Backbone-Determined Antiarrhythmic Structure-Activity Relationships for a Mirror- Image, Oligomeric Depsipeptide Natural Product
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Figure S101. ¹ H NMR (600 MHz, CDCl ₃) of 2.5
Figure S102. ¹³ C NMR (150 MHz, CDCl ₃) of 2.5
Figure S103. ¹ H NMR (600 MHz, CDCl ₃) of 3.1
Figure S104. ¹³ C NMR (150 MHz, CDCl ₃) of 3.1
Figure S105. ¹ H NMR (600 MHz, CDCl ₃) of 3.2161
Figure S106. ¹³ C NMR (150 MHz, CDCl ₃) of 3.2
Figure S107. ¹ H NMR (600 MHz, CDCl ₃) of 3.3
Figure S108. ¹³ C NMR (150 MHz, CDCl ₃) of 3.3
Figure S109. ¹ H NMR (600 MHz, CDCl ₃) of 3.4
Figure S110. ¹³ C NMR (150 MHz, CDCl ₃) of 3.4
Figure S111. ¹ H NMR (600 MHz, CDCl ₃) of 3.5167
Figure S112. ¹³ C NMR (150 MHz, CDCl ₃) of 3.5
Figure S113. ¹ H NMR (600 MHz, CDCl ₃) of 4.2
Figure S114. ¹³ C NMR (150 MHz, CDCl ₃) of 4.2
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Figure S116. ¹³ C NMR (150 MHz, CDCl ₃) of 4.3	172
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Experimental Section

General Information: Glassware was flame-dried under vacuum for all non-aqueous reactions. All reagents and solvents were commercial grade and purified prior to use when necessary. Toluene, THF and dichloromethane (CH₂Cl₂) were dried by passage through a column of activated alumina as described by Grubbs.¹ Flash column chromatography was performed using Sorbent Technologies 230-400 mesh silica gel with solvent systems indicated. Analytical thin layer column chromatography was performed using Sorbent Technologies 250 µm glass backed UV254 silica gel plates and were visualized by fluorescence upon 250 nm radiation and/or the by use of TLC stain. Solvent removal was effected by rotary evaporation under vacuum (~ 25-40 mm Hg). All extracts were dried with Na₂SO₄ unless otherwise noted. Preparative HPLC was performed on an Agilent 1260 system (column: Zorbax Eclipse XDB-C18; 21.2 mm x 150 mm, 5 µm, flow rate 8 mL/min) with 210 nm monitoring wavelength and acetonitrile/water (+0.1% TFA) gradient as indicated. Nuclear magnetic resonance spectra (NMR) were acquired on a Bruker AV-400 (400 MHz) or Bruker AV II-600 (600 MHz) instrument. Mass spectra were recorded by use of electron impact ionization (EI), or electro-spray ionization (ESI) on a high resolution TQ-Orbitrap 3 XL Penn or Orbitrap 2 Classic FPG in the Vanderbilt Mass Spectrometry Core Laboratory. IR spectra were recorded on a Nicolet Avatar 360 spectrophotometer and are reported in wavenumbers (cm⁻¹) as neat films on a NaCl plate (transmission). Melting points were measured using an OptiMelt automated melting point system (Stanford Research Systems) and are not corrected. Chiral HPLC analysis was conducted on an Agilent 1200 series Infinity instrument using a ChiralPak column. Optical rotations were measured on a Jasco P-2000 polarimeter.

Purity Statement: Although each macrocycle was purified by preparative HPLC, and detected by UV (210 nm), the compounds reported often lack a good chromophore. However, the use of HPLC in combination with NMR analysis led us to judge compounds to be a minimum of 90% pure, likely >95%. Images of NMR spectra of all new compounds are provided.

Biological Experiments: All animal studies were approved (protocol number M1900081) and carried out in accordance with the guidelines and procedures set forth by the Vanderbilt Division of Animal Care. Cardiomyocytes were isolated from C57BI/6J mice, as previously reported.² Calcium spark experiments were conducted, as previously reported.³ Briefly, ventricular cardiomyocytes were adhered to laminin-coated glass bottom culture dishes, permeabilized with saponin, and incubated with an internal solution containing the calcium sensitive dye, Fluo-4. Elementary spark recordings were made using an Olympus inverted microscope equipped with a solid-state diode laser at 488 nm for excitation, 40× silicone objective (1.25 NA), and Hamamatsu CMOS camera for detection. The internal solution contained vehicle or compound and all data were collected 15 – 30 minutes after drug application. Spark detection analysis was performed using SparkMaster2.⁴ Statistical analysis was carried out in R using a hierarchical clustering model to account for mouse-to-

¹ Pangborn, A. B.; Giardello, M. A.; Grubbs, R. H.; Rosen, R. K.; Timmers, F. J. Safe and convenient procedure for solvent purification. *Organometallics* **1996**, *15*, (5), 1518.

² Batiste, S. M.; Blackwell, D. J.; Kim, K.; Kryshtal, D. O.; Gomez-Hurtado, N.; Rebbeck, R. T.; Cornea, R. L.; Johnston, J. N.; Knollmann, B. C. Unnatural verticilide enantiomer inhibits type 2 ryanodine receptor-mediated calcium leak and is antiarrhythmic. *Proc. Natl. Acad. Sci. U. S. A.* **2019**, *116*, 4810.

³ Smith, A. N.; Blackwell, D. J.; Knollmann, B. C.; Johnston, J. N. Ring Size as an Independent Variable in Cyclooligomeric Depsipeptide Antiarrhythmic Activity. *ACS Med. Chem. Lett.* **2021**, *12*, (12), 1942.

⁴ Tomek, J.; Nieves-Cintron, M.; Navedo, M. F.; Ko, C. Y.; Bers, D. M. SparkMaster 2: A New Software for Automatic Analysis of Calcium Spark Data. *Circ Res* **2023**, *133*, (6), 450.

mouse variability provided by Sikkel et al.⁵ Bonferroni-adjusted P values are reported in the figure legends. A cutoff value of 0.05 was used as the threshold to reject the null hypothesis.

Safety: No unexpected or unusually high safety hazards were encountered.

Biological Experiment Data: Calcium spark parameters

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For all series, assay is run with 25 μ M solution of drug. Values are normalized to the vehicle (DMSO) condition and reported as mean ± standard deviation. Statistical comparisons were made using a hierarchical clustering model with Bonferroni correction. Adjusted p-values < 0.05 vs vehicle are indicated with an asterisk. Each mouse was an independent experiment, the replicates are listed as n (cells).

Series 1	Vehicle (DMSO)	<i>ent</i> -verticilide (1)	1.1	1.2	1.3	1.4	1.5
N (mice)	12	2	5	5	5	4	4
n (cells)	292	29	102	105	108	119	87
Frequency	1.00 ± 0.28	0.51 ± 0.36*	1.03 ± 0.27	0.92 ± 0.28	0.96 ± 0.29	0.90 ± 0.36	1.00 ± 0.31
Amplitude	1.00 ± 0.26	0.82 ± 0.43*	1.01 ± 0.25	0.99 ± 0.24	1.04 ± 0.28	0.93 ± 0.24	1.00 ± 0.25
Mass	1.00 ± 1.15	0.057 ± 0.47*	0.98 ± 0.59	0.95 ± 0.51	0.91 ± 0.48	0.84 ± 0.32	1.00 ± 0.32
Leak	1.00 ± 1.20	0.36 ± 0.38*	1.02 ± 0.71	0.91 ± 0.63	0.90 ± 0.60	0.77 ± 0.53	0.91 ± 0.46

Supplement Table 1. Calcium spark parameters from murine cardiomyocytes for ent-verticilide and series 1 analogs.

	1				
Series 2	2.1	2.2	2.3	2.4	2.5
N (mice)	2	2	2	2	2
n (cells)	38	46	54	50	50
Frequency	0.51 ± 0.25	0.83 ± 0.20	1.01 ± 0.20	0.97 ± 0.17	1.04 ± 0.20
Amplitude	0.93 ± 0.31	1.05 ± 0.24	0.92 ± 0.19	1.08 ± 0.22	1.17 ± 0.34
Mass	1.04 ± 0.64	0.95 ± 0.39	1.02 ± 0.43	0.98 ± 0.39	1.06 ± 0.35
Leak	0.60 ± 0.65	0.79 ± 0.44	1.03 ± 0.53	0.97 ± 0.56	1.09 ± 0.46

Supplement Table 2. Calcium spark parameters from murine cardiomyocytes for series 2 analogs.

1

Series 3	3.1	3.2	3.3	3.4	3.5
N (mice)	4	3	4	5	5
n (cells)	79	58	79	80	91
Frequency	0.79 ± 0.29	0.97 ± 0.28	0.96 ± 0.26	1.15 ± 0.26	0.94 ± 0.25
Amplitude	0.99 ± 0.20	1.02 ± 0.20	1.00 ± 0.16	0.99 ± 0.18	0.97 ± 0.19
Mass	0.81 ± 0.55	1.03 ± 0.50	0.96 ± 0.42	0.98 ± 0.45	1.07 ± 0.64
Leak	0.75 ± 0.63	0.99 ± 0.53	0.93 ± 0.45	1.20 ± 0.63	1.04 ± 0.71

Supplement Table 3. Calcium spark parameters from murine cardiomyocytes for series 3 analogs.

⁵ Sikkel, M. B.; Francis, D. P.; Howard, J.; Gordon, F.; Rowlands, C.; Peters, N. S.; Lyon, A. R.; Harding, S. E.; MacLeod, K. T. Hierarchical statistical techniques are necessary to draw reliable conclusions from analysis of isolated cardiomyocyte studies. *Cardiovasc. Res.* **2017**, *113*, (14), 1743.

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Series 4	4.1	4.2	4.3	4.4	4.5	4.6
N (mice)	4	2	2	2	2	2
n (cells)	66	42	37	38	43	37
Frequency	0.96 ± 0.28	0.87 ± 0.29	1.08 ± 0.27	1.02 ± 0.24	1.16 ± 0.30	1.08 ± 0.36
Amplitude	1.01 ± 0.31	1.24 ± 0.35	1.32 ± 0.31	1.16 ± 0.31	1.17 ± 0.22	1.15 ± 0.22
Mass	1.19 ± 1.03	1.57 ± 0.89	1.66 ± 0.94	1.42 ± 1.09	1.29 ± 0.40	1.10 ± 0.52
Leak	1.19 ± 1.10	1.35 ± 1.14	1.79 ± 1.49	1.38 ± 1.21	1.43 ± 0.66	1.15 ± 0.71

Supplement Table 4. Calcium spark parameters from murine cardiomyocytes for series 4 analogs.

Series 5	5.1	5.2	5.3	5.4	5.5	5.6
N (mice)	4	5	5	5	5	4
n (cells)	66	85	93	100	89	67
Frequency	0.96 ± 0.28	0.79 ± 0.35	0.94 ± 0.31	0.89 ± 0.29	1.02 ± 0.33	1.11 ± 0.29
Amplitude	1.01 ± 0.31	1.13 ± 0.23	1.08 ± 0.18	1.02 ± 0.21	1.06 ± 0.18	1.07 ± 0.19
Mass	1.19 ± 1.03	1.11 ± 0.47	1.17 ± 0.47	1.13 ± 0.69	0.96 ± 0.42	1.30 ± 0.75
Leak	1.19 ± 1.10	1.08 ± 0.66	1.23 ± 0.57	1.14 ± 0.69	1.14 ± 0.65	1.54 ± 1.17

Supplement Table 5. Calcium spark parameters from murine cardiomyocytes for series 5 analogs.

General Procedures

tert-Butyloxycarbonyl (Boc) Deprotection: A round-bottom flask was charged with the depsipeptide (1 equiv) and dissolved in either 4 M HCl/ethyl acetate (1 M in depsipeptide) or 20% TFA in DCM. The reaction was allowed to stir for 1-3 h at ambient temperature. The crude reaction mixture was concentrated, ether was added, and the mixture was then reconcentrated. This procedure was repeated 3 times with diethyl ether.

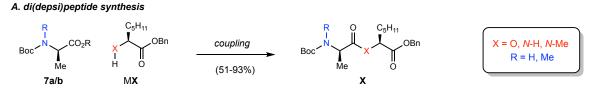
Benzyl Deprotection: A round-bottom flask was charged with the depsipeptide (1 equiv) dissolved in methanol or ethyl acetate (0.1 M) and treated with 10% Pd/C (20 mol%). The reaction flask was evacuated with a light vacuum (~40 Torr). Hydrogen (balloon) was added and then the flask was cycled through a light vacuum three times. The reaction was stirred for 1.5-3 h. After purging with argon, the crude reaction mixture was filtered through Celite and concentrated to afford the carboxylic acid.

PyBrop Coupling Reaction: A round-bottom flask was charged with the two depsipeptide coupling partners (1:1), along with dry DCM or DMF (0.1 M). The solution was cooled to 0 °C and placed under argon. PyBrop (1.5 equiv) and freshly distilled DIPEA (3 equiv) were added. The reaction was allowed to stir for 30 minutes at 0 °C and 1.5-18 h at ambient temperature. Reaction progression was monitored by TLC and/or TLC-MS. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography.

PyBOP Macrocyclization: The reaction mixture was added to a flame-dried round bottom flask. Dry DMF or DCM (0.005 M) was added and the reaction was cooled to 0 °C. Once at 0 °C, freshly distilled DIPEA (3 equiv) and PyBop (1.05 equiv) were added. The reaction was stirred at 0 °C for 30 minutes, then allowed to warm to ambient temperature and stir for an additional 1.5-3 h. Reaction progression was monitored by TLC and/or TLC-MS. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. The crude reaction mixture was purified via preparative HPLC.

HATU Coupling Reaction: A round-bottom flask was charged with the amine and acid (1:1), and dry DMF or DCM (0.1 M). The mixture was cooled to 0 °C and then freshly distilled DIPEA (3 equiv) and HATU (3 equiv) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 3-18 h. Reaction progression was monitored by TLC and/or TLC-MS. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography.

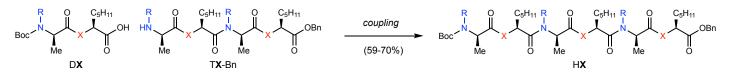
General Reaction Schemes



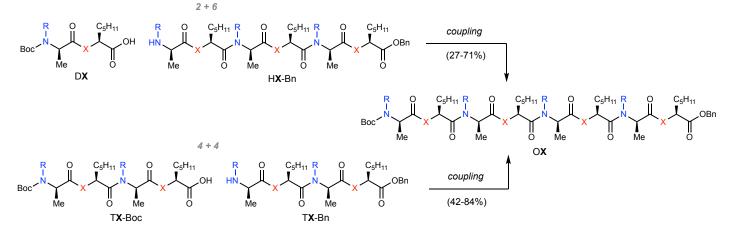
B. tetra(depsi)peptide synthesis



C. hexa(depsi)peptide synthesis

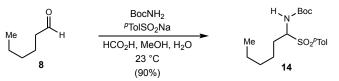


D. octa(depsi)peptide synthesis



Scheme 1. A-D. Generalized reaction schemes for the synthesis of various intermediates. Individual reaction conditions can be found in the Experimental Procedures section.

