

Annex to: EFSA's BIOHAZ Panel Scientific opinion "Public health aspects of *Vibrio* spp. related to the consumption of seafood in the EU". doi:10.2903/j.efsa.2024.8896

© European Food Safety Authority, 2024

# Annex D – Analytical methods for the detection and enumeration of potentially enteropathogenic *Vibrio* spp. in seafood

**Table D.1:** Overview of the analytical methods for the detection and enumeration of potentially enteropathogenic *Vibrio* spp. in seafood.

		•								
Standardised methods for detection										
Scope	Field of applicatio n / sample type	<i>Vibrio</i> species	Brief description of procedure	Genetic markers	Performance characteristics	Applicability / limitation				
ISO 21872-1:20			gy of the food chain – Horizontal method for the det s, Vibrio cholerae and Vibrio vulnificus. International							
Horizontal method for the detection of enteropathogeni c <i>Vibrio</i> spp. (human illness in or via the intestinal tract)			Enrichment: test portion up to 25 g or 25 mL, 1 medium, 2 consecutive enrichments and 2 temperatures Colony isolation: 2 selective media ID: biochemical tests ID confirmation: cPCR Pathogenicity: serology (O1/O139) or cPCR Enrichment: test portion up to 25 g or 25 mL, 1 medium, 2 consecutive enrichments and 2 temperatures Colony isolation: 2 selective media ID: biochemical tests ID confirmation: cPCR and rtPCR Pathogenicity: cPCR and rtPCR	Pathogenicity: ctx gene  ID confirmation: toxR	Matrix: cooked prawns LOD50: 11.34 CFU/g Sensitivity: 92.5% Specificity: 84%  ILS³  Matrix: raw oysters LOD50: 0.43 CFU/g Sensitivity: 72.5-100% for Vp, 86% for tdh+ Vp by cPCR, 94% for trh+Vp by cPCR, 73% for tdh+ Vp by rtPCR Specificity: 111% for Vp, 89% for tdh+ Vp by cPCR, 91% for trh+Vp by cPCR, 94% for tdh+ Vp by rtPCR Matrix: cooked prawns LOD50: unknown Sensitivity: 100% Specificity: 99%	Widely used method for the detection of Vp, Vv, Vc. Well standardised protocols. Primer sets for Vibrio spp. ID confirmation and Vp virulence associated genes. Subculture of a small number of colonies could lead to miss potentially pathogenic strains.				
		V. vulnificus	Enrichment: test portion up to 25 g or 25 mL, 1 medium, 2 consecutive enrichments and 1 temperature Colony isolation: 2 selective media ID: biochemical tests ID confirmation: cPCR and rtPCR	ID confirmation: vvhA gene (cPCR and rtPCR)	ILSa Matrix: raw oysters LOD50: 12.31 CFU/g Sensitivity: 37.5% Specificity: 77.5% Matrix: cooked prawns					



LOD50: 81.2 CFU/g Sensitivity: 61.25-71.5% Specificity: 60%

accuracy and linearity, repeatability.

FDA	-BAM (2004)	). Bacteriological	Analytical Manual Chapter 9: Vibrio. Food & Drug Ad	ministration, Bacteriolog	gical Analytical Manual, V	Vashington DC <sup>c</sup>
human pathogenic Vibrionaceae (V. cholerae, V. parahaemolyt icus and	Food and water	V. cholerae (V. mimicus)	<ul> <li>Enrichment: test portion 25 g, 1 medium, 1 or 2 consecutive enrichments and 2 temperatures</li> <li>Colony isolation: 2 selective media</li> <li>ID: biochemical tests, serological typing</li> <li>Pathogenicity: cell assay, cPCR</li> </ul>	Pathogenicity: ctx gene	NR	Presumptive identification o Vc (no molecular characterisation or confirmation). Biotyping and other tests recommended or optional.
	Seafood (molluscan shellfish, processed food)	, ,	Direct plating: test portion 0.1 g of homogenate, 1 selective medium, incubation at 1 temperature ID: colony hybridization (AP-probe) or multiplex cPCR Pathogenicity: colony hybridization (AP-probe and DIG-probe) or multiplex cPCR	cPCR) <b>Pathogenicity</b> : <i>tdh</i> gene		The use and specificity in given conditions of the <i>tlh</i> gene for Vp identification has been questioned (Yáñez et al., 2015)
	Seafood (molluscan shellfish, processed food)	V. vulnificus	<b>Direct plating:</b> test portion 0.1 g of homogenate, 1 selective medium, incubation at 1 temperature <b>ID:</b> colony hybridization (DIG-probe) and cPCR	ID: vvhA gene (DIG- probe, cPCR)	NR	

#### Standardised methods for enumeration

ISO/TS 21872-2:2020. Microbiology of the food chain – Horizontal method for the determination of *Vibrio* spp. Part 2: Enumeration of total and potentially enteropathogenic *Vibrio parahaemolyticus* in seafood using nucleic acid hybridization. International Organization for Standardization, Geneva, Switzerland.

