Agritrap project : Enhancing Buprestidae monitoring in Europe

Supplementary materials of the paper - detailed statistical analyses

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Contents

1 Introduction

This document contains the detailed analysis of our "Agritrap" project dataset. The report is entirely generated by R code (using Rmarkdown). If you want see the R code and reproduce our results, please refer to the R script with the same name as this document. All data and R code will be made available on publication in a public figshare repository : <https://doi.org/10.6084/m9.figshare.23982834> (DOI activated after publication)

Planed experimental design :

2021 :

- 5 sites per team (only Belgian teams in 2021)
	- **–** PC Fruit : pear Orchards
	- **–** CRA-W : Quercus forests
	- **–** Others = Quercus or Fagus forests/trees
- 3 tree per site with a combination of different traps on the same tree
- 4 type of traps :
	- **–** Bottle trap green
	- **–** Bottle trap green with decoys (most of the time on the same rope as the other bottle trap)
	- **–** MULTz yellow : on 2 trees only
	- **–** Multifunnel green : on 2 trees only

So 2 trees had a combination of 4 traps and 1 tree had only the two types of bottle traps. The multifunnel and MULTz arrived somewhat later in the season and it was not always possible to place them along with the bottle traps at the beginning of the season (however it Was OK for most of the sites with the highest abundance of Agrilus spp.)

2022 :

- 5 sites per team
	- **–** Belgium : same kind of distribution for the 4 Belgian partners
- 2 tree per site with a combination of different traps on the same tree (or very close)
- 6 types of traps :
	- **–** Fan-trap green
		- **–** Fan-trap green with decoys
		- **–** Fan-trap yellow
		- **–** Fan-trap yellow with decoys
		- **–** MULTz yellow
		- **–** Multifunnel green

There has been however some variations around this ideal protocol or some additions.

- Some Fan-traps were not yellow but "pale yellow" (see explanations below) \rightarrow these have been excluded from most analyses
- in some localities the samples from fan-traps with or without decoy were pooled together (called "Fan-traps mixed" in the dataset). This was an error made on the field caused by a communication problem between teams and personnel. This means that these samples are the sum of 2 traps and are not directly comparable to the other Fan-Traps. These traps samples were not used to test the effect of the decoys but were used with the other traps once the data from the traps with or without decoys has been pooled.
- ILVO tested a chemical lure on 3 multifunnel traps at 3 sites different sites \rightarrow these traps were not used in most analyses
- Some Belgian colleagues put also a few traps in Populus sites to see if it was possible to capture other species like Agrilus ater. The design was often less structured for these sites \rightarrow these data have been

excluded from most analyses.

- We sent Fan-Traps in Canada (team = RNCan) and France (Team = INRAE) where colleagues placed the fan-traps of different colors with or without decoys along with a variation of other traps commercial traps
- The French colleagues added some 3D decoys on some of the mutifunnel green traps. These traps have been excluded from most analyses because their low number made them not usable for global comparisons The decoys used on the Fan-Traps and Bottle traps where all dead A.planipennis

There were also some deviations from the planed design due to field or organisational constraints (some traps were destroyed or detached by the wind, some traps models were available later than expected, it was impossible to replace some traps when our stock was depleted, etc.).

NB: for the main statistical analyses and graphs, if a species has never been observed on a given tree in any of the traps, then the lines corresponding to this combination of treeID and burpestid species are removed from the dataset. This allows us to ignore the cases were a species is absent (the comparison between traps is useless in thi case). This means also that when we analyse the total number of buprestid caught, the trees with 0 captures are dropped.

The following table provides a summary of all the combinations of treatments, including the unusual variations that were most of the time excluded from the analyses.

 $NbIndiv = total number of individuals captured, $NbSites = number of sites \times bTraps = Number of traps,$$ NbSamples Number of samples (a sample is one catch in one trap during a certain period of time)

Another summary table showing only the main sites used in most analysis (eg excluding sites with 3Z lures, pale yellow traps, samples from Canada etc. . .) and grouping per country, year, tree species and type of trap. NbDays = average number of days each trap remained active on the field.

2 General overview of the data

2.1 Variability of total buprestidae captures between sites

NB : the "Vierzon" sites are from France and "Cranberry Lake" is the Canadian site.

2.1.1 Raw total catches per site

The following graph shows simply the sum of all individuals per site (without taking into account differences in trapping intensity). NB : Rochefort is the only site that has been sampled during the 2 years.

Figure 1:

2.1.2 Standardized total per trap and per site

Each point represent the captures in one trap standardized for a trapping duration of 90 days. The red dot and bars are the average and 95% bootstrap Confidence Interval. The scale is logarithmic !

Figure 2:

Conclusions

There is a very large variation between sites. In some sites we captured almost no Buprestidae probably because there were very few of them in that site. For pear orchards where the only captured species is Agrilus sinuatus, it is possible that this species in particular is not very much attracted by the traps.

It will be difficult to compare the traps types from sites where we had so few captures.

2.2 Specific composition

Full names of the Species

On request of one of the reviewers, we provide here the full names of the species, with their authority, as retrieved from the GBIF backbone taxonomy (through the taxize package).

2.2.1 Total number of individuals of different species per site

Comparison of the specific composition between countries. The Canadian fauna is of course completely different.

The French fauna is rather similar with a few additional species (A. curtulus, A. hastulifer,. . .). Agrilus sulcicollis seems to be much more frequent in Belgium.

Figure 3:

Same graph with only the European sites/fauna :

Figure 4:

2.2.2 Standardized number of individuals per trap and per species on different sites

Each point represent the number of individuals of a given species captured in a given trap (all trap types mixed).

Figure 5:

2.3 Specific composition per tree species

NB : the sampling effort is different between tree species. But the relative abundance can be compared between buprestid species within a given tree species.

Same graph with only the European sites/fauna :

Figure 6:

2.4 Maps

2.4.1 World map

Position of the sites in Belgium, France and Canada

2.4.2 Belgium

Map showing the distribution of the sites in Belgium for each tree species. All French sites were situated in 6 sites in the Vierzon forest and the Canadian samples came all from the same site near Cranberry Lake.

Figure 8:

Figure 9:

2.5 Summary table per site

the following table summarizes the data per site used to create these maps.

2.6 Conclusions

Because most species have been captured in very low quantities or in very few sites, most data items are "0" values and it will be difficult to exploit the data from these rare species (excepted in terms of species diversity).

We should probably limit ourselves to the 7 most common Agrilus species (all oak species) when we want a species by species analysis : "Agrilus sulcicollis", "Agrilus laticornis", "Agrilus angustulus", "Agrilus olivicolor", "Agrilus graminis", "Agrilus obscuricollis", "Agrilus biguttatus"

Overall, we captured 4814 Buprestidae specimens, including 4669 of the genus Agrilus. We collected 2220 samples from 382 traps spread over 46 sites in France and Belgium. On average, each trap remained active for 92.2 days. Most captures took place in June - July and declined in August. The number of catches varied a lot between sites (range of the number of Buprestidae captured per trap per 90 days: 0 to 400, median $= 0$, average $= 11.6$). Over the two years of study, the traps captured 23 different species (20 in Belgium and 12 in France, where the sampling effort was lower). The seven most abundant species (A.sulcicollis, A.laticornis, A.angustulus, A.olivicolor, A.graminis, A.obscuricollis, A.biguttatus) accounted for 92.7% of the total catches. In Belgium, the most abundant species was Agrilus sulcicollis with 1925 specimens captured (5.2 specimens/trap/90 days). This species occurred much less frequently in France (ranking 6th with 19 specimens captured, i.e. 1.03 specimens/trap/90 days). In France, A. laticornis was the most frequent species with 123 specimens (5.2 specimens/trap/90 days). The highest number of species was found in oak stands (21 species), while we found only Agrilus sinuatus in pear orchards. The catches were particularly poor in pear orchards: we only captured 17 specimens of Agrilus sinuatus in 2021 and no specimen at all in 2022 despite a confirmed presence by beating. For oak stands, the fauna from Belgium and France was quite similar, with a few species found only in traps from France (A. hastulifer, A. curtulus, Meliboeus fulgidicollis).

3 Main dataset: Comparison of the effect of different trap types on captures

3.1 Effect of the decoys on the number of catches

For 3 type of traps (bottle green, Fan-trap green, Fan-trap yellow), we had systematically the same trap at the same position (hanging on top of each other) with or without a decoy composed of a dead Agrilus planipennis.

However in certain sites the catches from the traps with or without decoy have been pooled and cannot be compared with the others.

This is why we are going to test here if there is an effect of the presence of the decoys. If not we will pool the catches of the same traps types with or without decoys for all sites.

