# Supporting Information

# *NMR and Mössbauer Studies Reveal a Temperature-Dependent Switch from S = 1 to S = 2 in a Nonheme Oxoiron(IV) Complex with Faster C–H Bond Cleavage Rates*

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# **1. Instrumentation**

NMR (nuclear magnetic resonance) spectra for iron(II) complexes as well as iron(IV) complexes were recorded on a Bruker 400 or 500 MHz spectrometer at temperatures stated. NMR spectra to characterize the ligands were obtained at 298 K. 8" Wilmad-LabGlass 528-PP-7-5 NMR tubes were used to collect NMR spectra for iron(IV) and iron(II) complexes. For all the variable temperature spectra, the lower temperatures were calibrated using a sealed NMR tube containing a solution of tetramethylsilane (TMS) and methanol as a standard.

Elemental analyses were carried out by Atlantic Microlab (Norcross, GA).

UV-vis spectra and reactivity studies were recorded on a HP8453A diode array spectrometer equipped with a cryostat from Unisoku Scientific Instruments (Osaka, Japan).

Mössbauer spectra were recorded with two spectrometers, using Janis Research (Wilmington, MA) SuperVaritemp dewars that allow studies in applied magnetic fields of up to 7.5 T in the temperature range from 1.5 to 200 K. A LakeShore Model 331A temperature controller was used to control the temperature in experiments. Mössbauer spectral simulations were performed using the WMOSS software package (SEE Co, Edina. Minnesota). Isomer shifts are quoted relative to α-Fe metal at 298 K. The spectra were plotted by SpinCount developed by Prof. Michael Hendrich at Carnegie Mellon University.<sup>1</sup>

# **2. Materials**

All materials were bought from Sigma Aldrich, Fischer Scientific or TCI Chemicals, unless otherwise noted. The substrates cyclohexane, cyclooctane, toluene, ethylbenzene, cyclooctane and cumene used in kinetic experiments were passed through a plug of silica before use. All experiments to determine rate constants were carried out under nitrogen. Commercially available solvents like anhydrous acetonitrile, 2,2,2-trifluoroethanol (TFE) and deuterated dichloromethane, acetone and acetonitrile were used without further purification, unless otherwise noted. All oxygen- and moisture-sensitive compounds were synthesized in a nitrogen-filled glovebox.



**Scheme S1.** Examples of *S* = 2 oxoiron(IV) complexes that are supported by trigonal arrangements of ligands

# **3. Synthesis and Characterization of Ligands, Oxidants, and Iron(II) Complexes**

### A) General synthetic procedures for ligands reported in this study



#### **1. Ligand names and structures**

**Scheme S2.** Names of TPA- and N4Py-based ligands examined in this study

Scheme S3 shows the BQPA-based ligands shown in Scheme S2 surrounding iron(IV) complexes and their corresponding names, in Scheme S2.



**Scheme S3.** Pyridine-ring-substituted variants of complex **2**

### **2. Synthesis of ligand BQP(5-Me)A**

2-(Aminomethyl)-5-methylpyridine<sup>6-8</sup> (0.61g, 5 mmol) was added to a suspension of 2chloromethylquinoline hydrochloride (2.14 g, 10 mmol) in anhydrous CH3CN (30 ml). The mixture was treated with anhydrous  $K_2CO_3$  (2.8 g, 20.2 mmol) and refluxed at 45 °C. After 3 days, the reaction mixture was cooled down to room temperature and filtered to remove any unreacted K<sub>2</sub>CO<sub>3</sub> and KCI salt formed and. The solvent was then evaporated under vacuum to yield yellow solids, which were recrystallized from diethyl ether (yield 83 %).

<sup>1</sup>H-NMR (400 MHz, CD2Cl2): 2.32 (3H, 5*-*C*H3*), 3.90 (2H, py*-*C*H2*), 4.10 (4H, Q*-*C*H2*), 7.50-7.57 (m, 4H), 7.71 (t, 2H), 7.78 (d, 2H), 7.84 (d, 2H), 8.00 (d, 2H), 8.18 (d, 2H), 8.38 (d, 1H). <sup>13</sup>C NMR (100 MHz,  $CD_2Cl_2$ ): 159.5 ppm, 159.3 ppm, 148.99 ppm, 146.19 ppm, 136.21 ppm, 136.02 ppm, 135.52 ppm, 131.44 ppm, 130.10 ppm, 128.64 ppm, 127.36 ppm, 126.35 ppm, 123.10 ppm, 121.94 ppm, 121.11 ppm, 60.93 ppm, 60.37 ppm, 21.22 ppm. ESI-MS (in MeOH): obsd (calcd) for  $[BQP^{3-Me}A + H]^+$ : m/z 405.3 (405.21).

### **3. Synthesis of ligand BQP(3-Me)A**

2-(Aminomethyl)-3-methylpyridine<sup>6-8</sup> (0.61g, 5 mmol) was added to a suspension of 2chloromethylquinoline hydrochloride (2.14 g, 10 mmol) in anhydrous CH3CN (30 ml). The mixture was treated with anhydrous  $K_2CO_3$  (2.8 g, 20.2 mmol) and refluxed at 45 °C. After 3 days, the reaction was cooled down to room temperature, and filtered to remove KCl salt and unreacted  $K_2CO_3$ . The solvent was then removed to yield a yellow solid that was recrystallized from diethyl ether (yield 87 %).

<sup>1</sup>H-NMR (400 MHz, CD2Cl2): 2.22 (3H, 3*-*C*H3*), 3.95 (2H, py*-*C*H2*), 4.00 (4H, Q*-*C*H2*), 7.08 (t, 1H), 7.41 (d, 1H), 7.53 (t, 2H), 7.61 (d, 2H), 7.70 (t, 2H), 7.82 (d, 2H), 8.04 (d, 2H), 8.12 (d, 2H), 8.37 (d, 1H). ESI-MS (in MeOH): obsd (calcd) for [BQP<sup>5-Me</sup>A +H]<sup>+</sup>: m/z 405.3 (405.21).

## **4. Synthesis of BQP\*A**

2-(Aminomethyl)-3,5-dimethyl,4-methoxypyridine<sup>6-8</sup> (1.66 g, 10 mmol) was added to a suspension of 2-chloromethylquinoline hydrochloride (4.28 g, 20 mmol) in anhydrous CH<sub>3</sub>CN (30 ml). The mixture was treated with anhydrous  $K_2CO_3$  (2.8 g, 20.2 mmol) and refluxed at 45  $\degree$ C. After 3 days, the reaction was cooled down to room temperature, and then filtered to remove KCI salt and unreacted  $K_2CO_3$ . The solvent was then removed to yield yellow solids, which was then recrystallized from diethyl ether (yield 90 %).