Method for the V. parahaemolytic Direct plating: test portion 0.1 q of homogenate, 1 non- ID: DIG toxR gene (DIG- Single-laboratory Accurate and specific but Seafood direct selective medium, incubation at 1 temperature, transfer probe) validation time consuming and enumeration of of colonies to nylon membrane Pathogenicity: tdh1, Matrix: bivalve shellfish, technically demanding **ID:** colony hybridization (DIG-probes) for total Vp and for tdh2, trh1, trh2 genes finfish, cephalopods, method; 0.1 g of sample total and tdh+ and/or trh+ Vp (DIG-probe) crustaceans. analysed only. potentially enteropathogeni Selectivity: (a) oligoprobes inclusivity: toxR 99.4%, tdh 96.7%, trh 97.8%; V. parahaemolyt icus in seafood (b) oligoprobes exclusivity: 100% for all targets Sensitivity: LOD 10 CFU/g, LOO 100 CFU/a (defined in accordance with ISO 7218) Performance parameters:

## FDM-BAM (2004) Bacteriological Analytical Manual Chapter 9: Vibrio. Food & Drug Administration, Bacteriological Analytical Manual, Washington DC<sup>c</sup>

Detection of V. parahaemolytic MPN format enrichment: test portion 1 g and following ID: tlh gene (AP-probe, NR Time consuming (MPN: 4-5 Seafood (molluscan us dilutions, 1 medium, incubation at 1 temperature cPCR) days, HGMF: 3-4 days). human pathogenic shellfish, Colony isolation: 1 selective medium Pathogenicity: tdh gene Identification is only Vibrionaceae processed **ID:** biochemical tests or colony hybridization (AP-probe) (AP-probe, cPCR), trh presumptive if no (Vibrio cholerae, food) or multiplex cPCR gene (cPCR)



## Public health aspects of Vibrio spp. related to the consumption of seafood in the EU

Vibrio parahaemolyticu s and Vibrio vulnificus) in seafood

Pathogenicity: colony hybridisation (AP-probe) or multiplex cPCR

**HGMF:** test portion 0.5 g, culture on filter on 1 selective medium, incubation at 1 temperature

ID: presumptive enumeration based on colony colour

**Direct plating:** test portion 0.1 g of homogenate, 1 non- **ID:** tlh gene (AP-probe, NR selective medium, incubation at 1 temperature),

ID: colony hybridisation (AP-probe) or multiplex cPCR Pathogenicity: colony hybridisation (AP-probe) or multiplex cPCR

MPN format enrichment: test portion 1 q and following ID: vvhA gene (DIGdilutions, 1 medium, incubation at 1 temperature

Colony isolation: 2 selective media

**ID:** biochemical tests or colony hybridisation (DIG-probe)

or cPCR

molecular confirmation is applied.

Not suitable for routine analysis.

The use and specificity of the tlh gene in given conditions for Vp identification has been questioned (Yáñez et al., 2015)

Other methods (national sta	andards)
-----------------------------	----------

Scope Field of Vibrio sp. applicatio n/ Sample type

Brief description of procedure

**Genetic markers** 

Pathogenicity: tdh gene

(AP-probe, cPCR), trh

cPCR)

gene (cPCR)

probe, cPCR)

**Performance** characteristics

NR

NR

Applicability / Limitations

### NMKL Method N°156, 2nd ed. 1997 (Scandinavian countries) Pathogenic Vibrio species. Detection and enumeration in foods

Detection and enumeration of cal control V. parahaemolyt of foods, icus, especially V. cholerae, shellfish, V. vulnificus and cravfish V. alginolyticus and fish. Microbial in food investigati

us, V. cholerae, V. vulnificus, V. alginolyticus

V. vulnificus

Microbiologi V. parahaemolytic **Detection**: enrichment (2 media, incubation at 1 temperature), colony isolation, colony identification (type based on colour, size, aspect), ID confirmation (biochemical tests, O1/O139 antisera Vc) Enumeration: direct plating (1 selective medium,

incubation at 1 temperature)

Not validated in a collaborative study Presumptive ID of colony (colony type description, colour, size, aspect) Enumeration based on colony type (aspect)

MFLP-37 (2019). Detection, isolation, and enumeration of Vibrio parahaemolyticus and/or Vibrio vulnificus in Seafood. Laboratory Procedures for the Microbiological Analysis of Foods. Volume 3. Government of Canada.