The area of the bottle and Fan-traps being much smaller than the area of the multifunnel and MULTz the resulting comparisons will probably be more "fair"

3.1.1 Do we capture more Buprestidae on comparable traps when we add a decoy ?

On the following graph the gray lines represent the total captures of a pair of similar traps from the same tree with or without decoy. The number of individual has been standardized to have a comparable number of trapping days (90 days). The red dots represent the average and the bars represent the 95% confidence interval (bootstrap).

Figure 10:

We can formally test the hypothesis of differences between traps with or without decoy (controlling for the type of Trap and their interaction $+2$ random effects : Site and Tree) :

We use a linear mixed model with the $log(x+1)$ transformed number of catches $*$ 90 days / nbr of Days. ie we use the corrected number of catches per trap.

lmer(log1p(Nb_Indiv_90_Days) ~ Trap * Trap_Decoy + (1|SiteID) + (1|TreeID), data = tmp)

The residual plots of this model looks very coherent with a gaussian model.

Figure 11:

The Anova table shows no interaction trap x decoy nor any main decoy effect while the trap has a higly significant effect

> Table 5: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

Note in the summary the high value for the random component of the Site : there is a large between site variation. . .

```
## Linear mixed model fit by REML ['lmerMod']
## Formula: log1p(Nb_Indiv_90_Days) ~ Trap * Trap_Decoy + (1 | SiteID) + (1 | TreeID)
## Data: tmp
##
## REML criterion at convergence: 339.5
##
## Scaled residuals:
                  Median 3Q Max
## -2.27983 -0.49617 -0.04304 0.60247 1.84696
##
## Random effects:
## Groups Name Variance Std.Dev.
## TreeID (Intercept) 0.1925 0.4387
## SiteID (Intercept) 1.1570 1.0756
   Residual
## Number of obs: 134, groups: TreeID, 48; SiteID, 22
##
## Fixed effects:
## Estimate Std. Error t value
## (Intercept) 0.863045 0.377993 2.283
## TrapFan-trap Green -0.171676 0.408680 -0.420
## TrapFan-trap Yellow 0.882763 0.408680 2.160
## Trap_DecoyNo decoy
## TrapFan-trap Green:Trap_DecoyNo decoy 0.192915 0.273448 0.705
## TrapFan-trap Yellow:Trap_DecoyNo decoy -0.002426 0.273448 -0.009
```
Another possibility would be to use a Poisson Mixed model with the raw total numbers per trap with the log of the number of days as an offset. :

glmer(Nb_Indiv ~ Trap * Trap_Decoy + offset(log(Nb_Days)) + (1|SiteID) + (1|TreeID), data = tmp, family = "poisson")

The residual plots are a bit weirder but the overdispersion is reasonable (1.8). The conclusions are similar.

Figure 12:

Table 6: Analysis of Deviance Table (Type II Wald chisquare tests)

	Chisa	Df $\mathrm{Pr}(\geqslant$ Chisq)
Trap	546.50	0.00
Trap Decoy	0.79	0.37
Trap:Trap Decoy	5.96	0.05

3.1.2 Are some species more attracted by decoys than others ?

The results of the previous section could be masked by specific responses to the decoys : some species might be attracted while others are not. We test this hypothesis here we a subset of species for which we have enough data (on Belgian and French Quercus sites only).

On the following graph the gray lines represent the total captures of a pair of similar traps from the same tree with or without decoy. The number of individual has been standardized to have a comparable number of trapping days (90 days). The red dots represent the average and the bars represent the 95% confidence interval (bootstrap).

Figure 13:

We test these hypothesies with the following model : $lmer(log1p(Nb_Indiv_90_Days)$ - Trap * Trap_Decoy * Species + (1|SiteID) + (1|TreeID), data = tmp)

NB : the residual plots show that the distribution is not as good as when we pool all species. However a Poisson mixed model of that complexity seems to systematically crash and on the other end, the amount of data is reasonable (central limit theorem) and the results are very clear (p-values either very high or very low).

Figure 14:

Interpretation The Anova table shows no interaction trap x decoy x species nor decoy x species \rightarrow we have no evidence that the effect of the decoy could be different for some of the species.

We have highly significant Trap and Species main effect \rightarrow the number of captures are different between trap types (whatever the presence of decoys) and also different between species (some species are more often captured than others whatever the trap type).

We also have a highly significant Trap x Species interaction : this means that the differences between trap types are not of the same magnitude for the different species. From what we see on the graph : the differences between trap types for rare species like A. biguttatus seem to be very limited (they are close to 0 for all trap types) while there are clearly more catches in yellow fan traps for more common species.

	F	Df	Df.res	$Pr(\geq F)$
Trap	52.31	2	69.61	0.000000
Trap Decoy	0.28	1	383.13	0.597745
Species	30.49	6	398.53	0.000000
Trap:Trap Decoy	0.10	$\overline{2}$	383.13	0.908431
Trap:Species	3.21	12	401.27	0.000209
Trap Decoy:Species	0.45	6	383.13	0.845888
Trap:Trap Decoy:Species	0.23	12	383.13	0.996717

Table 7: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

3.1.3 Are males or females differently attracted by decoys ?

NB : the sex has been identified only on the catches from CRAW (Belgium) and INRAE (France). These sites are the ones with the most numerous captures so this is OK but it means also that we remove a lot of 0 values when we work on this subset of data. This could explain some differences in the averages computed here and in the other sections.

3.1.3.1 Species by species We can test if the sex has an influence on the captures for our 7 target species. We get a rather complex model with 4 fixed effects (and all interactions because we are precisely interested by these interactions) and 2 random effects (Site and Tree within the site) : lmer(log1p(Nb) \sim Trap * Trap_Decoy * Species * Sex + $(1|StteID)$ + $(1|TreeID)$, data = tmp_sex)

Again the residual plots show that the distribution of the residuals deviates slightly from the needed Gaussian but we can make the same remark as in the previous section (too complex for a Poisson model, central limit theorem $+$ results very clear).

Figure 15:

None of the effects including the sex or the decoys are significant. So we cannot find any evidence in favor of a difference of attractivity to decoys between the sexes.

	F	\mathbf{D} f	Df,res	$Pr(\geq F)$
Trap	91.69	$\overline{2}$	67.92	0.000000
Trap_Decoy	0.10	1	749.01	0.755756
Species	55.89	6	763.52	0.000000
Sex	0.34	1	749.01	0.562239
Trap:Trap Decoy	0.13	$\overline{2}$	749.01	0.880534
Trap:Species	7.55	12	768.65	0.000000
Trap Decoy:Species	0.69	6	749.01	0.656957
Trap:Sex	0.42	$\overline{2}$	749.01	0.654183
Trap Decoy:Sex	0.01	1	749.01	0.932030
Species:Sex	1.88	6	749.01	0.081325
Trap:Trap Decoy:Species	0.20	12	749.01	0.998453
Trap:Trap Decoy:Sex	0.50	$\overline{2}$	749.01	0.607947
Trap:Species:Sex	1.01	12	749.01	0.441526
Trap Decoy:Species:Sex	0.33	6	749.01	0.923664
Trap:Trap Decoy:Species:Sex	0.26	12	749.01	0.994181

Table 8: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

We can make a graphical representation of the data. Again no obiousvisual differnces between the sexes or between traps with or without decoys.

3.1.3.2 All species grouped Here we use the data for all species grouped by sex (only for the CRAW and INRAE datasets and for pairs of bottle of Fan-Traps with or without decoy).

On the following graph the gray lines represent the total captures of a pair of similar traps from the same tree with or without decoy. The number of individual has been standardized to have a comparable number of trapping days (90 days). The red dots represent the average and the bars represent the 95% confidence interval (bootstrap).

Figure 17:

We use the following gaussian mixed model : m <- lmm 1mer($log1p(Nb)$ \sim Trap $*$ Trap_Decoy $*$ Sex + (1|SiteID) + (1|TreeID), data = tmp_sex)

Again, none of the effects including the sex or the decoys are significant. So we cannot find any evidence in favor of a difference of attractivity to decoys between the sexes.

Table 9: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

3.1.4 Conclusions

We could not find any evidence of the effect of decoys on the number of captures. Therefore, we consider that we can safely group the catches of the pairs of traps with and without decoys for the rest of the analysis. So,from now on the traps labelled as fan-traps or bottle traps are the pooled data of the pairs of traps with and without decoys.

3.2 Effect of trap type on the number of catches

3.2.1 What is the effect of trap type on the total number of individuals caught ?

We use a mixed model of the form $=$ lmer(log1p(Nb_Indiv_90_Days) \sim Trap_Type $+$ Tree_Species $+$ Year + $(1|SteID) + (1|TreeID)$, data = tmp)

The response is the total number of individuals per trap *90 / Number of days of trapping $(\log(x+1))$ transformed). We use The Trap Type, Tree species (Fagus or Quercus) and Year as fixed effects and the SiteID and TreeID (nested in SiteID) as random effects.