<sup>1</sup>H-NMR (400 MHz, CD2Cl2): 2.10 (3H, 3*-*C*H3*), 2.16 (3H, 5*-*C*H3*), 3.61 (3H, 4-MeO), 3.98 (2H, Py*-*C*H2*), 4.06 (4H, Q*-*C*H2*), 7.50 (t, 2H), 7.57 (d, 2H), 7.68 (t, 2H), 7.71 (d, 2H), 8.00 (m, 4H), 8.17 (d, 1H). <sup>13</sup>C NMR (100 MHz, CD<sub>2</sub>Cl<sub>2</sub>): 163.95 ppm, 160.3 ppm, 156.55 ppm, 148.34 ppm, 147.50 ppm, 135.81 ppm, 129.14 ppm, 129.03 ppm, 127.46 ppm, 127.27 ppm, 126.15 ppm, 126.01 ppm, 125.21 ppm, 121.63 ppm, 61.93 ppm, 59.60 ppm, 12.90 ppm, 10.75 ppm. ESI-MS (in MeOH): obsd (calcd) for [BQP\*A+H]+: m/z 449.3 (449.23).

## B) General synthetic procedures for iron(II) complexes

## **1) Synthesis of iron(II) complex [FeII(BQP(5-Me)A)](OTf)<sup>2</sup>**

To a solution of Fe(OTf)2•2CH3CN (97.0 mg, 0.220 mmol) in 1 mL anhydrous CH3CN inside an inert glovebox, 1 equiv. BQP5MeA (89.0 mg, 0.220 mmol) in 3 mL CH3CN was added, and the mixture was stirred for three hours to give a yellowish green solution. This solution was then filtered and poured into a diethyl ether bath to produce a yellowish green precipitate, which was filtered again and washed with diethyl ether, to give a yellowcolored solid. This solid was re-dissolved in acetonitrile and crystallized by vapor diffusion using diethyl ether to give yellow crystals. (125 mg, 75 % yield). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN, 25 °C): δ (ppm) 117.8 (1H; α-H Py), 100.9 (2H; CH<sub>2</sub> Py), 57.7 (4H; CH<sub>2</sub> quin), 53.5 (2H; 7-H quin), 48.3 (1H;  $\beta'$ -H py), 22.3 (2H; 5-H Qn), 11.43 (2H; 3-H Qn), 2.7 (2H; 6-H Qn), 0.6 (3H; 5-Me Py), -3.30 (2H; 4-H quin), -6.70 (1H;  $\gamma$ -H Py), -58.33 (2H; 8-H Qn). Anal. calcd. for  $C_{30}H_{28.5}F_6FeN_{4.5}O_{7.5}S_2$  corresponding to the complex  $Fe^{11}(BQP(5-$ Me)A)(OTf)2•1.5H2O•0.5CH3CN: C, 44.70; H, 3.56; N, 7.82. Found: C, 44.66; H, 3.53; N, 7.33. ESI-MS (in MeCN): obsd (calcd) for [Fe<sup>ll</sup>(BQP(5-Me)A)(OTf)]<sup>+</sup>: m/z 609.01 (609.09).

### **2) Synthesis of iron(II) complex [FeII(BQP(3-Me)A)(OTf)2]**

To a solution of  $Fe(OTf)<sub>2</sub>•2CH<sub>3</sub>CN$  (75.0 mg, 0.170 mmol) in 1 mL anhydrous  $CH<sub>3</sub>CN$ inside an inert glovebox was added 1 equiv. BQP3MeA (68.8 mg, 0.170 mmol) in 2 mL CH3CN and the mixture was stirred for three hours to give a yellowish green solution. This solution was then filtered and poured into a diethyl ether bath to produce a yellowish green precipitate, which was filtered again and washed with diethyl ether, to give a yellowcolored solid. This solid was re-dissolved in acetonitrile and crystallized by vapor diffusion using diethyl ether to afford yellow crystals. (100 mg, 78 % yield). <sup>1</sup>H NMR (400 MHz, CD3CN, 25 °C): δ (ppm) 109.3 (1H;  $\alpha$ -H py), 84.5 (2H; CH2 Py), 60, 65 (4H; CH2 quin),  $54.2$  (1H;  $\beta$ -H py),  $51.1$  (2H; 7-H quin), 22.6 (2H; 5-H quin), 7.6 (2H; 3-H quin), 3.1 (2H; 6-H quin), -1.7 (3H; 3Me Py) -4.77 (2H; 4-H quin), - 4.70 (1H; y-H py), -52.33 (2H; 8-H quin). Anal. calcd. for C<sub>29</sub>H<sub>26</sub>F<sub>6</sub>FeN<sub>4</sub>O<sub>7</sub>S<sub>2</sub> representing Fe<sup>ll</sup>(BQP(3-Me)A)(OTf)<sub>2</sub>·H<sub>2</sub>O: C, 44.86; H, 3.37; N, 7.22. Found: C, 44.66; H, 3.53; N, 7.33. ESI-MS (in MeCN): obsd (calcd) for [Fe<sup>ll</sup>(BQP<sup>3</sup>-MeA)(OTf)]<sup>+</sup>: m/z 609.30 (609.09).

## **3) Synthesis of iron(II) complex [FeII(BQP\*A)(OTf)2]**

To a solution of  $Fe(OTf)<sub>2</sub>•2CH<sub>3</sub>CN$  (195 mg, 0.442 mmol) in 2 mL anhydrous  $CH<sub>3</sub>CN$ inside an inert glovebox was added 1 equiv. BQP\*A (200 mg, 0.443 mmol) in 4 mL CH3CN, and the mixture was stirred for three hours to give rise to a yellow solution. This solution was then filtered and poured into a diethyl ether bath to produce a yellowish green precipitate, which was filtered again and washed with diethyl ether, to give a yellow-

colored solid. This solid was re-dissolved in acetonitrile and crystallized by vapor diffusion using diethyl ether to give yellow crystals. (302 mg, 85 % yield). <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>CN, 25 °C): δ (ppm) 111.5 (1H; α-H py), 92.3 (2H; CH2 Py), 61.1 (4H; CH2 quin), 51.7 (2H; 7-H quin), 22.2 (2H; 5 -H quin), 11.8 (2H; 3-H quin), 3.14 (2H; 6-H quin), 1.5 (3H; 4-MeO py), -0.7 (3H; 5-Me Py), -3.2 (3H; 3-Me Py), -51.3 (2H; 8-H quin) Anal. calcd. for  $C_{31}H_{31}F_6FeN_4O_{8.5}S_2$  representing  $Fe^{11}(BQP*A)(OTf)_{2}A.5H_2O$ : C, 44.88; H, 3.60; N, 6.75. Found: C, 48.30; H, 3.60; N, 6.66. ESI-MS (in MeCN): obsd (calcd) for [Fe<sup>ll</sup>(BQP<sup>\*</sup>A)(OTf)]<sup>+</sup>: m/z 609.01 (609.09).