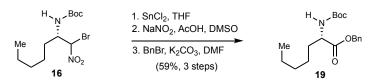
Experimental Procedures and Characterization Data for Reported Compounds



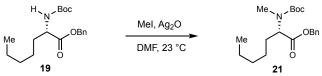
1-Tosylhexan-1-amine (14). A round-bottom flask was charged with hexanal (9.0 mL, 73 mmol), *tert*-butyl carbamate (5.7 g, 49 mmol), and MeOH (73 mL) and stirred until it became a homogenous solution. NaSO₂^{*p*}Tol (17.4 g, 97.6 mmol) was added, along with enough H₂O to dissolve the solids (100 mL). Then formic acid (3.68 mL, 97.6 mmol) was added, and the mixture was allowed to stir at ambient temperature under argon for 4 d. The reaction mixture was filtered and washed with H₂O and hexanes to afford the product as a white solid (15.5 g, 90%). Mp 107- 110 °C; R*f* = 0.26 (10% EtOAc/hexanes); IR (film) 3334, 2958, 2930, 2861, 1721, 1597, 1518, 1456, 1392, 1316, 1245, 1167, 1142, 1085 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) The small doubling of peaks is due to cis and trans amide rotamers. The largely favored isomer is listed here: δ 7.76 (d, *J* = 8.8 Hz, 2H), 7.30 (d, *J* = 7.9 Hz, 2H), 5.10 (d, *J* = 11.0 Hz, 1H), 4.79 (td, *J* = 10.9, 3.4 Hz, 1H), 2.38 (s, 3H), 2.24-2.16 (m, 1H), 1.74-1.65 (m, 1H), 1.55-1.23 (m, 6H), 1.19 (s, 9H) 0.86 (t, *J* = 6.5 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) ppm 154.0, 144.9, 134.0, 129.7, 129.4, 80.6, 31.2, 28.0, 27.7, 26.4, 25.0, 22.4, 21.7, 14.0; HRMS (EI): Exact mass calcd for C₁₈H₂₉NNaO₄S [M+Na]⁺ 378.1710, found 378.1719.



tert-Butyl ((2S)-1-bromo-1-nitroheptan-2-yl)carbamate (16). A round-bottom flask was charged with sulfone (6.00 g, 16.8 mmol), Cs₂CO₃ (27.5 g, 84.4 mmol), and toluene (160 mL). The mixture was allowed to stir at ambient temperature under argon for 6 h. The reaction mixture was filtered through a pad of Celite and concentrated. The crude oil was then dissolved in toluene (248 mL) and to it was added (S,S)-PBAM as its triflic acid salt (217.5 mg, 331 µmol). The reaction was cooled to -60 °C and bromonitromethane (1.74 mL, 24.8 mmol) was added. The reaction was allowed to stir for 48 h. The reaction mixture was quenched by running it through a short silica plug while still cold, with 100% EtOAc. The fractions were combined and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5% ethyl acetate in hexanes) to afford the α -bromo nitroalkane (1:1 dr (¹H NMR)) as a white solid (4.5 g, 88% 2-step); The diastereomers were determined to be 90% and 89% ee by chiral HPLC analysis (Chiralcel AD-H, 1% PrOH /hexanes, 1.0 mL/min, tr(d1, major) = 19.6 min, tr(d1, minor) = 21.1 min, tr(d2, minor) = 25.1 min, tr(d2, major) = 29.4 min). $[\alpha]_{D}^{23}$ -20 (c 0.53, CHCl₃); Mp 62-66 °C; Rf = 0.60 (10% EtOAc/hexanes); IR (film) 3332, 2958, 2931, 2861, 1706, 1567, 1501, 1458, 1392, 1367, 1248, 1165, 1045 cm⁻¹; ¹H NMR (600 MHz, CDCl₃, 1:1 mixture of diastereomers) δ 6.19 (d, J = 4.4 Hz, 1H), 6.17 (d, J = 3.0 Hz, 1H), 4.83 (d, J = 8.8 Hz, 1H), 4.73 (d, J = 9.1 Hz, 1H), 4.34 (dddd, J = 9.2, 9.2, 3.9, 3.9 Hz, 1H), 4.24 (dddd, J = 9.2, 9.2, 4.6, 4.6 Hz, 1H), 1.90-1.52 (series of m, 4H), 1.46 (s, 9H), 1.44 (s, 9H), 1.40-1.22 (m, 12H), 0.90 (t, J = 7.0 Hz, 3H), 0.88 (t, J = 6.9 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃, 1:1 mixture of diastereomers) ppm 155.2, 155.1, 84.5, 83.2, 80.9, 80.8, 55.0, 54.5, 31.32, 31.28, 30.5, 28.5, 28.4, 28.3, 25.6, 25.5, 22.52, 22.51, 14.0(2C); HRMS (EI): Exact mass calcd for C₁₂H₂₃BrN₂NaO₄ [M+Na]⁺ 361.0733, found 361.0737.



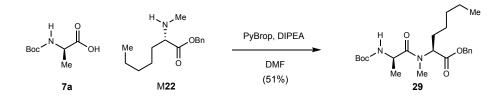
Benzyl (S)-2-((tert-butoxycarbonyl)amino)heptanoate (19). A flame-dried round-bottom flask was charged with halonitro alkane (383 mg, 1.13 mmol), tin chloride (428 mg, 2.26 mmol), and THF (11.3 mL). The mixture was allowed to stir at ambient temperature under argon for 12 h. The reaction mixture was then poured into H₂O, and diethyl ether was added into the separatory funnel. The organic layer was filtered through a pad of Celite, washed with H₂O (three times), dried, and concentrated to afford a pale-yellow oil. The crude oil was then dissolved in DMSO (8.7 mL) and to it was added NaNO₂ (234 mg, 3.39 mmol) and AcOH (885 µL, 17.0 mmol). The reaction was heated to 60 °C and allowed to stir for 12 h. The reaction mixture was allowed to cool to ambient temperature, guenched with 1 M ag HCl, poured into a separatory funnel, and extracted with EtOAc. The organic layers were washed with ice water (four times), dried, and concentrated. The crude reaction mixture was dissolved in DMF (2.1 mL), and to it was added K₂CO₃ (427 mg, 3.09 mmol) and BnBr (148 μL, 1.24 mmol). The reaction was allowed to stir at ambient temperature under argon for 12 h. The reaction was guenched with 1 M aq HCl and then extracted with EtOAc. The organic layers were then dried and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 10% ethyl acetate in hexanes) to afford the product as a colorless oil $(366 \text{ mg}, 59\% (3 \text{ steps})); [\alpha]_D^{23} - 5.39 (c 1.08, CHCl_3); Rf = 0.33 (10\% EtOAc/hexanes); IR (film) 3365, 2957, 2931, 2861, 1717, 10\% EtOAc/hexanes); IR (film) 3365, 2957, 2931, 2861, 28$ 1499, 1456, 1366, 1249, 1161, 1048 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.40-7.28 (m, 5H), 5.21 (d, J = 12.4 Hz, 1H), 5.12 (d, J = 12.4 Hz, 1H), 5.02 (d, J = 7.8 Hz, 1H), 4.35 (ddd, J = 11.7, 6.2, 6.2 Hz, 1H), 1.85-1.70 (m, 1H), 1.68-1.55 (m, 1H), 1.43 (s, 9H), 1.36-1.17 (m, 6H), 0.85 (t, J = 6.3 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃) ppm 172.9, 156.5, 136.6, 128.7, 128.4, 128.3, 79.8, 66.9, 53.7, 32.7, 31.4, 28.4, 24.9, 22.5, 14.0; HRMS (EI): Exact mass calcd for C₁₉H₂₉NNaO₄ [M+Na]⁺ 358.1989, found 358.1993.



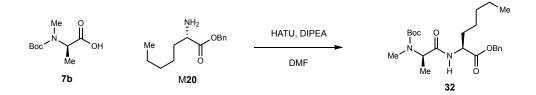
Benzyl (5)-2-(*(tert*-butoxycarbonyl)(methyl)amino)heptanoate (21). A flame-dried round-bottom flask was charged with the α-amino ester (266 mg, 793 µmol), MeI (789 µg, 12.69 mmol), Ag₂O (735 mg, 3.17 mmol) and DMF (7.9 mL). The reaction vessel was wrapped in aluminum foil, and the mixture was allowed to stir at ambient temperature under argon for 12 h. The reaction mixture was filtered through a pad of Celite and concentrated, and then dissolved in EtOAc, washed with H₂O, dried, and concentrated to afford a pale-yellow oil. The crude residue was subjected to flash column chromatography (SiO₂, 5% ethyl acetate in hexanes) to afford the product as a colorless oil (244 mg, 87%);[α]²³_D -20.6 (*c* 1.09, CHCl₃); Rf = 0.57 (10% EtOAc/hexanes); IR (film) 3033, 2931, 2860, 1743, 1697, 1455, 1391, 1366, 1327, 1149 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) Several peaks show a clear doubling for the *cis* and *trans* amide products. Other peaks are overlapping and appear only once. Those that are doubled have both values listed. δ 7.40-7.28 (m, 5H), 5.15 (broad s, 2H), 4.80/4.47 (dd, *J* = 10.5, 4.8 Hz, 1H), 2.82/2.75 (s, 3H), 1.97-1.86 (m, 1H), 1.79-1.63 (m, 1H), 1.45/1.40 (s, 9H), 1.36-1.23 (m, 6H), 0.88/0.87 (t, *J* = 6.5 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) ppm) The line listing for both *cis* and *trans* amide isomers is given, as a clear doubling of many peaks is evident. Whenever the isomer peaks overlap, it is stated that the particular peak is representing 2 or more carbons. 172.2, 172.0, 156.5, 155.8, 136.0, 135.8, 128.7, 128.6, 128.4, 128.2, 128.1, 128.0, 80.3, 80.0, 66.7, 66.6, 59.3, 57.9, 31.4(2C), 30.9, 30.5, 29.1, 28.7, 28.5, 28.4, 25.8(2C), 22.6(2C), 14.1, 14.0; HRMS (EI): Exact mass calcd for C₂₀H₃₁NNaO₄ [M+Na]⁺ 372.2145, found 372.2149.



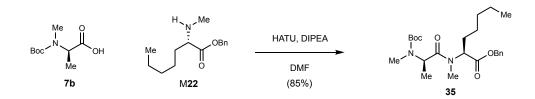
Benzyl (5)-2-(((*tert***-butoxycarbonyl)-***D***-alanyl)oxy)heptanoate (23). A round-bottom flask was charged with the amine (1.98 g, 9.73 mmol), alcohol (2.00 g, 8.46 mmol), and DCM (84.6 mL). The mixture was cooled to 0 °C and then EDCI (1.95 g, 10.2 mmol) and DMAP (51.7 mg, 424 µmol) were added. The reaction mixture was stirred at 0 °C for 30 min, and then allowed to warm to ambient temperature and stir for an additional 1.5 h before it was poured into water and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 2.5-20% ethyl acetate in hexanes) to afford the product as a white amorphous solid (2.54 g, 88%). [\alpha]_D^{24} –11.2 (***c* **0.43, CHCl₃); R_f = 0.29 (10% EtOAc/hexanes); IR (film) 2954, 2926, 2856, 1745, 1666, 1455, 1408, 1306, 1212, 1107, 1076 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) \delta 7.39-7.31 (series of m, 5H), 5.20 (d,** *J* **= 12.1 Hz, 1H), 5.14 (d,** *J* **= 12.1 Hz, 1H), 5.07 (t,** *J* **= 3.7 Hz, 1H), 5.02 (br m, 1H), 4.46-4.36 (br m, 1H), 1.89-1.82 (br m, 2H), 1.44 (s, 9H), 1.40 (br d,** *J* **= 7.3 Hz, 3H), 1.37-1.33 (br m, 2H), 1.28-1.24 (br m, 4H), 0.88 (br t,** *J* **= 6.8 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) ppm 172.7, 169.7, 155.9, 135.2, 128.6, 128.4, 128.3, 72.9, 70.5, 67.3, 67.0, 49.5, 31.2, 30.9, 28.3, 24.7, 18.6, 14.0; HRMS (EI): Exact mass calcd for C₂₂H₃₃NO₆Na [M+Na]⁺ 430.2206, found 430.2203. MPT-2-139.**



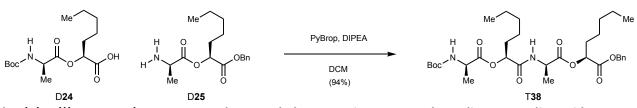
Benzyl (5)-2-((*R***)-2-((***tert***-butoxycarbonyl)amino)-***N***-methylpropanamido)heptanoate (29). A round-bottom flask was charged with the amine (1.20 g, 4.80 mmol), acid (908 mg, 4.80 mmol), and DMF (4.2 mL). The mixture was cooled to 0 °C and then DIPEA (2.52 mL, 14.4 mmol) and PyBrop (3.52 g, 7.20 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to room temperature and stirred overnight. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-30% ethyl acetate in hexanes) to afford the product as a white amorphous solid (1.07 g, 51%). [\alpha]_D^{24} + 7.2 (***c* **0.44, CHCl₃); R_f = 0.60 (25% EtOAc/hexanes); IR (film) 2930, 1744, 1698, 1456, 1390, 1156 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) \delta 7.38-7.32 (series of m, 5H), 6.85 (br d,** *J* **= 7.2 Hz, 1H), 5.25-5.21 (series of m, 2H), 5.03 (m, 1H), 4.70-4.21 (m, 1H), 3.16-2.62 (series of m, 3H), 1.84 (br m, 1H), 1.58 (br m, 2H), 1.45 (series of m, 9H), 1.36 (m, 2H), 1.28-1.23 (series of m, 6H), 0.86 (m J = 6.9 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) ppm 175.3, 171.0, 156.1, 135.3, 128.9, 128.4, 128.6, 128.5, 79.9, 72.8, 67.8, 52.8, 34.4, 31.9, 29.7, 28.4, 24.9, 22.3, 13.9; HRMS (EI): Exact mass calcd for C₂₃H₃₇N₂O₅ [M+H]⁺ 421.2702, found 421.2700. MPT-4-163.**



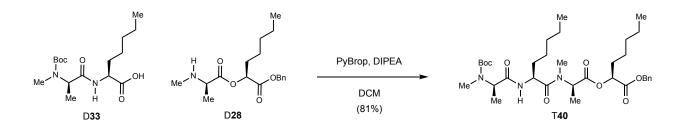
Benzyl (5)-2-((*R***)-2-((***tert***-butoxycarbonyl)(methyl)amino)propanamido)heptanoate (32). A round-bottom flask was charged with the amine (990 mg, 4.21 mmol), acid (855 mg, 4.21 mmol), and DMF (4.2 mL). The mixture was cooled to 0 °C and then DIPEA (2.20 mL, 12.6 mmol) and HATU (2.40 g, 6.32 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to room temperature and stirred overnight. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-30% ethyl acetate in hexanes) to afford the product as a white amorphous solid (1.15 g, 74%). [\alpha]_D^{24} + 29 (***c* **0.43, CHCl₃); R_f = 0.42 (30% EtOAc/hexanes); IR (film) 3330, 2957, 2932, 2861, 1739, 1692, 1525, 1456, 1368, 1154, 1091, 1047 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) \delta 7.37-7.32 (series of m, 5H), 6.72 (br m, 1H), 5.15-3.77 (series of m, 4H), 3.03-2.71 (series of m, 3H), 1.82 (br m, 1H), 1.65 (br m, 1H), 1.48-1.43 (series of m, 9H), 1.32-1.23 (series of m, 9H), 0.85 (br t,** *J* **= 6.9 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) ppm 172.8, 172.2, 159.9, 130.7, 129.1, 128.9, 128.7, 81.0, 67.6, 52.8, 46.3, 32.7, 31.9, 30.4, 25.6, 25.5, 24.9, 23.0, 14.5; HRMS (EI): Exact mass calcd for C₂₃H₃₇N₂O₅ [M+H]⁺ 421.2702, found 421.2684. MPT-4-135**



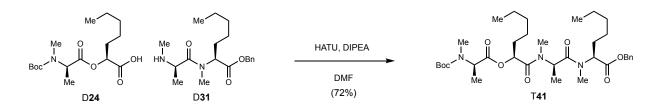
Benzyl (*S*)-2-((*R*)-2-((*tert*-butoxycarbonyl)(methyl)amino)-*N*-methylpropanamido)heptanoate (35). A round-bottom flask was charged with the amine (900 mg, 3.61 mmol), acid (734 mg, 3.61 mmol), and DMF (36.1 mL). The mixture was cooled to 0 °C and then DIPEA (1.89 mL, 10.8 mmol) and HATU (4.11 g, 10.8 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to room temperature and stirred overnight. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, ice water, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-20% ethyl acetate in hexanes) to afford the product as a white amorphous solid (1.33 g, 85%). $[\alpha]_D^{23}$ + 50 (*c* 0.34, CHCl₃); R_f = 0.20 (10% EtOAc/hexanes); IR (film) 2957, 2931, 2861, 1740, 1659, 1456, 1390, 1156 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.37-7.29 (series of m, 5H), 5.25-4.51 (series of m, 4H), 2.92-2.55 (series of m, 6H), 1.98 (m, 1H), 1.69 (m, 1H), 1.46 (s, 9H), 1.34-1.20 (series of m, 9H), 0.88 (t, *J* = 6.0 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) ppm 171.9, 171.3, 155.3, 135.6, 128.6, 128.4, 128.2, 80.0, 66.9, 56.9, 50.9, 31.3, 30.5, 29.0, 28.7, 28.4, 28.0, 25.7, 22.4, 13.9; HRMS (EI): Exact mass calcd for C₂₄H₃₈N₂NaO₅ [M+Na]⁺ 457.2678, found 457.2675. MPT-3-032



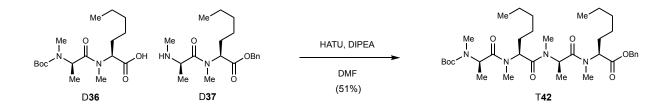
Benzyl (*R*)-2-(((*6R*,9*R*,12*R*)-2,2,5,6,11,12-hexamethyl-4,7,10-trioxo-9-pentyl-3,8-dioxa-5,11-diazatridecan-13-oyl)oxy) heptanoate (T38). A round-bottom flask was charged with the amine (402 mg, 1.31 mmol), acid (415 mg, 1.31 mmol), and DCM (12.0 mL). The mixture was cooled to 0 °C and then DIPEA (683 μL, 3.92 mmol) and PyBrop (638 mg, 1.31 mmol) were added. The reaction was stirred at 0 °C for 1 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a light yellow oil (745 mg, 94%). [*α*]_D²⁴ –20.1 (*c* 0.62, CHCl₃); R_f= 0.52 (30% EtOAc/hexanes); IR (film) 3312, 2956, 2930, 2861, 1747, 1692, 1530, 1455, 1367, 1255, 1160, 1069 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.40-7.35 (series of m, 5H), 7.05 (br d, *J* = 5.3 Hz, 1H), 5.27-5.16 (series of m, 3H), 5.11 (br t, *J* = 6.5 Hz, 1H), 4.99 (br d, *J* = 5.3 Hz, 1H), 4.69 (dd, *J* = 7.1, 7.1 Hz, 1H), 1.99-1.93 (b m, 1H), 1.88-1.80 (br m, 3H), 1.49-1.43 (series of m, 14H), 1.39-1.28 (series of m, 13H), 0.90-0.88 (series of m, 6H); ¹³C NMR (150 MHz, CDCl₃) ppm 173.4, 172.5, 170.3, 169.9, 156.0, 135.9, 129.2, 129.0, 128.9, 128.8, 80.9, 74.9, 73.5, 67.9, 67.6, 50.4, 48.7, 32.2, 31.9, 31.8, 31.6, 28.9, 25.3, 23.0, 22.9, 18.2, 18.0, 14.5; HRMS (EI): Exact mass calcd for C₃₂H₅₀N₂O₉Na [M+Na]⁺ 629.3414, found 629.3409. MPT-2-151.



