

Supplementary Table 1. A key resource table

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Antibodies		
Anti-SpAGS	Voronina & Wessel, 2006	N/A
Anti-Gai	Santa Cruz Biotech	# sc-56536
Anti- β -catenin	Yazaki et al., 2015	N/A
Anti-SpInsc	This article	N/A
Anti-SpNuMA	Poon et al., 2019	N/A
Anti- β -actin (8H10D10)	Cell Signaling Technology	# 3700S
Anti-Flag	Millipore-Sigma	# F1804
Alexa 488-conjugated goat anti-mouse IgG	Cell Signaling Technology	# 4412
Alexa 555-conjugated goat anti-mouse IgG	Cell Signaling Technology	# 4409
HRP-conjugated anti-Protein A antibody	Abcam	# ab7245
HRP-conjugated goat anti-mouse IgG	Cell Signaling Technology	# 7076
Anti-Digoxigenin-AP, Fab fragments	Roche	# 11093274910
Chemicals		
Hoechst 33342	Thermo Fisher Scientific	# 62249
DTT	Millipore-Sigma	# 10197777001
Tris buffered saline, with tween (TBST)	Millipore-Sigma	# T9039
Tris-MOPS-SDS Running Buffer	Genscript	# M00138
Transfer buffer powder	Genscript	# M00139
DIG RNA labeling mix	Roche	#11277073910
Critical Commercial Assays		
mMESSAGE mMACHINE SP6 Transcription Kit	Thermo Fisher Scientific	# AM1340
MEGAscript SP6 Transcription kit	Thermo Fisher Scientific	# AM1330
MEGAscript T7 Transcription kit	Thermo Fisher Scientific	# AM1333
In-Fusion HD Cloning	Clontech	# 639648

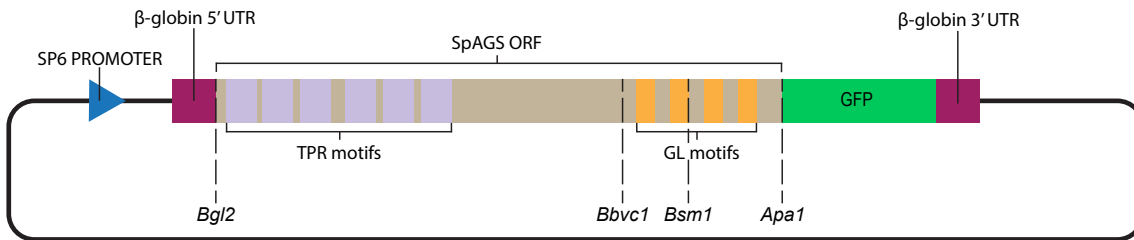
Duolink [®] In Situ Red Starter Kit Mouse/Rabbit	Millipore-Sigma	# DUO92101
Recombinant DNA		
Plasmid: SpAGS-GFP	Poon et al., 2019	N/A
Plasmids: SpAGS-dC-term-GFP, SpAGS-dGL1/2/3/4-GFP, SpAGS1111/2222/2134/4234-GFP	This article	N/A
Plasmids: SpAGS-dN-term-GFP	This article	N/A
Plasmid: SpAGS-mCherry	Poon et al., 2019	N/A
Plasmid: GFP-SpAGS, GFP-EtAGS, GFP-PmAGS	This article	N/A
Plasmid: GFP- SpAGS4444/GL1GL2/LGNGL/Dm GL/EtGL/AGS3GL/PmGL/S389A/A GS3GL-3S/A/AGS3GL-GL2GL3	This article	N/A
Plasmid: GFP-PmAGS-SpLinker	This article	N/A
Plasmid: GFP-SpDlg/PmDlg	This article	N/A
Plasmid: GFP-SpInsc/EtInsc/PmInsc	This article	N/A
Plasmid: GFP-NuMA	This article	N/A
Plasmid: mCherry-NuMA	This article	N/A
Plasmid: GFP-Par3	This article	N/A
Plasmid: Vasa-GFP	Yajima & Wessel, 2011	N/A
Plasmid: Vasa-mCherry	Uchida & Yajima, 2018	N/A
Plasmid : 3xFlag-GFP- SpAGS/SpDlg/SpNuMA, 3xFlag- Vasa-GFP	This article	N/A
Plasmid: 2x-mCherry-EMTB	Addgene	# 26742
Software and Algorithms		
Echinoderm protein sequences	EchinoBase	http://www.echinobase.org/Echinobase/
Protein motif search	NCBI blast	https://blast.ncbi.nlm.nih.gov/Blast.cgi
Protein sequence alignment	Clustal Omega	https://www.ebi.ac.uk/Tools/msa/clustalo/
Quantitative analysis	Image J	https://imagej.nih.gov/ij/
Statistical analysis	Graphpad PRISM 8	https://www.graphpad.com/scientific-software/prism/
Morpholino antisense oligos (Gene Tools: http://www.gene-tools.com/)		
NCBI reference sequence	Name	Sequence
NM_001040405.1	SpAGS	GGCCCGTTTCACAAAGCCTTTGTTT
Primers for ISH probes		
NCBI reference sequence / Source	Name	Sequence

