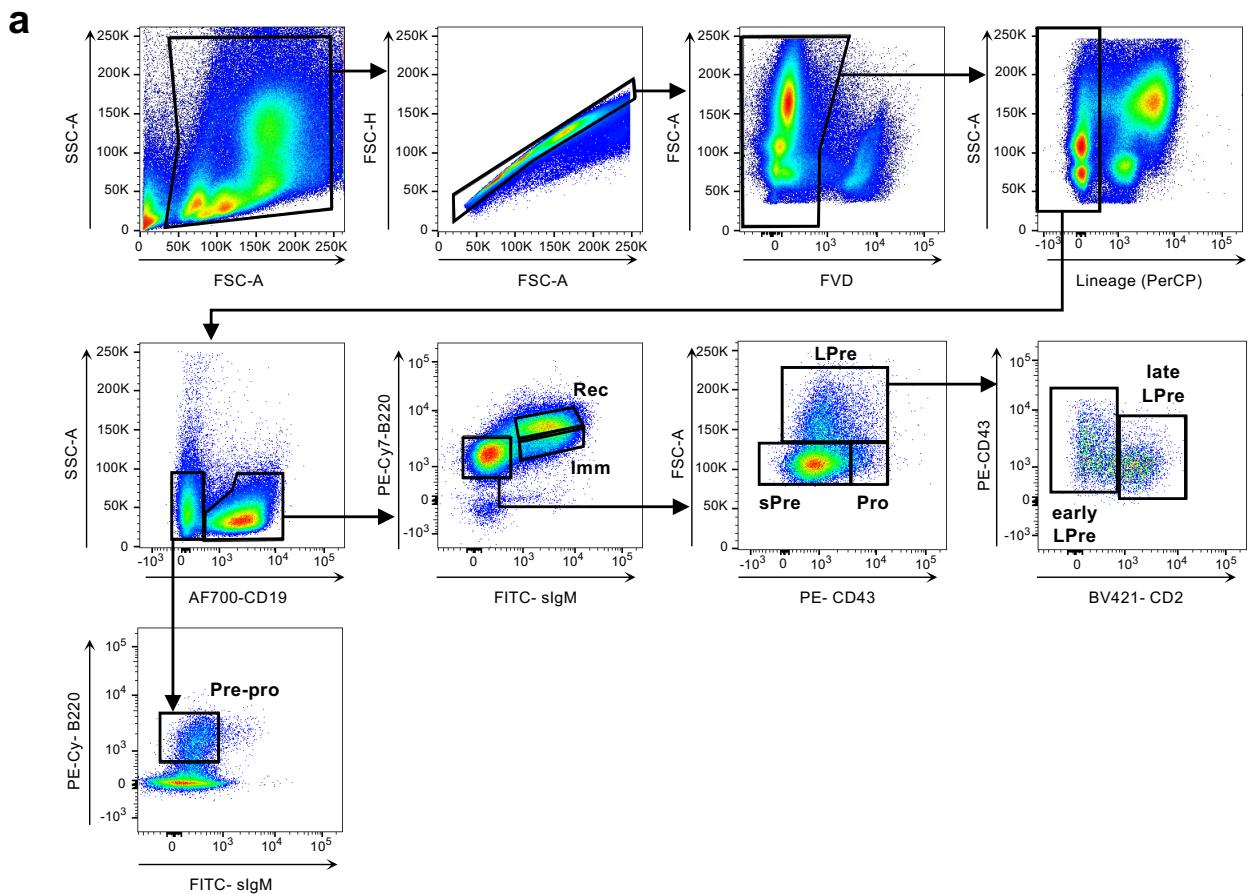
Supplementary Figure 6 Transcriptome analysis of *Upf1*-cKO/*Igh*^{B1-8hi} early LPre- and sPre-B cells

