

Figure S1

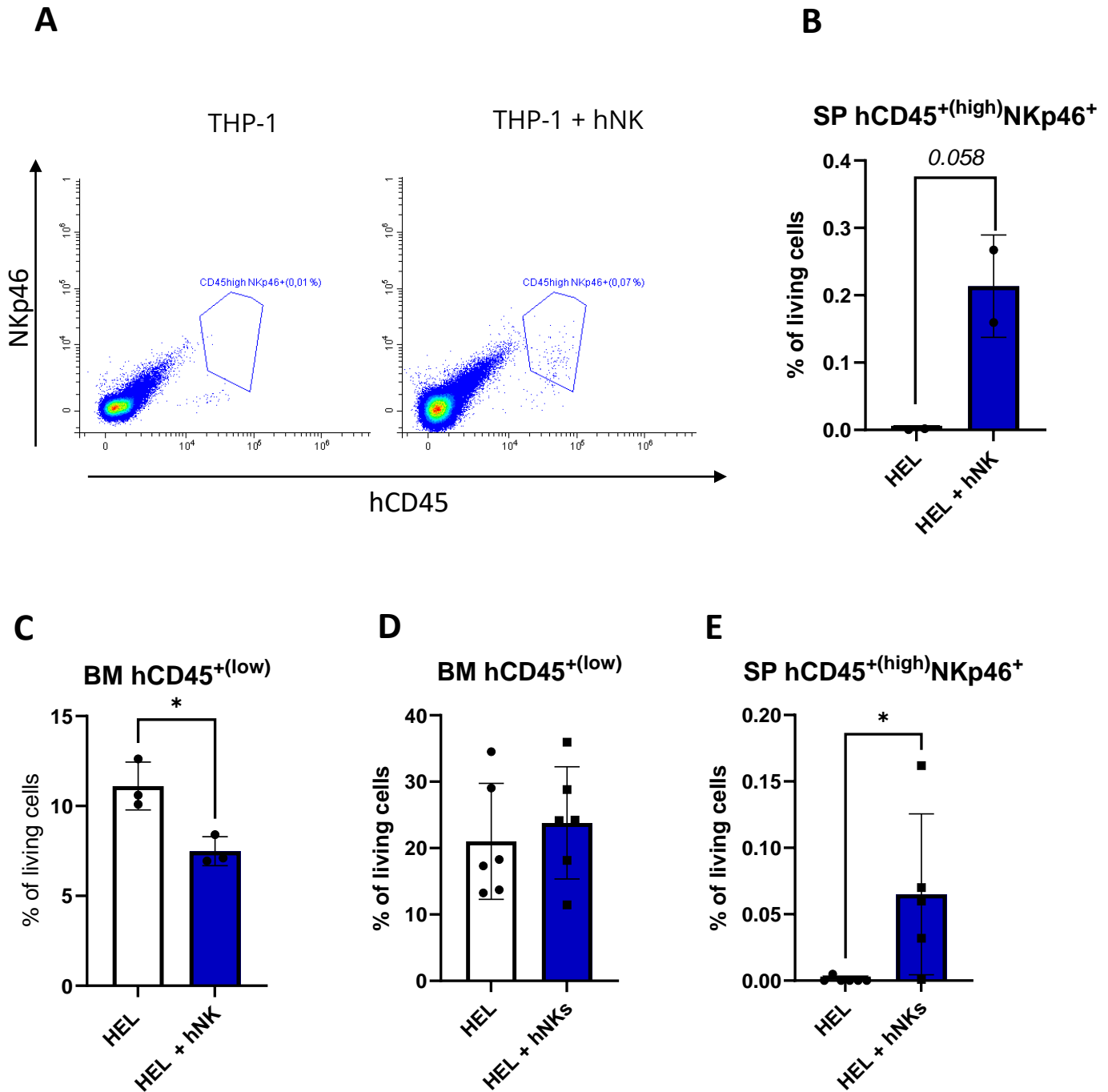


Figure S1. THP-1 and HEL cell line xenografts are killed by NK cells *in vivo*. (A-E) NSG-Tg(Hu-IL15) mice were injected with 0.5×10^6 (A) THP-1 cells or (B-E) HEL cells *i.v.*. In addition, the mice were injected with 10^6 human NK cells 24 h prior to leukemic cell inoculation or left untreated. (A) NK cells (hCD45^{+(high)}NKp46⁺) in the splenic single cell suspensions of THP-1-injected animals on day 28 post injection were analyzed by flow cytometry. (B,C): (B) NK cells (hCD45^{+(high)}NKp46⁺) in the splenic single cell suspensions or (C) leukemic cells (hCD45^{low}) in bone marrow of HEL-injected animals were analyzed by flow cytometry on day 28 post injection. (D, E): (D) leukemic cells (hCD45^{low}) in bone marrow or (E) NK cells (hCD45^{+(high)}NKp46⁺) in the splenic single cell suspensions of HEL-injected animals were analyzed by flow cytometry at the disease endpoint. (A) Representative flow cytometry plots. (B, C) Bar graphs represent mean \pm SD from 1 experiment (n=2-3 per group); (D, E) Bar graphs represent mean \pm SD from 2 independent experiments (n=6 per group); (B, C, E) Statistical analysis was performed using unpaired t-test. * p<0.05;