XM_011663478.2	SpAlx1	F: GGATATTTTCTCGACCGGGATC R: CGAGTAACCGTTCATCATCCCC
NM_214574.3	SpBlimp1b	F: ATGGGGTGCAACGACAACGCCGTG R: CTATGATTTGTTCTGTACGATTGAG
NM_214519.1	SpEndo16	F: GCAGAGTTCAACAGAATCGAC R: GCCAGTAGACGTAGCAGAAG
XM_030976919.1	SpEts1	F: TCAATCATGGCGTCTATGCACTG R: ACAGCTGCAGGGATAACAGG
NM_001079542.1	SpFoxA	F: ATGGCCAATAGTGCCATGATCTCG R: TCACATTGCATGGTTTGCTTG
XM_003731512.3	SpFoxQ2	F: ATGACTTTATTCAGCATTGACAAC R: TAGCAGGATCCTACAGAAGACCAG
NM_214610.3	SpSm50	F: ATGAAGGGAGTTTTGTTTATTGTGG CTAGTC R: GTTATGCCAACGCGTCTGCCTCTTG AAGC
XM_786173.5	SpTbr1	F: CCACCGCTGCACCAGACGAC R: CTGCCGGCTGGCGCCAATTGCG
NM_214667.1	SpWnt8	F: ATGGATGTTTTTACGGAATTTGTTG R: CTACAGCCTCGATCCAACGGGCTG

Supplementary Figures and Legends

Supplemental Figure S1

A



B

S. purpuratus AGS Protein Sequence

TPR1 TPR2
 MQSEASCMELALEGERLCKAGNCREGVRYLEAAVEVGTDDLKLSAIYSQLGNAYFYLEDYGKALTYHKHDL
 TPR3 117aa, before here removed in 1F
 TLATSIGDRLGEAKASGNIGNTLKVLGKFDEAICCCQRHLDISRELSSEKVGEGRALYNLGNVYHAKGKHAGRS
 TPR4
 GHQDPGDFPQEVTDCLKQAIQFYEANLAIVRELGDRAAQGRACGNLGNTHYLLGNFETAIQFHTERLAIKE
 222aa, before here removed in 2F
 TPR5 TPR6
 FGDKA AERRAYSNLGNACVFMVQFEVAAKYKSLHIARQLGELAMEAQACYSLGNTYLLREYKAVEYHS
 301aa, before here removed in 3F
 RHMEIAQQLNDRVGEGRACWSLGNHAHTSLGNHEKALHYATLHLQISREVDRTGEVTAKMNLQDLQTLFG
 ISTADLSEVSTTVQTPLQESKGARPRRRSMENLELVAMTPEKTEIVQVPRKIRPGSKLKLKNGGESKDKQS
 BbvC1 RE site GL1 (removed in Δ1)
 HKDSKESVSSAKNSVPAPSKAKSQTSARKGGEPQMLDDADNFFEALSRFQSNRMDEQRCSFGRLQKKLAD
 GL2 (removed in Δ2) Bsm1 RE site GL3 (removed in Δ3)
 EEGNGLPEKEELLDEIAKLQGSRLNEQRAF SVKRLPGLPLRANEDVVGKLLAKGERAEPD DDFEMIIIRCQG
 GL4 (removed in Δ4)
 ARIEDQRSTLPIQAPAPTVPDEDFFSLIQRIQSKRIEEQRSIAPWEKSGTSCVCFYDYDTSRTEGTACCVYSRP
 659aa
 VQESHL

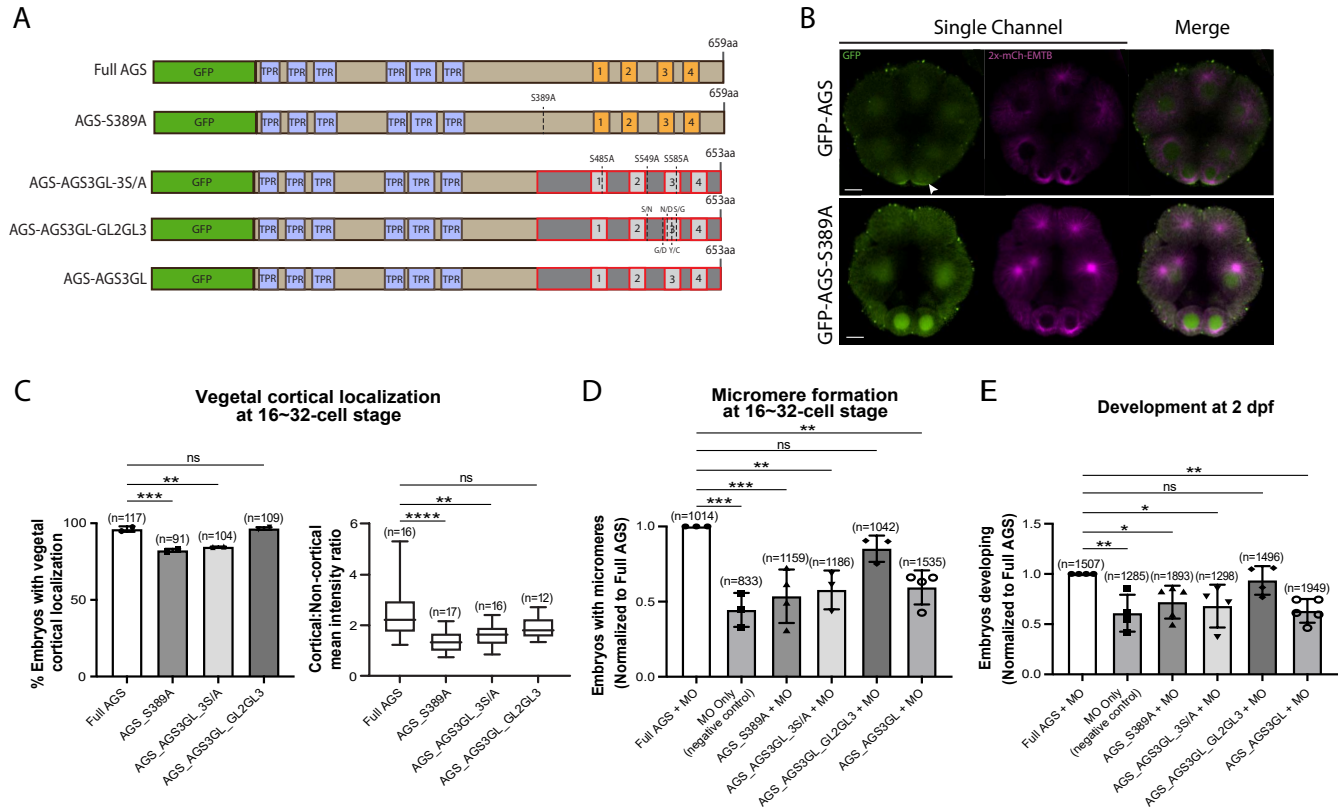
Supplemental Figure 1. Dissection of SpAGS motifs. **A**, The construct design of SpAGS-GFP, including the restriction enzyme sites used to prepare SpAGS mutants. **B**, The protein sequence of SpAGS. Predicted domains are labeled based on NCBI blast results, and the sequence portions deleted for each N-terminal construct are marked. The sequences for each GL motif used for deletion or swapping are indicated in orange. The internal restriction enzyme sites for *BbvCI* and *BsmI* are shown in green.