Supplementary Figure 7 Identifying UPF1-target RNAs

Supplementary Table 1 List of primers used for PCR

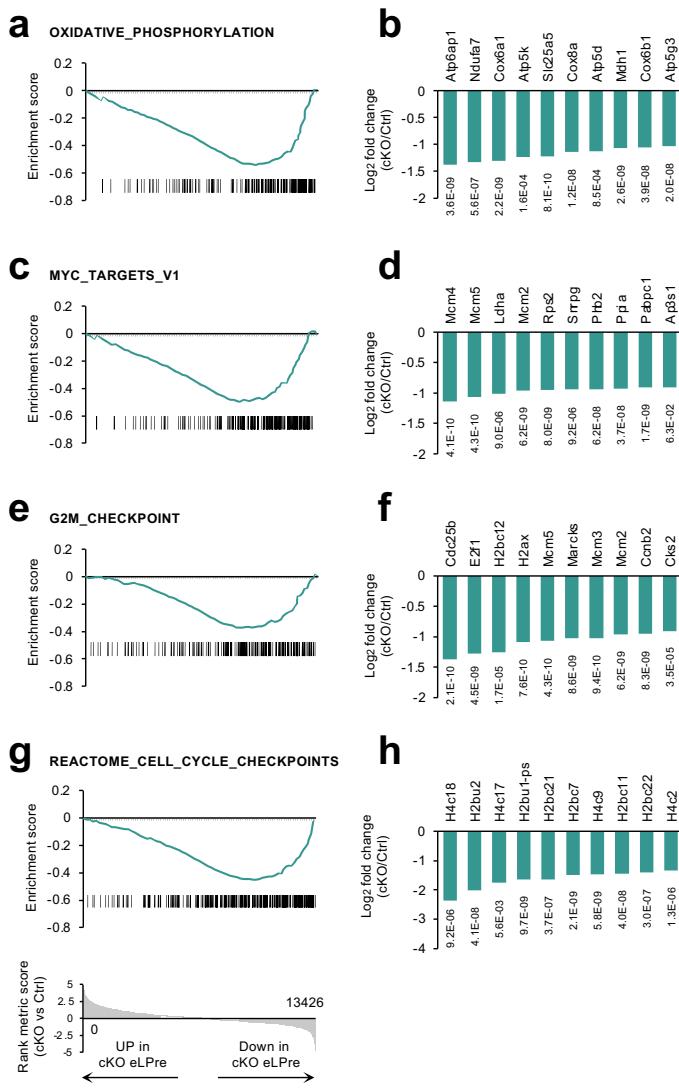
Supplementary Table 2 List of antibodies

Supplementary Table 3 List of primers used for RT-qPCR



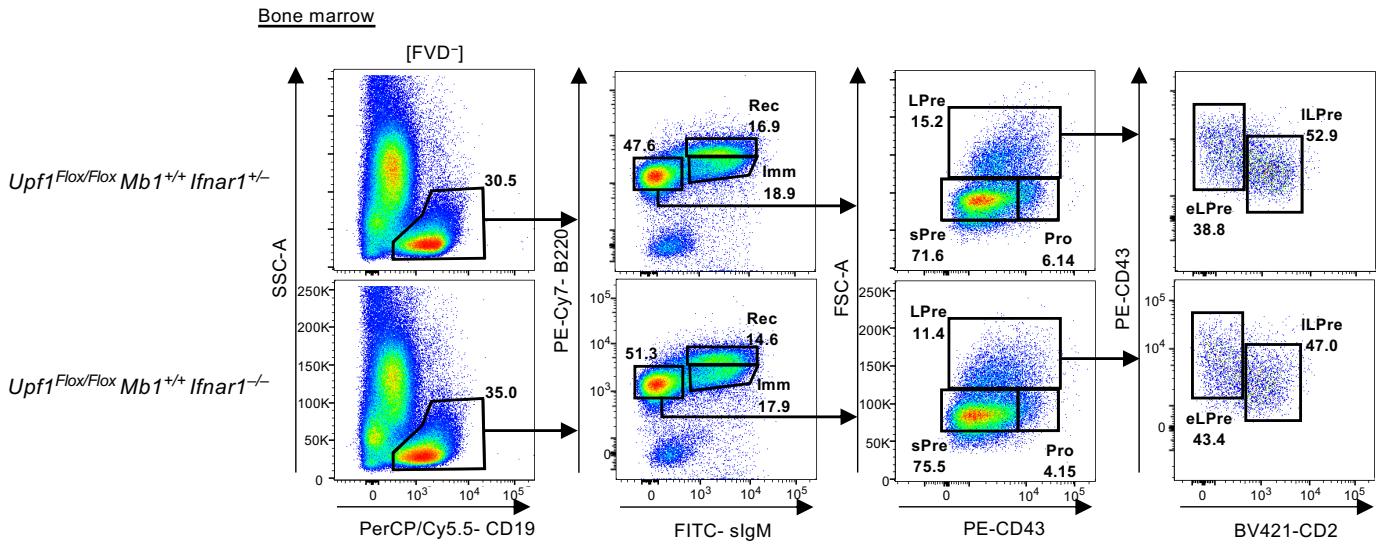
Supplementary Figure 1 Analysis and generation of B cell specific Upf1 deficient mice

(a) Gating strategy of BM B cells. Harvested cells were stained and separated with the indicated markers.
 (b) Structure of the target *Upf1* allele. Red arrows indicate the primers used for genotyping. (c) The schematic illustration of mouse UPF1 and predicted translation product from *Upf1*^{Flox} allele in *Upf1*-cKO.
 (d) PCR genotyping of genomic DNAs from *Upf1*^{+/+}, *Upf1*^{Flox/+}, or *Upf1*^{Flox/Flox} mice. Results are representative of at least three independent experiments.



