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Supplemental information

Hippocampal astrocytes induce sex-dimorphic

effects on memory

Samantha M. Meadows, Fernando Palaguachi, Minwoo Wendy Jang, Avital Licht-Murava, Daniel Barnett, Till S. Zimmer, Constance Zhou, Samantha R. McDonough, Adam L. Orr, and Anna G. Orr

Supplemental Figures



Figure S1. Further characterization of *Aldh111*-Cas9 mice and mGluR levels. Related to Figure 1.

(**A**) mGluR3 (red) and Cas9-eGFP (green) co-immunolabeling from the CA1 stratum radiatum (CA1sr, i) and DG molecular layer (DGmol) (ii) in sgRNA-injected *Aldh111*-CreER^{T2} single transgenic control mice. Scale bars: 400 μm, 40 μm (insets). (**B**) Cas9-eGFP (green) co-immunolabeling with cell type-specific markers (red), including GFAP for astrocytes (i, ii), Iba1 for microglia/macrophages (iii, iv), and NeuN for neurons (v, vi) in *Aldh111*-Cas9 double transgenic mice treated with tamoxifen (TAM). DAPI (blue) labeled cell nuclei. Scale bars: 400 μm, 40 μm (insets). (**C**) Immunofluorescence-based quantification of the percentage of GFAP-positive cells that were also Cas9-eGFP-positive in the CA1sr and DGmol regions of the hippocampus

of TAM-injected Aldh111-Cas9 mice. (D) mGluR3 (red) and Cas9-eGFP (green) co-immunolabeling in saline or sgRNA-injected Aldh111-Cas9 mice. Images highlight astrocytic loss of mGluR3 in Cas9-positive cells in the dentate gyrus (DG). Scale bar: 10 µm (E) Immunofluorescence-based quantification of the % DG area that was mGluR3-positive in saline-injected (Con) and sgRNA-injected Aldh111-Cas9 mice treated with TAM. Two-way ANOVA: F(1, 31) = 39.40, p < 0.001 for main effect of sqRNA; F(1, 31) = 0.59, p = 0.45 for interaction effect. (F) Immunofluorescence-based (protein) and RT-qPCR-based (mRNA) quantification of mGluR3 levels in the CA and DG regions of saline-treated male and female mice. Two-way ANOVA (Protein): F(1, 48) = 0.28, p = 0.60 for main effect of sex. Two-way ANOVA (mRNA): F(1, 33) = 0.03, p = 0.86 for main effect of sex. (G) RNA levels of different mGluR subtypes in the DG region of mice with or without mGluR3 knockdown, shown in the order of sequence homology to mGluR3. Three-way ANOVA: F(5, 173) = 1.01, p = 0.41 for interaction effect. (H) mGluR3 (magenta) and Cas9-eGFP (green) co-immunolabeling in saline- or sgRNA-injected Aldh1l1-Cas9 mice. Images highlight astrocytic loss of mGluR3 in Cas9-positive cells in the dentate gyrus (DG), but not in the neocortex or thalamus. Scale bar: 200 µm. (I) Immunofluorescence-based quantification of mGluR3 expression across brain regions, including the DG, neocortex (CTX), and thalamus (THL) in saline or sgRNA-injected Aldh111-Cas9 mice. Two-way ANOVA: F(2, 83) = 37.72, p < 0.0001 for interaction effect. (J) Experimental timeline for mice with astrocytic mGluR3 knockdown. Sidak's multiple comparisons post-hoc test (E, F, G, I): ###p < 0.001. Data are represented as mean ± SEM. See Table S3 for replicate details.



