# nature portfolio

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# **Reporting Summary**

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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| _                      | LЧ  | U   | J ( | ics                       |

| For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.   |       |
|---|-------|
| n/a Confirmed   |       |
| The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement   |       |
| A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly   |       |
| The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.  |       |
| A description of all covariates tested  |       |
| A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons   |       |
| A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) | ient) |
| For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.                       |       |
| For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings  |       |
| For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes  |       |
| $\square$ Estimates of effect sizes (e.g. Cohen's $d$ , Pearson's $r$ ), indicating how they were calculated  |       |
| Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.   |       |
| Software and code   |       |
| Policy information about <u>availability of computer code</u>   |       |

Data collection

Two-photon calcium imaging was performed using a commercially available multi-photon microscopy system from Bruker (formerly Prairie Technologies). Behavior data were collected using custom software implemented in Java via a microcontroller (Arduino DUE) on the treadmill. LFP data were collected using Intan RHD2000 recording system.

Data analysis

Initial motion correction of two-photon imaging data was performed using the SIMA software package, and ROI detection was performed manually. Subsequent imaging and LFP analysis was performed using custom-written code in Python.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

#### Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Data sets included in this study will be available upon publication.

#### Research involving human participants, their data, or biological material

Policy information about studies with <u>human participants or human data</u>. See also policy information about <u>sex, gender (identity/presentation)</u>, <u>and sexual orientation</u> and <u>race, ethnicity and racism</u>.

Reporting on sex and gender

Use the terms sex (biological attribute) and gender (shaped by social and cultural circumstances) carefully in order to avoid confusing both terms. Indicate if findings apply to only one sex or gender; describe whether sex and gender were considered in study design; whether sex and/or gender was determined based on self-reporting or assigned and methods used. Provide in the source data disaggregated sex and gender data, where this information has been collected, and if consent has been obtained for sharing of individual-level data; provide overall numbers in this Reporting Summary. Please state if this information has not been collected.

Report sex- and gender-based analyses where performed, justify reasons for lack of sex- and gender-based analysis.

Reporting on race, ethnicity, or other socially relevant groupings

Please specify the socially constructed or socially relevant categorization variable(s) used in your manuscript and explain why they were used. Please note that such variables should not be used as proxies for other socially constructed/relevant variables (for example, race or ethnicity should not be used as a proxy for socioeconomic status).

Provide clear definitions of the relevant terms used, how they were provided (by the participants/respondents, the researchers, or third parties), and the method(s) used to classify people into the different categories (e.g. self-report, census or administrative data, social media data, etc.)

Please provide details about how you controlled for confounding variables in your analyses.

Population characteristics

Describe the covariate-relevant population characteristics of the human research participants (e.g. age, genotypic information, past and current diagnosis and treatment categories). If you filled out the behavioural & social sciences study design questions and have nothing to add here, write "See above."

Recruitment

Describe how participants were recruited. Outline any potential self-selection bias or other biases that may be present and how these are likely to impact results.

Ethics oversight

Identify the organization(s) that approved the study protocol.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

### Field-specific reporting

| Please select the one belov   | w that is the best lit for your research. If you are not sure, read the appropriate sections before making your selection. |  |  |  |
|---|--|--|--|--|
| ☐ Life sciences   | Behavioural & social sciences Ecological, evolutionary & environmental sciences  |  |  |  |
| For a reference copy of the document with all sections, see <a href="mature.com/documents/nr-reporting-summary-flat.pdf">nature.com/documents/nr-reporting-summary-flat.pdf</a> |  |  |  |  |

# Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

No statistical methods were used to determine sample size a priori.

Data exclusions

Recording locations with a raw signal SNR < 10 based on shot-noise calibration were excluded.

Replication

Main effects were consistent across 11 mice.

Randomization

Randomization was not required as there is only one group.

Blinding

Blinding was not required as there is only one group.

#### Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

| Materials & experime   | ntal systems   | Methods   |  |
|--|--|---|--|
| n/a   Involved in the study  |  | n/a   Involved in the study   |  |
| Antibodies   |  | ChIP-seq  |  |
| Eukaryotic cell lines  |  | Flow cytometry  |  |
| Palaeontology and a  | rchaeology   | MRI-based neuroimaging  |  |
| Animals and other o  | rganisms   |   |  |
| Clinical data  |  |   |  |
| Dual use research o  | f concern  |   |  |
| ∑ Plants   |  |   |  |
| '  |  |   |  |
| Eukaryotic cell lin  | es   |   |  |
| Policy information about <u>ce</u>   | ell lines and Sex and Gen  | nder in Research  |  |
| Cell line source(s)  | HEK293A cells were obtained from Thermo Fisher (RRID:CVCL_6910).   |   |  |
| Authentication   |  | rformance metrics of voltage indicators in these cells are consistent with published results and with further eurons, cell lines were not authenticated.  |  |
| Mycoplasma contaminati   | on All cell lines were   | confirmed to be mycoplasma free before use in this study.   |  |
| Commonly misidentified lines (See ICLAC register)  No commonly misiden       |  | sidentified cell lines were used.   |  |
| (see <u>repre</u> register)  |  |   |  |
| Animals and othe   | r research orga  | nisms   |  |
| Policy information about <u>st</u><br>Research                               | udies involving animals;   | ARRIVE guidelines recommended for reporting animal research, and Sex and Gender in  |  |
| Laboratory animals   | Experiments were performed in adult (8-10 weeks) male and female wild-type C57BL/6J mice.                                      |   |  |
| Wild animals   | The present study did not involve wild animals.  |   |  |
| Reporting on sex   | The present study involve both male and female mice.   |   |  |
| Field-collected samples  | The present study did not involve Field-collected samples.   |   |  |
| Ethics oversight   | All experiments and procedures were conducted in accordance with the U.S. NIH Guide for the Care and Use of Laboratory Animals |   |  |
| and the Institutional Animal Care and Use Committees of Columbia University. |  |   |  |
| Note that full information on t  | he approval of the study pr  | rotocol must also be provided in the manuscript.  |  |
|  |  |   |  |
| Plants   |  |   |  |
| 6 1 1  |  |   |  |
| Seed stocks  |  | ll seed stocks or other plant material used. If applicable, state the seed stock centre and catalogue number. If lected from the field, describe the collection location, date and sampling procedures. |  |
| Novel plant genotypes  | Describe the methods by which all novel plant genotypes were produced. This includes those generated by transgenic approaches, |   |  |

 $number\ of\ independent\ lines\ analyzed\ and\ the\ generation\ upon\ which\ experiments\ were\ performed.\ For\ gene-edited\ lines,\ describe$ the editor used, the endogenous sequence targeted for editing, the targeting guide RNA sequence (if applicable) and how the editor

Authentication

was applied. Describe any authentication procedures for each seed stock used or novel genotype generated. Describe any experiments used to  $assess\ the\ effect\ of\ a\ mutation\ and,\ where\ applicable,\ how\ potential\ secondary\ effects\ (e.g.\ second\ site\ T-DNA\ insertions,\ mosiacism,$ off-target gene editing) were examined.