### **4) Synthesis of previously reported iron(II) complexes**

Previously reported iron(II) complexes include those supported by their known ligands TPA<sup>9</sup>, QBPA<sup>10</sup>, BQPA<sup>10</sup>, TQA<sup>11</sup>, N2Py2Q<sup>12</sup>, and N4Py.<sup>13</sup> In their syntheses, the only change was that iron(II) triflate was used instead of iron(II) perchlorate, in order to mitigate the risks associated with the explosive nature of perchlorates.

Other <sup>57</sup>Fe<sup>II</sup>-labeled complexes were synthesized using the same synthetic procedures as above, except that  ${}^{57}Fe^{11}(CH_3CN)_2(OTf)_2$  was used on a 10-20 mg scale as the iron(II) triflate starting material. Details for the preparation of oxoiron(IV) solutions for Mössbauer spectroscopy are detailed in the Mössbauer section.

## C)  ${}^{1}$ H-NMR spectra of iron(II) complexes







Figure S2. <sup>1</sup>H-NMR spectra of iron(II) complexes of substituted BQPA ligands obtained at 298 K in acetonitrile-*d*<sup>3</sup> (residual CH3CN peaks from the solvent denoted by #). Insets show selected magnified sections of the spectra. See Figure S1 for quin/py H labels

#### **Notes on assignments of <sup>1</sup>H NMR signals in solutions of iron(II) complexes:**

The β, β' and γ signals are assigned based on comparisons with the NMR spectra of [Fe<sup>ll</sup>(6MeTPA)(L)<sub>n</sub>]<sup>2+</sup> and [Fe<sup>ll</sup>(6Me<sub>3</sub>TPA)(L)<sub>n</sub>]<sup>2+</sup> complexes.<sup>9</sup> These signals can also be assigned based on changes observed upon substitution of these protons with methyl and methoxy groups. Quinoline H4 and pyridine γ-H protons give rise to similarly shifted signals, likely due to the similar paramagnetic effects experienced by these protons on their respective heterocycles. H5, H6 and H7 are the farthest from the paramagnetic center and thus have the sharpest signals, with H6 being the sharpest followed by H5 and then H7. This also follows from their expected relative proximity to the iron center. H8 is likely to be very broad and tentatively assigned to upfield signals from -50 to -60 ppm.



**Table S1.** <sup>1</sup>H NMR chemical shifts for iron(II) complexes at 298 K in CD<sub>3</sub>CN. FWHM given in parentheses. Italicized values are for methyl/methoxy protons.

In Table S1,  $\beta$ -H signals are broader compared to  $\beta$ -H signals, as indicated by their FWHMs in the parent iron(II) complex supported by BQPA. Similarly, the  $\beta$ -H signal of iron(II) complex supported by BQP(5-Me)A is broader than the  $\beta$ -H signal in iron(II) complex supported by BQP(3-Me)A. This is also true for *S* = 1 iron(IV) complexes, and would be expected to be true for *S* = 2 iron(IV) complexes, as FWHM has to do with proximity to the iron center, and  $\beta$ -H are generally closer to it than  $\beta$ -H (in Tables S5-6).

# **4. Synthesis & Characterization of Oxoiron(IV) Complexes**

## A) Synthesis and/or *in-situ* generation of oxoiron(IV) complexes

All oxoiron(IV) complexes supported by tripodal ligands in this study were generated at given temperatures by treatment of solutions of their iron(II) precursors of given concentration in appropriate solvents with 1-2 equivalents of oxidant ArIO. These solutions were used for reactivity studies or to obtain NMR, UV-visible or Mössbauer spectra. Details are given in the relevant section for sample preparation and each spectroscopic measurement. the sequential replacement of pyridines in  $[Fe^{IV}(O)(TPA)(MeCN)]^{2+}$  (0) with sterically bulkier quinolines to form the complexes  $[Fe^{IV}(O)(QBPA)(MeCN)]^{2+}$  (1),  $[Fe^{IV}(O)(BQPA)(MeCN)]^{2+}$  (2), and  $[Fe^{IV}(O)(TQA)(MeCN)]^{2+}$  (3). Of these complexes,  $^{14-17}$  only 2 has not been previously reported. The oxoiron(IV) complexes supported by pentadentate ligands were generated as previously reported.<sup>12, 18</sup>

# B) UV-vis spectroscopy of oxoiron(IV) complexes

All samples were generated in 1-cm quartz cells at appropriate temperatures using 1 mL of 1-mM iron(II) solutions in acetonitrile, to which 2 equivalents of ArIO dissolved in either 2,2,2-trifluoroethanol or dichloromethane-*d*<sup>2</sup> was added. Their generation was followed using UV-vis spectroscopy at given temperatures using the cryostat-equipped UV-visible spectrophotometer, to stabilize and lower the temperature using liquid from a dewar.



**Figure S3.** UV-vis spectra of **2**, **2a**, **2b** and **2\***, in acetonitrile at 233 K, obtained by adding 2 equiv ArIO in CD<sub>2</sub>Cl<sub>2</sub> to 1-mM iron(II) solutions. Instrument artifact at ≈ 660 nm.





# C) Reactivity and kinetic studies of oxoiron(IV) complexes

All experiments were conducted with 1-1.5 mL solutions of 1 mM iron(II) complexes in MeCN solutions at given temperatures for HAT (hydrogen-atom-transfer) substrates. The oxoiron(IV) complexes were generated using 1-2 eq. ArIO in 2,2,2-trifluoroethanol or DCM-*d*2. The pseudo-first order decay curves from absorption-time profiles were obtained by monitoring the decrease in absorbance for the oxoiron(IV) complexes with time, from their absorbance spectra, when a known concentration of substrate is introduced. These profiles at various concentrations of substrates were fit with a first order exponential decay equation. The rates obtained ( $k_{obs}$ ) were then plotted against different concentrations of substrates, and their slopes represent 2nd order rate constants (*k*2).



**Figure S4.**  $k_2$  plots for the reactions of various HAT substrates with  $[Fe<sup>IV</sup>(O)(TPA)(L)]<sup>2+</sup>$ (**0**). The slopes of the fitted red lines represent the second-order rate constants (*k*2) at 273 K. The oxoiron(IV) complex was generated by adding 1.2 eq s-ArIO in TFE to a MeCN solution of the iron(II) complex.



**Figure S5.** *k*<sup>2</sup> plots for the reactions of different HAT substrates with  $[Fe^{IV}(O)(QBPA)(L)]^{2+}$  (1) in MeCN. The slopes of the fitted red lines represent the secondorder rate constants (*k*2) at 273 K. The oxoiron(IV) complex was formed with 1.2 eq s-ArIO.