Supplemental Figure 2 Echinoderm AGS sequence alignment



Supplemental Figure 2. Echinoderm AGS sequence alignment. All AGS are similar in the N-terminus with the predicted conserved TPR motifs (blue) but are highly variable in the C-terminus with the predicted GL motifs (yellow).

Supplemental Figure 3



Supplemental Figure 3. The linker domain and GL2-GL3 region are important for AGS localization and function. **A**, Design of GFP-AGS mutant constructs tested in this study. TPR motifs are marked in blue, and GL motifs are in orange. The brown section shows the SpAGS portion and the red and grey boxes show the non-sea urchin (non-SpAGS) C-terminal sequence introduced. The dotted lines represent single amino acid mutations. **B**, Single Z-slice confocal images of sea urchin embryos at 8~16-cell stage showing localization of GFP-AGS-S389A mutant. Embryos were injected with 0.3 μ g/ μ l stock of GFP-AGS mRNA and 0.25 μ g/ μ l stock of 2x-mCherry-EMTB mRNA. The white arrowhead indicates vegetal cortical localization of GFP-AGS. **C**, % of the embryos with vegetal cortical localization of GFP-AGS mutants (left) and the ratio of the cortical-to-non-cortical mean intensity (right) in 16~32-cell embryos. Statistical analysis was performed against Full AGS by One-Way ANOVA. **D-E**, Embryos were injected with 0.15 μ g/ μ l stock of GFP-AGS mRNAs and 0.75mM SpAGS MO. The number of embryos making micromeres (D) and developing to gastrula or pluteus stage (E) were scored and normalized to that of the Full AGS. Statistical analysis was performed by One-Way ANOVA. n indicates the total number of embryos scored. * represents p-value<0.05, ** p-value < 0.01, *** p-value < 0.001, and **** p-value <0.0001. Each experiment was performed at least two independent times. Error bars represent standard error. Scale bars=10 μ m.

Supplemental Figure 4 SpAGS, EtAGS and PmAGS linker domain sequence alignment

A

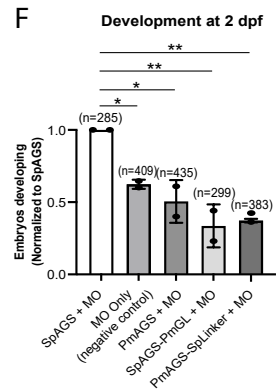
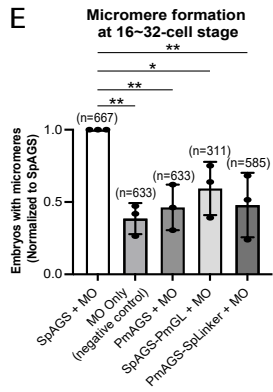
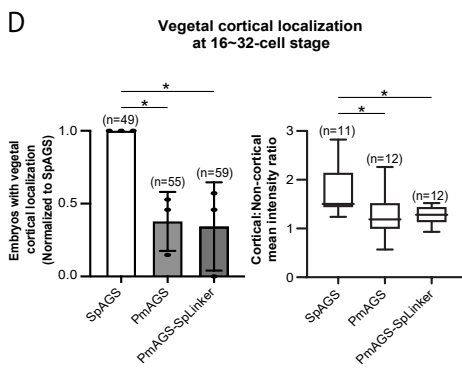
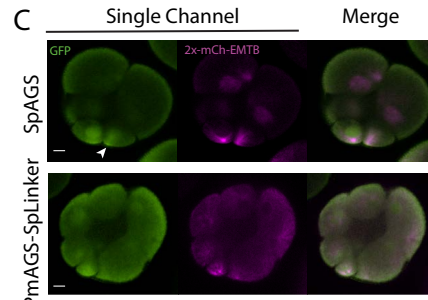
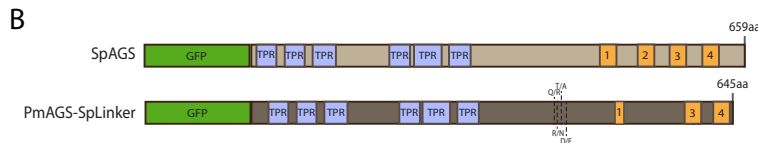
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SpAGS_linker  YHSRHMETAQQLNDRVGEGRACWSLGNHAHTSLGNHEKALHYATLHLQISREVGDRTEVET 346
EtAGS_linker  YHMRHMETAQQLQDRVGEGRACWSLGNHAHTSLGNHEKALHYATLHLQISREVGDRTEVET 346
PmAGS_linker  YHERHHRITAKLKDVRVGEGRACWSLGNHAHTALQNHKALHYATLHLQISREVGDKTELT 360
** ** .**:*:*****:*****:* *****:*****:***:*