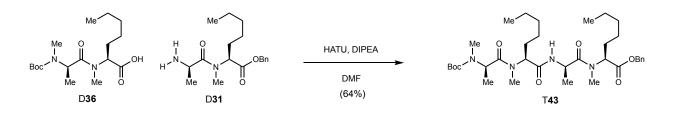
(*S*)-1-(benzyloxy)-1-oxoheptan-2-yl (*6R*,9*S*,12*R*)-2,2,5,6,11,12-hexamethyl-4,7,10-trioxo-9-pentyl-3-oxa-5,8,11-triazatridecan-13-oate (T40). A round-bottom flask was charged with the amine (122 mg, 378 µmol), acid (125 mg, 378 µmol), and DCM (7.0 mL). The mixture was cooled to 0 °C and then DIPEA (198 µL, 1.13 mmol) and PyBrop (277 mg, 567 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 2 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The rude residue was subjected to flash column chromatography (SiO₂, 5-30% ethyl acetate in hexanes) to afford the product as a light yellow oil (195 mg, 81%). [α]²⁴ +19 (c 0.83, CHCl₃); R_f = 0.36 (30% EtOAc/hexanes); IR (film) 2954, 2926, 2857, 1746, 1691, 1650, 1456, 1368, 1319, 1155, 1021 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.33-7.25 (series of m, 5H), 6.75 (br d, J = 7.3 Hz, 1H), 5.21-5.10 (series of m, 2H), 5.05 (m, 1H), 4.97 (br m, 1H), 4.86 (dt, J = 8.0, 7.9 Hz, 1H), 4.77 (m, 1H), 3.28-2.72 (series of m, 6H), 1.94 (br m, 1H), 1.81-1.72 (series of m, 3H), 1.69-1.61 (series of m, 2H), 1.44 (br s, 9H), 1.34-1.31 (br series of m, 4H) 1.27-1.17 (series of m, 12H), 0.90-0.88 (br t, J = 6.8 Hz, 3H), 0.90-0.88 (br t, J = 6.2 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) ppm 171.1, 170.9, 169.8, 169.3, 153.2, 135.2, 128.7, 128.5, 128.3, 80.6, 73.4, 73.0, 67.3, 67.0, 55.0, 52.6, 49.1, 32.8, 31.6, 31.2, 30.8, 29.7, 29.4, 28.4, 24.8, 24.6, 22.5, 22.3, 14.4, 13.9; HRMS (EI): Exact mass calcd for C₃₄H₅₉N₄O₈ [M+NH₄]⁺ 651.4327, found 651.4327. MPT-2-196



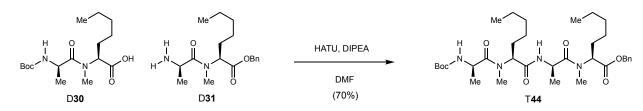
Benzyl (*S*)-2-((*6R*,9*S*,12*R*)-N,2,2,5,*6*,11,12-heptamethyl-4,7,10-trioxo-9-pentyl-3,8-dioxa-5,11-diazatridecan-13amido)heptanoate (T41). A round-bottom flask was charged with the amine (158 mg, 474 μmol), acid (157 mg, 474 μmol), and DMF (4.7 mL). The mixture was cooled to 0 °C and then DIPEA (248 μL, 1.42 mmol) and HATU (540 mg, 1.42 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-40% ethyl acetate in hexanes) to afford the product as a light yellow foam (222 mg, 72%). [*α*]_D²⁴ + 65 (*c* 0.87, CHCl₃); R_f = 0.33 (25% EtOAc/hexanes); IR (film) 2955, 2930, 2860, 1739, 1699, 1657, 1456, 1367, 1316, 1256, 1154 cm¹; ¹H NMR (600 MHz, CDCl₃) δ 7.38-7.32 (series of m, 5H), 5.47 (br m, 1H), 5.26 (br dd, *J* = 6.5, 6.5 Hz, 1H), 5.13 (m, 2H), 4.97 (dd, *J* = 8.0, 8.0 Hz, 1H), 2.92-2.60 (series of m, 9H), 2.03 (m, 1H), 1.91 (m, 1H), 1.73 (m, 1H), 1.61 (m, 1H), 1.49 (s, 9H) 1.40-1.19 (series of m, 18H), 0.90 (br t, *J* = 6.7 Hz, 3H), 0.88 (br t, *J* = 6.5 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃) ppm 172.2, 171.2, 170.8, 168.9, 156.1, 135.4, 128.9, 128.6, 128.4, 79.9, 71.3, 68.0, 67.2, 66.9, 57.9, 56.8, 54.4, 50.0, 31.3, 31.4, 30.3, 29.9, 29.1, 28.4, 27.7, 25.6, 24.9, 22.4, 21.9, 14.4, 14.0; HRMS (EI): Exact mass calcd for C₃₅H₅₈N₃O₈ [M+H]⁺ 648.4218, found 648.4216. MPT-3-006



Benzyl (6*R*,9*S*,12*R*,15*S*)-2,2,5,6,8,11,12,14-octamethyl-4,7,10,13-tetraoxo-9,15-dipentyl-3-oxa-5,8,11,14tetraazahexadecan-16-oate (T42). A round-bottom flask was charged with the amine (200 mg, 581 µmol), acid (200 mg, 581 µmol), and DMF (5.8 mL). The mixture was cooled to 0 °C and then DIPEA (304 µL, 1.74 mmol) and HATU (662 g, 2.76 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-40% ethyl acetate in hexanes) to afford the product as a light yellow oil (196 mg, 51%). $[\alpha]_D^{24} + 25$ (*c* 0.29, CHCl₃); R_f = 0.24 (30% EtOAc/hexanes); IR (film) 2956, 2931, 2860, 1742, 1691, 1650, 1456, 1390, 1321, 1156, 1078 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.39-7.27 (series of m, 5H), 5.50 (m, 1H), 5.39 (m, 1H), 5.20-5.06 (series of m, 3H), 4.65 (m, 1H), 3.20-2.66 (series of m, 12H), 1.99 (m, 1H), 1.81-1.60 (series of m, 5H), 1.49 (s, 9H), 1.37-1.21 (series of m, 16H), 0.90-0.86 (series of m, 6H); ¹³C NMR (150 MHz, CDCl₃) ppm 172.2, 171.8, 171.4, 170.9, 155.3, 135.5, 128.6, 128.2, 128.1, 80.3, 79.9, 66.8, 58.8, 56.5, 53.0, 52.1, 50.8, 49.6, 31.5, 31.2, 31.1, 30.4, 29.8, 29.3, 29.1, 28.9, 28.0, 25.6, 22.4, 22.3, 14.6, 13.9; HRMS (EI): Exact mass calcd for C₃₆H₆₁N₄O₇ [M+H]⁺ 661.4540, found 661.4537. MPT-4-300



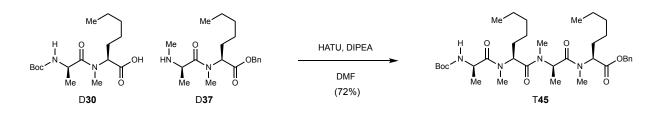
Benzyl (6*R*,9*S*,12*R*,15*S*)-2,2,5,6,8,12,14-heptamethyl-4,7,10,13-tetraoxo-9,15-dipentyl-3-oxa-5,8,11,14tetraazahexadecan-16-oate (T43). A round-bottom flask was charged with the amine (172 mg, 537 μmol), acid (185 mg, 537 μmol), and DMF (4.0 mL). The mixture was cooled to 0 °C and then DIPEA (281 μL, 1.61 mmol) and HATU (613 mg, 1.61 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h.. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-30% ethyl acetate in hexanes) to afford the product as a light yellow oil (221 mg, 64%). [α]_D²⁴ + 2.9 (*c* 0.68, CHCl₃); R_f = 0.23 (25% EtOAc/hexanes); IR (film) 3330, 2957, 2929, 2859, 1741, 1682, 1455, 1367, 1254, 1152, 1081 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.39-7.30 (series of m, 5H), 7.00 (br s, 1H), 5.44-3.74 (series of m, 6H) 3.23-2.69 (series of m, 9H), 1.98 (m, 1H), 1.74 (m, 1H), 1.56 (m, 1H), 1.46 (m, 9H), 1.36-1.22 (series of m, 19H), 0.89-0.86 (series of m, 3H); ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the 13C NMR spectrum; HRMS (El): Exact mass calcd for C₃₅H₆₂N₅O₇ [M+NH4]⁺ 664.4644, found 664.4642. MPT-3-056



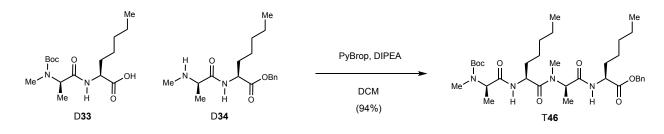
Benzyl

(6R,9S,12R,15S)-2,2,6,8,12,14-hexamethyl-4,7,10,13-tetraoxo-9,15-dipentyl-3-oxa-5,8,11,14-

tetraazahexadecan-16-oate (T44). A round-bottom flask was charged with the amine (400 mg, 1.21 mmol), acid (400 mg, 1.21 mmol), and DMF (3.0 mL). The mixture was cooled to 0 °C and then DIPEA (633 μL, 3.63 mmol) and HATU (1.38 g, 3.63 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-30% ethyl acetate in hexanes) to afford the product as a light yellow oil (536 mg, 70%). $[α]_D^{24}$ - 31 (*c* 0.57, CHCl₃); R_f = 0.13 (25% EtOAc/hexanes); IR (film) 3300, 2956, 2931, 2860, 1741, 1713, 1643, 1518, 1455, 1367, 1247, 1170, 1091 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.39-7.30 (series of m, 5H), 7.01 (series of m, 2H), 5.33-4.14 (series of m, 6H), 3.33-2.82 (series of m, 6H), 1.98 (m, 2H), 1.71 (m, 2H), 1.55 (m, 1H), 1.44 (s, 9H), 1.35-1.23 (series of m, 17H), 0.91-0.88 (series of m, 6H); ¹³C NMR (150 MHz, CDCl₃) ppm 173.2, 171.8, 171.6, 170.9, 151.7, 135.5, 128.7, 128.2, 127.8, 79.9, 79.5, 66.9, 59.6, 56.6, 52.3, 50.2, 47.0, 45.6, 45.6, 36.7, 31.4, 31.3, 31.1, 30.4, 29.7, 29.4, 28.3, 25.7, 25.4, 22.4, 14.0, 13.9; HRMS (EI): Exact mass calcd for C₃₄H₅₇N₄O₇ [M+H]⁺ 633.4227, found 633.4222. MPT-4-057



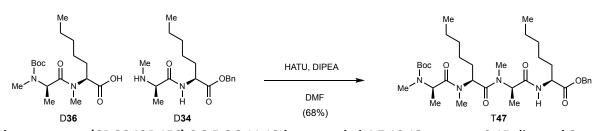
Benzyl (6*R*,9*S*,12*R*,15*S*)-2,2,6,8,11,12,14-heptamethyl-4,7,10,13-tetraoxo-9,15-dipentyl-3-oxa-5,8,11,14tetraazahexadecan-16-oate (T45). A round-bottom flask was charged with the amine (300 mg, 909 µmol), acid (300 mg, 909 µmol), and DMF (1.0 mL). The mixture was cooled to 0 °C and then DIPEA (475 µL, 2.72 mmol) and HATU (1.03 g, 2.72 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h.. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-30% ethyl acetate in hexanes) to afford the product as a light yellow foam (423 mg, 72%). [*α*]_D²⁴ + 7.8 (*c* 0.43, CHCl₃); *R*_{*f*} = 0.23 (25% EtOAc/hexanes); IR (film) 2956, 2930, 2860, 1739, 1712, 1643, 1456, 1405, 1247, 1079 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.36-7.30 (series of m, 5H), 5.50 (m, 2H), 5.22-5.06 (series of m, 3H), 4.61 (m, 1H), 3.00-2.75 (series of m, 9H), 1.98 (br m, 1H), 1.79-1.64 (br series of m, 4H), 1.42 (br s, 9H), 1.32-1.21 (series of m, 15H), 0.88 (br m, 6H); ¹³C NMR (150 MHz, (CDCl₃) ppm 173.1, 172.0, 171.1, 170.3, 155.1, 135.6, 128.0, 128.4, 128.2, 79.5, 67.0, 57.0, 56.7, 56.5, 52.9, 52.1, 49.9, 49.6, 31.5, 31.2, 30.2, 30.0, 29.3, 28.9, 28.0, 25.8, 22.5, 22.4, 18.8, 14.5, 13.9; HRMS (El): Exact mass calcd for C₃₅H₅₉N₄O₇ [M+H]⁺ 647.4384, found 646.4388. MPT-4-159



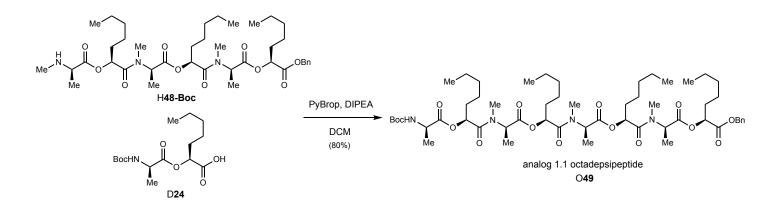
Benzyl

(6R,9S,12R,15S)-2,2,5,6,11,12-hexamethyl-4,7,10,13-tetraoxo-9,15-dipentyl-3-oxa-5,8,11,14-

tetraazahexadecan-16-oate (T46). A round-bottom flask was charged with the amine (254 mg, 793 μmol), acid (262 mg, 793 μmol), and DCM (7.0 mL). The mixture was cooled to 0 °C and then DIPEA (414 μL, 2.38 mmol) and PyBrop (581 mg, 1.19 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 2 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-30% ethyl acetate in hexanes) to afford the product as a light yellow oil (611 mg, 94%). $[\alpha]_D^{24}$ + 36 (*c* 0.86, CHCl₃); R_f = 0.26 (30% EtOAc/hexanes); IR (film) 3318, 2956, 2931, 2860, 1742, 1661, 1530, 1456, 1367, 1256, 1154, 1090 cm⁻¹; ¹H NMR (600 MHz, CDCl₃ δ 7.41-7.28 (series of m, 7H), 7.00 (br s, 1H), 6.70 (br s, 1H), 5.26 (br m, 1H), 5.13 (d, *J* = 12.1 Hz, 1H), 5.00 (d, *J* = 12.1 Hz, 1H), 4.71 (br m, 1H), 4.46 (m, 2H), 2.86-2.56 (series of m, 6H), 1.79 (br m, 1H), 1.64 (br m, 1H), 1.54 (br m, 1H), 1.43 (s, 9H), 1.37 (m, 3H), 1.27-1.17 (series of m, 16H), 0.83 (br t, *J* = 6.1 Hz, 3H), 0.78 (br t, *J* = 6.3 Hz, 3H); ¹³C NMR (150 MHz, CDCl₃ ppm 172.5, 172.4, 172.0, 170.6, 161.0, 134.9, 128.7, 128.6, 128.5, 79.9, 68.9, 52.5, 52.0, 50.0, 42.2, 32.0, 31.9, 31.6, 31.5, 31.2, 30.8, 30.7, 29.9, 29.7, 29.4, 25.4, 25.0, 22.4, 14.0, 13.4; HRMS (EI): Exact mass calcd for C₃₄H₆₀N₅O₇ [M+NH₄]⁺ 650.4493, found 650.4487. MPT-2-193