Supplementary Figure 2 Enrichment score plot of down-regulated genes in *Upf1*-deficient early LPre-B cells

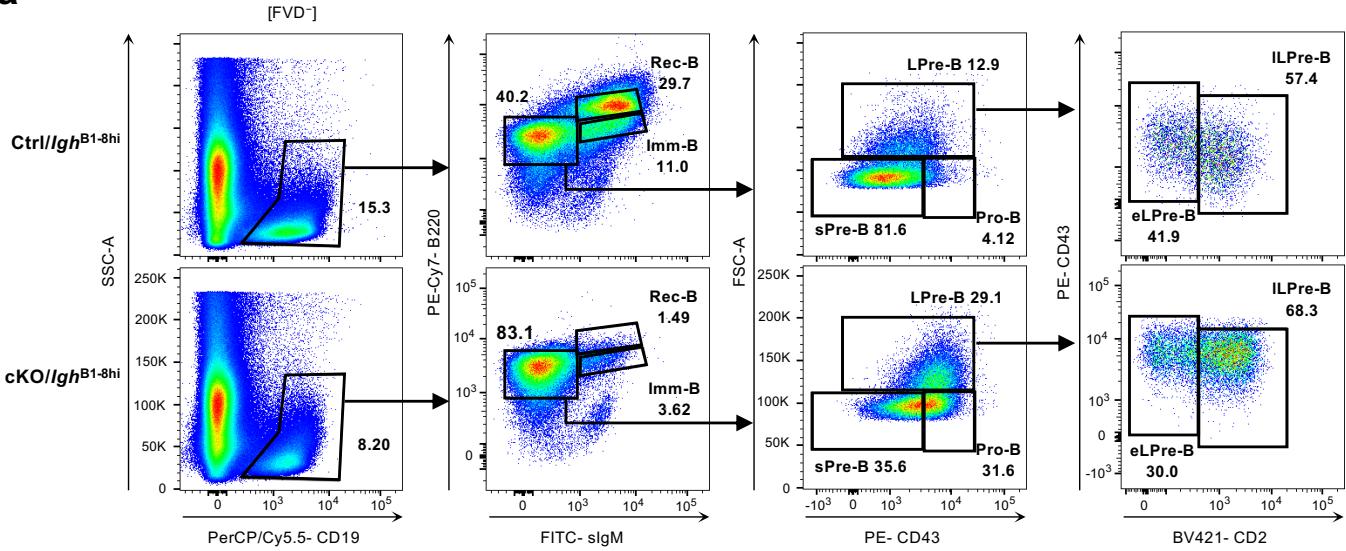
(a-h) Enrichment score plot (a, c, e, g) and comparison of each gene expression between *Upf1*-cKO- and Ctrl-early LPre-B cells (b, d, f, h) in indicated gene sets (a, b; OXIDATIVE PHOSPHORYLATION, c, d; MYC TARGETS V1, e, f; G2M CHECKPOINT, g, h; CELL CYCLE CHECKPOINTS). Bars indicate Log₂FC (Upf1-cKO vs Ctrl) of the top 10 genes with the largest difference between cKO and Ctrl, and the number adjacent to each bar indicates adj.*P*. The statistical analyses were performed using GSEA.



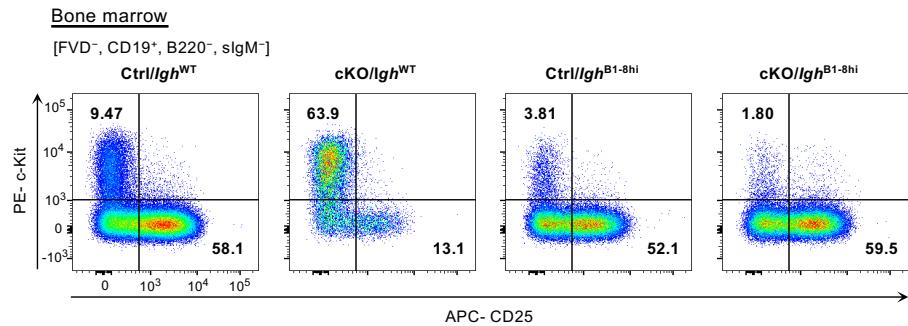
Supplementary Figure 3 Loss of IFN α R1 signaling does not impact B cell development.