Figure S2

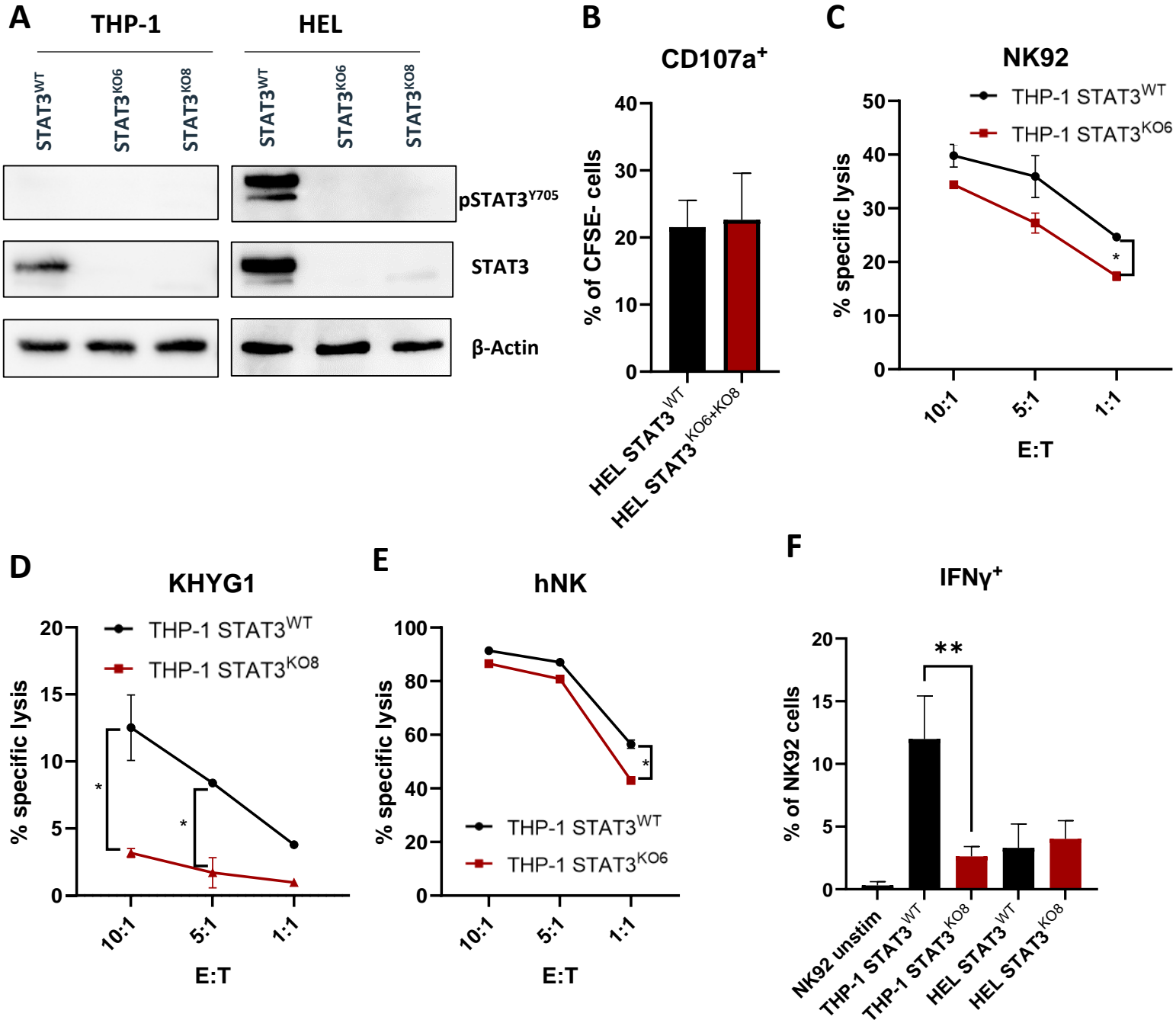


Figure S2. STAT3-deficient AML cells escape NK cell recognition. (A) Levels of pSTAT3, STAT3 and β -actin were assessed by Western blot in cell lysates of THP-1 and HEL STAT3^{WT}, STAT3^{KO6} and STAT3^{KO8} and a representative blot out of three independent experiments is shown. **(B)** CFSE stained HEL STAT3^{WT} and STAT3^{KO6} and STAT3^{KO8} cells were mixed at 1:1 ratio with human primary NK cells from three different healthy donors and the percentage of CD107a⁺ NK cells (CFSE⁻; in contrast to CFSE⁺ target cells) was analyzed by flow cytometry. Bar graphs represent mean \pm SD of n= 4 per group (3 donors, 1-2 experiments per donor). **(C, E)** CFSE-stained THP-1 STAT3^{WT} and STAT3^{KO6} cells were mixed at indicated effector : target (E:T) ratios with **(C)** NK92 for 4 h or **(E)** expanded primary human NK cells for 2 h. **(D)** CFSE-stained THP-1 STAT3^{WT} and STAT3^{KO8} cells were mixed at indicated effector : target (E:T) ratios with KHYG1 for 4 h. **(C-E)** Symbols and bars represent mean of technical duplicates \pm SD. Statistical analysis for each ratio was performed using unpaired t-test. **(F)** NK92 cells were stained for intracellular IFN γ upon incubation with STAT3^{WT} or STAT3^{KO8} THP-1 or HEL cell line. Bar graphs represent mean \pm SD of n= 4 per group from 2 independent experiments. Statistical analysis for each cell line separately was performed using unpaired t-test. * $p < 0.05$; ** $p < 0.01$