Benzyl (6*R*,9*S*,12*R*,15*S*)-2,2,5,6,8,11,12-heptamethyl-4,7,10,13-tetraoxo-9,15-dipentyl-3-oxa-5,8,11,14tetraazahexadecan-16-oate (T47). A round-bottom flask was charged with the amine (171 mg, 535 µmol), acid (184 mg, 535 µmol), and DMF (5.4 mL). The mixture was cooled to 0 °C and then DIPEA (283.3 µL, 1.61 mmol) and HATU (610 mg, 1.61 mol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-35% ethyl acetate in hexanes) to afford the product as a yellow foam (235 mg, 68%). [*α*]_D²⁴ + 24 (*c* 0.34, CHCl₃); R_f = 0.31 (25% EtOAc/hexanes); IR (film) 3448, 2956, 2929, 2810, 1745, 1701, 1686, 1473, 1389, 1259, 1182, 1087 cm⁻¹; ¹H NMR (600 MHz, (CD₃)₂CO) δ 7.45-7.27 (series of m, 5H), 7.04 (br m, 1H), 5.33-4.53 (series of m, 6H), 3.07-2.69 (series of m, 9H), 2.01-1.78 (series of m, 3H), 1.62 (m, 1H), 1.45 (m, 9H), 1.37-1.16 (series of m, 18H), 0.93-0.83 (series of m, 6H); ¹³C NMR (150 MHz, CDCl₃) ppm 173.2, 171.9, 171.2, 170.5, 155.3, 135.6, 128.7, 128.4, 128.1, 79.5, 67.0, 66.8, 56.9, 56.3, 53.1, 52.2, 49.6, 46.9, 31.5, 31.2, 30.2, 29.9, 28.9, 28.3, 27.9, 25.7, 25.4, 22.4, 18.9, 14.4, 13.9; HRMS (EI): Exact mass calcd for C₃₅H₅₈N₄NaO₇ [M+Na]⁺ 647.4384, found 647.4388. MPT-4-156



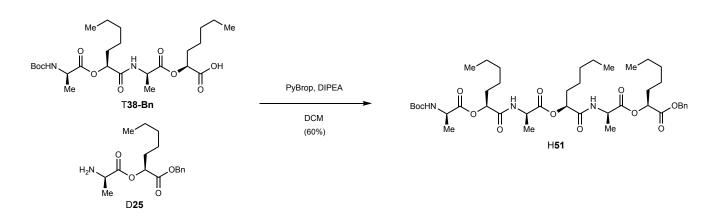
Benzyl (*S*)-2-(((*6R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*)-2,2,6,11,12,17,18,23,24-nonamethyl-4,7,10,13,16,19,22-heptaoxo-9,15,21-tripentyl-3,8,14,20-tetraoxa-5,11,17,23-tetraazapentacosan-25-oyl)oxy)heptanoate (O49). A round-bottom flask was charged with the amine (141 mg, 189 µmol), acid (66.0 mg, 189 µmol), and DCM (1.8 mL). The mixture was cooled to 0 °C and then DIPEA (98.8 µL, 567 µmol) and PyBrop (92.3 mg, 189 µmol) were added. The reaction was stirred at 0 °C for 30 min, then allowed to warm to ambient temperature and stir for an additional 1.5 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 10-60% ethyl acetate in hexanes) to afford the product as a light yellow oil (158 mg, 80%). $[\alpha]_D^{24} + 34$ (*c* 1.36, CHCl₃); R_f = 0.40 (60% EtOAc/hexanes); IR (film) 3302, 2930, 1743, 1663, 1456, 1186 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This

5 Ме Т**38-Вос**

HRMS (EI): Exact mass calcd for C₅₅H₉₄N₅O₁₅ [M+NH₄]^{*} 1064.6746, found [1064.6742]. MPT-2-166 $Me \xrightarrow{Me}_{Me} \xrightarrow{Me}_{Me}$

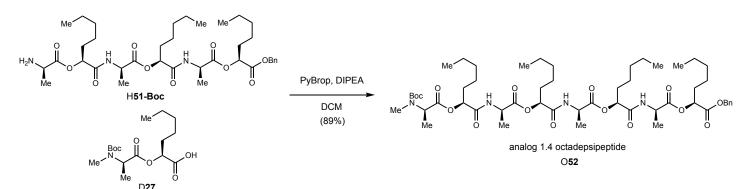
compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for $C_{55}H_{94}N_5O_{15}$ [M+NH₄]⁺ 1064.6746, found [1064.6742]. MPT-2-166

Benzyl (*S*)-2-(((*6R*,95,12*R*,15*S*,18*R*,21*S*,24*R*)-2,2,5,6,11,12,18,24-octamethyl-4,7,10,13,16,19,22-heptaoxo-9,15,21-tripentyl-3,8,14,20-tetraoxa-5,11,17,23-tetraazapentacosan-25-oyl)oxy)heptanoate (O50). A round-bottom flask was charged with the amine (93.0 mg, 184 µmol), acid (100 mg, 184 µmol), and DCM (1.8 mL). The mixture was cooled to 0 °C and then DIPEA (95.9 µL, 551 µmol) and PyBrop (108 mg, 220 µmol) were added. The reaction was stirred at 0 °C for 30 min, then allowed to warm to ambient temperature and stir for an additional 1.5 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 10-50% ethyl acetate in hexanes) to afford the product as a light-yellow oil (130 mg, 69%). [*α*]_D²⁴ –6.5 (*c* 0.53, CHCl₃); R_f = 0.21 (30% EtOAc/hexanes); IR (film) 3303, 2956, 2929, 2871, 2860, 1747, 1671, 1538, 1455, 1389, 1210, 1086 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.65 (br s, 1H), 7.40 (br d, *J* = 7.3 Hz, 1H), 5.21-5.07 (series of m, 7H), 4.91 and 4.07 (br q, *J* = 6.9 Hz, 1H), 4.67-4.63 (m, 1H), 4.25-4.20 (m, 1H), 3.19 (br m, 3H), 2.81 (br m, 3H), 1.85-1.68 (series of m, 8H), 1.48-1.43 (series of m, 18H), 1.38 (br s, 9H), 1.32-1.26 (series of m, 18H), 0.89-0.85 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) This compound is a mixture of rotamers causing significant peak broadening and overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₅₄H₈₈N₄NaO₁₅ [M+Na]⁺ 1055.6144, found 1055.6144. MPT-2-154

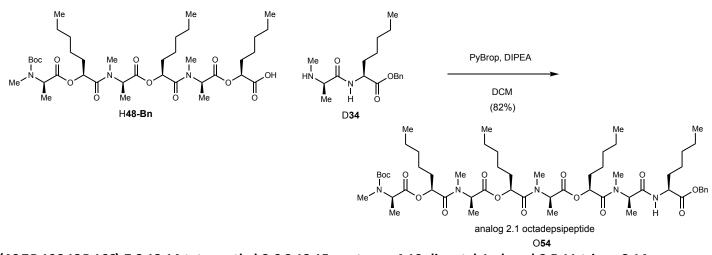


SI-22

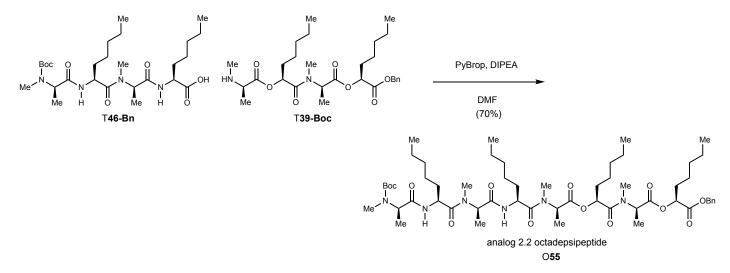
(S)-2-(((6R,9S,12R,15S,18R)-2,2,6,12,18-pentamethyl-4,7,10,13,16-pentaoxo-9,15-dipentyl-3,8,14-trioxa-Benzyl 5,11,17-triazanonadecan-19-oyl)oxy)heptanoate (H51). A round-bottom flask was charged with the amine (75.0 mg, 244 μmol), acid (126 mg, 244 μmol), and DCM (2.4 mL). The mixture was cooled to 0 °C and then DIPEA (128 μL, 732 μmol) and PyBrop (119 mg, 244 µmol) were added. The reaction was stirred at 0 °C for 30 min, then allowed to warm to ambient temperature and stir for an additional 1.5 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 10-60% ethyl acetate in hexanes) to afford the product as a light yellow oil (117 mg, 60%). $[\alpha]_D^{24} - 29$ (c 0.32, CHCl₃); R_f = 0.49 (30% EtOAc/hexanes); IR (film) 3346, 2956, 2927, 2858, 1751, 1662, 1534, 1457, 1157, 1069 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.37-7.31 (series of m, 5H), 5.23-5.14 (series of m, 3H), 5.09 (br t, J = 6.3 Hz, 1H), 5.00 (br d, J = 5.4 Hz, 1H), 4.66 (dd, J = 7.4, 7.4 Hz, 1H), 4.31 (dd, J = 6.8, 5.8 Hz, 1H), 4.14 (dd, J = 6.8, 6.8 Hz, 1H), 1.99-1.95 (series of m, 1H), 1.88-1.78 (series of m, 5H), 1.50 (br d, J = 7.6 Hz, 3H), 1.45 (br d, J = 7.4 Hz, 3H), 1.42 (s, 8H), 1.41 (br d, J = 7.6 Hz, 4H), 1.36-1.25 (series of m, 18H), 0.87 (t, J = 6.7 Hz, 3H), 0.86 (t, J = 7.0 Hz, 3H), 0.85 (t, J = 6.6 Hz, 3H) [three N-H peaks not observed]; ¹³C NMR (150 MHz, CDCl₃) ppm 173.5, 172.8, 172.6, 171.0, 170.3, 170.0, 156.4, 135.9, 129.1, 128.9, 128.8, 81.1, 74.9, 74.4, 73.4, 67.5, 50.5, 49.8, 48.6, 48.3, 32.2, 31.9, 31.8, 31.6, 30.5, 30.3, 28.8, 25.3, 25.2, 24.8, 23.0, 18.0, 17.5, 16.9, 14.7, 14.6, 14.52, 14.47 HRMS (EI): Exact mass calcd for C₄₂H₆₇N₃O₁₂Na [M+Na]⁺ 828.4623, found 828.4622. MPT-2-157



Benzyl (*S*)-2-(((*6R*,9*s*),1*2R*,15*s*),18*R*,21*s*,24*R*)-2,2,5,6,12,18,24-heptamethyl-4,7,10,13,16,19,22-heptaoxo-9,15,21-tripentyl-3,8,14,20-tetraoxa-5,11,17,23-tetraazapentacosan-25-oyl)oxy)heptanoate (O52). A round-bottom flask was charged with the amine (102 mg, 184 µmol), acid (47 mg, 184 µmol), and DCM (1.8 mL). The mixture was cooled to 0 °C and then DIPEA (95.0 µL, 540 µmol) and PyBrop (88.0 mg, 184 µmol) were added. The reaction was stirred at 0 °C for 30 min, then allowed to warm to ambient temperature and stir for an additional 1.5 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 10-60% ethyl acetate in hexanes) to afford the product as a light yellow oil (132 mg, 89%). [*α*]_D²⁴ -21 (*c* 0.93, CHCl₃); R_f = 0.28 (30% EtOAc/hexanes); IR (film) 3300, 2956, 2930, 2860, 1749, 1665, 1455, 1382, 1156, 1086 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.95 (br d, *J* = 4.0 Hz, 1H), 7.65 (br d, *J* = 4.0 Hz, 1H), 7.37-7.31 (series of m, 6H), 5.21-3.70 (series of m, 10H), 2.98 (br s, 3H), 1.88-1.77 (series of m, 8H), 1.52 (m, 1H), 1.51-1.43 (series of m, 19H), 1.37-1.32 (series of m, 7H), 1.30-1.24 (series of m, 18H), 0.89-0.85 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) This compound is a mixture of rotamers causing significant peak broadening and overlap. Refer to the image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₅₃H₈₆N₄NaO₁₅ [M+Na]⁺ 1041.5988, found 1041.5987. MPT-2-160

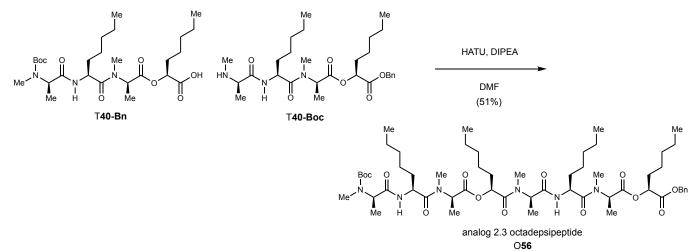


(4*S*,*7R*,10*S*,13*R*,16*S*)-7,8,13,14-tetramethyl-3,6,9,12,15-pentaoxo-4,10-dipentyl-1-phenyl-2,5,11-trioxa-8,14diazahenicosan-16-yl (*6R*,9*S*,12*R*)-2,2,5,6,11,12-hexamethyl-4,7,10-trioxo-9-pentyl-3-oxa-5,8,11-triazatridecan-13-oate (O54). A round-bottom flask was charged with the amine (88.0 mg, 120 µmol), acid (39.0 mg, 120 µmol), and DCM (1.2 mL). The mixture was cooled to 0 °C and then DIPEA (61.0 µL, 350 µmol) and PyBrop (86.0 mg, 180 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 2 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-40% ethyl acetate in hexanes) to afford the product as a light yellow oil (104 mg, 82%). $[\alpha]_D^{24} + 9.9$ (*c* 0.75, CHCl₃); R_f = 0.72 (30% EtOAc/hexanes); IR (film) 2925, 2856, 1746, 1668, 1456, 1377, 1215, 1103 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.31-7.24 (series of m, 5H), 6.89 (br s, 1H), 5.19-4.25 (series of m, 10H), 3.09-2.60 (series of m, 12H), 2.23 (m, 1H), 2.00 (m, 1H), 1.84-1.75 (br m, 3H), 1.61-1.56 (series of m, 3H), 1.43-1.40 (series of m, 9H), 1.34-1.30 (series of m, 3H), 1.27-1.24 (series of m, 22H), 1.23-1.20 (series of m, 11H), 0.90-0.88 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₅₆H₉₇N₆O₁₄ [M+NH₄]⁺ 1077.7063, found 1077.7058. MPT-2-189



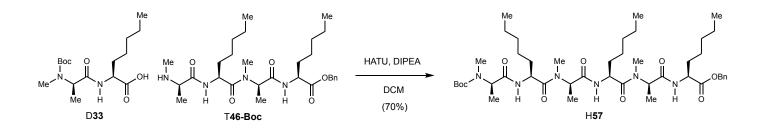
(S)-1-(((R)-1-(((S)-1-(benzyloxy)-1-oxoheptan-2-yl)oxy)-1-oxopropan-2-yl)(methyl)amino)-1-oxoheptan-2-yl

(6*R*,9*S*,12*R*,15*S*,18*R*)-2,2,5,6,11,12,17,18-octamethyl-4,7,10,13,16-pentaoxo-9,15-dipentyl-3-oxa-5,8,11,14,17pentaazanonadecan-19-oate (O55). A round-bottom flask was charged with the amine (110 mg, 206 µmol), acid (110 mg, 206 µmol), and DMF (2.0 mL). The mixture was cooled to 0 °C and then DIPEA (108 µL, 618 mmol) and PyBrop (638 mg, 1.31 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 2 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a light yellow oil (150 mg, 70%). $[\alpha]_D^{24}$ + 63 (*c* 0.64, CHCl₃); R_f = 0.25 (30% EtOAc/hexanes); IR (film) 3387, 2956, 2932, 2861, 1742, 1651, 1522, 1457, 1391, 1214, 1154, 1093 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.30-7.24 (series of m, 5H), 7.05 (br s, 1H), 5.18-4.98 (series of m, 6H), 4.73-4.36 (series of m, 4H), 2.99-2.71 (series of m, 12H), 1.82-1.73 (series of m, 3H), 1.66-1.56 (series of m, 6H), 1.47-1.41 (series of m, 9H), 1.40-1.34 (series of m, 14H), 1.28-1.18 (series of m, 20H), 0.90-0.88 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₅₆H₉₈N₇O₁₃ [M+NH₄]⁺ 1076.7223 found 1076.7217. MPT-2-279