Flow cytometry plots of indicated populations in the BM of indicated mice. Results are representative of two independent experiments.

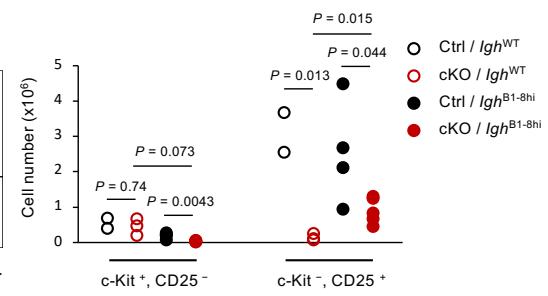
a Bone marrow



b

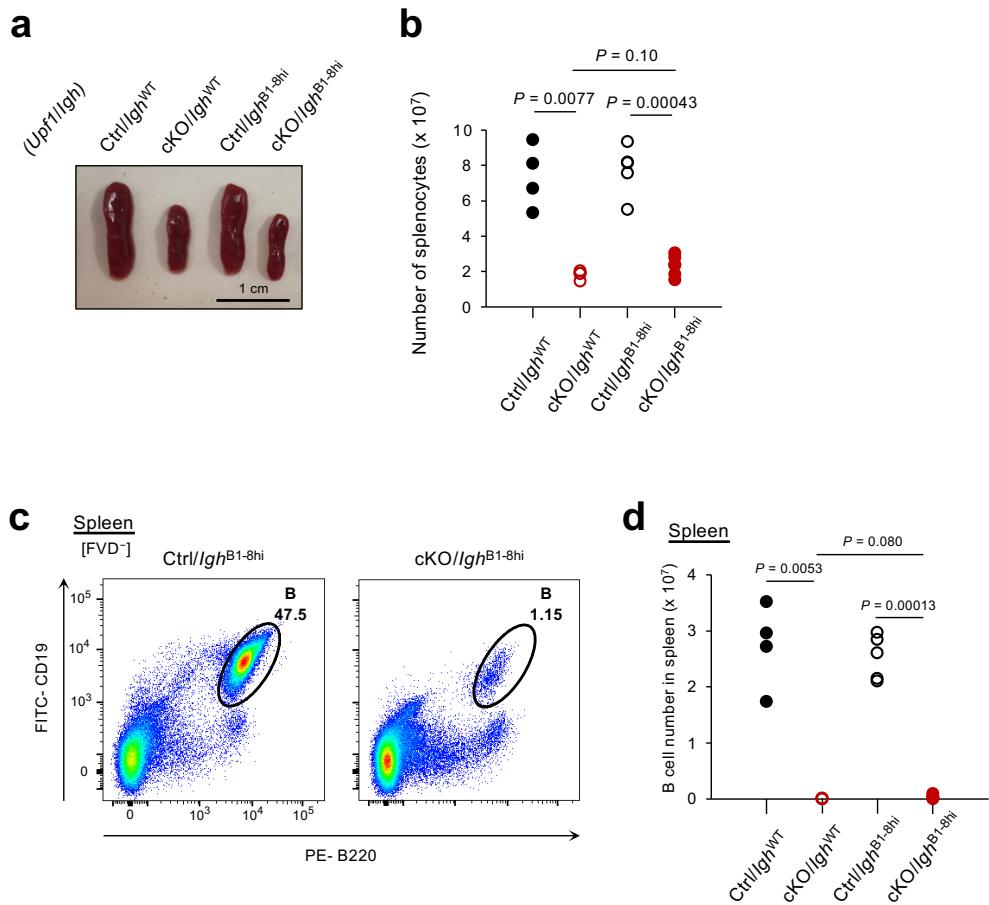


c



Supplementary Figure 4 Genetic pre-arrangement of *Igh* locus rescued early to late LPre B cell differentiation in *Upf1*-cKO mice