SpAGS_linker  AKMNLQDLQTLFGISTADLSEVSTT-V-QTPLQESKGARPRPRRSMENLELVAMTPEKKT 404
EtAGS_linker  AQMNLADLKTFLFGITGDLSDVTAAC-PPAMQPTKGATRPFRSMENLELVAMTPEKNT 405
PmAGS_linker  AQMNLADLRVTLGLDKDPCSQDGLLQNASKKTLEKELNQRFRSMERLELVMTPEKNA 420
*:* **:*:*. . * * *:*:***:***:***:

SpAGS_linker  EIVQVPKRKIRP-GSKLKLKNGGESKDKQSHKDSKESVSSAKNSVPAPS---KAKSQT 459
EtAGS_linker  DSA-VPNGKLPKTPGSKSKRRNGE---KLSRKGSNTSTGSSNSSSAPS---QASTQAS 457
PmAGS_linker  AAATANGPKQSGAKFKRK--GSNKEKLSRKSNTSSTSSSGESAACLRTENVNIQVS 478
. . . : * : * : . * : * : * * : . . . * . . . * : *

SpAGS_linker  ARKGGPEIQMLDDA 473
EtAGS_linker  SQKGGEPQEMLDDA 472
PmAGS_linker  P-QQRSSPEFMDDD 492
. . . : : : * *
    