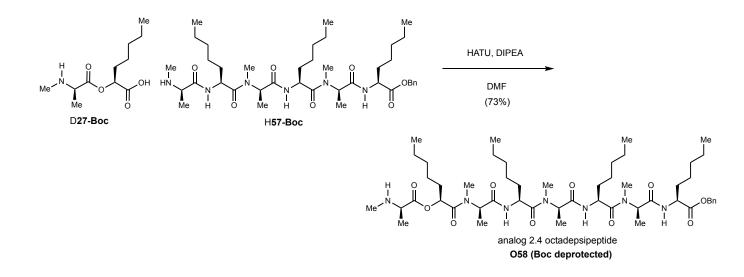


(4*S*,*7R*,10*S*,13*R*,16*S*)-7,8,13,14-tetramethyl-3,6,9,12,15-pentaoxo-4,10-dipentyl-1-phenyl-2,5-dioxa-8,11,14triazahenicosan-16-yl (6*R*,9*S*,12*R*)-2,2,5,6,11,12-hexamethyl-4,7,10-trioxo-9-pentyl-3-oxa-5,8,11-triazatridecan-13oate (O56). A round-bottom flask was charged with the amine (96.2 mg, 180 µmol), acid (98.0 mg, 180 µmol), and DCM (1.8 mL). The mixture was cooled to 0 °C and then DIPEA (95 µL, 541 µmol) and HATU (206 mg, 541 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a light yellow oil (97 mg, 51%). $[\alpha]_D^{24} + 51 (c 0.70, CHCl_3)$; $R_f = 0.16 (30\% EtOAc in hexanes); IR (film)$ $2929, 1747, 1652, 1514, 1473, 1212 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) <math>\delta$ 7.38-7.32 (series of m, 5H), 6.83 (br m, 2H), 5.37 (m, 1H), 5.22-5.15 (m, 2H), 5.13 (m, 3H), 5.02 (m, 1H), 4.91 (m, 1H), 4.86-4.74 (m, 2H), 3.32 (series of m, 12H), 1.86-1.61 (br m, 12H), 1.49 (br s, 9H), 1.38-1.23 (series of m, 32H), 0.90-0.88 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) This Johnston et al.

compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for $C_{56}H_{98}N_7O_{13}$ [M+NH₄]⁺ 1076.7223, found 1076.7217. MPT-2-247

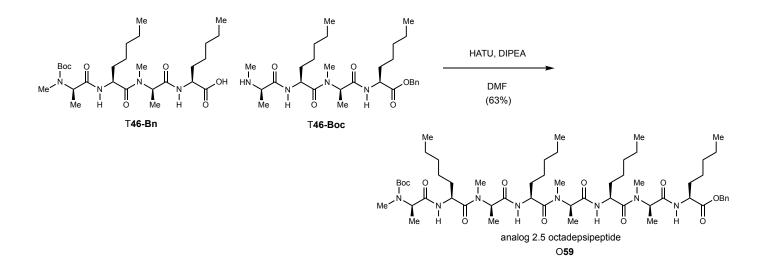


Benzyl (6R,9S,12R,15S,18R,21S)-2,2,5,6,11,12,17,18-octamethyl-4,7,10,13,16,19-hexaoxo-9,15,21-tripentyl-3-oxa-5,8,11,14,17,20-hexaazadocosan-22-oate (H57). A round-bottom flask was charged with the amine (112 mg, 338 μmol), acid (180 mg, 338 μmol), and DCM (2.1 mL). The mixture was cooled to 0 °C and then DIPEA (177 μL, 210 μmol) and HATU (385 mg, 1.01 mmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-60% ethyl acetate in hexanes) to afford the product as a light yellow oil (200 mg, 70%). $\left[\alpha\right]_{D}^{24}$ + 6.3 (c 0.48, CHCl₃); R_f = 0.23 (30% EtOAc/hexanes); IR (film) 3313, 2957, 2931, 2861, 1748, 1660, 1539, 1456, 1368, 1259, 1081 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.37-7.34 (series of m, 5H), 7.10 (br s, 1H), 6.91 (br s, 1H), 6.50 (br s, 1H), 5.20-5.14 (series of m, 3H), 5.10-5.17 (series of m, 2H), 4.66 (series of m, 2H), 4.35 (m, 1H), 3.28-2.69 (series of m, 9H), 1.96 (m, 1H), 1.80-1.73 (series of m, 2H), 1.66 (m, 1H), 1.42 (s, 9H), 1.40-1.38 (series of m, 4H), 1.37-1.35 (series of m, 4H), 1.29-1.26 (series of m, 2H), 1.24-1.21 (series of m, 9H), 1.21-1.18 (series of m, 10H), 0.89-0.85 (series of m, 9H); ¹³C NMR (150 MHz, CDCl₃) ppm 172.2, 172.1, 172.0, 171.6, 169.7, 169.3, 157.1, 135.4, 128.6, 128.4, 128.2, 80.9, 75.1, 74.6, 73.1, 72.8, 72.1, 67.2, 49.1, 48.9, 48.1, 47.9, 47.8, 31.7, 31.5, 31.3, 31.1, 31.0, 30.9, 30.8, 29.7, 29.6, 28.4, 28.3, 24.7, 24.6, 22.4, 22.3, 17.6, 17.4, 17.1; HRMS (EI): Exact mass calcd for C₄₅H₈₀N₇O₉ [M+NH₄]⁺ 862.6018, found 862.6019. MPT-2-287

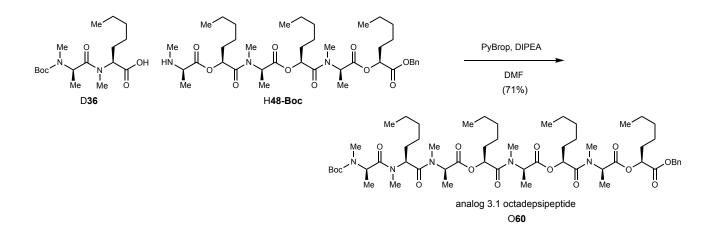


Benzyl (25,5*R*,85,11*R*,145,17*R*,205)-5,6,11,12,17,18-hexamethyl-20-((methyl-*D*-alanyl)oxy)-4,7,10,13,16,19-hexaoxo-2,8,14-tripentyl-3,6,9,12,15,18-hexaazapentacosanoate (O58, Boc-deprotected). A round-bottom flask was charged with the amine (162 mg, 217 µmol), acid (72.0 mg, 217 µmol), and DMF (2.2 mL). The mixture was cooled to 0 °C and then DIPEA (114 µL, 652 µmol) and HATU (248 mg, 652 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-60% ethyl acetate in hexanes) to afford the product⁶ as a light yellow oil (168 mg, 73%). [α]_D²⁴ + 22 (c 0.48, CHCl₃); R_f = 0.14 (50% EtOAc/hexanes); IR (film) 3333, 2956, 2930, 2860, 1747, 1667, 1532, 1456, 1381, 1187, 1156, 1082 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists as a mixture of rotamers causing significant peak broadening and overlap. Refer to the image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound is a mixture of rotamers causing significant peak broadening and overlap. Refer to the image of the ¹³C NMR spectrum; HRMS (El): Exact mass calcd for C₅₁H₉₁N₈O₁₀ [M+NH₄]⁺ 975.6858, found 975.6856. MPT-2-273

⁶ The product isolated, unexpectedly, was the Boc-deprotected O**58**. Therefore, it was characterized as such.



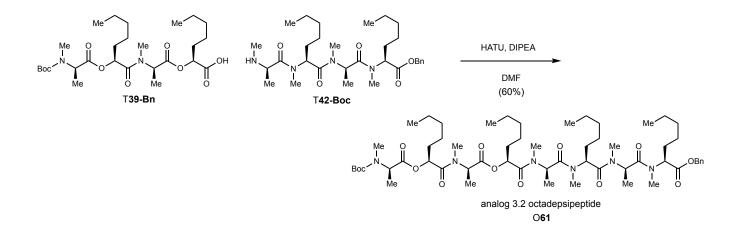
Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,5,6,11,12,17,18,23,24-decamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O59). A round-bottom flask was charged with the amine (93.3 mg, 175 µmol), acid (95.0 mg, 175 µmol), and DMF (1.8 mL). The mixture was cooled to 0 °C and then DIPEA (91.5 µL, 525 µmol) and HATU (200 mg, 525 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-40% ethyl acetate in hexanes) to afford the product as a light yellow oil (117 mg, 63%). [α]_D²⁴ + 37 (*c* 0.45, CHCl₃); R_f= 0.47 (30% EtOAc/hexanes); IR (film) 3405, 2956, 2929, 2859, 1741, 1652, 1540, 1457, 1391, 1260, 1155, 1092 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists as a mixture of rotamers causing significant peak broadening and overlap. Refer to the image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound is a mixture of rotamers causing significant peak broadening and overlap. Refer to the image of the ¹³C NMR spectrum; HRMS (El): Exact mass calcd for C₅₆H₁₀₀N₉O₁₁ [M+NH₄]⁺ 1074.7542, found 1074.7544. MPT-2-276



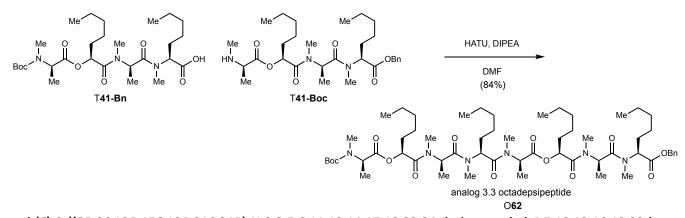
Benzyl (4*S*,7*R*,10*S*,13*R*,16*S*)-7,8,13,14-tetramethyl-3,6,9,12,15-pentaoxo-4,10-dipentyl-1-phenyl-2,5,11-trioxa-8,14-diazahenicosan-16-yl (6*R*,9*S*,12*R*)-2,2,5,6,8,11,12-heptamethyl-4,7,10-trioxo-9-pentyl-3-oxa-5,8,11-triazatridecan-13-

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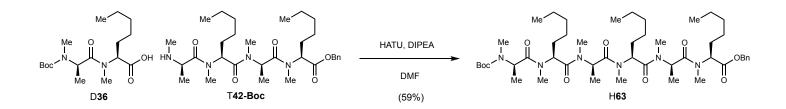
oate (O60). A round-bottom flask was charged with the amine (86.0 mg, 250 µmol), acid (186 mg, 250 µmol), and DMF (2.5 mL). The mixture was cooled to 0 °C and then DIPEA (130 µL, 750 µmol) and PyBrop (183 mg, 375 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a light yellow oil (189 mg, 71%). $[\alpha]_D^{24} + 22$ (*c* 0.46, CHCl₃); R_f = 0.27 (25% EtOAc/hexanes); IR (film) 2956, 2931, 2860, 1745, 1698, 1455, 1390, 1258, 1150, 1092 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists as a mixture of rotamers causing significant peak broadening and overlap. Refer to the image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₅₇H₉₉N₆O₁₄ [M+NH₄]⁺ 1091.7214, found 1091.7212. MPT-3-072



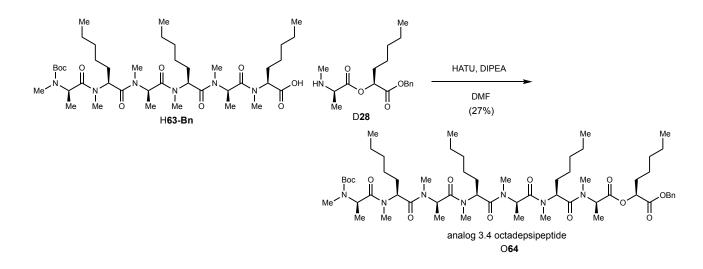
(25,5R,85,11R,145)-14-(((6R,95,12R)-2,2,5,6,11,12-hexamethyl-4,7,10-trioxo-9-pentyl-3,8-dioxa-5,11-Benzyl diazatridecan-13-oyl)oxy)-3,5,6,9,11,12-hexamethyl-4,7,10,13-tetraoxo-2,8-dipentyl-3,6,9,12-tetraazanonadecanoate (061). A round-bottom flask was charged with the amine (125 mg, 230 µmol), acid (125 mg, 230 µmol), and DMF (2.3 mL). The mixture was cooled to 0 °C and then DIPEA (120 μL, 690 μmol) and HATU (262 mg, 690 μmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a yellow oil (149 mg, 60%). $[\alpha]_D^{24}$ - 22 (c 0.25, CHCl₃); R_f = 0.37 (30% EtOAc/hexanes); IR (film) 2955, 2933, 2858, 1745, 1699, 1450, 1258, 1156, 1092 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.37-7.31 (series of m, 5H), 5.24-5.20 (series of m, 5H), 5.15 (m, 2H), 5.04 (m, 1H), 4.97 (m, 1H), 4.70 (m, 1H), 3.11-2.81 (series of m, 18H), 1.88-1.80 (series of m, 6H), 1.76-1.68 (series of m, 4H), 1.57-1.52 (m, 3H), 1.45 (s, 9H), 1.41-1.38 (series of m, 6H), 1.38-1.35 (series of m, 6H), 1.34-1.27 (series of m, 19H), 0.90-0.88 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) ppm 171.3, 171.1, 170.7, 170.5, 170.1, 169.9, 169.5, 169.4, 156.0, 135.3, 128.6, 128.5, 128.3, 79.7, 73.4, 73.0, 72.1, 71.8, 71.7, 71.6, 71.4, 67.2, 67.0, 55.0, 54.9, 54.5, 53.1, 52.4, 32.1, 31.6, 31.5, 31.4, 31.3, 31.2, 31.1, 30.9, 30.7, 30.2, 29.6, 28.4, 25.1, 24.9, 24.8, 24.5, 22.5, 22.4, 22.3, 15.6, 15.5, 14.4, 14.3, 14.0, 13.98, 13.94; HRMS (EI): Exact mass calcd for C₅₈H₁₀₂N₇O₁₃ [M+NH₄]⁺ 1104.7536, found 1104.7536. MPT-3-109



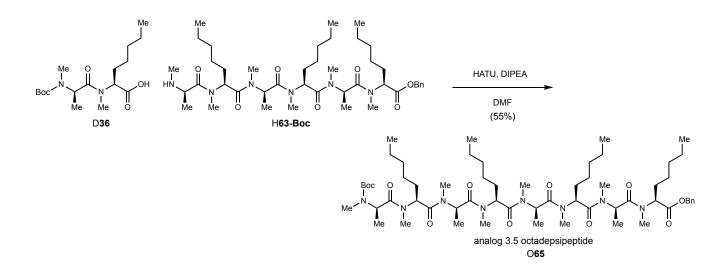
Benzyl (*S*)-2-((*6R*,9*5*,12*R*,15*5*,18*R*,21*5*,24*R*)-N,2,2,5,6,11,12,14,17,18,23,24-dodecamethyl-4,7,10,13,16,19,22-heptaoxo-9,15,21-tripentyl-3,8,20-trioxa-5,11,14,17,23-pentaazapentacosan-25-amido)heptanoate (O62). A round-bottom flask was charged with the amine (102 mg, 186 µmol), acid (104 mg, 186 µmol), and DMF (1.9 mL). The mixture was cooled to 0 °C and then DIPEA (100 µL, 559 µmol) and HATU (213 mg, 559 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-40% ethyl acetate in hexanes) to afford the product as a light yellow oil (170 mg, 84%). $[\alpha]_D^{24}$ - 13 (*c* 0.13, CHCl₃); R_f = 0.17 (25% EtOAc/hexanes); IR (film) 2927, 2858 2665, 1847, 1743, 1456, 1392, 1321, 1258, 1152, 1092 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) ppm 171.8, 171.6, 171.3, 170.7, 170.6, 167.9, 166.2, 165.6, 151.7, 135.6, 128.4, 128.2, 128.1, 80.9, 79.2, 77.5, 74.9, 71.3, 67.3, 67.2, 66.9, 66.7, 66.6, 60.7, 60.4, 60.1, 56.7, 56.5, 55.9, 54.7, 51.5, 34.7, 33.6, 32.5, 32.1, 31.8, 31.6, 31.4, 31.1, 30.4, 29.7, 29.1, 28.4, 28.2, 27.9, 25.9, 25.7, 25.2, 24.9, 24.7, 24.1, 22.4, 21.9, 21.7, 16.6, 14.6, 14.1, 13.9; HRMS (EI): Exact mass calcd for C₅₈H₁₀₂N₇O₁₃ [M+NH4]⁺ 1104.7536, found 1104.7532. MPT-3-020