NB : an equivalent Poisson model with number of trapping days as offset fails systematically. . .

Figure 19:

The Anova table shows a highly significant Trap type effect, and also a more marginal effect of the tree species (Fagus vs Quercus) but no year effect

	F.	Df	Df.res	$Pr(>\ F)$
Trap Type	12.01	4	110.29	0.000000
Tree Species	5.19		23.12	0.032223
Year	0.82		54.99	0.369305

In this model, the variance components of sites and tree random effects are rather high \rightarrow there is a lot of variation between sites but also between the trees on which the traps are placed

The following graph provides a graphical summary of the model.

The gray lines represent traps from the same tree. In red : the average and 95% bootstrap Confidence Interval. The compact letter display shows which pairwise comparisons are significantly different from each other (with p-value correction). The fan-traps or bottle traps with and without decoys have been pooled together.

Note that there is no significant difference between the bottle traps and the fan-traps but this could be due to the fact that these two type of traps where not present the same year so it is difficult to compare them with confidence

Figure 20:

Here is the detail of the pairwise comparisons :

```
##
## Simultaneous Tests for General Linear Hypotheses
##
## Multiple Comparisons of Means: Tukey Contrasts
##
##
## Fit: lmer(formula = log1p(Nb_Indiv_90_Days) ~ Trap_Type + Tree_Species +
## Year + (1 | SiteID) + (1 | TreeID), data = tmp)
##
## Linear Hypotheses:
## Estimate Std. Error z value Pr(>|z|)
## Fan-trap yellow - Fan-trap green == 0 0.9526 0.2027 4.700 < 0.001 ***
## MULTz yellow - Fan-trap green == 0 1.2042 0.1977 6.090 < 0.001 ***
\# Multifunnel green - Fan-trap green == 0 0.7593 0.1961 3.871
## Bottle green - Fan-trap green == 0 0.3095 0.3028 1.022 0.83789
## MULTz yellow - Fan-trap yellow == 0 0.2517 0.1936 1.300 0.67984
## Multifunnel green - Fan-trap yellow == 0.
## Bottle green - Fan-trap yellow == 0 -0.6430 0.3002 -2.142 0.19316
## Multifunnel green - MULTz yellow == 0 -0.4450 0.1668 -2.667 0.05566 .
## Bottle green - MULTz yellow == 0 -0.8947 0.2558 -3.498 0.00404 **<br>## Bottle green - Multifunnel green == 0 -0.4497 0.2611 -1.722 0.40681
## Bottle green - Multifunnel green == 0 -0.4497 0.2611 -1.722 0.40681
#### Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Adjusted p values reported -- single-step method)
```
Note also that there are plenty of sites where we have very few individuals and this probably tend to decrease artificially the differences between traps because if there are really very few individuals present the traos will be very limited in the total captures possible. (NB : remember that we have already removed all trees in which no individuals of each species were captured).

This is for example what we obtain if we keep only the trees with more than 5 individuals captured (a rather arbitrary choice). The results are similar if we chose other arbitrary thresholds like 3, 5, 7 or 10.

Now the Hungarian MULTz traps have significantly more captures than the multifunnel green while the yellow fan-traps are intermediate (no significant difference between fan-traps yellow and multifunnel green or

MULTz). The green fan-traps have significantly less captures than these 3 type of traps. The green bottle traps captures are only significantly lower than the MULTz. However we had very few traps with captures in bottle traps and the fan traps have been placed on the field during a different year which makes the comparison not totally fair.

Figure 21:

Detail of the pairwise comparisons with p-value correction :

```
##
     Simultaneous Tests for General Linear Hypotheses
##
## Multiple Comparisons of Means: Tukey Contrasts
##
##
## Fit: lmer(formula = log1p(Nb_Indiv_90_Days) ~ Trap_Type + Tree_Species +
## Year + (1 | SiteID) + (1 | TreeID), data = tmp)
##
## Linear Hypotheses:
## Estimate Std. Error z value Pr(>|z|)
## Fan-trap yellow - Fan-trap green == 0 1.2662
## MULTz yellow - Fan-trap green == 0 1.6356 0.2651 6.169 < 0.001 ***
## Multifunnel green - Fan-trap green == 0 1.0165 0.2618 3.882 < 0.001 ***
## Bottle green - Fan-trap green == 0 0.1974 0.4788 0.412 0.99341
## MULTz yellow - Fan-trap yellow == 0
## Multifunnel green - Fan-trap yellow == 0 -0.2498 0.2361 -1.058 0.81715<br>## Bottle green - Fan-trap yellow == 0 -1.0688 0.4650 -2.299 0.13488
## Bottle green - Fan-trap yellow == 0 -1.0688 0.4650 -2.299 0.13488
## Multifunnel green - MULTz yellow == 0 -0.6191 0.2153 -2.876 0.02979 *<br>## Bottle green - MULTz yellow == 0 -1.4382 0.4277 -3.363 0.00631 **
\## Bottle green - MULTz yellow == 0 -1.4382 0.4277 -3.363<br>\## Bottle green - Multifunnel green == 0 -0.8191 0.4277 -1.915
## Bottle green - Multifunnel green == 0 -0.8191 0.4277 -1.915 0.29238
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Adjusted p values reported -- single-step method)
```
3.2.2 What is the relative importance of trap type, site and tree to explain the number of catches ?

We use a model with only random effects for an easy decomposition of the variance. We use the log of the number of individuals per 90 days as response and the trap type, site and tree as random effects.

The variation between sites is by far more imporrtant than the variation between trees or between trap types


```
## Linear mixed model fit by REML ['lmerMod']
## Formula: log1p(Nb_Indiv_90_Days) ~ (1 | Trap_Type) + (1 | SiteID) + (1 | TreeID)
## Data: tmp
##
## REML criterion at convergence: 458.9
##
## Scaled residuals:
                    Median 3Q Max
## -2.11972 -0.64660 -0.04528 0.56909 3.06772
##
## Random effects:
## Groups Name Variance Std.Dev.
            (Intercept) 0.1579
## SiteID (Intercept) 1.6983 1.3032
## Trap_Type (Intercept) 0.2251 0.4744
## Residual 0.5658 0.7522
## Number of obs: 158, groups: TreeID, 54; SiteID, 26; Trap_Type, 5
##
## Fixed effects:
## Estimate Std. Error t value
## (Intercept) 1.7467 0.3444 5.071
```
3.2.3 What is the impact of the area of the trap ?

The traps used have rather different catching areas and we can suppose that this can have an impact on the number of catches. However we have only limited ways to test explicitly this hypothesis with our dataset. To do that we would need similar trap types with varying areas in our experimental setup. We can standardize the catches by the area of the trap but by doing that we would "force" or "impose" on our data a linear effect of the area without really knowing its real effect. So this section is just showing what happens to our results when we correct of the trap size but this should not be used as is.

Dimensions of the traps :

- Bottle traps \sim trapeze with 15cm base x 8cm base x 15 cm height x 2 sides x 2 pairs of traps on top of each other $= 690 \text{ cm}^2$
- Fan-traps = $15x20$ cm rectangle x 2 sides x 2 pairs of traps hanging on top of each other = 1200 cm²
- MULTz traps: $23x15$ cm x 2 sides x 4 pieces = 2760 cm²
- Multifunnel green traps: they are a cylinder but we will consider a projection as if they were "flat" –> 19 cm diameter x 88cm height (11 pieces x 8cm) and x 2 sides $=$ 3344 cm²

NB : Imrei et al 2020 DOI: 10.1093/forestry/cpz071. used an area of 11 160 cm2 for 12 funnel of Lindgren funnel traps

We use a statistical analysis approach identical to the previous section. We just divided the catches by the area of the trap and then multiplied that value by 2000 cm^2 to have a common trapping area reference

The Anova table shows a highly significant Trap type effect and of the tree species (Fagus vs Quercus) but no year effect

Table 12: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

	F.	Df	Df.res	$Pr(>\ F)$
Trap Type Tree Species	9.51 5.36	л.	23.15	4 110.18 0.000001 0.029868
Year	0.19		54.63	0.666615

The following graph provides a graphical summary of the model.

Now the yellow traps are capturing significantly more than the other types. The yellow fan traps capture the most but these catches are not significantly different from the MULTz.