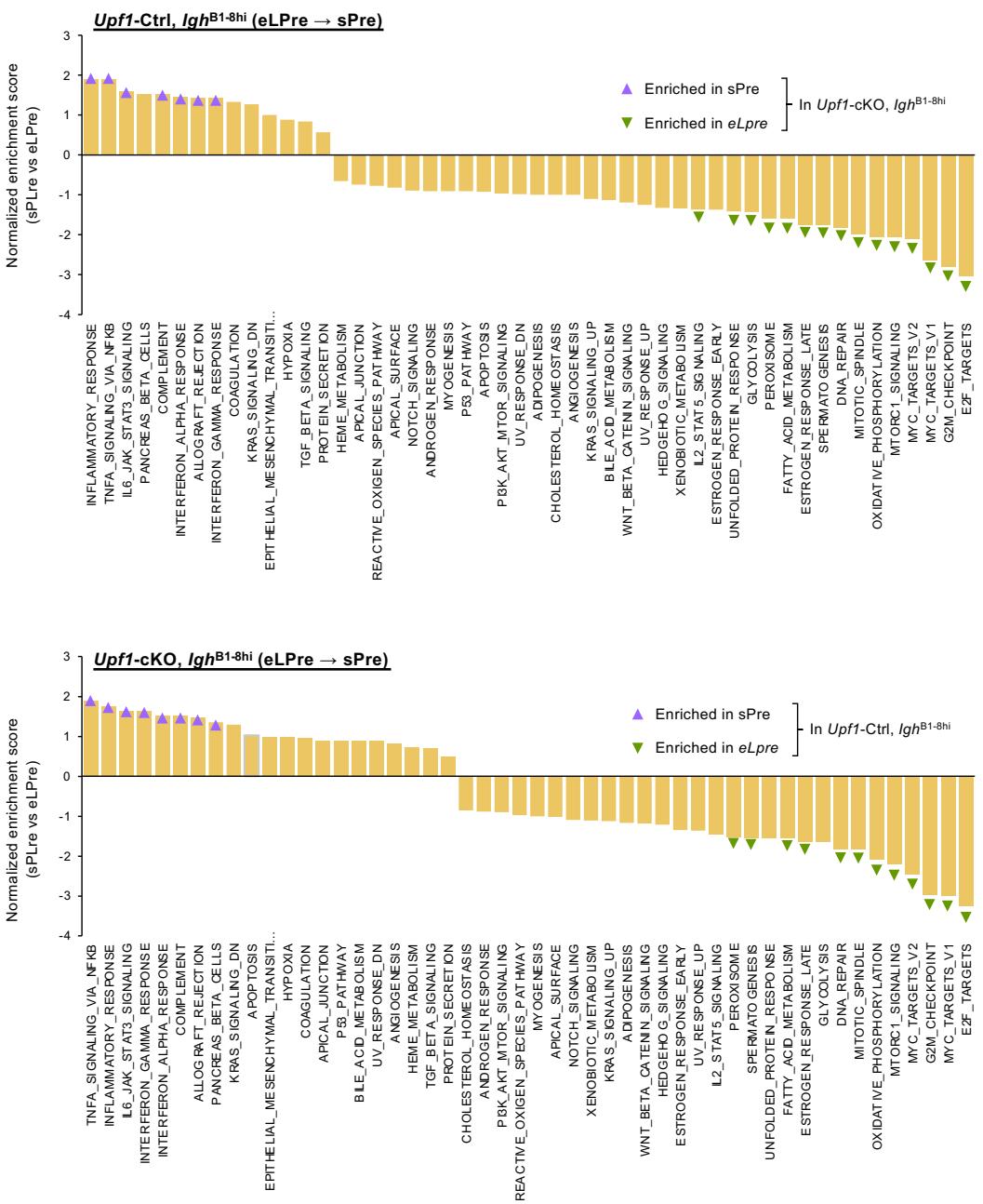
(a) Flow cytometry of indicated populations in the BM of indicated mice. (b, c) Flow cytometry plots (b) and cell number quantification (c) of FVD⁻, CD19⁺, B220⁺, sIgM⁻ cells derived from BM of indicated mice. BMs were harvested from bones of both hind legs. The *p*-values were calculated using two-sided Student's t-test. Results are representative of at least three independent experiments ($N=3$ (Ctrl/*Igh*^{WT}, cKO/*Igh*^{WT}), $N=4$ (Ctrl/*Igh*^{B1-8hi}), $N=5$ (cKO/*Igh*^{B1-8hi}).