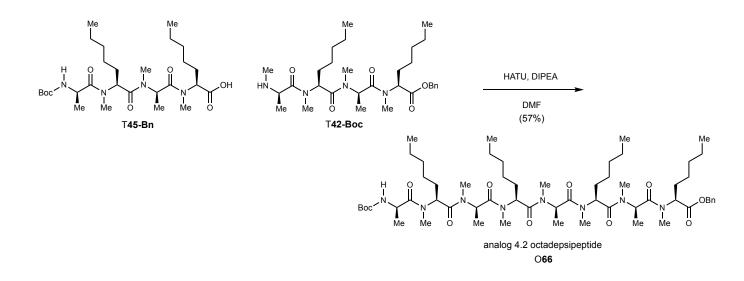
Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*)-2,2,5,6,8,11,12,14,17,18,20-undecamethyl-4,7,10,13,16,19-hexaoxo-9,15,21-tripentyl-3-oxa-5,8,11,14,17,20-hexaazadocosan-22-oate (H63). A round-bottom flask was charged with the amine (180 mg, 343 µmol), acid (118 mg, 343 µmol), and DMF (3.4 mL). The mixture was cooled to 0 °C and then DIPEA (179 µL, 1.03 mmol) and HATU (392 mg, 1.03 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h.. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a yellow oil (179 mg, 59%). [α]_D²⁴ + 14 (*c* 0.33, CHCl₃); R_f = 0.18 (30% EtOAc/hexanes); IR (film) 2954, 2931, 2860, 1744, 1699, 1649, 1455, 1390, 1322, 1259, 1156 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.39-7.30 (series of m, 5H), 5.26-5.20 (series of m, 4H), 5.12 (m, 1H), 5.05 (m, 1H), 4.98 (m, 1H), 4.69 (m, 1H), 3.15-2.81 (series of m, 18H), 1.86 (m, 2H), 1.73 (m, 2H), 1.59 (m, 2H), 1.47 (s, 9H), 1.40-1.35 (series of m, 8H), 1.34-1.25 (series of m, 19H), 0.90-0.88 (series of m, 9H); ¹³C NMR (150 MHz, CDCl₃) ppm 171.1, 170.6, 170.1, 169.9, 169.5, 169.4, 156.0, 135.3, 128.6, 128.5, 128.3, 79.7, 73.4, 73.0, 72.1, 71.6, 67.2, 67.0, 54.9, 54.4, 53.0, 52.4, 32.0, 31.6, 31.4, 31.3, 31.2, 31.1, 30.9, 30.7, 30.2, 29.7, 29.6, 28.4, 25.1, 24.8, 24.5, 22.4, 15.6, 15.3, 14.6, 14.3, 14.0, 13.9; HRMS (EI): Exact mass calcd for C₄₈H₈₆N₇O₉ [M+NH₄]⁺ 904.6487, found 904.6484. MPT-4-204



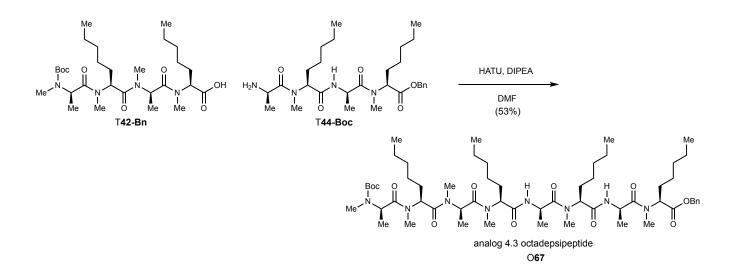
(*S*)-1-(benzyloxy)-1-oxoheptan-2-yl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*)-2,2,5,6,8,11,12,14,17,18,20,23,24-tridecamethyl-4,7,10,13,16,19,22-heptaoxo-9,15,21-tripentyl-3-oxa-5,8,11,14,17,20,23-heptaazapentacosan-25-oate (O64). A round-bottom flask was charged with the amine (32.0 mg, 100 µmol), acid (80.0 mg, 100 µmol), and DMF (1.0 mL). The mixture was cooled to 0 °C and then DIPEA (60.0 µL, 300 µmol) and HATU (114 mg, 300 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-40% ethyl acetate in hexanes) to afford the product as a light yellow oil (30 mg, 27%). [α]_D²⁴ + 2.3 (c 0.53, CHCl₃); R_f = 0.13 (30% EtOAc/hexanes); IR (film) 2956, 2928, 2858, 1742, 1651, 1456, 1393, 1218, 1155, 1083 cm⁻¹; ⁻¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) ppm 172.6, 171.8, 171.1, 170.6, 170.0, 169.8, 169.5, 169.3, 156.0, 135.2, 128.6, 128.4, 128.3, 80.0, 79.8, 73.4, 73.0, 72.9, 71.5, 71.3, 71.2, 71.0, 70.9, 67.2, 67.1, 67.0, 54.9, 54.4, 54.3, 53.4, 53.3, 53.1, 52.8, 52.6, 52.5, 51.4, 31.6, 31.5, 31.4, 31.2, 30.9, 30.2, 29.7, 29.5, 28.9, 28.7, 28.3, 25.3, 25.0, 24.7, 22.6, 15.5, 14.7, 14.3, 14.0; HRMS (EI): Exact mass calcd for C₅₉H₁₀₅N₈O₁₂ [M+NH₄]* 1117.7852, found 1117.7848. MPT-5-168



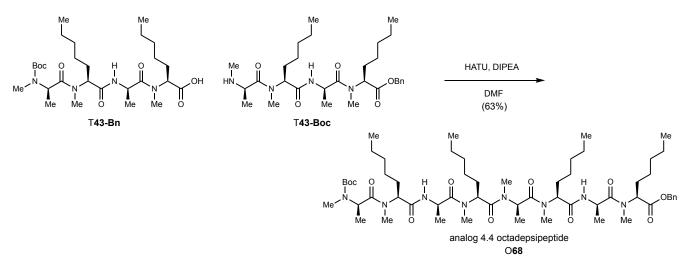
Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,5,6,8,11,12,14,17,18,20,23,24,26-tetradecamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaozaoctacosan-28-oate (O65). A round-bottom flask was charged with the amine (95.2 mg, 121 µmol), acid (41.7 mg, 121 µmol), and DMF (1.2 mL). The mixture was cooled to 0 °C and then DIPEA (63.3 µL, 363 µmol) and HATU (238 mg, 363 µmol) were added. The reaction was stirred at 0 °C for 1 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-60% ethyl acetate in hexanes) to afford the product as a yellow oil (74.1 mg, 55%). [α]_D²⁴ + 9.0 (*c* 0.33, CHCl₃); R_f = 0.23 (40% EtOAc/hexanes); IR (film) 2955, 2930, 2860, 1743, 1695, 1667, 1456, 1389, 1315, 1185 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.37-7.31 (series of m, 5H), 5.23-5.17 (m, 4H), 5.12 (d, *J* = 4.8 Hz, 1H), 5.09 (d, *J* = 4.8 Hz, 1H), 5.06 (series of m, 2H), 4.98 (m, 1H), 4.69 (m, 1H), 3.20-2.81 (series of m, 24H), 1.87-1.80 (series of m, 6H), 1.74-1.69 (m, 2H), 1.54 (m, 3H), 1.47 (s, 9H), 1.42-1.39 (series of m, 4H), 1.38-1.35 (series of m, 10H), 1.31-1.24 (series of m, 19H), 0.90-0.84 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) pm 172.5, 171.7, 171.1, 170.5, 170.1, 169.9, 169.5, 169.3, 156.0, 135.3, 128.6, 128.5, 128.4, 80.0, 79.7, 73.4, 73.0, 72.0, 71.6, 71.3, 71.1, 67.2, 67.0, 55.0, 54.9, 54.4, 53.1, 52.5, 31.6, 31.4, 31.3, 31.2, 31.1, 31.0, 30.9, 30.6, 30.4, 30.2, 29.7, 29.6, 29.5, 28.4, 25.1, 24.8, 24.7, 24.5, 24.4, 22.5, 22.4, 22.3, 15.4, 15.3, 14.6, 14.3, 14.0, 13.9; HRMS (EI): Exact mass calcd for C₆₀H₁₀₈N₉O₁₁ [M+NH₄]⁺ 1130.8168, found 1130.8166. MPT-5-184



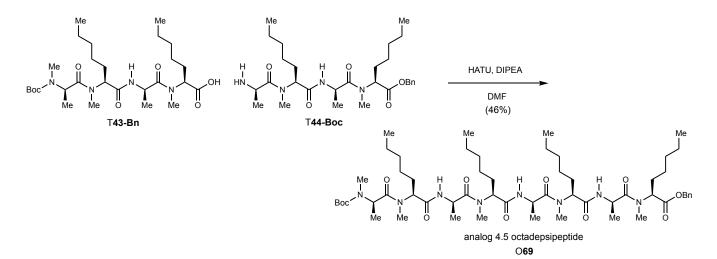
(6R,9S,12R,15S,18R,21S,24R,27S)-2,2,6,8,11,12,14,17,18,20,23,24,26-tridecamethyl-4,7,10,13,16,19,22,25-Benzvl octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O66). A round-bottom flask was charged with the amine (124 mg, 193 µmol), acid (124 mg, 193 µmol), and DMF (1.9 mL). The mixture was cooled to 0 °C and then DIPEA (83.0 μL, 579 μmol) and HATU (177 mg, 559 μmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a yellow oil (121 mg, 57%). [α]²⁴_D - 19 (c 0.31, CHCl₃); R_f = 0.19 (30% EtOAc/hexanes); IR (film) 3650, 2958, 2932, 2861, 1731, 1660, 1466, 1408, 1381, 1141 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.31-7.25 (series of m, 6H), 5.17-5.13 (m, 5H), 5.06 (d, J = 4.6 Hz, 1H), 5.05 (d, J = 4.6 Hz, 1H), 4.97 (m, 1H), 4.92 (m, 1H), 4.64 (m, 1H), 3.12-2.64 (series of m, 21H), 1.81-1.74 (series of m, 6H), 1.70-1.62 (m, 4H), 1.48 (m, 3H), 1.41 (s, 9H), 1.36-1.33 (series of m, 4H), 1.32-1.29 (series of m, 8H), 1.26-1.16 (series of m, 19H), 0.85-0.73 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) 171.7, 171.4, 171.1, 170.8, 170.3, 170.1, 169.9, 169.5, 156.0, 135.3, 128.7, 128.4, 128.3, 79.7, 73.4, 73.0, 72.1, 72.0, 71.7, 71.6, 71.4, 67.6, 67.2, 67.0, 55.0, 54.9, 53.3, 52.4, 46.1, (29) 31.6, 31.5, 31.4, 31.3, 31.2, 31.1, 30.8, 30.7, 30.6, 30.7, 30.2, 29.6, 28.9, 28.3, 25.3, 25.1, 24.8, 24.5, 22.6, 22.5, 22.4, 22.3, 15.4, 14.7, 14.3, 14.0; HRMS (EI): Exact mass calcd for C₅₉H₁₀₆N₉O₁₁ [M+NH₄]⁺ 1116.8012, found 1116.8007 MPT-4-178



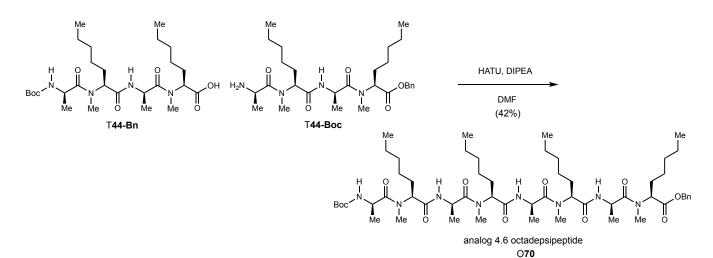
Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,5,6,8,11,12,14,18,20,24,26-dodecamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O67). A round-bottom flask was charged with the amine (63.6 mg, 111 µmol), acid (59.3 mg, 111 µmol), and DMF (1.1 mL). The mixture was cooled to 0 °C and then DIPEA (49.4 µL, 333 µmol) and HATU (105 mg, 333 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a yellow oil (64.1 mg, 53%). [α]_D²⁴ - 11 (*c* 0.27, CHCl₃); R_f = 0.22 (30% EtOAc/hexanes); IR (film) 3322, 2921, 2862, 1748, 1669, 1539, 1456, 1392, 1158, 1092 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.40-7.32 (series of m, 7H), 5.31 (m, 1H), 5.22-5.12 (series of m, 5H), 5.06 (m, 1H), 4.99 (m, 1H), 4.70 (m, 2H), 3.36-2.80 (series of m, 18H), 1.82 (m, 2H), 1.73 (m, 2H), 1.63 (m, 2H), 1.56-1.51 (series of m, 6H), 1.46 (m, 9H), 1.41-1.37 (series of m, 6H), 1.32-1.29 (series of m, 6H), 1.27-1.20 (series of m, 20H), 0.90-0.83 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) ppm 171.8, 171.1, 170.5, 170, 169.9, 169.8, 169.5, 169.3, 156.0, 135.3, 128.6, 128.4, 128.3, 79.8, 73.4, 73.0, 71.4, 67.2, 67.0, 54.1, 52.7, 52.5, 31.7, 31.6, 31.55, 31.5, 31.4, 31.37, 31.3, 31.2, 31.1, 31.0, 30.9, 30.8, 30.4, 30.3, 30.2, 30.1, 29.7, 29.6, 29.4, 28.3, 25.0, 24.8, 24.7, 24.4, 22.42, 22.4, 22.32, 22.3, 15.3, 14.6, 14.2, 14.0; HRMS (El): Exact mass calcd for C₅₈H₁₀₄N₉O₁₁ [M+NH₄]⁺ 1102.7855, found 1102.7850. MPT-4-246



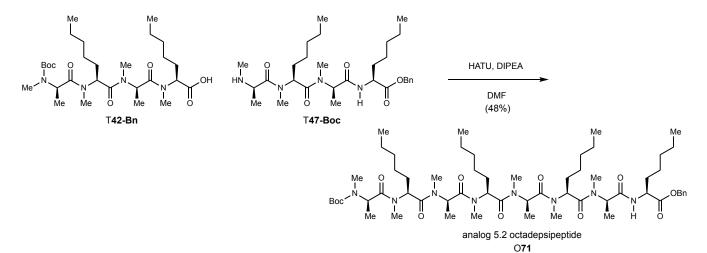
Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,5,6,8,12,14,17,18,20,24,26-dodecamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O68). A round-bottom flask was charged with the amine (83.0 mg, 149 µmol), acid (81.3 mg, 149 µmol), and DMF (1.5 mL). The mixture was cooled to 0 °C and then DIPEA (80.0 µL, 448 µmol) and HATU (171 mg, 559 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-40% ethyl acetate in hexanes) to afford the product as a light yellow oil (102 mg, 63%). [α]_D²⁴ - 18 (*c* 0.16, CHCl₃); R_f = 0.10 (25% EtOAc/hexanes); IR (film) 3418, 2958, 2925, 2854, 1740, 1644, 1557, 1540, 1375, 1260, 1080 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.40-7.29 (series of m, 6H), 7.09 (m, 1H), 5.29 (m, 1H), 5.22-5.08 (series of m, 4H), 5.03 (m, 1H), 4.96 (m, 1H), 4.90-4.72 (series of m, 3H), 3.03-2.71 (series of m, 18H), 2.02-1.88 (series of m, 3H), 1.65 (br s, 9H), 1.48-1.44 (series of m, 10H), 1.35-1.23 (series of m, 28H), 0.90-0.83 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₅₈H₁₀₄N₉O₁₁ [M+NH₄]⁺ 1102.7855, found 1102.7850. MPT-3-087