Figure 24:

Here is the detail of the pairwise comparisons :

```
##
     Simultaneous Tests for General Linear Hypotheses
##
## Multiple Comparisons of Means: Tukey Contrasts
##
##
## Fit: lmer(formula = log1p(Nb_Indiv_90_Days) ~ Trap_Type + Tree_Species +
## Year + (1 | SiteID) + (1 | TreeID), data = tmp)
##
## Linear Hypotheses:
## Estimate Std. Error z value Pr(>|z|)
## Fan-trap yellow - Fan-trap green == 0 1.0213 0.2038 5.012 < 0.001 **<br>## MULTz yellow - Fan-trap green == 0 0.6778 0.1988 3.409 0.00559 **
## MULTz yellow - Fan-trap green == 0 0.6778 0.1988 3.409 0.00559<br>## Multifunnel green - Fan-trap green == 0 0.1064 0.1972 0.539 0.98219
## Multifunnel green - Fan-trap green == 0 0.1064 0.1972 0.539 0.98219
## Bottle green - Fan-trap green == 0## MULTz yellow - Fan-trap yellow == 0 -0.3435 0.1946 -1.765 0.38129
## Multifunnel green - Fan-trap yellow == 0 -0.9149 0.1930 -4.740<br>## Bottle green - Fan-trap yellow == 0 -0.7701 0.3017 -2.552
## Bottle green - Fan-trap yellow == 0 -0.7701 0.3017 -2.552 0.07512.
## Multifunnel green - MULTz yellow == 0 -0.5715 0.1677 -3.408 0.00557 **
## Bottle green - MULTz yellow == 0## Bottle green - Multifunnel green == 0 0.1449 0.2625 0.552 0.98060
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Adjusted p values reported -- single-step method)
```
3.2.4 Are some species more attracted by certain types of traps ?

We use a mixed model of the form $=$ lmer(log1p(Nb Indiv 90 Days) ~ Trap Type*Species + $(1|SiteID) + (1|TreeID)$, data = tmp)

The response is the total number of individuals of each species per trap *90 / Number of days of trapping $(\log(x+1)$ transformed). We use The Trap Type, Agrilus species and their interaction as fixed effects and the SiteID and TreeID as random effects. The tree species was not included here because we kept only the Quercus sites from the dataset (beacuse the target species are living mostly on Quercus). The year was excluded to simplify the model

NB : an equivalent Poisson model with number of trapping days as offset fails systematically. . .

Figure 25:

The Anova table shows a highly significant Trap type x species effect. This means that there are differences in the amount of captures between trap types but that these differences are not the same depending on the Agrilus species.

The following graph provides a graphical summary of the model.

On the following graph the gray lines represent the total captures for each species in a given trap from the same tree. So the lines are relating directly comparable captures from different types of traps. The number of individuals has been standardized to have a comparable number of trapping days (90 days). The red dots represent the average and the bars represent the 95% confidence interval (bootstrap).

Trap types sharing the same letter are not significantly different. The lettres should only be compared within a species (you cannot compare the trap types between species using these lettres). To obtain these letters we recomputed a separate mixed model for each species then performed all pairwise comparisons with p-value corrections (for multiple testing). We used this approach because it is much more easy to obtain the letters display in an automated manner with this approach. Some people might however criticize this approach (see below).

Interpretation : The differences between trap types are more or less similar for the 6 most common species but A. biguttatus show a slightly different pattern. For the 6 most common species, the captures tend to be more abundant in the 2 yellow traps (Fan-Traps and MUTLTz) and lower in the green fan-traps and to a lower extend in the multifiunnel green ones. The captures in the green bottle traps are often very low but most of the time the differences with the other trap types is not significant (probably due to several reasons : the number of replicates is low, they were never places in the same sites as the Fan-Traps,. . .). The differences between traps tend to be less obvious as the global abundance of the species tend to decrease.

In A. biguttatus however the highest captures occured in the multifunnel green trap and the green and yellow Fan-traps captured significantly less individuals, while the difference with the yellow MULTz is not significant.

When we compute the pairwise comparisons with a separate model for each species, the p-value correction is done only for these 10 pairwise comparison within each species while in reality these 10 comparisons are repeated for the 7 species.

It is possible to write manually a contrast matrix comparing the traps within each species and based on the full model with Trap_type * Species as fixed effects. In that case the p-value correction is done on all 70 pairwise comparisons together and some people might consider this as "more correct". The higher the number of comparison the stronger the p-value correction and the lower the statistical power.

One should keep in mind that the choice of the p-value correction method (here a "Tukey like" single-step method) is rather arbitrary and choosing other methods will provide other results. The choice is always a matter of which kind of error you favor : do you prefer more false negative (type II error) or more false positives (type I error) ? For example, with a Bonferoni correction, the correction will be even stronger and you will have more false negatives. So, in the end, the previous approach with a separate model for each

species might be considered as a reasonable compromise in my opinion and I think that few people with bother to tediously build manually a 35 x 70 contrast matrix (as I did below) with a relatively high risk of errors. . .

The following table shows the results of all 70 pairwise comparisons computed with a manually build contrast matrix. It could be possible to derive a compact letter display from this table but this is rather tedious and must be done entirely manually.

You can see that with this approach we "lost" all significant differences for the 4 most rare species : A.olivicolor, graminis, obscuricollis, biguttatus.

```
##
## Simultaneous Tests for General Linear Hypotheses
##
## Fit: lmer(formula = log1p(Nb_Indiv_90_Days) ~ -1 + Trap_Type:Species +
\# (1 | SiteID) + (1 | TreeID), data = tmp)
##
## Linear Hypotheses:
## Estimate Std. Error z value Pr(>|z|)
## A.sulcicollis : Fan-trap green - Fan-trap yellow == 0
## A.sulcicollis : Fan-trap green - MULTz yellow == 0 -1.235100 0.284896 -4.335 <0.01 ***
## A.sulcicollis : Fan-trap green - Multifunnel green == 0 -0.290217 0.286630 -1.013 1.0000
## A.sulcicollis : Fan-trap green - Bottle green == 0 -0.232114 0.369969 -0.627 1.0000
## A.sulcicollis : Fan-trap yellow - MULTz yellow == 0 -0.007103 0.283543 -0.025 1.0000
## A.sulcicollis : Fan-trap yellow - Multifunnel green == 0
## A.sulcicollis : Fan-trap yellow - Bottle green == 0 0.995883 0.369198 2.697 0.3075
## A.sulcicollis : MULTz yellow - Multifunnel green == 0 0.944883 0.255374 3.700 0.0131 *
## A.sulcicollis : MULTz yellow - Bottle green == 0.
## A.sulcicollis : Multifunnel green - Bottle green == 0 0.058103 0.339309 0.171 1.0000
## A.laticornis : Fan-trap green - Fan-trap yellow == 0 -1.159831 0.274511 -4.225 <0.01 **
## A.laticornis : Fan-trap green - MULTz yellow == 0 -1.501973 0.256962 -5.845 <0.01 ***<br>## A.laticornis : Fan-trap green - Multifunnel green == 0 -0.644206 0.256962 -2.507 0.4586
## A.laticornis : Fan-trap green - Multifunnel green == 0 -0.644206 0.256962 -2.507 0.4586
## A.laticornis : Fan-trap green - Bottle green == 0 -0.203260 0.376509 -0.540 1.0000
## A.laticornis : Fan-trap yellow - MULTz yellow == 0 -0.342142 0.257891 -1.327 0.9995
## A.laticornis : Fan-trap yellow - Multifunnel green == 0 0.515625 0.257891 1.999 0.8732
## A.laticornis : Fan-trap yellow - Bottle green == 0 0.956572 0.377495 2.534 0.4360
## A.laticornis : MULTz yellow - Multifunnel green == 0 0.857767 0.234174 3.663
## A.laticornis : MULTz yellow - Bottle green == 0 1.298714 0.354271 3.666 0.0148 *
## A.laticornis : Multifunnel green - Bottle green == 0 0.440947 0.354271 1.245 0.9998
## A.angustulus : Fan-trap green - Fan-trap yellow == 0 -1.231383 0.322761 -3.815 <0.01 **
## A.angustulus : Fan-trap green - MULTz yellow == 0 -2.004215 0.301027 -6.658 <0.01 ***
## A.angustulus : Fan-trap green - Multifunnel green == 0 -0.761689 0.303014 -2.514 0.4544
## A.angustulus : Fan-trap green - Bottle green == 0 -0.443340 0.430128 -1.031 1.0000
## A.angustulus : Fan-trap yellow - MULTz yellow == 0 -0.772832 0.293207 -2.636
## A.angustulus : Fan-trap yellow - Multifunnel green == 0 0.469694 0.295213 1.591 0.9911
## A.angustulus : Fan-trap yellow - Bottle green == 0 0.788043 0.425090 1.854 0.9397
## A.angustulus : MULTz yellow - Multifunnel green == 0 1.242526 0.266546 4.662 <0.01 ***
## A.angustulus : MULTz yellow - Bottle green == 0 1.560875 0.397063 3.931 <0.01 **
## A.angustulus : Multifunnel green - Bottle green == 0 0.318349 0.401618 0.793<br>## A.olivicolor : Fan-trap green - Fan-trap yellow == 0 0.607829 0.342260 -1.776
## A.olivicolor : Fan-trap green - Fan-trap yellow == 0 -0.607829 0.342260 -1.776 0.9628
## A.olivicolor : Fan-trap green - MULTz yellow == 0 -1.110158 0.332972 -3.334 0.0488 *
## A.olivicolor : Fan-trap green - Multifunnel green == 0 -0.604265 0.332972 -1.815 0.9523
## A.olivicolor : Fan-trap green - Bottle green == 0 -0.383248 0.590154 -0.649 1.0000
## A.olivicolor : Fan-trap yellow - MULTz yellow == 0
## A.olivicolor : Fan-trap yellow - Multifunnel green == 0 0.003563 0.319345 0.011 1.0000
## A.olivicolor : Fan-trap yellow - Bottle green == 0 0.224581 0.582988 0.385 1.0000
## A.olivicolor : MULTz yellow - Multifunnel green == 0 0.505893 0.305853<br>## A.olivicolor : MULTz yellow - Bottle green == 0 0.726910 0.573958
## A.olivicolor : MULTz yellow - Bottle green == 0 0.726910 0.573958 1.266 0.9998
## A.olivicolor : Multifunnel green - Bottle green == 0 0.221017 0.573958 0.385 1.0000
## A.graminis : Fan-trap green - Fan-trap yellow == 0 -0.891825 0.328448 -2.715 0.2944
## A.graminis : Fan-trap green - MULTz yellow == 0 -0.693701 0.317001 -2.188 0.7408
## A.graminis : Fan-trap green - Multifunnel green == 0 -0.457971 0.317001 -1.445 0.9979
## A.graminis : Fan-trap green - Bottle green == 0 -0.561225 0.586827 -0.956 1.0000
## A.graminis : Fan-trap yellow - MULTz yellow == 0 0.198124 0.303456 0.653 1.0000
## A.graminis : Fan-trap yellow - Multifunnel green == 0 0.433855 0.303456 1.430 0.9983
## A.graminis : Fan-trap yellow - Bottle green == 0 0.330600 0.579997 0.570 1.0000
## A.graminis : MULTz yellow - Multifunnel green == 0 0.235730 0.289308 0.815 1.0000
## A.graminis : MULTz yellow - Bottle green == 0 0.132476 0.567672 0.233 1.0000
## A.graminis : Multifunnel green - Bottle green == 0 -0.103254 0.567672 -0.182 1.0000
## A.obscuricollis : Fan-trap green - Fan-trap yellow == 0 -0.840495 0.353429 -2.378 0.5745
```