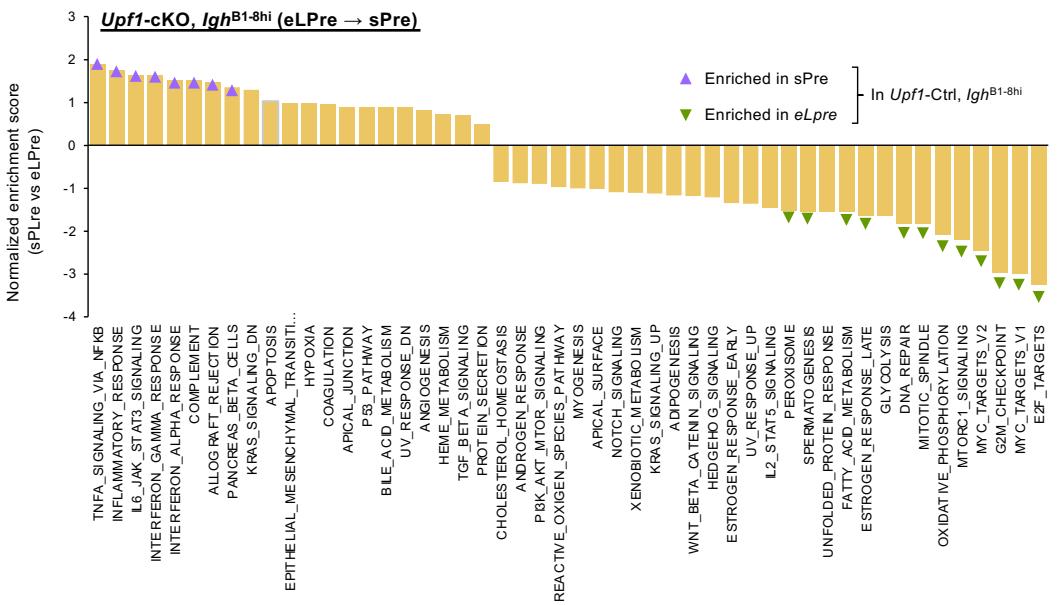
Supplementary Figure 5 Genetic pre-arrangement of *Igh* locus did not rescue the splenic B cell population in *Upf1*-cKO mice

(a) Appearance of spleens derived from indicated mice. Results are representative of at least three independent experiments. (b) Numbers of splenocytes derived from indicated mice. $N = 4$ (Ctrl/*Igh*^{WT}, cKO/*Igh*^{WT}), $N = 5$ (Ctrl/*Igh*^{B1-8hi}), $N = 6$ (cKO/*Igh*^{B1-8hi}). (c and d) Flow cytometry plots (c) and quantifications (d) of splenic B cells (B220⁺, CD19⁺) of indicated mice. $N = 4$ (Ctrl/*Igh*^{WT} cKO/*Igh*^{WT}), $N = 5$ (Ctrl/*Igh*^{B1-8hi}), $N = 6$ (cKO/*Igh*^{B1-8hi}). The *p*-values were calculated using two-sided Student's t-test.