Figure S2. Additional behavioral characterization of *Aldh111*-Cas9 mice with or without mGluR3 knockdown in hippocampal astrocytes. Related to Figure 1. (A) Initial trajectory errors in male and female mice during training. Three-way ANOVA: F(19, 820) = 1.07, p = 0.38 for interaction effect. (B, C) Search strategies in male and female mice during training. (D, E) Swim speeds in probe trials conducted one (D) and nine (E) days post-training. Two-way ANOVA: F(1, 38) = 0.03, p = 0.85 for interaction effect (D); F(1, 40) = 1.17, p = 0.29 for interaction effect (E). (F) Target platform crossings as compared to other analogous

locations during a probe trial performed nine days after training. Target preferences were compared using Student's t-tests:*p < 0.05. Index values were analyzed using a two-way ANOVA with Sidak's multiple comparisons post-hoc test. Two-way ANOVA (crossings index): F(1, 39) = 1.59, p = 0.215 for interaction effect. (**G**) Exploration in the elevated plus maze (EPM). Two-way ANOVA: F(1, 37) = 10.88, p = 0.0022 for treatment/sex interaction effect. (**H**) Open-arm entries in the EPM. Two-way ANOVA: F(1, 39) = 0.23, p = 0.63 for interaction effect. Three-way (A) or two-way ANOVA with Sidak's multiple comparisons or Tukey's post-hoc tests: #p < 0.05; Sidak's post-hoc test (vs. male/saline group): ^^p < 0.01. NS: not significant. Data in panels A and D–H are represented as mean ± SEM. See Table S3 for replicate details.



Figure S3. Hippocampal mGluR3 expression levels and validation of AAV vector encoding HA-tagged mGluR3. Related to Figure 2. (**A**) Experimental timeline for mice with increased astrocytic mGluR3 expression. (**B**) mGluR3 (green) and GFAP (red) co-immunolabeling in the hippocampus of saline-injected (control) *Aldh111*-CreER^{T2} male and female mice imaged with longer exposure (as shown in insets included in

main Figure 2). DAPI (blue) labeled cell nuclei. Scale bars: 400 µm, 40 µm (insets). (C) mGluR3 (green) and NeuN (red) co-immunolabeling in the CA1 and DG of AAV vector-injected Aldh111-CreER^{T2} male and female mice. Scale bars: 400 µm, 40 µm (insets). (D) HA-mGluR3 (green) and GFAP or Iba1 (red) coimmunolabeling in the DG of Aldh111-CreER^{T2} mice injected with either saline or mGluR3-encoding AAV vector. DAPI (blue) labeled cell nuclei. Scale bar: 10 µm. (E) Total mGluR3 expression (green) and GFAP (astrocyte, red) or NeuN (neuron, red) co-immunolabeling in the DG of Aldh111-CreER^{T2} mice injected with either saline or mGluR3-encoding AAV vector. Inherent differences in mGluR3 intensities required the use of different image acquisition and processing settings. Insets show images of saline-injected mice in which mGluR3 signal was processed similarly to mice with AAV injections. Scale bar: 10 µm. (F) Representative Western blot and guantification of mGluR3 dimers and monomers in primary cultured astrocytes untransduced (UT) or transduced with a Cre-dependent AAV vector encoding mGluR3. Experiments were performed in two independent cultures. Student's t-test: ***p < 0.001. (G) Representative Western blots and quantification of phosphorylated Akt and total Akt levels following mGluR3 stimulation with LY354740 (LY, 1 µM, 10 min) in cultured astrocytes that were untransduced (UT) or transduced with a Cre-dependent AAV vector encoding mGluR3. Experiments were performed in two independent cultures. Two-way ANOVA of pAkt: F(1, 19) = 8.16, p = 0.01 for interaction effect. Sidak's multiple comparisons post-hoc test (vs Veh): #p < 0.01. (H) Co-immunolabeling for mGluR3 (green) and astrocyte marker GFAP (magenta) in the hippocampus of nontransgenic male and female mice at 4 and 12 months of age. DAPI (blue) labeled cell nuclei. Scale bar: 100 µm. (I) Quantification of mGluR3 immunolabeling in the indicated regions of the hippocampus of nontransgenic males and females at three different ages. Data were normalized to 4-monthold males per brain region. Student's t-test (males vs. females per age group): *p < 0.05. Two-way ANOVA (DGmol): F(2, 63) = 66.88, p < 0.0001 for age effect. Sidak's multiple comparisons post-hoc test: *p < 0.05. $^{\#\#}$ p < 0.0001 (vs. 4 months of age per sex). Two-way ANOVA (CA1rad): F(2, 65) = 21.64, p < 0.0001 for age effect. Sidak's multiple comparisons post-hoc test: ###p < 0.0001 (vs. 4 months of age per sex). Data in panels F, G, and I are represented as mean ± SEM. See Table S3 for replicate details.