**Figure S6.** *k*<sup>2</sup> plots for the reactions of different HAT substrates with  $[Fe^{IV}(O)(BQPA)(L)]^{2+}(2)$ . The slopes of the fitted red lines represent the second-order rate constants (*k*2) at 233 K. The oxoiron(IV) complex was generated using 2 eq s-ArIO followed by addition of substrate, except for cyclooctane and cyclohexane where the substrate was dissolved first before adding the oxidant.

**Table S3.** Summary of 2nd-order rate constants (*k*2) for oxoiron(IV) complexes, as derived from the plots in Figures S4 through S6. HAT rates were measured under  $N_2$  at given temperatures.



Table S4. Summary of 2nd-order rate constants ( $k_2'$ , normalized per equivalent C–H bond present on substrate) for TPA-based complexes, as derived from the plots in Figures S4 through S6. HAT rates were measured under N<sup>2</sup> at given temperatures.





**Figure S7.**  $k_2$  plots for the reactions of ethylbenzene with  $[Fe<sup>IV</sup>(O)(TPA)(L)]<sup>2+</sup>$  and  $[Fe^{IV}(O)(BQPA)(L)]^{2+}$ . The slopes of the fitted red lines represent the second-order rate constants (*k*2) at 233 K. The oxoiron(IV) complex was generated using 2 eq s-ArIO followed by addition of substrate.

## D) NMR spectroscopy of oxoiron(IV) complexes

### **1) Generation of the ferryl complexes for NMR spectroscopic analysis**

4-mM stock solutions of starting iron(II) precursors were made within an air-free nitrogenfilled glovebox in acetonitrile-*d*<sub>3</sub> or acetone-*d*<sub>6</sub>. Samples of the oxoiron(IV) complexes for NMR analysis were generated by transferring 0.5 mL of a precursor solution into an NMR tube (in glove box) and cooling it to the desired temperature in the NMR instrument for analysis; at this point a <sup>1</sup>H NMR spectrum of the iron(II) precursor was obtained, after locking to the residual proton signal in the deuterated solvent, tuning to the intended nucleus, and then shimming to obtain a uniform magnetic field around the sample. The following three steps were then performed in quick succession: 1) the NMR tube was ejected from the instrument, 2) 2 equivalents of ArIO in either 2,2,2-trifluoroethanol (TFE) or DCM-*d*<sup>2</sup> were quickly injected to generate the oxoiron(IV) intermediate; the tube was recapped with either a septum or its plastic cap (to minimize decay of the species) and 3) the tube was immediately inserted back into the instrument for data collection. Because the instrument had been locked to the solvent as well as tuned to the nucleus of interest, only quick shimming was performed again before collection of NMR data for the oxoiron(IV) species. To obtain the spectrum of the decayed species, the complex was warmed to room temperature by ejecting the NMR tube from the instrument, and after the species had decayed, it was inserted back into the instrument and NMR data was obtained again at a lowered temperature.

### **2) General parameters for data collection and processing**

The following parameters were used to acquire the <sup>1</sup>H NMR data for paramagnetic compounds: acquisition time =  $0.064$  s; relaxation delay =  $0.03$  s; sweep width = 400 ppm offset (centered) at 6 ppm; line broadening factor = 10-30 Hz. Chemical shifts (ppm) were referenced to residual protic solvent peaks. The NMR spectra obtained were processed using the NMR processing software MestReNova 12.0 or Bruker's Top-Spin 3.5pl7.

#### **3) Consequences of having different structural configurations of pyridines around the Fe=O unit.**

For the 6-coordinate complex **0**, there are two possible configurations with respect to the oxo unit, either it is trans to the tertiary amine (more commonly observed) or trans to one of the three pyridines and cis to the other two pyridines with planes oriented perpendicular to the  $Fe<sup>N</sup>=O$  unit. The DFT-predicted <sup>1</sup>H-NMR shifts for the pyridine-H signals of the latter isomer span a range of only 20 to -10 ppm, a prediction confirmed in the <sup>1</sup>H-NMR spectrum of  $[Fe<sup>IV</sup>(O)(BnTPEN)<sup>2+</sup> (BnTPEN = N-benzyl-N,N,N-tris(2-pyridylmethyl)-1,2- $W-N$$ diaminoethane) in which the pyridine with its plane perpendicular to the Fe=O unit exhibits resonances at 8 ppm for the γ-H and at -0.2 and -2.0 ppm for the β-H's. However, the <sup>1</sup>H-NMR spectrum of complex **0** obtained at 233 K does not exhibit such a small range of shifts predicted for this hypothetical isomer, thus ruling out this possible isomer. Instead, a set of peaks similar to that seen for  $[Fe<sup>IV</sup>(O)(N4Py)]<sup>2+</sup>$  is observed, indicating that there is no pyridine in **0** that is bound with its plane perpendicular to the Fe=O unit.



**Figure S8.**  <sup>1</sup>H-NMR spectra of various complexes that establish the solution state structure of complex **0** (top panel) at 233 K in CD<sub>3</sub>CN. The red highlighted peaks belong to the red pyridines. The NMR shift patterns for any pyridine in **0** do not match the NMR patterns observed for the pyridine perpendicular to the oxoiron(IV) unit in [FeIV(O)(BnTPEN)]2+ (bottom panel), thus ruling out any solution-state configuration for **0** where a pyridine is perpendicular to the oxoiron(IV) unit. However the peaks observed do match shift patterns found in  $[Fe<sup>IV</sup>(O)(N4Py)]<sup>2+</sup>$  (middle panel) where all the pyridines are aligned roughly parallel to the oxoiron(IV) unit.

**4) NMR spectra for [FeIV(O)(N2Py2Q)] 2+**



**Figure S9.** <sup>1</sup>H-NMR spectra of  $[Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>$  in acetonitrile- $d_3$  at 298 K (panel A) and 233 K (panel B). Quinoline peaks are highlighted in red, and denoted. <sup>1</sup>H-NMR spectra of  $[Fe^{IV}(O)(N4Py)]^{2+}$  in acetonitrile- $d_3$  at 298 K are shown in panel C and at 233 K in panel D for comparison. See Table S5 for signal assignments in CD<sub>3</sub>CN at 298 K.

Table S5. <sup>1</sup>H NMR signals of [Fe<sup>lV</sup>(O)(N2Py2Q)]<sup>2+</sup> obtained in acetonitrile-*d*<sub>3</sub> at 298 K.