```



Supplemental Figure 4. The Dlg recognition sequence in the linker domain is important for the AGS function. **A**, Alignment of linker domain between echinoderms including sea urchin (SpAGS_linker), pencil urchin (EtAGS_linker), and sea star (PmAGS_linker). Bold letters represent the conserved core of the linker domain. The green highlight indicates conserved amino acids in the consensus kinase recognition motif. The red highlight represents amino acid mutated to match SpAGS residues in the PmAGS-SpLinker mutant. **B**, Design of GFP-AGS constructs tested in this study from *S. purpuratus* (Sp) and *P. miniata* (Pm). TPR motifs are marked in blue, and GL motifs are in orange. The dotted lines represent single amino acid mutations. **C**, Single Z-slice confocal images of sea urchin embryos at 8~16-cell stage showing localization of GFP-AGS. Embryos were injected with 0.3µg/µl stock of GFP-AGS mRNAs and 0.25µg/µl stock of 2x-mCherry-EMTB mRNA. The white arrowhead indicates vegetal cortical localization of GFP-AGS. **D**, The number of embryos with vegetal cortical localization of GFP-AGS mutants in 16~32-cell embryos was scored and normalized to that of the SpAGS (left graph). Right graph shows the ratio of the cortical-to-non-cortical mean intensity. Statistical analysis was performed against SpAGS by One-Way ANOVA. **E-F**, Embryos were injected with 0.3µg/µl stock of GFP-AGS mRNAs and 0.75mM SpAGS MO. The number of embryos making micromeres (E) and developing to gastrula or pluteus stage (F) were scored and normalized to that of the SpAGS. Statistical analysis was performed by One-Way ANOVA. n indicates the total number of embryos scored. * represents p-value<0.05, and ** p-value < 0.01. Each experiment was performed at least two independent times. Error bars represent standard error. Scale bars=10µm.

Supplemental Figure S5 GoLoco Domain sequence alignment

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SpAGS_GL 461 RKGGEPIQLDDADNFFFEALSRLFQSNRMDEQRCSFGRLQKCLA-----
DmPins_GL 455 EKLRAQATRKAADDFFEMLSRSQSKRMDDQRCSIKVNPAGAPAVATGATRKPL----
PmAGS_GL 479 PQQRSSEPFMDDDDSFVDVLSRFQGRMDEQRCSFNRMQDKQRERE-----
EtAGS_GL 459 QKGGEPQEMLDDADNFFFEALSRLFQSNRMDEQRCSFGRMQRKLQE-----
HsLGN_GL 468 NSQRKISADTI GDEGFFDLSRLFQSNRMDDQRCCLEKNCHTASTTTSSTPPKMLKT
HsAGS3_GL 460 PRTSIPRAPSSDECEFFDLLTKFQSSRMDDQRCLDDGQAGAAEATAAPTLEDRI--A
      : **: **: *..**:* ** :

SpAGS_GL DEEENGLPEKEELLDEIAKLQGSRLNEQRAFVSKRPLPGLPGLR-ANE---DVVGK---LL
DmPins_GL -----VQNSLFVDPTNLPGLKSPSSANPSAIGHGGLAR
PmAGS_GL NDEDRDYPMKEELMNQIASFQRSRLNEQRSS----ICNLPGLKTNNE---EVLGQ---LL
EtAGS_GL DEEANGLPKEKEELLDEIAKLQGSRLNEQRAFSAQRLPGLPGLR-ANE---NVLGK---LL
HsLGN_GL SSVPVVSPNTDEFFDLLASSQSRRLDDQRAS----FSNLPGLRLTQNSQ-SVLS---HL
HsAGS3_GL QPSMTASPQTEEFFDLIASSQSRRLDDQRAS----VGLPGLRITHSNA-GHLR---G-
      * : .****: . :

SpAGS_GL AKGERAEDDDDFFEMIIRCQGARIEDQRSTLP-I-----