Figure S4. Additional behavioral characterization of *Aldh111*-CreER^{T2} mice with or without enhancement in astrocytic mGluR3 expression. Related to Figure 2. (A) Initial trajectory errors during training in male and female *Aldh111*-CreER^{T2} mice after injection with mGluR3-encoding AAV vector or saline (Con). Three-way ANOVA: F(19, 1045) = 1.27, p = 0.19 for interaction effect. (B, C) Search strategies in male and female mice during training. (D, E) Swim speeds in male and female mice during probe trial at one day (D) and nine days (E) post-training. (F) Distance traveled in the elevated plus maze (EPM). Two-way ANOVA: F(1, 49) = 1.04, p = 0.31 for interaction effect. (G) Open-arm entries in the EPM. Two-way ANOVA: F(1, 50) = 0.19, p = 0.66 for interaction effect. Data in (A) were analyzed with a three-way ANOVA and Tukey's post-hoc test. Data in (D, E, F, G) were analyzed with a two-way ANOVA and Sidak's multiple comparisons post-hoc test. NS: not significant. Data in panels A and D–G are represented as mean ± SEM. See Table S3 for replicate details.



Figure S5. Lack of effects from AAV vector injections on behavioral readouts in nontransgenic control littermates of *Aldh1l1*-CreER^{T2} mice. Related to Figure 2. (A) Distance to reach the platform during training in male and female nontransgenic mice injected with saline or Cre-dependent mGluR3encoding AAV vector. Three-way ANOVA: F(9, 432) = 0.72, p = 0.69 for interaction effect. (B) Initial trajectory errors of male and female mice during training. Three-way ANOVA: F(19, 896) = 1.008, p = 0.45 for interaction effect. (C, D) Search strategies of male and female mice during training. (E) Quadrant durations in target and non-target quadrants (two-way ANOVA: F(1, 46) = 1.9, p = 0.18 for interaction effect); number of target and non-target platform crossings (two-way ANOVA: F(1, 45) = 0.03, p = 0.87 for interaction effect); swim speeds during probe trial. (**F**) Quadrant durations in target and non-target quadrants (two-way ANOVA: F(1, 46) = 0.04, p = 0.84 for interaction effect); number of target and non-target platform crossings (two-way ANOVA: F(1, 46) = 0.002, p = 0.97 for interaction effect); swim speeds during probe trial. (**G**) Distance traveled in the EPM by nontransgenic male and female mice with or without (Con) injection with the mGluR3-encoding AAV vector. Two-way ANOVA: F(1, 41) = 2.09, p = 0.16 for interaction effect. (**H**) Open-arm entries in the EPM by nontransgenic male and female mice with or without AAV vector injections. Two-way ANOVA: F(1, 41) = 0.18, p = 0.68 for interaction effect. Data were analyzed with a three-way ANOVA and Tukey's post-hoc test (A, B), two-way ANOVA and Sidak's multiple comparisons post-hoc test (G, H), or a Student's t-test (E, F): *p < 0.05, **p < 0.01, ***p < 0.001, NS: not significant. Data in all panels except C and D are represented as mean ± SEM. See Table S3 for replicate details.



