

- - Control K₃Citrate

Supplementary Figure 1. Plasma chemistry is stable with K₃Citrate

supplementation in LG/J mice. (A) Albumin, **(B)** Calcium, **(C)** Chloride, **(D)** Glucose, and **(E)** Phosphorus were analyzed using an IDEXX BioAnalytics custom blood chemistry panel (Control: n=5 mice (1F, 4M); K₃Citrate: n=6 mice (2F, 4M)). **(F)** Fetuin-A, an inhibitor of mineralization, showed no differences across groups (Control: n=7 mice (2F, 5M); K₃Citrate: n=7 mice (3F, 4M)). **(G)** GlycA, **(H)** Protein, **(I)** total branched chained amino acids (BCAA), **(J)** Leucine, **(K)** Isoleucine, **(L)** Valine, **(M)** Alanine, **(N)** Acetoacetate, **(O)** Acetone, **(P)** Total Ketone Bodies, **(Q)** Beta Hydroxybutyrate, **(R)** Chelatable Mg2+, **(S)** Citrate, **(T)** ApoA-1, **(U)** ApoB, **(V)** Total Trigliyceride, **(W)** Total Cholesterol, **(X)** total calibrated low-density lipoprotein particle (cLDLP), and **(Y)** total calibrated high-density lipoprotein particle (cHDLP) were measured using NMR at LabCorp Global Research Services, and showed no changes with K₃Citrate supplementation (Control: n=6 mice (1F, 5M); K₃Citrate: n=7 mice (3F, 4M)). Sample size varied between assays based on the volume of plasma required and the volume of plasma collected from each mouse. Significance was determined using an unpaired t-test or Mann-Whitney test, as appropriate.



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Theme





Supplementary Figure 2. Analysis of upregulated DEGs and most enriched

downregulated transcripts. (A) RNA-sequencing analysis shows downregulated DEGs have high thematic enrichment. (A') Transcripts within each super cluster that have the highest entity scores (cutoff of 100). (B) RNA-sequencing analysis shows upregulated DEGs have high thematic enrichment. (Control: n=4 mice (1F, 3M); K₃Citrate: n=7 mice (2F, 2M))



Supplementary Figure 3. K₃Citrate supplementation minimally impacts the caudal vertebrae in LG/J mice. (A-A') Representative µCT reconstructions of the hemi-section caudal motion segments., and (B) vertebral length was unchanged. Trabecular properties of (C) bone volume fraction (BV/TV), (D) trabecular separation (Tb. Sp.), (E) trabecular number (Tb. N.), (F) trabecular thickness (Tb. Th.), and (G) trabecular bone mineral density were unchanged in K₃Citrate mice. (H-H') Representative µCT reconstructions central cross sections of the caudal vertebrae. Analysis of the cortical properties (I) bone volume (BV), (J) cortical tissue mineral density, (K) cross sectional thickness (Cs. Th.), (L) mean cross-sectional bone area (B. Ar.), (M) closed porosity, and (N) bone perimeter (B. Pm.) revealed cortical thinning of K₃Citrate caudal vertebrae. (Control mice: n=7 mice (2F, 5M); K₃Citrate mice: n=7 mice (3F, 4M); 2 vertebrae/mouse; 28 discs, 14 vertebrae/treatment) Data are shown as mean ± SD. Significance was determined using an unpaired t-test or Mann-Whitney test, as appropriate. Multiplex assay analysis showed no significant changes in the plasma concentrations of (O) IFN-γ, (P) IL-1β, (Q) IL-2, (R) IL-4, (S) IL-5, (T) IL-6, (U) IL-10, (V) IL-12/p70, (W) IL-15, (X) IL-17A/F, (Y) IL-27/p28/IL-30, (Z) IL-33, (AA) IP-10, (BB) KC/GRO, (CC) MCP-1, (DD) MIP-1α, (EE) MIP-2, (FF) TNF-α (Control: n=6 mice (1F, 5M); K₃Citrate: n=7 mice (3F, 4M)). Data are shown as mean ± SD. Significance was determined using an unpaired t-test or Mann-Whitney test, as appropriate.



Supplementary Figure 4. K₃Citrate alters knee calcification without impact on cartilage and bone morphology. (A-A') Representative μ CT reconstructions of the knees of control and K₃Citrate mice. Quantification of calcification nodules in the (B) synovium, (C) meniscus, and (D) patellar. (E-E') Representative hematoxylin and eosin (H&E) staining and (F-F') toluidine (Tol.) blue staining of the lateral tibial plateau. Summed (G) ACS score, (H) toluidine blue score, (I) osteophyte score, and (J) synovial hyperplasia score show no difference between cohorts. Additionally, K₃Citrate supplementation did not result in changes to the area or thickness of (K-K') articular cartilage, (L-L') calcified cartilage, or (M-M') subchondral bone. (Control: n=4 mice (1F, 3M); K₃Citrate: n=5 mice (3F, 2M)). Data are shown as mean ± SD. Significance was determined using an unpaired t-test or Mann-Whitney test, as appropriate.



Supplementary Figure 5. Validation of accelerated differentiation protocol in ATDC5 cells. CT and Diff. CT treatment groups were compared for all chondrogenic markers assessed to verify accelerated differentiation in ATDC5 cells receiving β -glycerophosphate and ascorbic acid: (A) Sox9, (B) Acan, (C) Col2a1, (D) Runx2, (E) Col10a1, (F) Mmp13, (G) Col1a1, (H) Ihh, (I) Alpl, (J) Bglap, (K) Sp7, and (L) Fgfr3, (M) Ank, and (N) Enpp1. (n=8 sets/timepoint, 2 averaged replicates/set) Data are shown as mean \pm SD. Significance was determined using an unpaired t-test or Mann-Whitney test, as appropriate.



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Supplementary 6. K₃Citrate supplementation does not impact glycolytic or oxidative metabolism in ATDC5 cells cultured for 7 days. (A) OCR and (B) ECAR

traces for ATDC5 cells cultured with or without K₃Citrate (C) to evaluate glycolytic capacity and glycolytic reserve. (D) OCR and (E) ECAR traces for ATDC5 cells cultured with or without K₃Citrate (F) to evaluate glycolytic and oxidative ATP production rates. (G) OCR and (H) ECAR traces for ATDC5 cells cultured with or without K₃Citrate (I) for the classical Mito Stress test to evaluate key parameters of mitochondrial function. (n = 3 sets, 3-4 replicates/set) Data are shown as mean \pm SD. Significance was determined using an ANOVA or Kruskal-Wallis test, as appropriate.