DmPins_GL SATTTQQPDDDFLDMLMRCQGSRLEEQRSELPRPNVTMDAEAEAPPRSVPEAAVPGAPRG
PmAGS_GL QKGNQGVDDDDFFDMLMRCQGSRLINDQRSEPP-V-----
EtAGS_GL AKGDLAVDDDDFFEMLMRCQGARIEDQRSTLP-M-----
HsLGN_GL MTNDNKEADEFFDLLVKCGSRLDDDQRCAPPPA-----
HsAGS3_GL -HGEPQEPDDFFMLIKQSRLDDDQRCPPDV-----
      .:****: : *.:**:* *

SpAGS_GL -QAPAPTVPDEDFSLIQRIQSKRIEEQRSIAPWEKSGT-----SCVCFYDYDTSR
DmPins_GL QTGRGATVPDEDFSLIMKVQSGRMEDQRASIPFRNANNNNNSRS-----NNNG
PmAGS_GL -LQPAPTVPDEDFFALIQRVQSKRMDAQRSDKGPQEQTKT-----SD-----
EtAGS_GL -PTPAPTVPDEDFSLIQRIQSRRIEEQRTIPPWEKGPKP-----SDK-----
HsLGN_GL -TTKGPTVPDEDFSLILRSQGRMDEQRVLLQRDQNRDDFGLKDFLQNNALLEFKNSG
HsAGS3_GL -LPRGPTMPDEDFSLIQRVQAKRMDEQRVLDLAGGPEQAGGPPEPQQQC-----QPG
      . * :*****:** : * . **: **

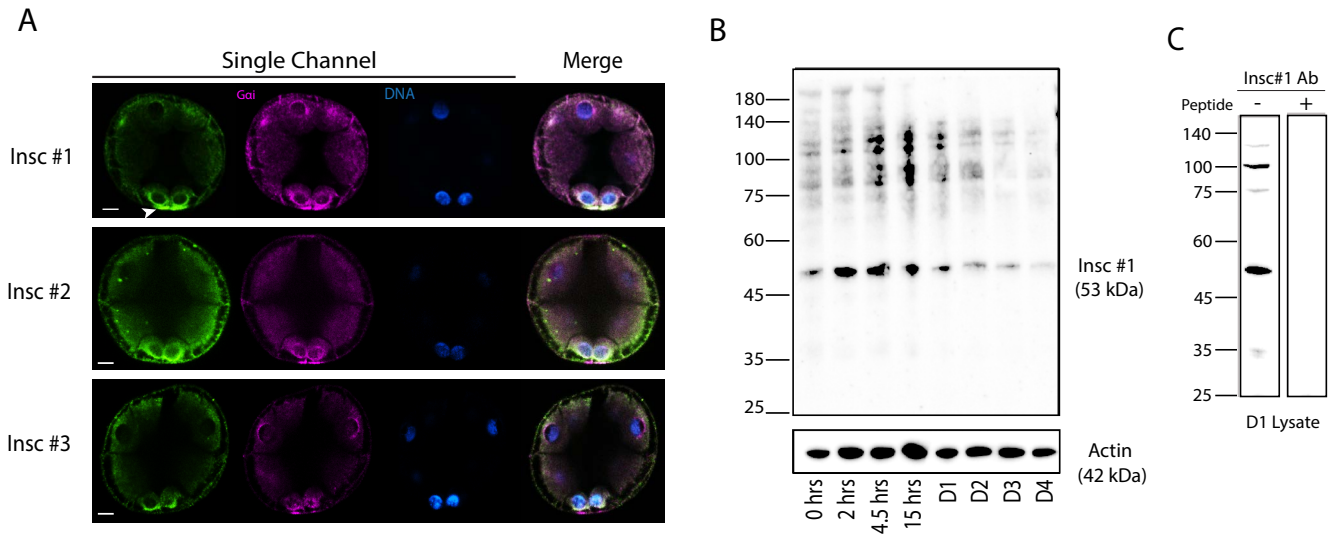
SpAGS_GL TEGTACCVYSRPVQESHL
DmPins_GL SAGGAGK-----
PmAGS_GL -----
EtAGS_GL -----
HsLGN_GL KKSADH-----
HsAGS3_GL AS-----

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Supplemental Figure 5. Alignment of C-terminus GoLoco domain sequences used for chimeric mutants.

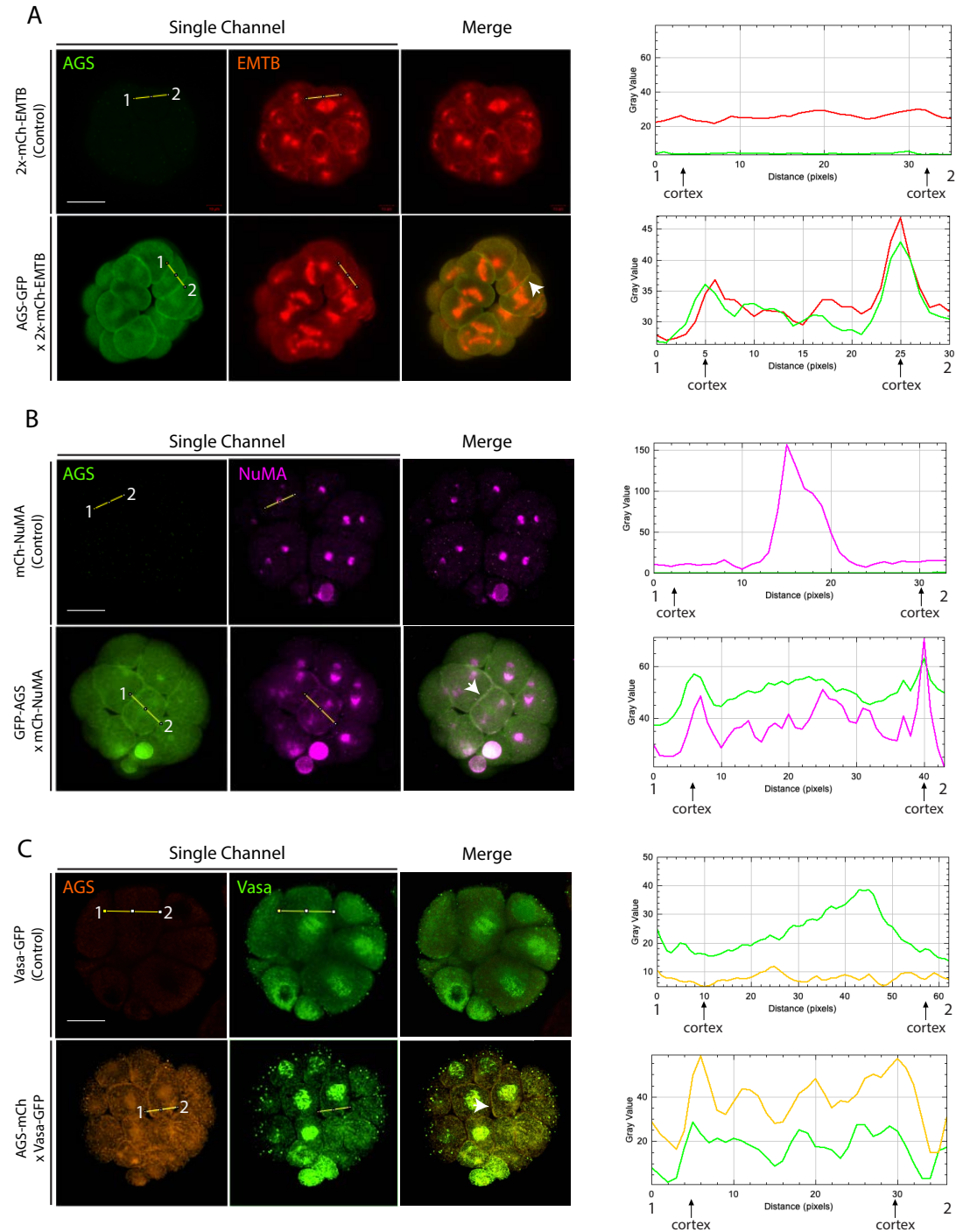
Sea urchin *S. purpuratus* (SpAGS_GL), *Drosophila* (DmPins_GL), sea star *P. miniata* (PmAGS_GL), pencil urchin *E. tribuloides* (EtAGS_GL), human *H. sapiens* LGN (HsLGN_GL) and human *H. sapiens* AGS3 (HsAGS3_GL). Bold letters indicate GoLoco motif sequences. The green highlight indicates additional serine amino acid present uniquely in HsAGS3_GL and mutated to Alanine in AGS_AGS3GL_3S/A mutant. The highlighted amino acids between GL2 and GL2 and within GL3 are those mutated to match HsLGN_GL in AGS_AGS3GL_GL2GL3 mutant.

Supplemental Figure 6



Supplemental Figure 6. Insc protein expression during embryonic development. **A**, Endogenous Insc protein localization by immunofluorescence. Embryos were stained with three Insc antibodies (green) designed for different Insc amino acid sequence sections. Embryos were stained with Gai antibody (magenta) and Hoechst dye (blue). During the 16-cell stage, all antibodies show signal enriched at the vegetal pole. With #2 and #3 antibodies, some non-specific cortex signals were also observed around the entire embryo. **B**, Insc immunoblot analysis. Embryos were collected at 0, 2, 4.5, 15, 24, 48, 72, and 96 hours post fertilization and subjected to immunoblot with Insc #1 antibody. Actin (42kDa) was used as a loading control. The expected size of Insc is 53kDa. **C**, Peptide competition assay with Insc #1 antibody. The 24-hour lysate was used. Each experiment was performed at least two independent times. Error bars represent standard error. Scale bars=10 μ m.

Supplemental Figure S7