3.2.5 Are males or females differently attracted by certain type of traps ?

3.2.5.1 Species by species We use the following model to test for the effect of sex : lmer(log1p(Nb)) \sim Trap_Type * Species * Sex + (1|SiteID) + (1|TreeID), data = tmp_sex)

Figure 27:

All effects involving the sex are not significant except for Species x Sex. In theory this would mean that there are differences in captures between males and females depending on the Agrilus species (but independent of the Trap Type). However when comparing males vs females catches for all combinations of Species and trap type (with p-value correction for multiple testing, see below), none are significantly different.

So we found no evidence that one type of trap might attract more the males or the females.

F	Df	Df.res	$Pr(\geq F)$
56.43	4	776.80	0.000000
50.28	6	830.21	0.000000
0.76	1	814.42	0.384891
4.05	24	824.47	0.000000
1.30	$\overline{4}$	814.42	0.266531
3.06	6	814.42	0.005708
0.81	24	814.42	0.720595

Table 14: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

Graphical visualization of the data

Figure 28:

Multiple comparisons :

```
##
    Simultaneous Tests for General Linear Hypotheses
##
## Fit: lmer(formula = log1p(Nb) ~ -1 ~ + ~ Sex:Trap\_Type: Species ~ + (1 ~ |## SiteID) + (1 | TreeID), data = tmp_sex)
##
## Linear Hypotheses:
## Estimate Std. Error z value Pr(>|z|)
## A.sulcicollis - Fan-trap green : Female - Male == 0 -0.153065 0.301084 -0.508 1.000
## A.sulcicollis - Fan-trap yellow : Female - Male == 0 0.142421 0.301084 0.473 1.000
## A.sulcicollis - MULTz yellow : Female - Male == 0 -0.382334 0.242742 -1.575 0.986
## A.sulcicollis - Multifunnel green : Female - Male == 0 -0.315277 0.242742 -1.299 0.999
## A.sulcicollis - Bottle green : Female - Male == 0 -0.166311 0.343289 -0.484 1.000
## A.laticornis - Fan-trap green : Female - Male == 0 -0.105783 0.271394 -0.390 1.000
## A.laticornis - Fan-trap yellow : Female - Male == 0 -0.494489 0.280294 -1.764<br>## A.laticornis - MULTz yellow : Female - Male == 0 -0.372152 0.231445 -1.608<br>## A.laticornis - Multifunnel green : Female - Male == 0 0.0949
## A.laticornis - MULTz yellow : Female - Male == 0 -0.372152 0.231445 -1.608 0.982<br>## A.laticornis - Multifunnel green : Female - Male == 0 0.094953 0.231445 0.410 1.000
## A.laticornis - Multifunnel green : Female - Male == 0 0.094953 0.231445 0.410 1.000
## A.laticornis - Bottle green : Female - Male == 0 0.213335 0.361858 0.590 1.000
## A.angustulus - Fan-trap green : Female - Male == 0 0.218391 0.301084 0.725 1.000
## A.angustulus - Fan-trap yellow : Female - Male == 0 -0.374768 0.290132 -1.292 1.000
## A.angustulus - MULTz yellow : Female - Male == 0 -0.676265 0.249048 -2.715 0.207
## A.angustulus - Multifunnel green : Female - Male == 0 0.135758 0.249048 0.545 1.000
## A.angustulus - Bottle green : Female - Male == 0 -0.106976 0.443184 -0.241 1.000
## A.olivicolor - Fan-trap green : Female - Male == 0 0.178190 0.327313 0.544 1.000
## A.olivicolor - Fan-trap yellow : Female - Male == 0 0.524092 0.301084 1.741 0.949
## A.olivicolor - MULTz yellow : Female - Male == 0
## A.olivicolor - Multifunnel green : Female - Male == 0 0.063142 0.280294 0.225 1.000
## A.olivicolor - Bottle green : Female - Male == 0 0.480276 0.626757 0.766 1.000
## A.graminis - Fan-trap green : Female - Male == 0 0.085341 0.301084 0.283 1.000
## A.graminis - Fan-trap yellow : Female - Male == 0 0.459480 0.280294 1.639 0.976
## A.graminis - MULTz yellow : Female - Male == 0
## A.graminis - Multifunnel green : Female - Male == 0 0.573887 0.255872 2.243 0.586
## A.graminis - Bottle green : Female - Male == 0 0.653365 0.626757 1.042 1.000
## A.obscuricollis - Fan-trap green : Female - Male == 0 0.119590 0.313378 0.382 1.000
## A.obscuricollis - Fan-trap yellow : Female - Male == 0 -0.232419 0.290132 -0.801 1.000
## A.obscuricollis - MULTz yellow : Female - Male == 0.
## A.obscuricollis - Multifunnel green : Female - Male == 0 0.009936 0.271394 0.037 1.000
## A.obscuricollis - Bottle green : Female - Male == 0 0.000000 0.767617 0.000 1.000
## A.biguttatus - Fan-trap green : Female - Male == 0.
## A.biguttatus - Fan-trap yellow : Female - Male == 0 0.367406 0.361858 1.015 1.000
## A.biguttatus - MULTz yellow : Female - Male == 0 -0.159298 0.327313 -0.487 1.000
## A.biguttatus - Multifunnel green : Female - Male == 0 -0.745248 0.327313 -2.277 0.554
## A.biguttatus - Bottle green : Female - Male == 0 -0.320927 0.767617 -0.418 1.000
## (Adjusted p values reported -- single-step method)
```
3.2.5.2 All species grouped We compare here the captures between the sexes of all species grouped (only for the CRAW and INRAE samples, ie only in Quercus sites, because sex is not available for other sub datasets).

We use a mixed model of the form $=$ lmer(log1p(Nb_Indiv_90_Days) ~ Trap_Type $*$ Sex+ (1|SiteID) + (1|TreeID), data = tmp)

The response is the total number of individuals per trap *90 / Number of days of trapping $(\log(x+1))$ transformed). We use The Trap Type, Sex and their interaction as fixed effects and the SiteID and TreeID as random effects.

Figure 29:

The Anova table shows no significant effect of the sex nor the interaction of the tree species (Fagus vs Quercus) but no year effect

	F	Df	Df.res	
Trap Type	30.57			4 145.72 0.000000
Sex	1.33	-1.	130.46	0.250743
Trap Type:Sex	0.77		4 130.46	0.547311

Table 15: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

The following graph provides a graphical summary of the model.