Figure S6. Trajectories and spatial search strategies in early probe trials after knockdown or enhancement of astrocytic mGluR3 levels. Related to Figure 3. Initial trajectory errors (A, C) and search strategies (B, D) in probe trials performed one day after training in male and female mice with either reduced (A, B) or enhanced (C, D) mGluR3 in hippocampal astrocytes. Data in (A, C) were analyzed with two-way ANOVA and Sidak's post-hoc tests. Data in (B, D) were analyzed with Fisher's exact test. NS: not significant. Data in panels A and C are represented as mean ± SEM. See Table S3 for replicate details.



Figure S7. Characterization of astrocyte-targeted hM4Di- and rM3Ds-encoding AAV vectors and CNO pharmacokinetics. Related to Figure 4. (A) Astrocyte marker GFAP (green) and HA-hM4Di (red) coimmunolabeling in hippocampal sections from *Aldh111*-Cre male and female mice injected with AAV vector encoding HA-tagged hM4Di. DAPI (blue) labeled cell nuclei. Scale bars: 400 µm, 40 µm (insets). (B) HAhM4Di (red) immunolabeling in hippocampal sections from *Aldh111*-Cre mice. HA labeling was robust in the hippocampal formation, the region where the AAVs were injected, but minimal in brain regions outside of the hippocampus. Scale bar: 400 µm. (C) Immunofluorescence-based quantification of the percentage of GFAPpositive, Iba1-positive, and NeuN-positive cells that were also HA-positive in the hippocampus of *Aldh111*-Cre mice injected with AAV vectors encoding HA-hM4Di. One-way ANOVA: *F*(2, 15) = 23467, p < 0.0001 for cell marker effect. Dunnett's multiple comparisons test: ###p < 0.0001, GFAP+ vs. others. (D) Phospho-Akt levels following hM4Di stimulation with CNO (5 µM) in primary astrocytes. Data collected from one culture. (E) CNO does not cause significant increases in Akt or CREB phosphorylation levels in control astrocytes without AAV transduction. Data collected from two independent cultures. Two-way ANOVA and Sidak's multiple comparisons post-hoc test. NS: not significant. (F) Quantification of c-Fos immunolabeling in HA-positive cells in Aldh111-Cre mice injected with AAV vectors encoding HA-hM4Di and then injected i.p. with CNO (1 h; 5 mg/kg) or saline (Con). (G) Pharmacokinetic analyses of clozapine and CNO in the hippocampal formation following peripheral CNO injection (i.p.) in adult mice at indicated doses. In concurrence with previous findings, CNO was not detectable in the brain, likely due to rapid conversion to clozapine [S1, S2]. (H) Simplified diagram of several previously characterized signaling cascades engaged by Gi/o-coupled and Gscoupled GPCRs and their mutual regulation. (I) Co-immunolabeling for GFAP (green) and HA-rM3Ds (red) in mouse hippocampal sections from Aldh111-Cre female and male mice injected with AAV vector encoding HAtagged rM3Ds. Scale bar: 400 µm, 40 µm (insets). (J) HA-rM3Ds (red) immunolabeling in hippocampal sections from Aldh111-Cre mice. HA labeling was robust in the hippocampal formation, the region where the AAVs were injected, but minimal in brain regions outside of the hippocampus. Scale bar: 400 µm. (K) Immunofluorescence-based quantification of the percentage of GFAP-positive, Iba1-positive, and NeuNpositive cells that were also HA-positive in the hippocampus of Aldh111-Cre mice injected with AAV vectors encoding HA-rM3Ds. One-way ANOVA: F(2, 21) = 32370, p < 0.0001 for cell marker effect. Dunnett's multiple comparisons test: ###p < 0.0001, GFAP+ vs. others. (L) Levels of phosphorylated CREB following rM3Ds stimulation with CNO (5 µM) in cultured astrocytes. Data collected from one culture. Student's t-tests: ***p< 0.001, **p < 0.01, *p < 0.05. (M) Quantification of c-Fos immunolabeling in HA-positive cells in Aldh111-Cre mice injected with AAV vectors encoding HA-rM3Ds and then injected i.p. with CNO (1 h; 5 mg/kg) or saline (Con). Data are represented as mean ± SEM, except violin plots in F and M show medians. See Table S3 for replicate details.