Figure S3

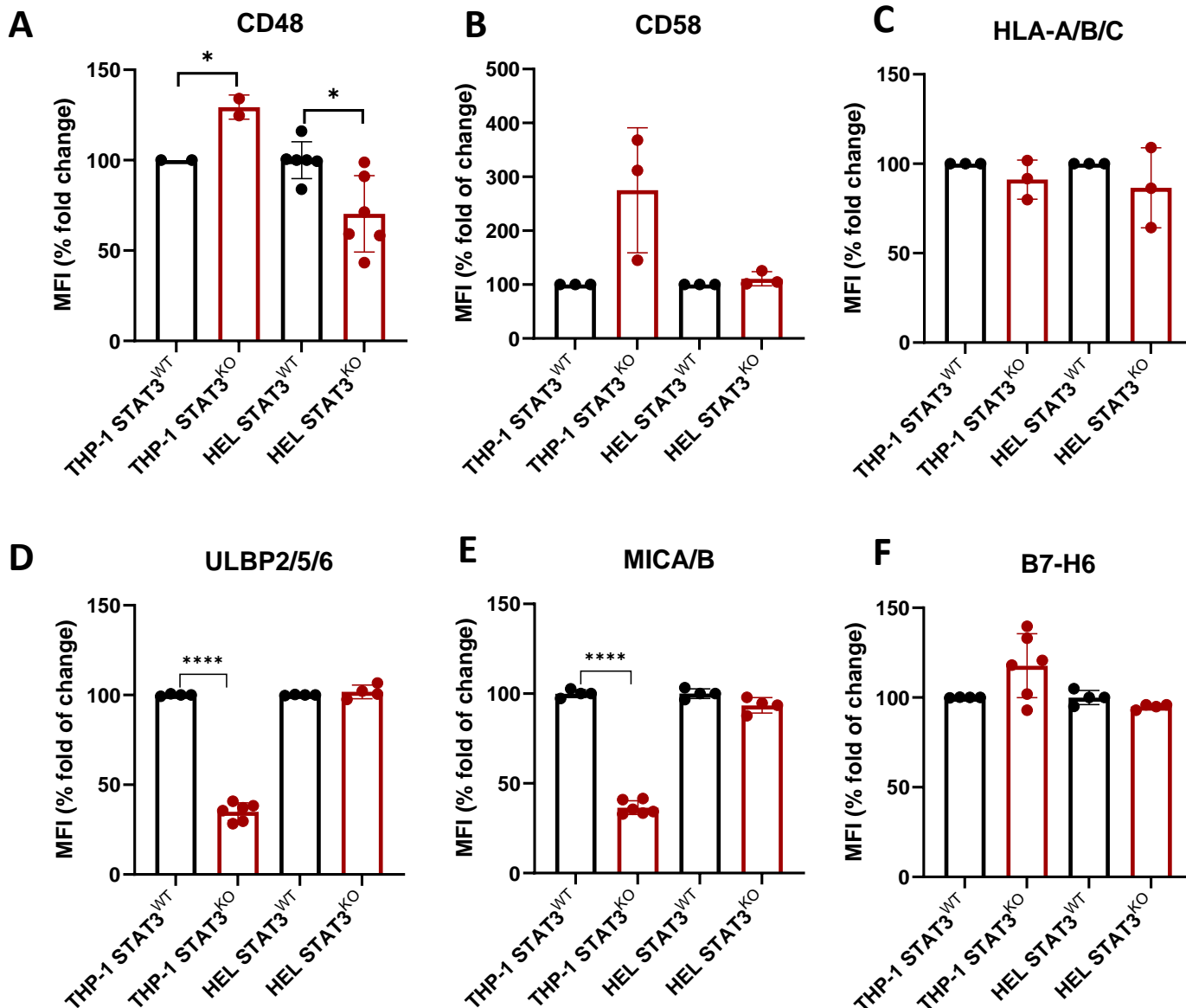


Figure S3. Surface expression of NK cell ligands. (A, B) STAT3^{WT} and STAT3^{KO8} THP-1 and HEL were analyzed for surface expression of CD48, CD58, HLA-A/B/C, ULBP2/5/6, MICA/B, B7-H6 via flow cytometry. Bar graphs showing mean fluorescence intensity (MFI) as percentage of fold change of (A) CD48 (B) CD58, (C) HLA-A/B/C, (D) ULBP2/5/6, (E) MICA/B and (F) B7-H6 +/- SD are shown for each cell line (n=3-6 per group from 2-3 independent experiments). Statistical analysis was performed using unpaired t test for THP-1 and HEL cells separately; * $p < 0.05$; **** < 0.0001