The gray lines represent traps from the same tree. In red : the average and 95% bootstrap Confidence Interval.

Figure 30:

3.3 Effect of trap type on number of species caught

3.3.1 Number of species vs number of individuals or trapping duration

There is a clear asymptotic between the number of species and the number of individuals captured.

Figure 31:

With a log scale on the x axis the relation becomes linear...

Figure 32:

The pearson correlation coefficient value is large. NB the test here is not strictly speaking valid because the traps are not independent (grouped by sites, tree,...)

```
##
    Pearson's product-moment correlation
##
## data: log1p(tmp$Nb_Indiv) and tmp$Nb_Sp
## t = 34.244, df = 156, p-value < 2.2e-16
## alternative hypothesis: true correlation is not equal to 0
## 95 percent confidence interval:
## 0.9179765 0.9554486
## sample estimates:
## cor
## 0.9394609
```
We could fit a linear mixed model but this is a bit overkill as the correlation is very obvious here... NB the random component of the tree is estimated to be 0 and causes boundary warnings \rightarrow removed from this model.

```
## Linear mixed model fit by REML ['lmerMod']
\text{#}\# Formula: Nb_Sp ~ log1p(Nb_Indiv) + (1 | SiteID)<br>\text{#}\# Data: tmp
     Data: tmp
##
## REML criterion at convergence: 443
##
## Scaled residuals:
               1Q Median 3Q Max
## -2.4192 -0.4368 0.0920 0.1892 3.9538
##
## Random effects:
## Groups Name Variance Std.Dev.
## SiteID (Intercept) 0.05246 0.2290
## Residual 0.89046 0.9436
## Number of obs: 158, groups: SiteID, 26
##
## Fixed effects:
                  Estimate Std. Error t value
## (Intercept) -0.11613 0.12911 -0.899
## \log 1p(Nb_Indiv) 1.52998
##
  Correlation of Fixed Effects:
## (Intr)
## lg1p(Nb_In) -0.723
## Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)
##
## Response: Nb_Sp
                       F Df Df.res Pr(>F)
## log1p(Nb_Indiv) 853.73 1 36.321 < 2.2e-16 ***
#### Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```
But there is no clear correlation with the number of trapping days.

Figure 33:

3.3.2 Number of species vs trap type

We use a mixed model of the form $=$ lmer(log1p(Nb_Sp) \sim Trap_Type + Tree_Species + Year + $(1|SiteID) + (1|TreeID), data = tmp)$

The response is the total number of species per trap $(log(x+1)$ transformed). We use The Trap Type, Tree species (Fagus or Quercus) and Year as fixed effects and the SiteID and TreeID as random effects.

NB : an equivalent Poisson model with number of trapping days as offset fails systematically. . .

Figure 34:

The Anova table shows a highly significant Trap type effect, and also a more marginal effect of the tree species (Fagus vs Quercus) but no year effect

Table 16: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

The following graph provides a graphical summary of the model.

The gray lines represent traps from the same tree. In red : the average and 95% bootstrap Confidence Interval. The compact letter display shows which pairwise comparisons are significantly different from each other (with p-value correction).

The green fantraps captured less species on average than the yellow fan-traps, MULTz and Multifunnel green. Note that there is no significant difference between the bottle traps and the fan-traps but this could be due to the fact that these two type of traps where not present the same year so it is difficult to compare them with confidence

Figure 35:

Here is the detail of the pairwise comparisons :

```
##
## Simultaneous Tests for General Linear Hypotheses
##
## Multiple Comparisons of Means: Tukey Contrasts
##
##
## Fit: lmer(formula = log1p(Nb_Sp) ~ Trap_Type + Tree_Species + Year +
\texttt{##} (1 | SiteID) + (1 | TreeID), data = tmp)
##
## Linear Hypotheses:
## Estimate Std. Error z value Pr(>|z|)
## Fan-trap yellow mixed - Fan-trap green mixed == 0 0.32982 0.10410 3.168 0.01248 *<br>## MULTz yellow - Fan-trap green mixed == 0 0.38048 0.10150 3.748 0.00156 **## MULTz yellow - Fan-trap green mixed == 0 0.38048 0.10150 3.748 0.00156 *<br>## Multifunnel green - Fan-trap green mixed == 0 0.31784 0.10070 3.156 0.01294 *## Multifunnel green - Fan-trap green mixed == 0 0.31784 0.10070 3.156 0.01294<br>## Bottle green mixed - Fan-trap green mixed == 0 -0.03965 0.15565 -0.255 0.99901
## Bottle green mixed - Fan-trap green mixed == 0## MULTz yellow - Fan-trap yellow mixed == 0 0.05066 0.10041 0.505 0.98613
## Multifunnel green - Fan-trap yellow mixed == 0 -0.01198 0.09962 -0.120 0.99995<br>## Bottle green mixed - Fan-trap yellow mixed == 0 -0.36947 0.15495 -2.385 0.11330
## Bottle green mixed - Fan-trap yellow mixed == 0 -0.36947## Multifunnel green - MULTz yellow == 0 -0.06264 0.08650 -0.724 0.94819
## Bottle green mixed - MULTz yellow == 0
## Bottle green mixed - Multifunnel green == 0 -0.35749 0.13452 -2.658 0.05712.
## ---
## Signif. codes: 0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
## (Adjusted p values reported -- single-step method)
```
Summary table with the average number of species caotured per trap type :

3.4 Effect of trap type on the probability of detection of each species

Instead of working with the abundance, we use the presence/absence of each species in each trap over the whole season.

We use a generalized linear mixed model with binomial distribution of the form $=$ glmer(Presence \sim Trap_Type*Species + (1|SiteID) + (1|TreeID) ,family = binomial)

We had convergence problems. So the solution we found was 1) to remove the bottle traps form the analysis and 2) change the optimizer and number of iterations

Note that the sampling effort was not uniform for all traps so we should ideally add the number of trapping days as covariate. But when we do that, the model never converges. So we didn't include this co-variate. The previous graphs have shown that the number of sapling days does not seem to be correlated with number of species anyways.

Figure 36:

The analysis of deviance table shows highly significant Trap type and species main effects but no interaction. This results tends to mean that some traps have higher probability to capture the species present and that some of the species are more frequent. But the difference between traps does not depend on the species

The flowing graph shows a summary of this model with all pairwise comparisons between traps whatever the species. Each gray line corresponds to one of the 7 most common species. The values are the % of traps in which each of these species has been detected (present at least once during the whole season).

NB : the bottle traps have no letter because we had to remove this treatment from the analysis to avoid model convergence problems. . .

Figure 37:

If we perform all pairwise comparison between traps separately for each species most comparisons are not significant anymore but this is most likely due to a lack of statistical power relative to the grouped model and this is not really necessary because the trap type x species interaction was not significant.

Figure 38:

3.5 How robust are our conclusions if we change the statistical approach or the data ?

Reviewer#1 of our manuscript was worried by the fact that our experimental design was an incomplete randomized bloc design (i.e. not all trap types were present in each tree at each period). He/she also suggested that it might be better to use models with Poisson distribution.

Our dataset is complex and there are plenty of valid ways to analyse it. The most important question is to understand if a different statistical approach would change the general conclusions of the paper.

We perform in this section a robustness analysis : we change the statistical approach, use a subset of the full dataset and see how much it impacts our conclusions.

As suggested by the review we used a subset of the data, keeping only the trees and the dates in which all trap types available for a given year were present (ie we dropped all the trees and dates for which we had missing data). Then we applied various statistical approaches : repeating the analysis of the paper, dropping the bottle traps, performing a separate analysis for each year,. . . We also replicated the analysis with Poisson or Negative binomial models when possible (these more complex models struggle to converge in some situations).

tl;dr : our conclusions are robust to the statistical approach and to the subset of the data used.

3.5.1 Gaussian Mixed Model on a subset of the data with only the complete blocs

3.5.1.1 Total number of individuals caught .

This is just a replicate of th analysis of the paper but on a subset of the data. We use a mixed model of the form $=$ lmer(log1p(Nb_Indiv_90_Days) ~ Trap_Type + Tree_Species + Year + (1|SiteID) + $(1|TreeID)$, data = tmp)

The residual plots show that a gaussian model is a very good approximation of this data

Figure 39:

The Anova table shows a highly significant Trap type effect, and also a more marginal effect of the tree species (Fagus vs Quercus) but no year effect

	Table 19: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger		
df)			

The following graph provides a graphical summary of the model.

3.5.1.2 Total number of individuals caught without the bottle traps .

What happens if we drop the bottle traps ?

NB : the comparison between the bottle traps and the fan-traps has higher uncertainty because these traps were never present in the same year and same blocs at the same time. Also the bottle traps captured very few individuals, so we check here what happens if we drop them from the pairwise comparisons.

	F.	Df	$_{\rm Df,res}$	$Pr(>\)$
Trap Type	10.31	3	73.21	0.000010
Tree Species Year	-3.17 0.01	\mathbf{L} 1.	15.47 32.94	0.094792 0.922441

Table 20: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

Figure 41:

 $->$ same results

3.5.1.3 Total number of individuals caught and each year analysed separately .

OK but what happens if we analyse both years separately in order to avoid having both the bottle traps and fan-traps involved in the pairwise comparisons while these type of traps were placed only in different years and different sites ?

Figure 42:

3.5.2 Poisson/Negative binomial model on a subset of the data with only the complete blocs

Instead of a gaussian model on log transformed counts standardized on 90 days, we could use a Poisson model on the raw counts, with an offset on the log number of trapping days. With the full dataset this kind of model was systematically failing (convergence problem etc). Here we managed to obtain a converging model but only after removing the "year" effect.

3.5.2.1 Total number of individuals caught The model has the following structure :

glmer(Nb_Indiv ~ Trap_Type + Tree_Species + (1|SiteID) + (1|TreeID) + offset(log(Nb_Days)), family = poisson) However the overdesipersion is rather high : 4.85

We also managed to fit with this subset of the data a negative binomial model which incorporates an overdispersion parameter. The model has the following form :