**Figure S10.** <sup>1</sup>H COSY NMR spectrum of  $[Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>$  in acetonitrile- $d_3$  at 233 K from 0 to 35 ppm. Only the peaks at 19 and 20.8 ppm are found to have relaxation properties at this temperature that allow cross-talk with each other. The lower temperature was chosen to minimize the self-decay of  $[Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>$  over the course of data collection time (over 12 h) on a 10-mM sample.



**5) Shift patterns of** *S* **= 1 oxoiron(IV) complexes containing heterocycles**

**Figure S11.** Stacked <sup>1</sup>H NMR spectra of complexes (top)  $[Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>$ , (middle)  $[Fe^{IV}(O)(N4Py)]^{2+}$  and (bottom)  $[Fe^{IV}(O)(N2Py2B)]^{2+}$  in CD<sub>3</sub>CN at 298 K<sup>21</sup> from –75 to 75 ppm, along with assignments. The dotted lines cross through the common γ proton signal in each of the spectra.

In these oxoiron(IV) complexes, the quinoline H3 and H4 protons appear at similar chemical shifts to the pyridine γ and β' protons, respectively, indicating that these sets of protons experience similar paramagnetic effects from the  $S = 1$  Fe<sup>IV</sup> center as pyridines. This is not surprising given that a quinoline can be thought of as a pyridine having a fused phenyl ring.

H5/H6 in  $[Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>$  and H4/H5 in  $[Fe<sup>IV</sup>(O)(N2Py2B)]<sup>2+21</sup>$  belong to the fused phenyl ring on the respective heterocycles and have the sharpest resonances, showing that they are likely to have relaxation properties that make them amenable to COSY spectroscopy, allowing cross-peaks to be observed in their COSY spectra obtained at lower temperature. Because H7 in [Fe<sup>IV</sup>(O)(N2Py2**B**)]<sup>2+</sup> and H8 in [Fe<sup>IV</sup>(O)(N2Py2**Q**)]<sup>2+</sup> are too close to their respective iron centers, their signals are too broad to be observed. The broadest yet observable fused heterocyclic proton peaks in the NMR spectra of these ferryl complexes belong to H7 in  $[Fe<sup>N</sup>(O)(N2Py2Q)]<sup>2+</sup>$  and H6 in  $[Fe<sup>N</sup>(O)(N2Py2B)]<sup>2+</sup>$ respectively. However, H7 in [FeIV(O)(N2Py2**Q**)]2+ is more downfield-shifted than H6 in  $[Fe^{IV}(O)(N2Py2B)]^{2+}$ . H6 in  $[Fe^{IV}(O)(N2Py2B)]^{2+}$  is also 0.5 Å farther from the iron(IV) center than H7 in  $[Fe<sup>IV</sup>(O)(N2Py2**Q**)]<sup>2+</sup>$ , and thus its peak is less paramagnetically shifted and sharper compared with H7 in [Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>. When pyridines, benzimidazoles and quinolines are viewed together, the assignments of quinoline protons in the  $S = 1$ iron(IV) center are greatly facilitated by the parallels observed among pyridines, benzimidazoles and quinolines.



Table S6.<sup>1</sup>H NMR spectroscopic properties for complexes containing benzimidazoles or quinolines in the N4Py framework, obtained at 298 K.<sup>21</sup>

**6) <sup>1</sup>H-NMR spectra of oxoiron(IV) complexes and their corresponding decay products**



**Figure S12.** Stacked <sup>1</sup>H-NMR spectra of substituted variants of complex **2**, identifying the peaks belonging to methyl groups (insets), and other peaks between 0 to 10 ppm. Residual CH2Cl<sup>2</sup> arises from CD2Cl<sup>2</sup> used to dissolve the oxidant. The # indicates the *tert*butyl group signal in the oxidant as well residual solvent signals (including CH<sub>3</sub>CN and H2O). Dashed lines cross a common peak through the spectra.



**Figure S13.** Evidence of  $C_1$  symmetry in S = 2 complex 3 is portrayed by the DFT optimized structures of **3** (top two figures, shown as ball and stick diagrams or its combination with wireframe views of some atoms for clarity), with mean planes (green and red) formed by quinolines not aligned in the DFT structure.<sup>11</sup> It is also obvious from the iron(II) crystal structure (bottom) with thermal ellipsoids of non-hydrogen atoms at 30% (triflate atoms shown as yellow balls). The planes of the two trans quinolines are not aligned across the N–Fe–N bonds in both iron(II) and DFT-optimized iron(II) structures. N atoms  $=$  blue, iron  $=$  orange, carbon  $=$  gray, triflate ion atoms (carbon, oxygen, sulfur, fluorine) = yellow. Hydrogen atoms were hidden for clarity in all cases.



**Figure S14.**  <sup>1</sup>H-NMR spectra of complex **2**, prepared by adding 2 equiv. ArIO in a 2,2,2 trifluoroethanol solution to a 4-mM solution of its iron(II) precursor in acetonitrile-*d*<sup>3</sup> at 233 K (top) and after its thermal decay in acetonitrile-*d*<sup>3</sup> at 233 K (bottom).



**Figure S15.**  <sup>1</sup>H-NMR spectra of complex **2a** prepared by adding 2 equiv. ArIO to a 4-mM solution of its iron(II) precursor in acetonitrile-*d*<sup>3</sup> at 233 K (top) and after its thermal decay in acetonitrile-*d*<sup>3</sup> at 233 K (bottom).



Figure S16. <sup>1</sup>H-NMR spectra of complex 2b, prepared by adding 2 equiv ArIO to a 4-mM solution of its iron(II) precursor in acetonitrile-*d*<sup>3</sup> at 233 K (top) and after its thermal decay at 233 K (bottom).



**Figure S17.**  <sup>1</sup>H-NMR spectra of complex **2\***, prepared by adding 2 equiv ArIO to a 4-mM solution of its iron(II) precursor in acetonitrile- $d_3$  at 233 K (top) and after its thermal decay (bottom).



**Figure S18.**  <sup>1</sup>H-NMR spectra of **2** and its variants from -50 to 250 ppm, showing that there are no observable peaks in the region past 120 ppm.