Supplemental Figure 7. SpAGS colocalizes with micromere-specific fate determinants. **A**, Embryos were co-injected with 2x-mCherry-EMTB (0.5 μ g/ μ l stock) mRNA with or without GFP-AGS (0.5 μ g/ μ l stock) mRNA. **B**, Embryos were co-injected with mCherry-NuMA (0.15 μ g/ μ l) mRNA with or without GFP-AGS (0.5 μ g/ μ l stock). **C**, Embryos were co-injected with Vasa-GFP (1 μ g/ μ l stock) mRNA with or without AGS-mCherry (0.5 μ g/ μ l stock) mRNA. The intensity of each signal, from one cortex to the other, was measured and plotted from point 1 to 2 on the corresponding graph (right) using *ImageJ*. White arrows indicate the cortical colocalization of each construct. All images represent over 50% of the embryos observed (n=30 or larger) per group. Scale bars=20 μ m.

Supplemental Figure 8 Sea urchin SpDlg and sea star PmDlg sequence alignment

SpDlg	-----	0
PmDlg	MN NIEEWEYE DIILNRGGAGLGFS IAGGVNDNPHV PDDPSIFITKLI PGGAAAVGGRRLRLM	60
SpDlg	-----MVLRRWPRNPSTG KTLSELIKSGKGLGFS	30
PmDlg	DV IIRVNETEIHNVPHQNAVDALKKTKGSIVNLVV RRRRQRRTGD VIRIKLVKGTKGLGFS	120
	: : * * : * * . : * : * : * * * * * * * *	
SpDlg	IAGGQGNQHVPGDNGIFVTKIIDGGAAEQDGR LQVGDKITAVGDN NLVEVTHE DAVAVLK	90
PmDlg	IAGGCGNQHVAGDNGIFVTKIIDGGAAQMDGNLIGDKIIAVGNDKLEEVTHE NAVAVLK	180
	**** *	
SpDlg	ATSNRVL LTVIKN ATLLAGTQMAPYSQNDENAQP PDVSNVDMSAANFVYAT-----	141
PmDlg	ATSEVVL LTVIKG AVISYSP PASI-----TSQPDVSTMQPSSPPPPYPEAIVSSVAGV	234
	*** : * * * * * * * * * * : * : * : *	
SpDlg	--MQPTLMQEP RSD LQELDGAFGTNVAFGTNVANGANGANGSNGEPPSYNRVTNSQPP	198
PmDlg	SLQSEPAL-QRPPTPPHQEMSNMNP LTYHQPPQDTAYGSM-----KMKHVPPSP	284
	: * : * * * : * * : * * . : . : : : * * : : : * *	
SpDlg	NAFNNTPAKIFIQNDGIPRE PRQVVLNKGATGLGFNIVGGEDGEGIFISFILAGGVAD	258
PmDlg	TANYEGNGVKPYQNDDFPRE EARTVVLNKGATGLGFNIVGGEDGEGIFISFILAGGVAD	344
	. * : . * : *	
SpDlg	LSGALRRGDQILAVNSKDLVNATHEDAA LKAGQVVTI DAQYKPEEYNCFEAKIQSLR	318
PmDlg	LSGELKRGDQILSVNKKDLRGATHEDAA LKGTQTVLIEAQYKPEEYNRFEAKIQSLR	404
	*** : *	
SpDlg	EEMMQNSVS--PTGSLRTSAKRTLFVRTLFDYDKSRD SGLPSQGLSFD FGDIIHV TNASD	376
PmDlg	EEMNNSVSSSTT GSLRTSAKRSLYVRA LFDYDKTKDSGLPSQGLS FNYGDI LHV TNASD	464
	*** : *	
SpDlg	DEWVQARHILPENGEEGEIGIIPSKR RVERKEKARLR SVKFSGNKDRNNIQEPAGTVNSKG	436
PmDlg	DEWVQAKQVLPTEEGEMGVIPSKR RVEKREKARLR SVKFSGRGGS---LESKGS MNEKR	521
	*** : *	
SpDlg	KKGFF-RKFKNRSDQETSDFEPFTSNASDSESSYRN EEVILSYEAVVQQEIKYTRPVI	495
PmDlg	KKGFFSRKFKKGRSDLDTS DAEHITSNASDSESSYRNDEMI LSYEGVVQQEVKYTRPVI	581
	*** *	
SpDlg	ILGPGKDRINDDLISEMPDEFGSCVPHTTRARPHEVDGRDYHFVESREQMEKDIQNHLE	555
PmDlg	ILGSGKDRVNDLISEMPEKFGSCVPHTTRPRRPHEVDKRDYHFVESREQMEKDIQDHLE	641
	*** *	
SpDlg	IEAGQYENENLYGTSVASVKEVSEKGGKHCILDVSGNAIKRLQVAQLYPIAIFLKP KSVESI	615
PmDlg	IEAGQYENENLYGTSVASVREVADKGGKHCILDVSGNAIRRLQVANLHPAIAIFIRPKSVESV	701
	*** : *	
SpDlg	MEMNRVSEDAQKTYERALKLEQEFLEYFTA IILGDTIDDIYQQSKEI IHEQ SGPTIWI	675
PmDlg	MEMNRMSREQAEKTYERAIKLEIEFGEFFTA IISGDTLDEIYYKKEVIHDQ SGSTIWI	761
	: *	
SpDlg	PAKEKL 681	
PmDlg	PAKDKL 767	
	*** : * *	

PDZ
SH3
GUK

Supplemental Figure 8. Sea urchin (SpDlg) and sea star (PmDlg) sequence alignment. Blue, yellow, and green highlights indicate the PDZ, SH3, and GUK domains, respectively.