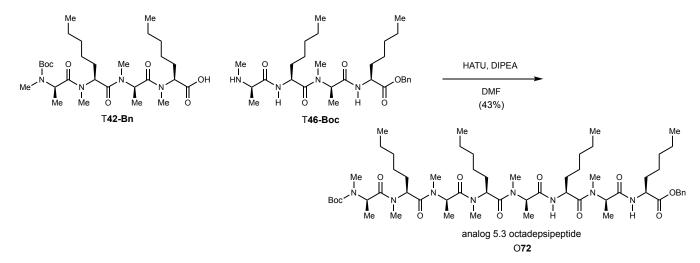
Benzvl (6R,9S,12R,15S,18R,21S,24R,27S)-2,2,5,6,8,12,14,18,20,24,26-undecamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O69). A round-bottom flask was charged with the amine (85.8 mg, 154 µmol), acid (82.0 mg, 154 µmol), and DMF (1.5 mL). The mixture was cooled to 0 °C and then DIPEA (48.7 µL, 462 µmol) and HATU (240 mg, 462 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-65% ethyl acetate in hexanes) to afford the product as a light yellow oil (76 mg, 46%). $\left[\alpha\right]_{D}^{24}$ - 29 (c 0.39, CHCl₃); R_f = 0.15 (40% EtOAc/hexanes); IR (film) 3419, 2960, 2923, 2853, 1747, 1652, 1540, 1456, 1378, 1261, 1100 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.32-7.24 (series of m, 5H), 7.20-7.09 (*N*-H, series of m, 3H), 5.22-5.09 (series of m, 6H), 5.05 (m, 1H), 4.97 (m, 1H), 4.86 (m, 1H), 4.63 (m, 1H), 3.29-2.69 (series of m, 15H), 1.81-1.75 (series of m, 4H), 1.66 (m, 3H), 1.54 (s, 6H), 1.49 (m, 3H), 1.38-1.28 (series of m, 18H), 1.25-1.17 (series of m, 19H), 0.83-0.77 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) ppm 172.0, 171.4, 171.1, 170.8, 170.2, 170.0, 169.5, 169.3, 155.0, 135.3, 128.9, 128.6, 128.3, 80.0, 79.7, 73.5, 73.0, 72.3, 71.7, 71.4, 68.6, 67.3, 60.5, 58.8, 55.0, 53.0, 52.5, 35.1, 34.7, (29) 32.1, 31.7, 31.5, 31.4, 31.3, 31.2, 31.1, 30.9, 30.8, 30.7, 29.7, 29.6, 28.2, 28.1, 25.1, 24.8, 24.5, 24.1, 22.5, 22.3, 15.5, 14.3, 14.0, 13.9; HRMS (EI): Exact mass calcd for C₅₇H₁₀₂N₉O₁₁ [M+NH₄]⁺ 1088.7699, found 1088.7696. MPT-5-237



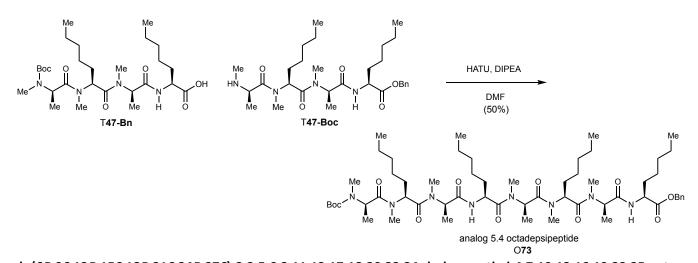
Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,6,8,12,14,18,20,24,26-decamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O70). A round-bottom flask was charged with the amine (170 mg, 316 µmol), acid (170 mg, 316 µmol), and DMF (3.0 mL). The mixture was cooled to 0 °C and then DIPEA (136 µL, 948 µmol) and HATU (291 mg, 559 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-50% ethyl acetate in hexanes) to afford the product as a light yellow oil (139 mg, 42%). [α]_D²⁴ - 16 (*c* 0.22, CHCl₃); R_f = 0.11 (40% EtOAc/hexanes); IR (film) 3444, 2960, 2931, 2863, 1717, 1694, 1682, 1443, 1326, 1206, 1137 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists as a mixture of rotamers causing significant peak broadening and overlap. Refer to the image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) ppm 172.5, 172.0, 170.9, 170.2, 169.5, 167.3, 166.7, 166.0, 156.0, 134.9, 128.7, 128.6, 128.5, 79.8, 71.5, 67.3, 61.6, 61.1, 55.0, 54.5, 53.4, 51.7, 46.1, 32.2, 31.8, 31.5, 31.4, 31.3, 31.1, 31.0, 30.8, 30.7, 30.6, 30.3, 30.2, 29.8, 29.6, 28.4, 26.2, 25.0, 24.9, 24.8, 24.7, 24.0, 22.4, 15.6, 15.3, 14.7, 14.4, 14.1, 14.0, 13.9; HRMS (EI): Exact mass calcd for C₅₆H₁₀₀N₉O₁₁ [M+NH₄]⁺ 1074.7542, found 1074.7538. MPT-4-067



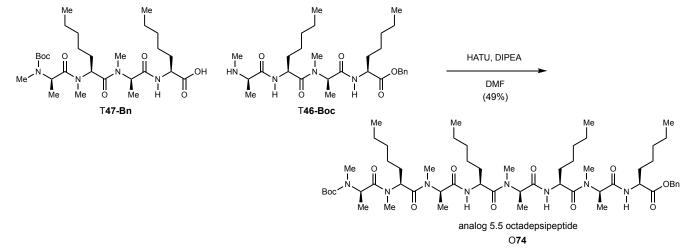
Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,5,6,8,11,12,14,17,18,20,23,24-tridecamethyl-4,7,10,13,16,19,22,25octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O71). A round-bottom flask was charged with the amine (103 mg, 181 µmol), acid (99.0 mg, 181 µmol), and DMF (1.8 mL). The mixture was cooled to 0 °C and then DIPEA (57.2 µL, 543 µmol) and HATU (282 mg, 543 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-60% ethyl acetate in hexanes) to afford the product as a light yellow oil (95.9 mg, 48%). [*α*]²⁴_D - 13 (*c* 0.86, CHCl₃); R_f = 0.16 (50% EtOAc/hexanes); IR (film) 3317, 2956, 2930, 2859, 1739, 1644, 1525, 1458, 1369, 1275, 1090 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.57 and 6.57 (br m, 1H), 7.41-7.32 (series of m, 5H), 5.23-4.53 (series of m, 10H), 3.16-2.66 (series of m, 21H), 1.83 (m, 3H), 1.77-1.62 (series of m, 5H), 1.52-1.36 (series of m, 20H), 1.38-1.24 (series of m, 25H), 0.91-0.84 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) ppm 172.4, 172.2, 172.0, 171.5, 171.2, 170.5, 169.8, 169.6, 156.2, 135.3, 128.5, 128.3, 128.2, 80.9, 74.5, 74.2, 73.7, 72.8, 66.9, 57.4, 49.5, 49.1, 47.9, 34.9, 31.6, 31.4, 31.35, 31.27, 31.2, 31.0, 30.9, 30.8, 29.7, 29.6, 29.3, 28.4, 28.3, 28.2, 24.61, 24.57, 24.50, 24.4, 24.2, 23.9, 22.6, 22.4, 22.3, 22.3, 17.4, 16.4, 14.7, 13.92, 13.89, 13.87, 13.83; HRMS (EI): Exact mass calcd for $C_{59}H_{106}N_9O_{11}$ [M+NH₄]⁺ 1116.8012, found 1116.8009. MPT-4-105



Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,5,6,8,11,12,14,17,18,23,24-dodecamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O72). A round-bottom flask was charged with the amine (104 mg, 196 µmol), acid (112 mg, 196 µmol), and DMF (2.0 mL). The mixture was cooled to 0 °C and then DIPEA (62.0 µL, 588 µmol) and HATU (305 mg, 588 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-60% ethyl acetate in hexanes) to afford the product as a light yellow oil (90.2 mg, 43%). [*α*]²⁴_D - 12 (*c* 0.56, CHCl₃); *R*_f = 0.13 (50% EtOAc/hexanes); IR (film) 3300, 2957, 2933, 2861, 1742, 1650, 1458, 1408, 1275, 1086 cm⁻¹; ¹H NMR (600 MHz, (CD₃)₂CO) δ 7.29-7.18 (series of m, 5H), 7.07 (br d, *J* = 7.9, 1H), 7.02 (br d, *J* = 7.6, 1H), 5.22-3.55 (series of m, 10H), 2.96-2.61 (series of m, 18H), 1.88-1.63 (series of m, 6H), 1.60-1.42 (series of m, 3H), 1.34-1.09 (series of m, 44H), 0.85-0.69 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) ppm 172.4, 171.1, 170.7, 170.1, 169.9, 169.7, 169.5, 169.3, 155.0, 135.3, 128.6, 128.5, 128.3, 79.6, 73.4, 73.0, 71.7, 71.6, 67.0, 55.0, 52.5, 49.3, 46.1, 42.7, 32.1, 31.6, 31.5, 31.43, 31.38, 31.3, 31.2, 31.1, 30.9, 30.8, 30.7, 29.7, 29.5, 28.3, 25.1, 24.9, 24.9, 24.8, 24.5, 22.42, 22.39, 22.3, 18.7, 18.4, 15.6, 15.4, 14.3, 14.0, 13.9; HRMS (EI): Exact mass calcd for C₄₆H₈₅N₈O₉ [M-Boc-Bn]⁻ 893.6445, found 893.6446. MPT-4-008

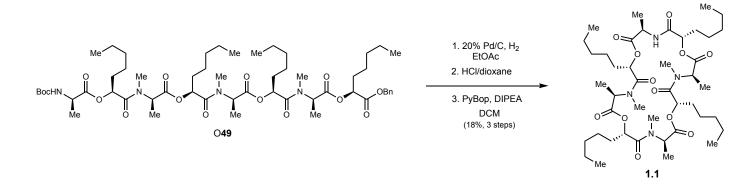


Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,5,6,8,11,12,17,18,20,23,24-dodecamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O73). A round-bottom flask was charged with the amine (68.8 mg, 124 µmol), acid (68.0 mg, 124 µmol), and DMF (1.2 mL). The mixture was cooled to 0 °C and then DIPEA (39.2 µL, 372 µmol) and HATU (193 mg, 462 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-60% ethyl acetate in hexanes) to afford the product as a light yellow oil (66.9 mg, 50%). [α]_D²⁴ - 27 (*c* 0.34, CHCl₃); R_f = 0.152 (25% EtOAc/hexanes); IR (film) 3384, 2955, 2932, 2861, 1742, 1651, 1522, 1417, 1392, 1215, 1093 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) ppm 171.7, 171.6, 171.3, 171.3, 170.7, 170.5, 170.0, 169.3, 155.6, 135.2, 128.6, 128.4, 128.3, 79.9, 79.6, 73.3, 72.9, 72.7, 71.4, 71.3, 71.2, 66.9, 57.5, 56.7, 55.7, 54.5, 38.8, 37.6, 37.2, 37.0, 36.6, 31.4, 31.3, 31.1, 30.9, 30.8, 30.3, 29.8, 29.6, 28.3, 24.9, 24.7, 24.5, 24.4, 23.3, 23.2, 22.8, 22.3, 22.2, 21.0, 14.0, 13.9, 13.84, 13.82; HRMS (El): Exact mass calcd for C₅₈H₁₀₄N₉O₁₁ [M+NH₄]⁺ 1102.7855, found 1102.7855. MPT-5-190



SI-39

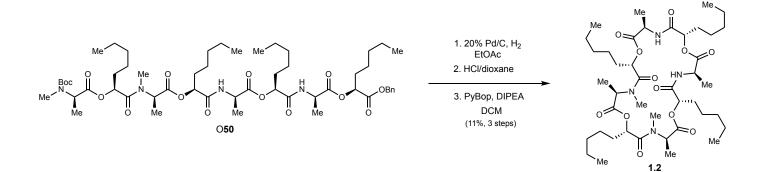
Benzyl (6*R*,9*S*,12*R*,15*S*,18*R*,21*S*,24*R*,27*S*)-2,2,5,6,8,11,12,17,18,23,24-undecamethyl-4,7,10,13,16,19,22,25-octaoxo-9,15,21,27-tetrapentyl-3-oxa-5,8,11,14,17,20,23,26-octaazaoctacosan-28-oate (O74). A round-bottom flask was charged with the amine (104 mg, 187 µmol), acid (100 mg, 187 µmol), and DMF (1.9 mL). The mixture was cooled to 0 °C and then DIPEA (59.1 µL, 561 µmol) and HATU (292 mg, 561 µmol) were added. The reaction was stirred at 0 °C for 30 min, then warmed to 23 °C for 18 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, dried, and concentrated. The crude residue was subjected to flash column chromatography (SiO₂, 5-60% ethyl acetate in hexanes) to afford the product as a light yellow oil (98.0 mg, 49%). [α]_D²⁴ - 12 (*c* 0.25, CHCl₃); R_f = 0.22 (40% EtOAc/hexanes); IR (film) 3429, 2960, 2931, 2862, 1694, 1651, 1633, 1443, 1326, 1206, 1137 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) ppm 172.4, 171.1, 170.8, 170.4, 169.8, 169.5, 168.9, 168.6, 156.0, 135.2, 128.6, 128.5, 128.3, 80.1, 73.4, 73.1, 73.0, 72.2, 71.9, 71.6, 71.2, 71.1, 67.1, 54.9, 54.4, 53.5, 53.4, 50.8, 49.3, 36.0, 35.9, 32.2, 32.1, 31.3, 31.2, 30.9, 30.7, 30.4, 30.1, 29.7, 28.3, 25.0, 24.9, 24.8, 24.8, 24.6, 24.5, 22.4, 22.3, 15.5, 14.6, 14.2, 13.9; HRMS (EI): Exact mass calcd for C₅₇H₁₀₈N₉O₁₁ [M+NH₄]⁺ 1088.7699, found 1088.7695. MPT-4-181



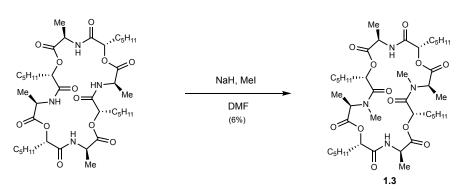
(3*R*,65,9*R*,12*S*,15*R*,18*S*,21*R*,24*S*)-3,4,9,10,15,16,21-heptamethyl-6,12,18,24-tetrapentyl-1,7,13,19-tetraoxa-4,10,16,22-tetraazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (1.1). A round-bottom flask was charged with the depsipeptide (108 mg, 103 µmol), dissolved in EtOAc (1.0 mL), and treated with 10% Pd/C (2.2 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added HCl/dioxane (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DCM (20.8 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (40.0 µL, 228 µmol) and PyBop (56.7 mg, 108 µmol) were added. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 20.2 m) afforded the 24-membered macrocycle (16.8 mg, 18%) as a light yellow oil. [α]²⁴₂+25 (c 0.18, CHCl₃); R_f = 0.34 (60% EtOAc in hexanes); IR (film) 3331, 2927, 2855, 2359, 1747, 1653, 15457, 1196 cm⁻¹; ¹H NMR

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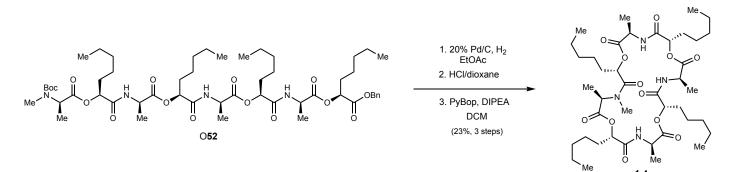
(600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₃H₇₆N₄O₁₃ [M+CH₃OH]⁺ 856.5409 found [856.5443]. MPT-3-261

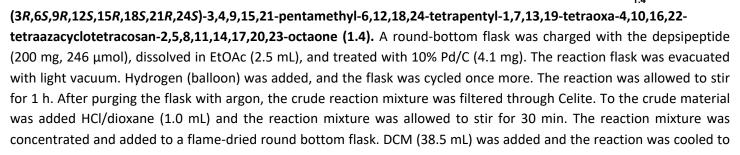


(3R,65,9R,125,15R,185,21R,245)-3,4,9,10,15,21-hexamethyl-6,12,18,24-tetrapentyl-1,7,13,19-tetraoxa-4,10,16,22tetraazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (1.2). A round-bottom flask was charged with the depsipeptide (130 mg, 126 µmol), dissolved in EtOAc (1.3 mL), and treated with 10% Pd/C (2.7 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added HCl/dioxane (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DCM (25.0 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (48.0 μL, 275 μmol) and PyBop (68.4 mg, 131 μmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 20.2 m) afforded the 24-membered macrocycle (24.6 mg, 11%) as a light yellow oil. $\left[\alpha\right]_{D}^{24}$ +19 (c 0.27, CHCl₃); R_f = 0.42 (60% EtOAc in hexanes); IR (film) 3300, 2954, 2928, 2859, 1744, 1648, 1546, 1460, 1219, 1082 cm⁻ ¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₂H₇₂N₄O₁₂Na [M+Na]⁺ 847.5045, found 847.5046. MPT-2-169