	$[FeIV(O)(N2Py2Q)]2+$				$\overline{2}$				3		
H	exp 6 233 K	calc δ $S = 1$	calc δ $S = 2$	н	exp 6 233 K	calc δ $S = 2$	calc δ $S = 1$	н	exp 6 233 K	calc δ $S = 2$	calc δ $S = 1$
		pyridine				pyridine			pyridine		
α	broad	$-43.9$ $-43.9$	141.9 141.9	α	not observed <sup>[b]</sup>	71.9	$-51.7$	No pyridine-H signals			
β	31	35.8	155.6	β	112	170.3	48.7		quinoline		
		35.8	155.6						30 <sup>[a]</sup>	48.0	$-21.2$
ß'	$-18$	$-18.9$	72.7	β'	not	48.1	$-26.1$	H <sub>3</sub>	93 <sup>[a]</sup>	93.3	$-20.7$
		$-18.9$	72.7		observed <sup>[b]</sup>				$54^{[a]}$	53.0	$-17.5$
γ	7.4	10.4	$-7.1$	٧	$-26$	$-31.8$	7.4		$-5.3^{[a]}$	1.7	11.9
		$-7.1$ 10.4					H4	$7.2^{[a]}$	8.1	13.9	
		quinoline				quinoline			$-5.3^{[a]}$	$-29.3$	11.4
H3	$-19$	$-20.8$	72.1	H3	76, 77 or	103.5	$-19.7$		$84^{[a]}$	72.4	16.7
		$-20.8$	72.1		83 <sup>[a]</sup>	103.5	$-19.7$	H <sub>5</sub>	86 <sup>[a]</sup>	90.8	24.4
H <sub>4</sub>	15	15.6	1.9	H4	27, 25 or	13.6	16.2		$20^{[a]}$	27.1	6.7
		15.6	1.9		23 <sup>[a]</sup>	13.6	16.2		$23^{[a]}$	33.9	34.1
<b>H5</b>	21	26.0	80.2	H <sub>5</sub>	76.77 or	89.7	21.0	H <sub>6</sub>	20 <sup>[a]</sup>	32.9	25.5
		26.0	80.2		$83^{[a]}$	89.7	21.0		$18^{[a]}$	26.0	18.7
H <sub>6</sub>	19	27.3	27.8	H <sub>6</sub>	27, 25 or	28.7	30.1	H7	$27^{[a]}$	32.7	31.6
		27.3	27.8		23 <sup>[a]</sup>	28.7	30.1		$26^{[a]}$	25.9	25.5
<b>H7</b>	23	30.3	32.7	<b>H7</b>	27, 25 or	33.7	28.0		$-5.3^{[a]}$	1.6	4.6
		30.3	32.7		23 <sup>[a]</sup>	33.7	28.0		not	230.6	113.8
H <sub>8</sub>	broad	72.8	105.5	H <sub>8</sub>	not	132.4	111.5	H <sub>8</sub>		270.7	166.3
		72.8	105.5		observed <sup>[b]</sup>	132.4	111.5		observed <sup>[b]</sup>	251.6	95.6

Table S7. Observed and predicted chemical shifts (in ppm) for [Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>, 2 and **3** at 233 K

[a] Peaks assigned based on DFT calculations. [b] Signals not observed because they are likely to be too broad due to their proximity to the iron center

### E) Mössbauer spectroscopy of oxoiron(IV) complexes

#### **1) Generation of the ferryl complexes for NMR spectroscopic analysis**

1-mM acetonitrile solutions of labeled (<sup>57</sup>Fe) iron(II) complexes were precooled in a quartz cell at 233 K. To this stirred solution, 2 equivalents of the oxidant s-PhIO in 2,2,2 trifluoroethanol were added. The formation of the oxoiron(IV) complex was instantaneous in each case. Each sample was then transferred using cryo-cooled 9" glass pipettes into a precooled Mössbauer cup suspended on top of liquid nitrogen in a dewar.

Mössbauer spectra of a sample containing complex **1** and a sample containing the decay product of **1** were measured at 4.2 K with zero applied field (Figure S19). The decay product exhibited a quadrupole doublet with isomer shift (δ) of 0.47 mm/s and quadrupole splitting (ΔEQ) of 1.59 mm/s, which are typical parameters from a diferric species. Such a diferric species was also observed in the sample containing **1** with almost identical Mössbauer parameters (δ = 0.46 mm/s,  $\Delta E_Q$  = 1.65 mm/s), which represents ~ 30% of total iron in the sample. An additional quadrupole doublet was also observed in the sample containing **1**, which exhibited Mössbauer parameters  $\delta$  = 0.05 mm/s,  $\Delta E$ <sub>Q</sub> = 0.70 mm/s, which should be assigned to **1** and represents ~ 60% of total iron. Judging by the isomer shift, **1** should still have an *S* = 1 ground spin state, although its isomer shift is 0.04 mm/s higher than that of complex **0**. The remaining spectral features belong to mononuclear ferric species as indicated by the black arrows in Figure S19.

Mössbauer spectra of a sample containing complex **2** were also measured. At 4.2 K, the spectrum exhibited two quadrupole doublets close to 1:1 ratio (Figure S19). The spectral simulation by using two quadrupole doublets led to two different solutions with equal quality of fit judging by the  $\chi^2$  value of the least square fitting. In solution I, the two quadrupole doublets have the following parameters:  $\delta_1(I) = 0.03$  mm/s,  $\Delta E_{Q1}(I) = 0.83$ mm/s, and  $Γ_1(1) = 0.34$  mm/s;  $δ_2(1) = 0.53$  mm/s,  $ΔE_2(1) = 1.52$  mm/s,  $Γ_2(1) = 0.32$  mm/s. In solution II, the parameters of the two quadrupole doublets are  $\delta_1(1) = 0.10$  mm/s,  $\Delta E_{Q1}(II) = 0.66$  mm/s, and  $\Gamma_1(II) = 0.29$  mm/s;  $\delta_2(II) = 0.44$  mm/s,  $\Delta E_{Q2}(II) = 1.70$  mm/s, Γ2(II) = 0.38 mm/s. In both solutions, site 1 should represent complex **2**, while site 2 is from the diferric decay product. Due to the similarity of the supporting ligands for **1** and **2**, it is reasonable to expect that the Mössbauer parameters of the diferric decay product of **2** should be similar with those of the decay product of **1**. Based on this, solution II should be the correct solution. However, since both solutions produce a similar quality of fit to the experimental data, we sought additional experimental support for solution II. Variable field and temperature Mössbauer analysis was then carried out. In Figure S20, the spectral features belonging to the diamagnetic (*S* = 0) diferric species are indicated by black arrows. The magnitude of magnetic splitting of an *S* = 0 species depends on magnitude of quadrupole splittings and the magnitude of externally applied magnetic field. In solutions I and II described above, the quadrupole splittings of the diferric species are very different. Thus, the spectral features of the diamagnetic diferric species in the 7-T spectrum should provide strong evidence to distinguish the two spectral simulation solutions. Indeed, the use of  $\Delta E_Q = 1.70$  mm/s can nicely reproduce the spectral features of the diferric species in the 7-T spectrum, while the use of  $\Delta E_{\rm Q} = 1.52$  mm/s cannot.