Supplemental Figure 9 Sea urchin SpInsc, pencil urchin EtInsc and sea star PmInsc sequence alignment

PmInsc	-----MNGFNES-ALAGWLADDVIEHMTDEDSVVRWLQDLRTMTCECECMCILQG	49
SpInsc	-----MSDN---KNTESFP-EKKLMQRCDVDSVQQWLIDIRLMPEQECTVVMET	45
EtInsc	MTCLYGVVRTMYGPPAPVICEEKLPSTSRFQLTEVRSVQQWLSDLQGMMECECN-ILET	59
	* . : . :: : ** : ** * : * * * * : :	
PmInsc	KPIRTSAHDMKAVTSSARDNIYILRQRASSISADFAKLYKLDSGSWDQVH-WLCLSLTI	108
SpInsc	KPIHNSPLNLENLAATVKE SVYRLQSIKDGFLSLSQIYMCLDLSLQ-----	92
EtInsc	KPIHNSPLNLENVAATKENIYRLRNRAENLARELQQLYHKLTSGTYSQRLRPAVLALT	119
	***:. * : : : : : : : * * : . . : : : * * .	
PmInsc	RLRFLHECTS YVPTMPLHLFQRQQTVLEECGRLVKHAESFEELNGTLPALPTLNVLSS	168
SpInsc	-----SVK--LFIQSLTT	103
EtInsc	HVRSCLEECNEYCPDF-LHMKEKQQVVIDAMATLVKNFEMYDNSKGCELD--SLIHSLTS	176
	: : * : :	
PmInsc	LGKAFNDLVHMVQGI LIQT VVRSIHESASEYNIKIAINS IHCLSEDDSEIRRLLAKEGAI	228
SpInsc	VAKAIKAYLEMVQ GALVVRKLVDMVQGAATELNVKFAVLTVHSLAQDGVWLRRLLIQEDTM	163
EtInsc	LGNALNSIRQTVQSALVRKLVMMIQDSASELNVKFAVITAHSLSQDSIWLRRILIQENAM	236
	: . : * : . * * : : * : : * : : * : : * : : * : : * : : * : : * : : *	
PmInsc	QALLQLCQQASL--NAMKPMALKTVANLNCNVAEGRQLEERASGVEYLSLSSPTATESV	286
SpInsc	RALLAICRI TTFSIRSIPTALEIVSILC TEVEGRAELEKVGVGALSEILASPTILEAV	223
EtInsc	GALFAVCRI TTLSIPSIVTALQAI STLCAEPEGRVELEKIGGLGFLGELSSPSTSEAV	296
	** : * : : : : : ** : : * * * * * : . * : * * * * * : * : *	
PmInsc	KAEAA SVIAQITSPNIDNYHLLGFIEYMEDLLKDLTALSGSASSSSVFIATSAIANIT	346
SpInsc	KKEAASVLA EITS PDADIFHRMLCFIEHMEDLLRNLTVLC DQTS SPGVF LVATTAIANIT	283
EtInsc	KLEAASVLA EITS PDINSYHRMLCFIEHMEDLLKNLTALCDQTSNPLVFLVSASAIANIT	356
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PmInsc	FMDSMACDC LASFHTARALVKGCNNGMANTLYCKDQVATILANMAVTDQCRKEIMVSGGT	406
SpInsc	FMDAMASDY LAEFQTA EVLIQGYLKGKAPSIYSKHQVITIIANISCSKSCREQIVSSGGM	343
EtInsc	FMDTMSDF LAMFNATILIDGYTSGKAQNMYSKDQVVTIIANMSGSEKREQI IKAGGM	416
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PmInsc	ELLVDFLHTRALVHPSSAK--KGLKGSGLSGGCSGGKQDVVMSSSCEKVHQKAAIALA	463
SpInsc	NILAE LLQGV PSTSRPTSLSRKLSFDGN-----VSLGQMGEESAYEDIYQKAATALG	395
EtInsc	KVLVELLHTRAPFTTVKTNQVN---QRA-----DSKAACGMASTCEHVHQKSAIALG	465
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PmInsc	RLARDQDSAE LILELDAI PRLVELCCNRRERNDSEFVLVACLAALRKLHNLGSPVFKSS	523
SpInsc	RLCQDYETCKMAVDLKVVSRLADLCKHARERNHRSDVLVACLAALRRIHSHIGSSPLKEN	455
EtInsc	RLCQDHD TAMMAVDLGVIPRLVDMCRHRRERNDSEFVLVACLAALRKLH SVVGASKFEPK	525
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PmInsc	DVDQLIKPRLIDSYLMCSATK--ESFV	548
SpInsc	DVEQLIQPRLMESFLHCSTTVRRETFV	482
EtInsc	ALQQLIRPRLIESFVTCSSIGSTETFV	552
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LBD (LGN-binding domain)

Supplemental Figure 9. Sea urchin (SpInsc), pencil urchin (EtInsc), and sea star (PmInsc) sequence alignment. The blue highlight indicates the LBD domain.