Detection, isolation and enumeration of V. parahaemolyt icus and/or s) V. vulnificus in

seafood

Seafood (fish, shellfish, crustacean

ons in connection with disease outbreaks

V. parahaemolytic **Enumeration**: MPN format enrichment (test portion 1g and following dilutions, 1 medium, incubation at 1 temperature)

Colony isolation: 1 selective medium **ID**: biochemical tests, multiplex cPCR (described in method MLFP-23, March 2006<sup>d</sup>), multiplex rt PCR

Pathogenicity: multiplex cPCR or multiplex rt PCR (as in the identification)

(described in method MLFP-102, July 2017e)

**ID**: R72H fragment, *tlh* NR

Pathogenicity: tdh and trh genes (multiplex cPCR or multiplex rtPCR)





V. vulnificus **Enumeration**: MPN format enrichment (test portion 1a) and following dilutions, 1 medium, incubation at 1

temperature)

Colony isolation: 1 selective medium

ID: biochemical tests

Presumptive identification of Vv (no molecular characterisation or confirmation)

#### GB 4789.7-2013. National Food Safety Standard - Microbiological Examination of Food Hygiene - Examination of Vibrio parahaemolyticus. National Standard of the People's Republic of China

Method for the inspection of V. parahaemolyt ds, icus in food crustacean

V. parahaemolytic **Detection**: enrichment (test portion 25 g, 1 medium, Fish, cephalopo us shellfish,

V. vulnificus

incubation at 1 temperature), colony isolation (1 selective medium), colony identification (biochemical tests, serological typing)

Enumeration: MPN format enrichment (test portion of 2.5 g and continuous dilutions, 1 medium, incubation at 1 temperature), colony isolation (1 selective medium), colony identification (biochemical tests, serological typing)

Presumptive identification of Vp (no molecular characterisation or confirmation)

#### GB 4789.44-2020. National Food Safety Standard - Microbiological Examination of Food Hygiene - Examination of Vibrio vulnificus. National Standard of the People's Republic of China

Method for the Fish, examination of V. vulnificus in aquatic products shellfish,

shrimp, crab, cephalopo **Detection**: enrichment (1 selective medium, incubation **ID**: vvhA gene (cPCR) NR at 1 temperature), PCR testing and colony isolation (2 selective media), colony identification (biochemical tests 16S rRNA A/B, SerE and cPCR identification (optional) and

typing/pathogenicity (optional)

Typing/pathogenicity: gene, Bt2 gene, vcgC/E genes cPCR)

Abbreviations: AP-probe, alkaline phosphatase labelled probe; cPCR, conventional PCR; DIG-probe, digoxigenin labelled probe; HGMF, hydrophobic grid membrane filtration; ID, identification; ILS, inter-laboratory study; ITS, internal transcribed spacer; LOD, limit of detection; LOO, limit of quantification; MPN, most probable number; NR, not reported; PCR, polymerase chain reaction; rtPCR, real-time PCR; Va, Vibrio alginolyticus; Vc, V. cholerae; Vm, V. mimicus; Vp, V. parahaemolyticus; Vv. V. vulnificus.

<sup>a</sup>An international inter-laboratory study, ILS (Hartnell et al., 2019) using the method specified in the ISO 21872:2017 was carried out for detection of V. parahaemolyticus, V. vulnificus and V. cholerae in raw and cooked seafood (Hartnell et al., 2019). Real-time PCR for identification of V. cholerae and detection of trh genes of *V. parahaemolyticus* was tested in this international ILS, but the data were not reliable and were therefore excluded from the final ILS dataset.

<sup>b</sup>The specificity calculated for *V. parahaemolyticus* (111%) was due to positive signals/identification obtained by cPCR in control samples.

https://www.fda.gov/food/laboratory-methods-food/bam-chapter-9-vibrio, accessed on the 14 May 2024.

<sup>d</sup>MFLP-23, March 2006: Specific detection of Vibrio parahaemolyticus strains using a multiplex polymerase chain reaction (PCR) based on R72H taxonomic marker and the haemolysin genes TDH and TRH.

<sup>e</sup>MFLP-102, July 2017: Identification of *Vibrio parahaemolyticus* colonies by real-time polymerase chain reaction.

#### References

Hartnell, R. E., Stockley, L., Keay, W., Rosec, J. P., Hervio-Heath, D., Van den Berg, H., Leoni, F., Ottaviani, D., Henigman, U., Denayer, S., Serbruyns, B., Georgsson, F., Krumova-Valcheva, G., Gyurova, E., Blanco, C., Copin, S., Strauch, E., Wieczorek, K., Lopatek, M., ... Baker-Austin, C. (2019). A pan-European ring trial to validate an international standard for detection of Vibrio cholerae, Vibrio parahaemolyticus and Vibrio vulnificus in seafoods. International Journal of Food Microbiology, 288, 58-65, https://doi.org/10.1016/j.jifoodmicro.2018.02.008

Yáñez, R., Bastías, R., Higuera, G., Salgado, O., Katharios, P., Romero, J., Espejo, R., & García, K. (2015), Amplification of tlh gene in other Vibrionaceae specie by speciespecific multiplex PCR of Vibrio parahaemolyticus. Electronic Journal of Biotechnology, 18(6), 459-463. https://doi.org/10.1016/j.ejbt.2015.09.007