```
glmer.nb(Nb_Indiv ~ Trap_Type + Tree_Species + (1|SiteID) + (1|TreeID) + offset(log(Nb_Days)))
```
This model, the Trap_Type effect is highly significant and the Tree_Species effect is not significant (but close to the usual 0.05 significance level) The year effect could not be tested (including it makes the model crash)

We can then compute all pairwise multiple comparisons as done before and summarize the results on the graph :

Figure 43:

3.5.2.2 Total number of individuals caught without the bottle traps OK but what happens if we do not use the bottle traps in the pairwise comparisons ?

This model, the Trap_Type effect is highly significant and the Tree_Species effect is not significant (but close to the usual 0.05 significance level) The year effect could not be tested (including it makes the model crash)

Table 22: Analysis of Deviance Table (Type II Wald chisquare tests)

	Chisa		Df $\mathrm{Pr}(\geqslant C$ hisq)
Trap Type	60.10	-3-	0.0000
Tree Species	2.99		0.0836

We can then compute all pairwise multiple comparisons as done before and summarize the results on the graph :

Figure 44:

3.5.2.3 Total number of individuals caught and each year analysed separately .

OK, but what happens if analyse each year separately ??

We tried many approaches but none of the models converged whatever the optimizer, the number of iterations and even by simplifying the model to the bare minimum (no TreeID random effects, no offset, etc.)

3.5.3 Trap catches for different species

3.5.3.1 Gaussian model on a subset of the data with only the complete blocs .

We repeat our approach from the main paper for the comparison of the 7 most common species but on the subset of the data

We use a mixed model of the form (on the subset of data) = $lmer(log1p(Nb_Indiv_90_Days)$ ~ Trap_Type*Species + (1|SiteID) + (1|TreeID), data = tmp)

Figure 45:

The Anova table shows a highly significant Trap type x species effect as with the full dataset

	Table 23: Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger		
df)			

The following graph provides a graphical summary of the model. The general pattern is very similar to the one obtained with the full dataset, in particular the difference between A.biguttatus and the other species.

3.5.3.2 Poisson / Negative binomial model on a subset of the data with only the complete blocs .

A Poisson model fails to converge or is nearly unidentifiable and is overdispersed anyways (overdisp ~ 6).

A negative binomial model takes ages to compute and finally fails to converge even when changing the optimizer and increasing the number of iteration.

We didn't tested the models for each species separately

3.5.4 General conclusions of the robustness analysis

Globally, whatever the statistical approach and even on a small subset of the full dataset, the patterns are very similar to the ones obtained with the full dataset.

Our conclusions are very robust.

For the total catches :

- Green fan-traps catches are always significantly lower than the yellow fan traps, MULTz and multifunnel
- Yellow fan-traps and MULTz catches are very similar and never significantly different
- Bottle traps captured significantly lower numbers than MULTz

The only case where we can have small differences in the statistical tests depending on the analysis is for the multifunnel traps. The pattern is however always the same : the multifunnel tend to capture slightly less than the MULTz (and to a lower extend than the yellow fan-traps) but with a lot of variation. As a consequence the difference is statistically significant with some analyses and not with others. In the paper we took a conservative approach and simply declared the the catches tended to be lower but generally not significant.

For the species level analysis the conclusions are also very similar to the gloabl analysis. Most species follow a similar pattern (more captures in the yellow traps) with the notable exception of A. biguttatus which seems to be clearly more attracted by the multifunnel green traps.

4 Examination a few miscellaneous special cases

4.1 "Pale Yellow" Fan-traps

Beside the two main trap colors : vivid "yellow" and "green", a few traps have a "Pale yellow" color. This is due to the fact that the wrong yellow color was used to paint the first traps. So at first the pale yellow and green fan-traps were placed on each tree (2 trees per site) of the 10 sites for ULB team. When the correct yellow paint has become available an additional trap with the same vivid yellow color as for the other teams was placed in all trees along with the pale yellow and green ones.

The pale yellow traps captured only 3 specimens over 8 trapping periods of \sim 2 weeks for all 10 traps. \sim to simplify the experimental design, we have simply dropped these traps in the rest of the analysis

4.2 Does (3Z)-hexanol chemical lure have an effect on the captures ?

We performed a small preliminary test with this chemical lure (not a pheromone though) supposed to be attractive for *Agrilus planipennis*.

This is a small separate experiment on sites different from the rest of the study.

Three multifunnel traps have been baited with (3Z)-hexanol. They have been placed in 3 different sites and paired with a non baited multifunnel trap on the same site (but a different tree). All 6 traps catches have been collected 4 times.

On the following graph, each line relates the trap catches from the same site (sub-graphs) at the same date (ie comparable catches).

There is no obvious differences. However it is rather difficult to conclude with only 3 traps of each. One of the sites had also 0 captures.

Figure 47:

Very basic linear mixed model of the $log(x+1)$ transformed catches vs type of lure + TrapID as random effect (all more complicated model failed to converge). No significant difference but statistical power very low. . . .

 $Model: Imer(log1p(Nb_Indiv) ~ Trap_Lure + (1|TrapID), data = tmp)$

Analysis of Deviance Table (Type II Wald F tests with Kenward-Roger df)

Response: log1p(Nb_Indiv)

- $##$ F Df Df.res Pr($>F$)
- ## Trap_Lure 0.0238 1 4 0.8849

Figure 48:

4.3 Captures in Pear trees orchards

In Pear orchards we captured as expected only the target species : *Agrilus sinuatus.* However we captured only 17 individuals in 2021 and 0 in 2022 despite their presence being attested by visual observations (beating tray).

Note that most of the captures comes from only one of the 5 sites of this year and that many captures happened rather late in the season (end August - September).

The multifunnel traps were received later and trapped during 38 days relative to 101 days for bottle traps and 45-101 days for the MULTz

In 2022 the design was perfectly balanced with 85 days of trapping between early June and end August. The sampling finished only ~ 15 days earlier relative to 2021

The differences between trap types seem to be limited. Based on the number of individuals captured corrected by the number of days of trapping , the bottle traps seems less efficient but this is based on very few real catches.

Conclusions : based on these 2 years of captures, the usage of traps for the surveillance of *Agrilus sinuatus* in pear orchards seems to be of limited interest while the small size of the trees allows the use of the beating tray for standardized counts.

Because this *Pyrus* dataset is very different form the rest and with very few catches, these pear sites have not not been included in the rest of the analysis.

4.4 Captures on Poplars

Poplars have a rather different fauna (and the sites were more open, not forests).

As expected the 2 most frequent species are poplars specialists : Agrilus pratensis and Agrilus ater. Anthaxia manca is a rare species living on Ulmus spp. (all 10 specimens collected in the same sample)., Agrilus convexicollis is living on Fraxinus excelsior.

Traps were placed on 11 sites but with very uneven sampling intensities. In most of the sites only one or two MULTz or Multifunnel have been placed for a short period of time.

One site in Oud-Heverlee received a full design with fan traps with or without decoys MULTz and multifunnel and the vast majority of catches are coming from that site. Agrilus ater has just been captured in 2 other sites (8 individuals)

Nb catches on Poplars

Figure 50:

4.5 Captures in Canada

As expected the Canadian species composition is rather different than the European one. The number of replicates was also not very high in Canada. As a consequence it was not always desirable to include the Canadian data in the global analysis. It is nevertheless very interesting to see how the fan-traps compare to the traditional multifunnel green with a completely different fauna.

There was only 3 different traps in 3 different trees (9 traps in total).

For the total number of captures, the results are more or less in line with what was observed in Europe : the green fan-traps capture less while the yellow fan-traps are equivalent to the green multifunnel. So the yellow fan-traps are also clearly attractive for the non European fauna.

Figure 51:

The total number of captures per species is not very large so the species by species comparison is not easy.

Figure 52:

Figure 53:

5 Phenology

5.1 Total catches

Each gray line represents the same trap

Figure 54:

5.2 Phenology of the most common species

Each line represent the average for each site of the number of beetles captured per 14 days (average of all the traps for this site at a given date).

A.sulcicollis and A.biguttatus seem to fly a bit earlier than the other species

Figure 55:

The previous graph might be not so easy to read and we are more accustomed to bar graphs to represent the phenology. This is not so easy here for 2 reasons : 1) the sampling effort is not constant over time so we need to correct for that but there are different ways to do so and 2) we need to regroup the observations per bins (ie we need to discretise the the sampling date). The problem is that each sample is itself a time period of ~14 days but they are not all the same on all sites.

We make a tentative on the following graph :

- 1) We divided the number of individuals captures for each species by the number of trapping days, then we averaged that over all sites and all traps for each period
- 2) We took the median of the sampling date (mid date between the end and start of the sampling session) and attributed it to a period corresponding to the beginning or the end of each month (2 periods per month)

We can clearly see that A.sulcicollis and A.biguttatus were already at their peak (or had even passed it) when we started our sampling while the other species peaked during our sampling period.

Figure 56:

NB : In 2022, the sampling started 15 days earlier in the oak tree sites from Wallonia (the Belgian sites where we captured the most beetles) than in France. This could explain why the relative number of A.sulcicollis is much lower in France : we probably miss the peak of this early species

Country	SiteID	Year	Date Start	Date End
Belgium	Sclassin	2021	2021-05-20	2021-09-16
Belgium	Libin	2021	2021-06-04	2021-09-16
Belgium	Rochefort	2021	2021-06-04	2021-09-16
Belgium	Haute Bodeux	2021	2021-06-07	2021-09-15
Belgium	Spa	2021	2021-06-07	2021-09-15
Belgium	Tenneville E	2021	2021-06-08	2021-09-23
Belgium	Uccle Forêt de Soignes	2022	2022-05-10	2022-09-12
Belgium	Tenneville	2022	2022-05-11	2022-09-13
Belgium	Finneveaux	2022	2022-05-12	2022-09-02
Belgium	Jamblinne	2022	2022-05-12	2022-09-02
Belgium	Rochefort	2022	2022-05-12	2022-09-02
Belgium	Custinne	2022	2022-05-13	2022-09-02
Belgium	Houyet	2022	2022-05-13	2022-09-02
France	Vierzon 11	2022	2022-05-31	2022-07-28
France	Vierzon 179	2022	2022-05-31	2022-07-28
France	Vierzon 236	2022	2022-05-31	2022-07-28
France	Vierzon 249	2022	2022-05-31	2022-07-28
France	Vierzon 70	2022	2022-05-31	2022-07-28
France	Vierzon 71	2022	2022-05-31	2022-07-28
France	Vierzon 234	2022	2022-06-14	2022-07-28

6 Spectral analysis of the traps color (reflectance spectra)

Notes :

- Reflectance is a unit less ratio between the quantity of light reflected and the incident quantity
- Measured on a ASD FieldSpec4 spectrometer (Malvern Panalytical, Malvern, United Kingdom)
- The measures have been performed in 2023, on traps that have been placed during several season outside. So their color might have changed a little bit relative to brand new traps.
- We present the average of 5 reflectance measures
- The yellow color used on the graphs is different from the true yellow color of the traps because the true color is too bright and difficult to see on the graphs.
- The green color used on the graphs is relatively similar to the one of the traps
- We added a small color gradient scale showing the theoretical correspondence between wavelengths and "standard" human vision colors

The Multz and Multifunnel are commercial traps with a predetermined color. The fan-traps and bottle traps were painted with two different paints/colors :

- Green paint similar to the the multifunnel green (RAL 6038)
- The yellow paint (yellow fluor spray layer Motip article number 04022, EAN 8711347040223) was applied over a first layer of plastic primer and a second layer of matt white (all products from Motip, Wolvega, The Netherlands).

Raw values from the spectrometer :

Wavelength corresponding to the maximum Reflectance :

Figure 57:

Corrected values.

We added the absolute value of the minimum when it is negative, then we divided by the maximum reflectance value.

NB: this is similar to the options fixneg $=$ "addmin" and opt $=$ "maximum" in the function pavo::procspec (but we didn't used that package to reduce unnecessary dependances).

Figure 58:

The variability between replicates is very low (one line $= 1$ replicate)

Figure 59:

7 Session Info