Figure S4

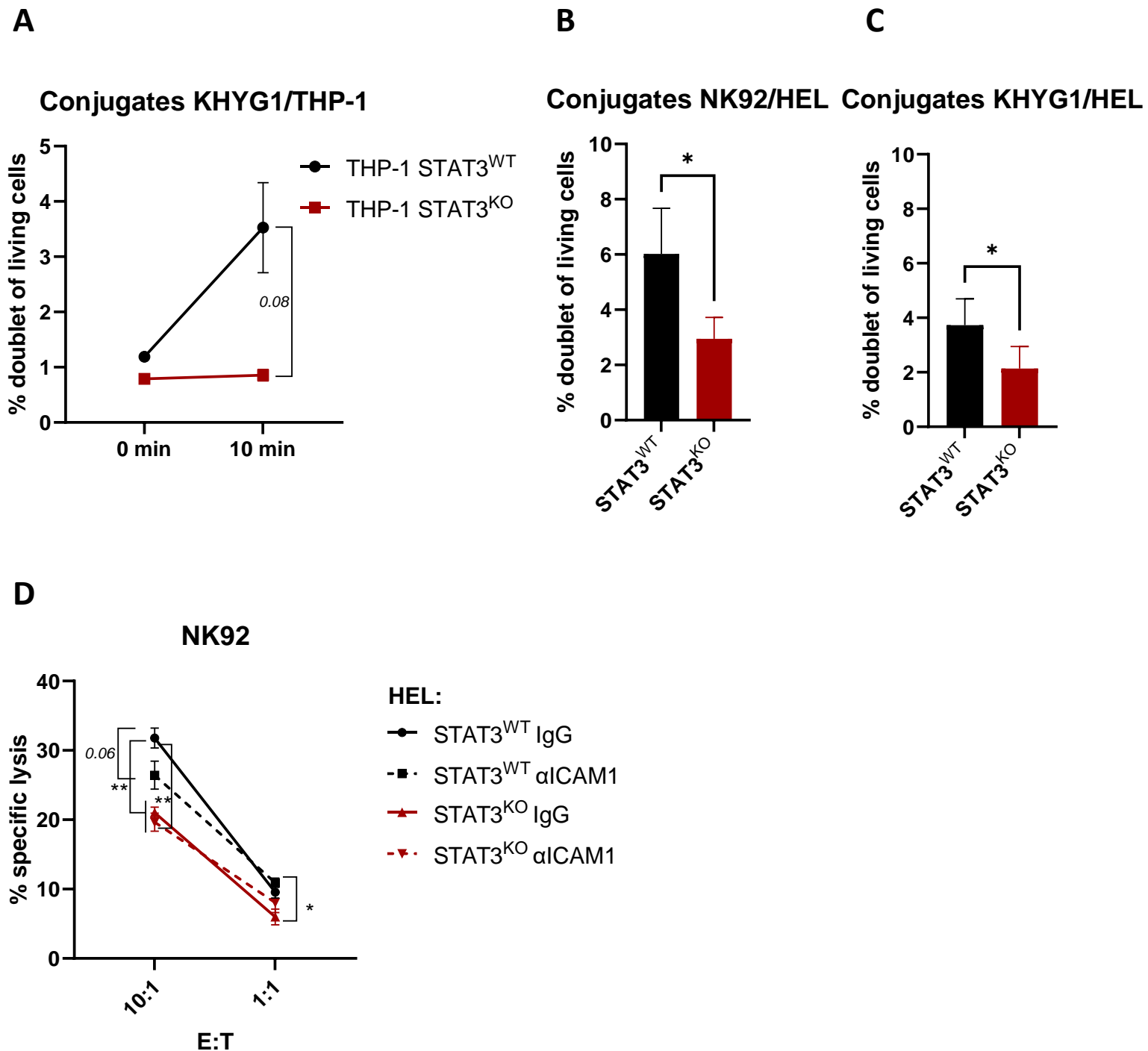


Figure S4. STAT3-deficient AML cells form fewer synapses. (A) THP-1 STAT3^{WT} and STAT3^{KO8} or (B, C) HEL STAT3^{WT} and STAT3^{KO8} were incubated with (A, C) KHYG1 or (B) NK92 cells at 1:1 ratio and the amount of NK-target doublets was analyzed after (A) 10min or (B, C) 20 min. (A) Representative time course out of two independent experiments. For timepoint 10 min dots represent the mean percentage of doublets +/- SD (n=2). (B, C) Bar graphs represent the mean percentage of doublets +/- SD (n=2-4 per group from one to two independent experiment); (A-C) Statistical analysis was performed using unpaired t-test. * $p < 0.05$. (D) CFSE-stained HEL STAT3^{WT} and STAT3^{KO8} cells were preincubated with isotype control (IgG) or ICAM-1 blocking antibody and mixed at indicated effector : target (E:T) ratios with NK92 cells for 4 h. The specific lysis of target cells was assessed by flow cytometry. One representative out of two independent experiments is shown. Symbols and bars represent the mean of technical duplicates +/- SD. Statistical analysis was performed using one-way ANOVA with Tukey post-test for each ratio. Vertical lines indicate that the same significance level refers to all conditions covered by the line. ** $p < 0.05$; ** $p < 0.01$

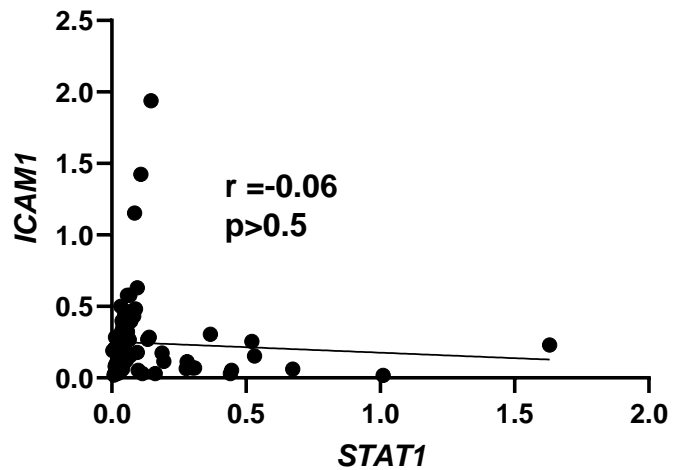
Figure S5

A

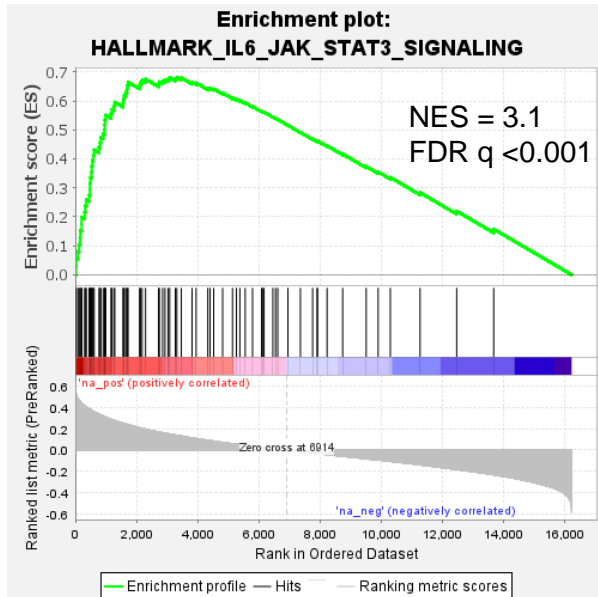
Pearson correlation coefficient (r)

	ICAM-1	STAT3
Affymetrix ID	202638_s_at	225289_at
ICAM-1	202638_s_at	1 (p<1E-04)
STAT3	0.2175 (p=2e-04)	1 (p<1E-04)