Therefore, solution II is a better solution to describe the spectra measured on the sample containing **2** with the quadrupole doublet having  $\delta_1(II) = 0.10$  mm/s,  $\Delta E_{Q1}(II) = 0.66$  mm/s to represent **2**. In addition, by using the variable field and temperature Mössbauer data, we determined that **2** still retained an *S* = 1 ground spin state with a large and positive *D*  $(D \sim 35 \text{ cm}^{-1})$  and an axial <sup>57</sup>Fe hyperfine tensor  $(A_x = A_y = -23 \text{ T})$  (**Figure S21**). Interestingly, the isomer shifts of complexes **0**, **1**, and **2** exhibit a clear pattern, where an increase in the number of quinoline donors on the supporting ligand leads to higher isomer shift. This is consistent with the correlation between isomer shift and iron-ligand bond length initially identified by Neese, $22$  namely longer iron-ligand bonds give rise to higher isomer shift. Therefore, the current isomer shift trend suggests that in going from complex **0** to **2**, the average iron-ligand bond length increases due to an increase in the number of quinoline donors.



**Figure S19.** Zero field Mössbauer spectra of a sample containing complex **1** (top) and a sample containing the diferric decay product of **1** (bottom) measured at 4.2 K. The experimental data are shown as vertical black bars, the overall simulations of the spectra are shown as the grey solid lines, and the spectral simulation of **1** is indicated as the red solid line. The spectral features indicated by the black arrows belong to mononuclear high-spin ferric species. The simulation parameters are listed in **Table S8**.



**Figure S20.** 4.2 K and 7 T Mössbauer spectra of a sample containing **2**. The black arrows indicate the spectral features belonging to the diamagnetic diferric decay product. The experimental data are shown as the vertical black bars and the simulations of the diamagnetic diferric decay product are shown in blue, which are generated by the simulation parameters from solution II (top) and from solution I (bottom). It is clearly shown that the simulated spectrum generated by the parameters of solution I does not match the diferric spectral feature well, while the parameters of solution II do.

Temp (K)	<b>Species</b>	<b>Simulation</b> Solution	$\Delta^{[\mathsf{a}]}$ (mm/s)	ΔE <sub>Q</sub> (mm/s)	$\Gamma$ (mm/s)	%A
			0.05	0.70	0.34	48
4.2	Di-ferric decay product of 1		0.46	1.65	0.32	45
4.2	Di-ferric species[b]		0.47	1.59	0.33	100
4.2	$\overline{2}$	Solution I	0.03	0.83	0.34	48
		Solution II	0.10	0.66	0.29	45
4.2	Di-ferric decay	Solution I	0.53	1.52	0.32	45
	product of 2	Solution II	0.45	1.70	0.38	50
150	2	Solution II	0.08	0.63	0.36	46
	Di-ferric decay product of 2		0.42	1.76	0.50	53

**Table S8.** Mössbauer simulation parameters of various species.[a]

[a]The change in isomer shifts at different temperatures originates from the second order Doppler effect on isomer shift due to the difference on the source temperature and the sample temperature.

[b]The di-ferric species obtained from the fully decay of complex **1**.

# **5. DFT calculations for oxoiron(IV) complexes**

We optimized the structures of complexes [Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>, 2, and 3 in Gaussian 16 (Rev. B01)<sup>23</sup> in both the  $S = 1$  and the  $S = 2$  states with modified B3LYP<sup>24-26</sup> (labeled as B3LYP\* in the following) functionals (Iop(3/76=1000001500) Iop(3/77=0720008000) Iop(3/78=0810010000)), a 6-31G(d',p') basis set and a built-in acetonitrile solvation model (scrf=(solvent=acetonitrile)).<sup>27</sup> The empirical dispersion using the D3 version of Grimme's dispersion with the original D3 damping function was also included in the geometry optimization.<sup>28</sup> To obtain a more precise result, we ran the optimization with a small step size (opt=maxstep=10), while the default value of *maxstep* was 30. For complex **2**, a C<sup>s</sup> symmetry was enforced during the geometry optimization. Then we performed <sup>1</sup>H-NMR shift calculations by using the same functional but with a Dunning correlation-consistency basis set cc-pVTZ by following the protocols described by Bagno and coworkers.29-32

The calculated <sup>1</sup>H-NMR shifts were determined by the following formula:  $33, 34$ 

$$
\sigma = \sigma_{ref} - (\sigma_{orb} + \sigma_{FC} + \sigma_{PC})
$$

where *σref* = 31.03 ppm for TMS was obtained at the same theoretical level as those for the Fe(IV) complexes included in this study. *σorb* is the orbital contribution to the proton, which is equivalent to the shielding for diamagnetic systems. The Fermi contact term, *σFC*, which originates from the scalar interaction between magnetic field from unpaired electrons and the magnetic momentum of target proton, dominates the paramagnetic component in the <sup>1</sup>H-NMR shift arising from the paramagnetic center. The Fermi contact term can be calculated from Fermi's hyperfine interaction parameters as the following:

$$
\sigma_{FC} = \frac{2\pi}{\gamma_I} g_{iso} \mu_B A \frac{S(S+1)}{3kT}
$$

in which *γ<sup>I</sup>* is the magnetogyric ratio of the nucleus I and *giso* is the isotropic g factor of the spin system, *μ<sup>B</sup>* is the Bohr magneton and *A* is the Fermi hyperfine interaction parameter. For  $[Fe^{IV}(O)(N2Py2Q)]^{2+}$ , the g values have been determined by high field EPR measurements,<sup>12</sup> therefore for this complex we used the experimentally determined *g* values to determine  $q_{iso}$ , which is 2.05. We then used this value for all the  $S = 1$  state NMR calculations. For the S = 2 complex **3**, no experimentally determined *g* values are available. However, the literature reported  $q$  values for two other  $S = 2$  oxoiron(IV) complexes, [Fe<sup>IV</sup>(O)H<sub>3</sub>buea]<sup>-4</sup> and [Fe<sup>IV</sup>(O)(tpa<sup>Ph</sup>)]<sup>-</sup>,<sup>2</sup> are all close to 2, therefore, for all the S = 2 state NMR calculations, *g*iso is fixed at 2. The contribution from pseudo-contact term *σ<sub>PC</sub>*, can be approximated by the theory developed by Hrobárik et al.<sup>35</sup> Usually it is small and can be ignored compared with  $\sigma$ <sub>FC</sub>. In this work we estimate the <sup>1</sup>H-NMR shift only by the orbital term and Fermi-contact term. All calculations were done at 233 K, the same temperature at which most of the <sup>1</sup>H-NMR measurements were performed in this work.