```
## R version 4.3.2 (2023-10-31)
## Platform: x86_64-pc-linux-gnu (64-bit)
## Running under: Ubuntu 22.04.3 LTS
##
## Matrix products: default
## BLAS: /usr/lib/x86_64-linux-gnu/openblas-pthread/libblas.so.3
## LAPACK: /usr/lib/x86_64-linux-gnu/openblas-pthread/libopenblasp-r0.3.20.so; LAPACK version 3.10.0
##
## locale:
## [1] LC_CTYPE=en_GB.UTF-8 LC_NUMERIC=C LC_TIME=en_GB.UTF-8 LC_COLLATE=en_GB.UTF-8
## [5] LC_MONETARY=en_GB.UTF-8
## [9] LC_ADDRESS=C LC_TELEPHONE=C LC_MEASUREMENT=fr_BE.UTF-8 LC_IDENTIFICATION=C
##
## time zone: Europe/Brussels
## tzcode source: system (glibc)
##
## attached base packages:
              graphics grDevices utils datasets methods base
##
## other attached packages:
## [1] patchwork_1.1.3 dplyr_1.1.3 tidyr_1.3.0 ggspatial_1.1.9 sf_1.0-14 taxize_0.9.100 car_3.1-2
## [8] carData_3.0-5 multcomp_1.4-25 TH.data_1.1-2 MASS_7.3-60 survival_3.5-7 mvtnorm_1.2-3 lme4_1.1-34
## [15] Matrix_1.6-1.1 ggplot2_3.4.4 kableExtra_1.3.4 pander_0.6.5 knitr_1.44
##
## loaded via a namespace (and not attached):
## [1] DBI_1.1.3 gridExtra_2.3 sandwich_3.0-2 rlang_1.1.1 magrittr_2.0.3 e1071_1.7-13
                                         maps_3.4.1 systemfonts_1.0.5 vctrs_0.6.4
## [13] stringr_1.5.0 httpcode_0.3.0 pkgconfig_2.0.3 crayon_1.5.2 fastmap_1.1.1 backports_1.4.1
## [19] labeling_0.4.3 utf8_1.2.4 rmarkdown_2.25 nloptr_2.0.3 purrr_1.0.2 xfun_0.40
## [25] jsonlite_1.8.7 uuid_1.1-1 broom_1.0.5 parallel_4.3.2 cluster_2.1.5 R6_2.5.1
## [31] stringi_1.7.12 boot_1.3-28.1 rpart_4.1.19 Rcpp_1.0.11 bookdown_0.35 iterators_1.0.14
## [37] zoo_1.8-12## [43] rstudioapi_0.15.0 abind_1.4-5 yaml_2.3.7 codetools_0.2-19 curl_5.1.0 lattice_0.22-5
## [49] tibble_3.2.1 withr_2.5.1 evaluate_0.22 foreign_0.8-85 units_0.8-4 proxy_0.4-27
                                         KernSmooth_2.23-22 checkmate_2.2.0
## [61] munsell_0.5.0 scales_1.2.1 minqa_1.2.6 class_7.3-22 glue_1.6.2 Hmisc_5.1-1
                       \begin{tabular}{llll} data.table\_1.14.8 & webshot\_0.5.5 & \quad for cats\_1.0.0 & \quad grid\_4.3.2 \\ urltools\_1.7.3 & \quad colorspace\_2.1-0 & \quad nlme\_3.1-163 & \quad htmlTable\_2.4.1 \\ \end{tabular}## [73] ape_5.7-1 urltools_1.7.3 colorspace_2.1-0 nlme_3.1-163 htmlTable_2.4.1 conditionz_0.1.0
                                                          virtualistite_0.4.2 svglite_2.1.1 gtable_0.3.4
## [85] digest_0.6.33 classInt_0.4-10 pbkrtest_0.5.2 crul_1.4.0 htmlwidgets_1.6.2 farver_2.1.1
## [91] htmltools_0.5.6.1 lifecycle_1.0.3 httr_1.4.7
```
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