B



C



D

	Multivariate analysis		
	Hazard ratio	95% CI	p value
Age	1.01	1.004-1.024	0.004
Gender	0.98	0.75-1.21	n.s.
Karyotype	2.08	1.90-2.26	7.1E-07
	Univariate analysis		
	Hazard ratio	95% CI	p value
Age	1.03	1.022-1.030	1.2E-06
Gender	0.96	0.79-1.13	n.s.
Karyotype	1.85	1.71-1.99	6.7E-07

Figure S5. STAT3 correlates with ICAM1 in AML patients. (A, B) The correlation between **(A)** STAT3 and ICAM1 or **(B)** STAT1 and ICAM1 expression was analyzed using Pearson test in **(A)** publicly available dataset (kmplot.com) and **(B)** in a cohort of 79 AML patients as described in Fig. 5. **(C)** Enrichment plot of the hallmark IL6_JAK_STAT3_SIGNALING. Red and blue colors represent, respectively, positive and negative Spearman's correlations with ICAM1. Analysis was performed on normalized gene expression RNA-seq data of human AML patients (TCGA-LAML) from The Cancer Genome Atlas (TCGA). Normalized enrichment score (NES) and false discovery rate (FDR) q value are indicated. **(D)** Uni- and multivariate Cox regression analysis was performed to correlate survival and ICAM1 expression, age, gender, karyotype. The multivariate analysis was performed by pairing ICAM1 with the clinical variables and the hazard ratio, 95% confidence interval (CI) and p value are shown for each variable.

Table S1

Total number of patients	79
Males	50 (63.3%)
Females	29 (36.7%)
Age (years)	
Median	66
Range	23-89
WBC at diagnosis (10⁹/l)	
Median	6
Range	0-388
BM blasts at diagnosis (%)	
Median	68
Range	15-100
Pathogenic/likely pathogenic mutations (number of samples)	
KRAS	12
NRAS	14
RUNX1	13
IDH2	8
WT1	2
ASXL1	14
TET2	24
DNMT3A	13
NPM1	11
CEBPA	13
TP53	17
FLT3 / FLT3-IDH	0
Other Mutations (IDH1, CRSF2, STAG2, KIT, etc.)	48
Therapy	
Intensive incl. allogeneic hematopoietic stem cell transplantation	45
Non-intensive	28
Best supportive care	6

Table S1. Characteristics of 79 AML patients.

Table S2

<i>GAPDH</i> Fw	TCTCCTCTGACTTCAACAGCG
<i>GAPDH</i> Rev	ACCACCCTGTTGCTGTAGCC
<i>ULBP1</i> Fw	TTTCCTTAAAGGGCAACTGCT
<i>ULBP1</i> Rev	AGGAACTGCCAAGATCCTCT
<i>MICA</i> Fw	GAATCCGGCGTAGTCCTGAG
<i>MICA</i> Rev	TCCGGGGATAGAAGCTGGAA
<i>B7H6</i> Fw	TTTTCCATTATTGGTGGCCT
<i>B7H6</i> Rev	TGCCCGAGTGCAAAAGAATATG
<i>HLA_A</i> Fw	AGATACACCTGCCATGTGCAGC
<i>HLA_A</i> Rev	GATCACAGCTCCAAGGAGAACC
<i>ICAM1</i> Fw	AGCGGCTGACGTGTGCAGTAA T
<i>ICAM1</i> Rev	TCTGAGACCTCTGGCTTCGTCA
<i>CD48</i> Fw	GCTTGAAACCACCCTTATGCCAC
<i>CD48</i> Rev	CGTGACCACTAGCCAACTTGCA
<i>CD58</i> Fw	GACACTGTGTCAGGTAGCCTCA
<i>CD58</i> Rev	GCACAAGTTAGTGTGGGAGATGG
<i>STAT3</i> Fw	CTGACCCAGGTAGCGCTGCCCCATACC
<i>STAT3</i> Rev	TCACAATGGGGGAGGTAGCGCACTCCG

Table S2. Primer list used for RT-qPCR analysis.