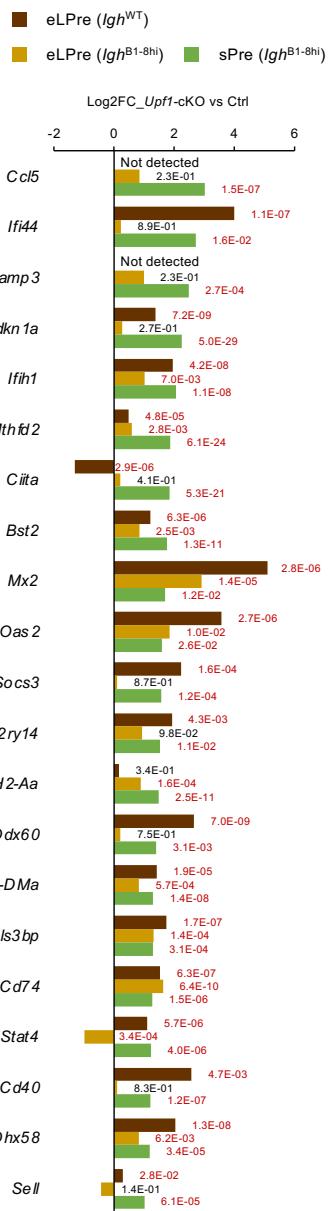
a



b

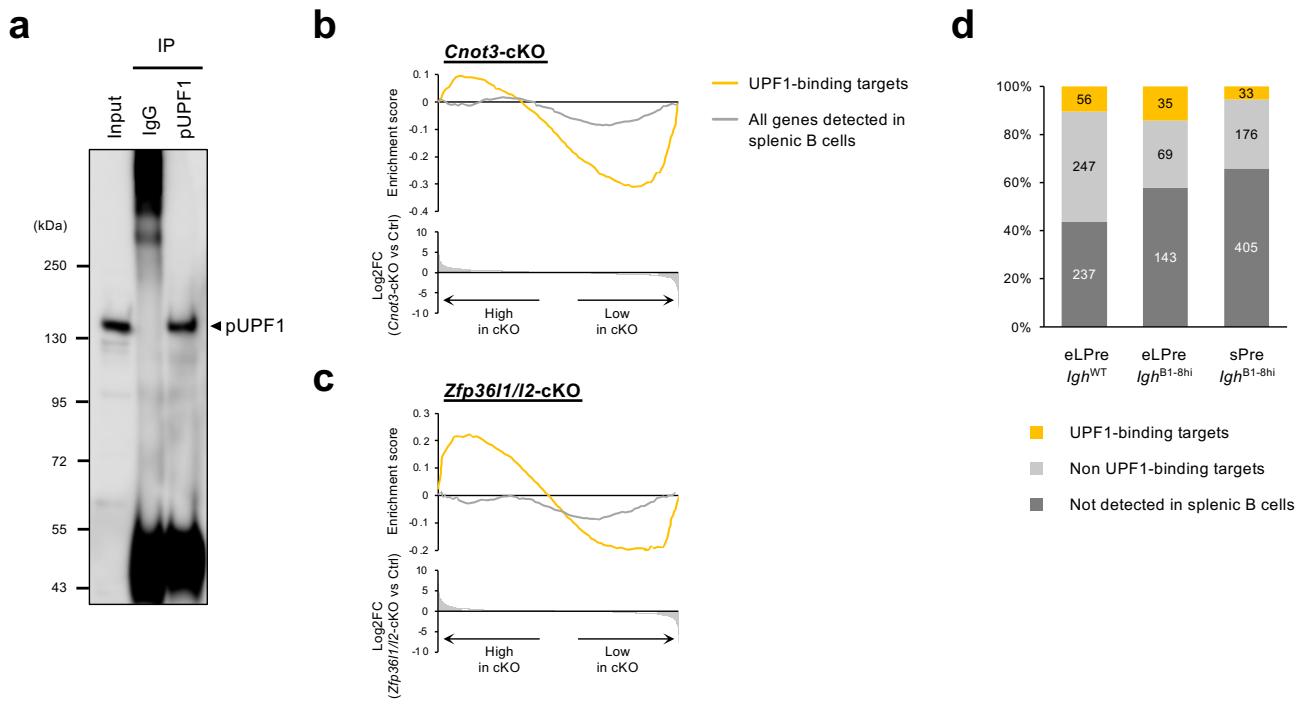


c



Supplementary Figure 6 Transcriptome analysis of *Upf1*-cKO/*Igh*^{B1-8hi} early LPre- and sPre-B cells

(a, b) Normalized enrichment score of GSEA (sPre- vs early LPre (eLPre)-B cells) in *Upf1*-Ctrl/*Igh*^{B1-8hi} (a) and *Upf1*-cKO/*Igh*^{B1-8hi} (b). Green and purple arrows indicate the gene sets enriched in early LPre- and sPre-B cells in antagonistic genotype (*Upf1*-Ctrl/*Igh*^{B1-8hi} for *Upf1*-cKO/*Igh*^{B1-8hi} and vice versa), respectively. (c) Comparison of IFNA/IFNG RESPONSE gene expression between *Upf1*-cKO vs Ctrl in indicated genotypes and cell types. The number adjacent to each bar indicates adj.*P* (red; adj.*P* < 0.05). Only the genes with significantly high expression in *Upf1*-cKO/*Igh*^{B1-8hi} sPre B cells (Log2FC ≥ 1, adj.*P* < 0.05) are shown. The statistical analyses were performed using limma (*Igh*^{WT}), and edgeR (*Igh*^{B1-8hi}).



Supplementary Figure 7 Identifying UPF1-target RNAs

(a) Immunoblot analysis of p-UPF1 precipitated with anti-p-UPF1 antibody or normal rIgG1 (negative control). Arrow indicates p-UPF1. Results are representative of two independent experiments. **(b, c)** Enrichment score plot of UPF1-binding targets (yellow) and all genes detected in splenic B cells (gray) in *Cnot3*-cKO vs Ctrl **(b)** and *Zfp36l1/l2*-cKO vs Ctrl **(c)**. **(d)** Percentage of detected (yellow; target of UPF1, gray; non-target of UPF1) and undetected (dark gray) RNAs in RIP-seq using splenic B cells among the genes with significantly high expression in *Upf1*-cKO in indicated genotypes and cell types. The absolute number of the genes is shown in each bar.