Figure S8. Further behavioral characterization of DREADD-expressing mice. Related to Figure 4. (**A**) Probe trials performed one day after training of hM4Di-expressing mice. Target quadrant durations compared to other analogous locations. Target preferences were compared using Student's t-tests. Student's t-test: ***p < 0.001. (**B**, **C**) Swim speeds in mice expressing hM4Di in hippocampal astrocytes during day one (B) and day three (C) probe trials. Two-way ANOVA: F(1, 62) = 0.01, p = 0.91 for interaction effect (B), F(1, 61) = 0.007, p = 0.93 for interaction effect (C). (**D**) Probe trials performed one day after training of rM3Ds-expressing mice. Target quadrant durations compared to other analogous locations. Target preferences were compared using Student's t-tests. Student's t-test: **p < 0.01. NS: no significant preference for target. (**E**, **F**) Swim speeds in mice expressing rM3Ds in hippocampal astrocytes during day one (E) and day three (F) probe trials. Two-way ANOVA and Sidak's post-hoc test: F(1, 43) = 0.03, p = 0.87 for interaction effect (E), F(1, 43) = 0.095, p = 0.76 for interaction effect (F). NS: not significant. Data are represented as mean ± SEM. See Table S3 for replicate details.



Figure S9. Lack of CNO effects in nontransgenic control mice. Related to Figure 4. (A) GFAP (green) and DREADD (red) co-immunolabeling in AAV vector-injected nontransgenic mice used for testing off-target effects of CNO. DREADD labeling was absent in nontransgenic mice. DAPI (blue) labeled cell nuclei. Scale bars: 400 µm, 40 µm (insets). (B) Distance to reach the platform during training by vehicle (Con)- or CNO-injected non-transgenic mice. Three-way ANOVA: F(2, 150) = 0.09, p = 0.92 for interaction effect (time, sex, CNO). (C) Target quadrant durations and platform crossings compared to other analogous locations during a probe trial one day after training. Mice were injected with vehicle (Con) or CNO 1 h prior to the training trials and tested off-treatment during probe trials. (D) Target quadrant durations and platform crossings compared to other analogous locations during a probe trial three days after training. Data in (C, D) were analyzed with a two-way ANOVA and Student's t-tests to determine target preferences. *p < 0.05, **p < 0.01, ***p < 0.001. (E, F) Swim speeds during indicated probe trials in nontransgenic mice injected with vehicle or CNO during training. Two-way ANOVA and Sidak's post-hoc test: F(1, 75) = 1.67, p = 0.20 for interaction effect (E), F(1, 75) = 0.73, p = 0.40 for interaction effect (F). NS: not significant. Data are represented as mean ± SEM. See Table S3 for replicate details.

Supplemental Tables

Study	Receptor(s)	Sex	Sex effects reported?
Gs			
Orr et al., 2015	Rs1	Males and Females	No assessment of sex differences
Oe et al., 2020	rM3Ds	Males and Females	No assessment of sex differences
G _{i/o}			
Jones et al., 2018	hM4Di	Males	N/A
Nagai et al., 2019	hM4Di	Only males in behavior	No assessment of sex differences
Nam et al., 2019	hM4Di	Males	N/A
Kol et al., 2020	hM4Di	Males	N/A
Yu et al., 2020	hM4Di	Only males in behavior	No assessment of sex differences
Akter et al., 2023	hM4Di	Males	N/A
Reitman et al., 2023	hM4Di	Males and Females	No assessment of sex differences
G _{i/o} and G _q			
Yang et al., 2015	hM4Di, hM3Dq	Males	N/A
Kim et al., 2021	hM4Di, hM3Dq	Males	N/A
Vaidyanathan et al., 2021	hM4Di, hM3Dq	Males and Females	No assessment of sex differences
Van Den Herrewegen et al., 2021	hM4Di, hM3Dq	Males	N/A
Refaeli et al., 2024	hM4Di, hM3Dq	Males	N/A
Gq			
Adamsky et al., 2018	hM3Dq	Males	N/A
Nagai et al., 2021	hM3Dq	Males and Females	No assessment of sex differences
Delepine et al., 2023	hM3Dq	Males and Females	No assessment of sex differences