		Bond Length (Å)					
<b>Spin State</b>	Complex	$\mathsf{Fe}\text{--}\mathsf{Neq}^{[\operatorname{a}]}$	$\mathsf{Fe}\text{--}\mathsf{N}_{\mathsf{ax}}{}^{\text{[b]}}$	$Fe = O$	$Fe-Navq$ <sup>[c]</sup>		
		1.974, 1.997, 2.057, 1.975	2.103	1.648	2.021		
$S = 1$	2	1.971, 1.978, 2.069, 2.069	2.089	1.647	2.035		
		2.075, 2.075, 2.064, 1.988	2.088	1.646	2.058		
		2.095, 2.145, 2.118, 2.237	2.132	1.635	2.145		
$S = 2$	$\mathbf{2}$	2.199, 2.199, 2.088, 2.194	2.120	1.631	2.160		
	3	2.111, 2.166, 2.196, 2.206	2.090	1.637	2.154		

**Table S9.** Selected bond lengths of the DFT optimized structures for complexes **1** - **3**.

[a] The Fe–N bond lengths of the ligands in the equatorial plane that is perpendicular to the Fe=O axis. [b] The Fe–N bond length of the ligand trans to the Fe=O moiety.

[c] The averaged Fe–N bond lengths.

**Table S10.** Calculated & experimental <sup>1</sup>H NMR parameters for  $S = 1$  [Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup> at 233 K in CD<sub>3</sub>CN.



**Table S11.** Calculated & experimental <sup>1</sup>H NMR parameters for *S* = 2  $[Fe<sup>IV</sup>(O)(N2Py2Q)]<sup>2+</sup>$  at 233 K in CD<sub>3</sub>CN.



Table S12. Calculated & experimental <sup>1</sup>H NMR parameters for a *C*<sub>s</sub>-symmetric structure of *S* = 1 complex **2** ([FeIV(O)(BQPA)(CD3CN)]2+) at 233 K in CD3CN.

protons	$\sigma^{\rm orb}$	$A$ (MHz)	$\sigma$ <sub>FC</sub>	$\sigma^{FC}$ (total)	<b>O</b> calculated	<b>O</b> experimental
рy						
β	23.5444	0.45571	-42.24478441	-18.70038441	49.7803844	118
V	23.2122	$-0.0047$	0.435694821	23.64789482	7.43210518	$-26$
$\beta'$	23.8816	$-0.36858$	34.16774405	58.04934405	-26.969344	unassigned
$\alpha$	20.7705	$-0.68677$	63.66428341	84.43478341	-53.354783	unassigned
quin						
H <sub>3</sub>	23.3565	$-0.30443$	28.22097325	51.57747325	$-20.497473$	76, 77, or
	23.3569	$-0.30417$	28.19687098	51.55377098	-20.473771	83
H <sub>4</sub>	22.3234	0.08221	-7.620951322	14.70244868	16.3775513	27, 25, or
	22.3238	0.08256	<u>-7.653396681</u>	14.67040332	16.4095967	23
H <sub>5</sub>	22.6686	0.13958	-12.93920917	9.729390835	21.3506092	76, 77, or
	22.6688	0.14033	-13.00873493	9.660065066	21.4199349	83
H6	22.6918	0.2397	-22.22043586	0.471364143	30.6086359	27, 25, or
	22.6919	0.24017	-22.26400534	0.427894661	30.6521053	23
H7	22.4242	0.21465	-19.89827516	2.525924837	28.5540752	27, 25, or
	22.4242	0.21532	-19.96038485	2.46381515	28.6161849	23
H8	19.5847	1.1068	<u>-102.6014952</u>	-83.01679523	114.096795	<b>Not</b>
	19.5826	1.10902	-102.8072915	-83.2246915	114.304692	observed

Table S13. Calculated & experimental <sup>1</sup>H NMR parameters for a *C*<sub>s</sub>-symmetric structure of *S* = 2 complex **2** ([FeIV(O)(BQPA)(CD3CN)]2+) at 233 K in CD3CN.





**Table S14.** Calculated & experimental <sup>1</sup>H NMR parameters for *S* = 1 complex **3**  $([Fe<sup>IV</sup>(O)(TQA)(CD<sub>3</sub>CN)]<sup>2+</sup>)$  at 233 K in CD<sub>3</sub>CN.

**Table S15.** Calculated & experimental <sup>1</sup>H NMR parameters for *S* = 2 complex **3**   $([Fe<sup>IV</sup>(O)(TQA)(CD<sub>3</sub>CN)]<sup>2+</sup>)$  at 233 K in CD<sub>3</sub>CN.



Table S16. The calculated free energy difference between Fe<sup>(IV)=</sup>O complexes and their corresponding one-electron reduced complexes a



 $a \Delta G$ (Felll/Fel<sup>V</sup>) = G(Felll)-G(Fel<sup>V</sup>)

 $b$  ΔΔG(Fe<sup>III</sup>/Fe<sup>IV</sup>) is the relative free energy differences between complexes 1, 2 (S = 2), 2 (S = 1) and 3 to complex 0 by setting  $\Delta G$ (Fe<sup>III</sup>/Fe<sup>IV</sup>) of complex 0 as the reference (setting the value to be zero).

<sup>c</sup>  $ΔE$  is the relative reduction potential differences between complexes 1, 2 (S = 2), 2 (S = 1) and 3 to complex 0 by setting the reduction potential of complex 0 as the reference.

# **6. DFT Coordinates**





#### **Coordinates of** *S* **= 2 [FeIV(O)(N2Py2Q)]2+:**





 $\begin{array}{cccc} \text{C} & 1.18313000 & 2.46617200 & -1.01794100 \\ \text{C} & 1.13795200 & 2.17824200 & 0.38373200 \\ \text{C} & 0.48831600 & -2.44759000 & 1.88297800 \\ \text{C} & & 1.74099000 & -2.01657400 & 1.18293200 \end{array}$ C 1.13795200 2.17824200 0.38373200 C 0.45831600 -2.44759000 1.88297800 C 1.74099000 -2.01657400 1.18293200 C 3.01165200 -2.34003800 1.73516900<br>C 4.16057000 -1.99056400 1.02993700 C 4.16057000 -1.99056400 1.02993700



#### **Coordinates of** *S* **= 1 [FeIV(O)(TQA)NCCH3] 2+:**



# **6. DFT Coordinates**



#### **Coordinates of** *S* **= 1 [FeIV(O)(BQPA)NCCH3] 2+:**





N 0.89871500 -0.47074400 1.60599600 C -0.14704800 -1.38749400 2.19047700 H -0.17133200 -1.32396000 3.29807300<br>H 0.11194400 -2.43104100 1.91932200 H 0.11194400 -2.43104100 1.91932200 C 2.29959200 -1.03778400 1.68827300 H 2.23164300 -2.13439600 1.53469900 H 2.75442600 -0.86247400 2.68480900



#### **Coordinates of** *S* **= 2 [FeIV(O)(QBPA)NCCH3] 2+:**



# **6. DFT Coordinates**



# **7. References**

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