Supplementary Table 1 List of primers used for PCR

Genotyping for <i>Upf1</i> -Flox forward	GAATTGGTGGCATGGGCCCCATATCCCAG
Genotyping for <i>Upf1</i> -Flox reverse	GACTCTCCTGGCCTGAGCCTGGATG
V _H J558 forward	CGAGCTCTCCARCACAGCCTWCATGCARCTCARC
V _H 7183 forward	CGGTACCAAGAASAMCCTGTWCCTGCAAATGASC
D _H forward	TTCAAAGCACAATGCCTGGCT
J _H 3 reverse	GTCTAGATTCTACAAGAGTCGATAGACCCTGG
C _μ forward	TGGCCATGGGCTGCCTAGCCCGGGACTT
C _μ reverse	GCCTGACTGAGCTCACACAAGGAGGA
V _k forward	GGCTGCAGSTTCAGTGGCAGTGGRTCWGGRAC
J _k reverse	CTCATTCTGTTGAAGCTTGTGACAATGGG

Supplementary Table 2 List of antibodies

Antibody	Cat#	Company	Clone	Application	Dilution
anti- <i>Upf1</i>	NBP1-05967	Novus biologicals	Polyclonal	Immunoblot	1:1000
anti-phospho- <i>Upf1</i>	07-1016	Merck	Polyclonal	Immunoblot	1:1000
anti-β-actin	sc-1615	Santa Cruz	Polyclonal	Immunoblot	1:2000
anti-Mouse IgG	NA9310	Cytiva	Polyclonal	Immunoblot	1:2000
anti-Rabbit IgG	NA9340	Cytiva	Polyclonal	Immunoblot	1:2000
Biotin anti-mouse CD2	100103	Biolegend	RM2-5	Flow cytometry	1:300
PerCP/Cy5.5 anti-mouse CD19	152405	Biolegend	1D3	Flow cytometry	1:200
FITC anti-mouse CD19	115506	Biolegend	6D5	Flow cytometry	1:200
Alexa Fluor 700 anti-mouse CD19	115527	Biolegend	6D5	Flow cytometry	1:200
APC Rat Anti-Mouse CD19	550992	BD Pharmingen	1D3	Flow cytometry	1:200
APC anti-mouse CD25	102011	Biolegend	PC61	Flow cytometry	1:200
PE Rat Anti-Mouse CD43	561857	BD Pharmingen	S7	Flow cytometry	1:200
FITC anti-mouse IgM	406505	Biolegend	RMM-1	Flow cytometry	1:200
APC anti-mouse IgM	406509	Biolegend	RMM-1	Flow cytometry	1:200
PE-Cy7 Rat Anti-Mouse CD45R/B220	552772	BD Pharmingen	RA3-6B2	Flow cytometry	1:200
FITC anti-mouse/human CD45R/B220	103206	Biolegend	RA3-6B2	Flow cytometry	1:200
PE anti-mouse/human CD45R/B220	103208	Biolegend	RA3-6B2	Flow cytometry	1:200
PE anti-mouse CD117 (c-Kit)	105807	Biolegend	2B8	Flow cytometry	1:200
PerCP Hamster Anti-Mouse CD3e	553067	BD Pharmingen	145-2C11	Flow cytometry	1:100
PerCP/Cy5.5 anti-mouse/human CD11b	101228	Biolegend	M1/70	Flow cytometry	1:100
PerCP anti-mouse CD11c	117326	Biolegend	N418	Flow cytometry	1:100
PerCP anti-mouse F4/80	123125	Biolegend	BM8	Flow cytometry	1:100
PerCP/Cy5.5 anti-mouse TER-119	116227	Biolegend	TER-119	Flow cytometry	1:100
Brilliant Violet 421 Streptavidin	405225	Biolegend	—	Flow cytometry	1:400

Supplementary Table 3 List of primers used for RT-qPCR

	Forward	Reverse
18S rRNA	CGGACAGGATTGACAGATTG	CAAATCGCTCCACCAAGTAA
mouse Upf1	GCTGAACCTCGAGGAAGATG	CTTCCTTGATTTGCCCTC
mouse Gas5	CAGGTATTAATGGGTACACCTC	CTTCTATTGAGCCTCCATCC
mouse Mx1	GATCCGACTTCACTCCAGATGG	CATCTCAGTGGTAGTCAACCC
mouse Mx2	TTCACCAGGCTCGAAAAGAG	ACAAACCCCTGGCAATTCTCG