N/A: Sex differences could not be assessed, only one sex included.

Table S1. Summary of males and females assessed in previous *in vivo* studies using chemogeneticmanipulations and behavioral measurements. Related to Figures 1–4. Citations in the table are specifiedin the Supplemental reference list [S3–S19].

Fig #	Panel	Condition	Sex		
			Male	Female	
		GFAP	15	12	
	C (CA1, DG)	lba1	17	15	
		NeuN	6	6	
		Con	6	6	
	D (CA, DO)	sgRNA	5	5	
1	E (CA)	Con	7	11	
		sgRNA	8	7	
		Con	8	9	
	L (DG)	sgRNA	5	10	
		Con	12	12	
	6-1	sgRNA	10	11	
2	С	GFAP, NeuN	9	9	
	D (CA, DG)	Con	12	11	
		mGluR3	13	15	
	E	Con	12	10	
	L	mGluR3	13	14	
	E-I	Con	16	12	
	1-1	mGluR3	15	15	
3	ACE	Con	12	12	
	Λ, Ο, Ε	mGluR3	10	11	
		Con	16	12	
	D, D, I	mGluR3	15	14	
	С	NA	10	17	
4	DE	Con	19	15	
	D-r	CNO	19	15	
	I	NA	8	8	
		Con	12	13	
	J-L	CNO	12	13	

Table S2. Numbers of mice used in each experiment shown in the main figures. Related to Figures 1–4.

Image: second	Supplemental Fig #	Panel	anel Condition		Sex	
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$\begin{array}{c c c c c c c c c c c c c c c c c c c $	S4	F-G	Con	15	13	
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$			mGluR3	11	15	
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S9 ^{В-Н} CNO 21 19	00	D C	Con	21	19	
	59	B-F	CNO	21	19	

Table S3. Numbers of mice used in each experiment shown in the supplemental figures. Related to Figures S1–S9.

Gene	Forward primer	Reverse primer
Grm1	GCAGCGAGCCTTGCTTAAA	AGATCCAGCAGCAGCTCAC
Grm2	GGACTTCGTGCTCAATGTCA	CCATCTCCAAAGCGGTCAAA
Grm3	ACCTCAACAGGTTCAGTGTCA	TTGCACACTGTCGGGACATA
Grm4	TCAAGAAGGGAAGCCACATCAA	ACCTTCCCCTCCTGTTCGTA
Grm5	TGCAGTGAACCGTGTGAGAA	AAGGTGTGCAGGTCCAACAA
Grm7	AAGGAGCCATCACCATCCAA	TCAAGTGTCCGGGATGTGAA
Grm8	CCACTGGACCAATCAACTTCAC	GGGTGCGTGTGCTCTCTATTA
Actb	CCCTAAGGCCAACCGTGAAA	AGCCTGGATGGCTACGTACA
Gapdh	CAAGGTCATCCCAGAGCTGAA	CAGATCCACGACGGACACA
Gusb	AGTATGGAGCAGACGCAATCC	ACAGCCTTCTGGTACTCCTCA
Tbp	CCTTGTACCCTTCACCAATGAC	ACAGCCAAGATTCACGGTAGA

 Table S4. Primer sequences used for RT-qPCR. Related to STAR Methods.

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