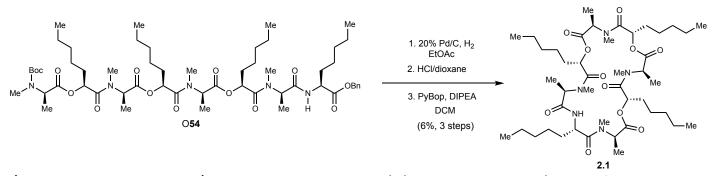


(3*R*,6*S*,9*R*,12*S*,15*R*,18*S*,21*R*,24*S*)-3,4,9,15,16,21-hexamethyl-6,12,18,24-tetrapentyl-1,7,13,19-tetraoxa-4,10,16,22tetraazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (1.3). A flame-dried round-bottom flask was charged with N-H depsipeptide (30.0 mg, 37.6 µmol) and dry DMF (527 µL) at 0 °C. Methyl iodide (94 µL, 1.51 mmol) was then added to the reaction mixture, and NaH (9.0 mg, 380 µmol in DMF (225 µL)) was added in 3 aliquots of 75 µL over 15 m. The reaction was allowed to stir at 0 °C for 20 m, and it was then quenched by the dropwise addition of satd aq NH₄Cl. The aqueous layer was extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, satd aq Na₂S₂O₃, water, brine, dried, concentrated. The residue was subjected to flash column chromatography (SiO₂, 1-10% methanol in dichloromethane) to afford 2 mg (6%) of the desired macrocycle as a colorless foam, as well as 9.3 mg (31%) of *ent*verticilide. $[\alpha]_D^{24}$ +11 (*c* 0.06, CHCl₃); R_f = 0.40 (4% MeOH/DCM); IR (film) 3298, 2926, 2856, 1739, 1644, 1556, 1455, 1261, 1092 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the 1H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₂H₇₆N₅O₁₂ [M+NH₄]⁺ 842.5490, found [842.5482]. MPT-5-040

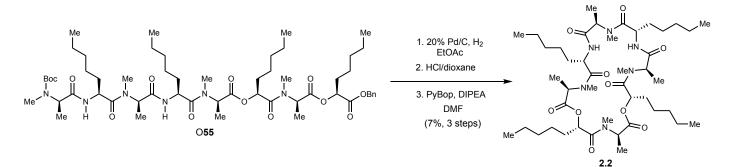




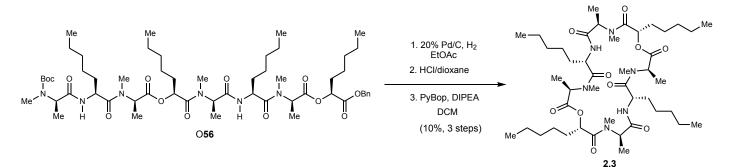
0 °C. Once at 0 °C, DIPEA (73.8 μL, 422 μmol) and PyBop (105 mg, 201 μmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, $R_t = 20.2$ m) afforded the 24-membered macrocycle (29.0 mg, 23%) as a white amorphous solid $[\alpha]_D^{24}$ +25 (*c* 0.98, CHCl₃); R_f = 0.33 (60% EtOAc in hexanes); IR (film) 3284, 2929, 2860, 2359, 1744, 1649, 1555, 1456, 1377, 1210 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) δ 7.70 (br d, 1H), 7.17 (br m, 2H), 5.09 (br m, 2H), 5.00 (dd, *J* = 8.9, 6.0 Hz, 1H), 4.88 (dd, *J* = 5.8, 5.8 Hz, 1H), 4.61 (br m, 2H), 4.06 (br s, 1H), 3.80 (br s, 1H), 3.17 (s, 3H), 2.02-1.93 (m, 2H), 1.88-1.60 (series of m, 8H), 1.52-1.46 (series of m, 8H), 1.42 (br d, *J* = 6.5 Hz, 4H), 1.34-1.24 (series of m, 22H), 0.93-0.86 (series of m, 12H); ¹³C NMR (150 MHz, CDCl₃) ppm 171.9, 171.7, 170.7, 170.5, 170.4, 170.0, 169.7, 169.2, 74.4, 74.3, 58.9, 49.2, 48.0, 47.7, 35.7, 31.7, 31.6, 31.5, 31.4, 31.35, 31.33, 31.25, 31.18, 31.0, 30.3, 29.7, 29.6, 25.3, 25.1, 25.0, 24.4, 22.6, 22.5, 22.4, 17.7, 16.7, 16.2, 14.0, 13.9, 13.9, 13.5; HRMS (EI): Exact mass calcd for C₄₁H₇₄N₅O₁₂ [M+NH₄]⁺ 828.5334, found [828.5316]. MPT-3-237



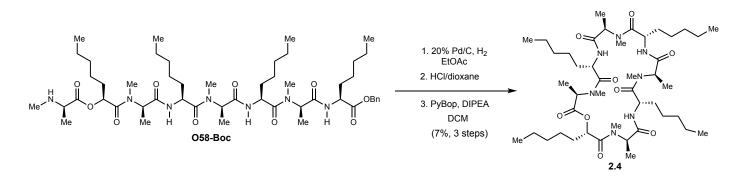
(3R,6S,9R,12S,15R,18S,21R,24S)-3,4,9,10,15,16,21,22-octamethyl-6,12,18,24-tetrapentyl-1,7,13-trioxa-4,10,16,19,22pentaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (2.1). A round-bottom flask was charged with the depsipeptide (100 mg, 94.4 µmol), dissolved in EtOAc (944 µL), and treated with 10% Pd/C (20.0 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/dioxane (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DCM (18.9 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (49.3 µL, 283 µmol) and PyBop (51.8 mg, 99.1 µmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with DCM. The combined organic layers were washed with satd ag NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 20.5 m) afforded the 24-membered macrocycle (6.24 mg, 6.0%) as a light yellow oil. $[\alpha]_D^{24} - 6.2$ $(c 0.29, CHCl_3); R_f = 0.50 (30\% EtOAc in hexanes); IR (film) 3381, 2955, 2925, 2854, 1744, 1660, 1540, 1260, 1093 cm⁻¹; ¹H$ NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₄H₇₈N₅O₁₁ [M+H]⁺ 852.5698, found 852.6592. MPT-2-201



(3R,65,9R,125,15R,185,21R,245)-3,4,9,10,15,16,21,22-octamethyl-6,12,18,24-tetrapentyl-1,7,13-trioxa-4,10,16,19,22pentaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (2.2). A round-bottom flask was charged with the depsipeptide (150 mg, 142 µmol), dissolved in EtOAc (1.9 mL), and treated with 10% Pd/C (30 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/dioxane (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DMF (24.0 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (66.0 μL, 426 μmol) and PyBop (111 mg, 213 μmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 24.2 m) afforded the 24-membered macrocycle (8.43 mg, 7.0%) as a light yellow oil. $[\alpha]_D^{24}$ - 13 (c 0.15, CHCl₃); R_f = 0.50 (50% EtOAc in hexanes); IR (film) 3443, 2959, 2920, 2851, 1650, 1633, 1556, 1455 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for $C_{44}H_{79}N_6O_{10}$ [M+H]⁺ 851.5858, found 851.5853. MPT-2-293

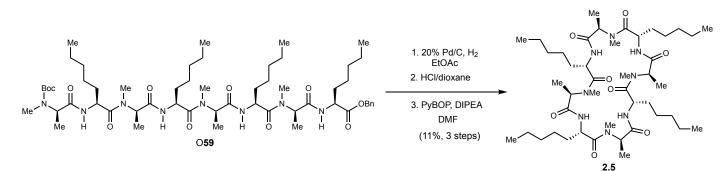


(3R,65,9R,125,15R,185,21R,245)-3,4,9,10,15,16,21,22-octamethyl-6,12,18,24-tetrapentyl-1,13-dioxa-4,7,10,16,19,22hexaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (2.3). A round-bottom flask was charged with the depsipeptide (97.0 mg, 92.0 µmol), dissolved in EtOAc (2.0 mL), and treated with 10% Pd/C (2.0 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCI/dioxane (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DCM (18.0 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (35.0 μL, 200 μmol) and PyBop (50.0 mg, 97.0 μmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with DCM. The combined organic layers were washed with satd ag NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 22.2 m) afforded the 24-membered macrocycle (7.8 mg, 10.0%) as a light yellow oil. $\left[\alpha\right]_{D}^{24}$ - 16 (c 0.23, CHCl₃); R_f = 0.43 (50% EtOAc in hexanes); IR (film) 3478, 2922, 2851, 1740, 1633, 1455, 1260 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the 1 H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₄H₈₂N₇O₁₀ [M+NH₄]⁺ 868.6123, found 868.6119. MPT-2-267

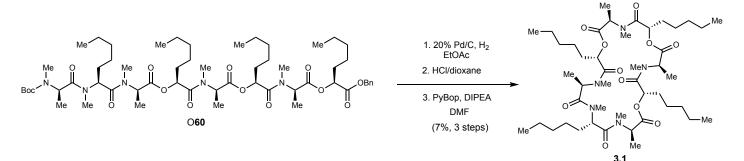


(3R,6S,9R,12S,15R,18S,21R,24S)-3,4,9,10,15,16,21,22-octamethyl-6,12,18,24-tetrapentyl-1-oxa-4,7,10,13,16,19,22heptaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (2.4). A round-bottom flask was charged with the depsipeptide (204 mg, 193 μmol), dissolved in EtOAc (1.9 mL), and treated with 10% Pd/C (4.90 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material

was added HCl/dioxane (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DCM (38.6 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (73.9 μ L, 424 μ mol) and PyBop (105 mg, 202 μ mol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with DCM. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 24.2 m) afforded the 24-membered macrocycle (12.0 mg, 7.3%) as a yellow oil. [α]_D²⁴ + 6.0 (*c* 0.14, CHCl₃); R_f = 0.41 (60% EtOAc in hexanes); IR (film) 3389, 2952, 2851, 1743, 1633, 1455, 1165cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₄H₈₃N₈O₉ [M+NH4]⁺ 867.6283, found 867.6283. MPT-3-015

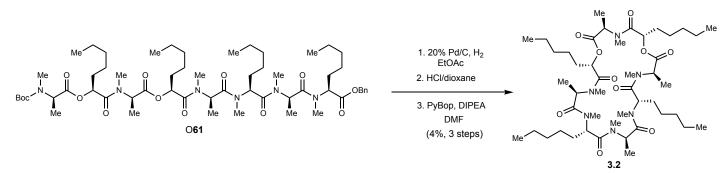


(35,6R,9S,12R,15S,18R,21S,24R)-1,6,7,12,13,18,19,24-octamethyl-3,9,15,21-tetrapentyl-1,4,7,10,13,16,19,22octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (2.5). A round-bottom flask was charged with the depsipeptide (116 mg, 110 µmol), dissolved in EtOAc (1.1 mL), and treated with 10% Pd/C (2.4 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/dioxane (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DMF (24.0 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (43.4 µL, 330 µmol) and PyBop (60.0 mg, 115 µmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd ag NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 23.1 m) afforded the 24-membered macrocycle (10.2 mg, 11.0%) as a light yellow oil. $\left[\alpha\right]_{D}^{24}$ - 11 (c 0.29, CHCl₃); R_f = 0.33 (50% EtOAc in hexanes); IR (film) 3292, 2929, 1654, 1637, 1560, 1458 cm⁻¹; ¹H NMR (600 MHz. CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₄H₈₄N₉O₈ [M+NH₄]⁺ 866.6443, found 866.6440. MPT-2-290



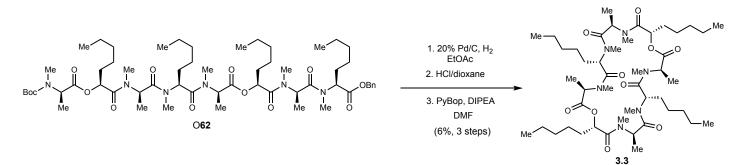
(3R,6S,9R,12S,15R,18S,21R,24S)-3,4,9,10,15,16,19,21,22-nonamethyl-6,12,18,24-tetrapentyl-1,7,13-trioxa-

4,10,16,19,22-pentaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (3.1). A round-bottom flask was charged with the depsipeptide (189 mg, 176 µmol), dissolved in EtOAc (1.8 mL), and treated with 10% Pd/C (37.8 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/dioxane (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DMF (49.5mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (49.5 µL, 284 µmol) and PyBop (70.6 mg, 129 µmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 23.0 m) afforded the 24-membered macrocycle (7.6 mg, 5.1%) as a yellow oil. $[\alpha]_D^{24}$ + 5.0 (c 0.19, CHCl₃); R_f = 0.48 (50% EtOAcEtOAc in hexanes); IR (film) 2930, 2861, 1747, 1715, 1669, 1539, 1157 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₅H₈₃N₆O₁₁ [M+NH₄]⁺ 883.6120, found 883.6122. MPT-3-078



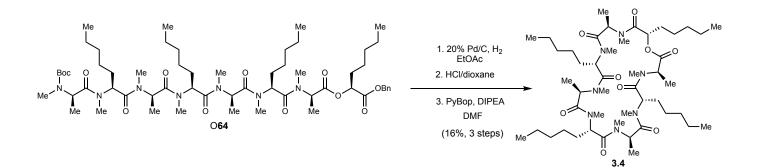
(3R,6S,9R,12S,15R,18S,21R,24S)-3,4,9,10,13,15,16,19,21,22-decamethyl-6,12,18,24-tetrapentyl-1,7-dioxa-4,10,13,16,19,22-hexaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (3.2). A round-bottom flask was charged with the depsipeptide (149 mg, 138 μmol), dissolved in EtOAc (1.4 mL), and treated with 10% Pd/C (29.8 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/dioxane (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction

mixture was concentrated and added to a flame-dried round bottom flask. DMF (48.0 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (55.0 µL, 414 µmol) and PyBop (76.0 mg, 150 µmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 23.0 m) afforded the 24-membered macrocycle (4.80 mg, 4.0%) as a yellow oil. $[\alpha]_D^{24}$ - 13 (*c* 0.15, CHCl₃); R_f = 0.40 (60% EtOAc in hexanes); IR (film) 2930, 2850, 1744, 1698, 1456, 1156 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₆H₈₆N₇O₁₀ [M+NH₄]⁺ 896.6436, found 896.6433. MPT-4-037



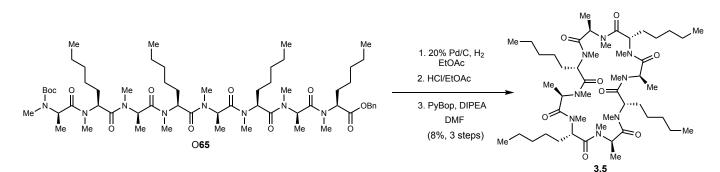
(3R,6S,9R,12S,15R,18S,21R,24S)-3,4,7,9,10,15,16,19,21,22-decamethyl-6,12,18,24-tetrapentyl-1,13-dioxa-

4,7,10,16,19,22-hexaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (3.3). A round-bottom flask was charged with the depsipeptide (154 mg, 142 µmol), dissolved in EtOAc (1.4 mL), and treated with 10% Pd/C (30.8 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/dioxane (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DMF (28.4 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (49.5 µL, 284 µmol) and PyBop (70.6 mg, 136 µmol) were added. The reaction was stirred at 0 °C for 1.5 h, then allowed to warm to ambient temperature and stir for an additional 1 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, $R_t = 23.0$ m) afforded the 24-membered macrocycle (7.0 mg, 6.1%) as a yellow oil. $[\alpha]_D^{24} - 23$ (c 0.22, CHCl₃); R_f = 0.44 (50% EtOAc in hexanes); IR (film) 2932, 2871, 1745, 1667, 1456, 1190 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₆H₈₆N₇O₁₀ [M+NH₄]⁺ 896.6436, found 896.6435. MPT-3-035



(3R,6S,9R,12S,15R,18S,21R,24S)-3,4,7,9,10,13,15,16,19,21,22-undecamethyl-6,12,18,24-tetrapentyl-1-oxa-

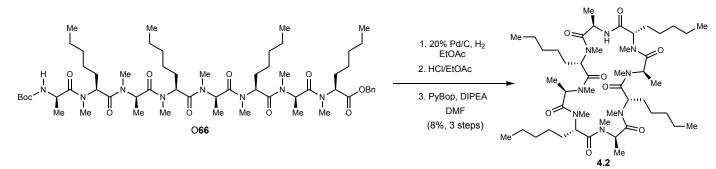
4,7,10,13,16,19,22-heptaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (3.4). A round-bottom flask was charged with the depsipeptide (30 mg, 27 µmol), dissolved in EtOAc (270 µL), and treated with 10% Pd/C (6.0 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/dioxane (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DMF (5.4 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (15 µL, 81 µmol) and PyBop (15 mg, 28 µmol) were added. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, Rt = 23.1 m) afforded the 24-membered macrocycle (2.9 mg, 16%) as a yellow oil. [α]²⁴_D + 4.0 (c 0.11, CHCl₃); R_f = 0.40 (60% EtOAc in hexanes); IR (film) 2931, 2860, 1743, 1663, 1456, 1185 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₇H₈₉N₈O₉ [M+NH₄]* 909.6753, found 909.6748. MPT-5-181



(3R,6S,9R,12S,15R,18S,21R,24S)-1,3,4,7,9,10,13,15,16,19,21,22-dodecamethyl-6,12,18,24-tetrapentyl-1,4,7,10,13,16,19,22-octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (3.5). A round-bottom flask was charged with

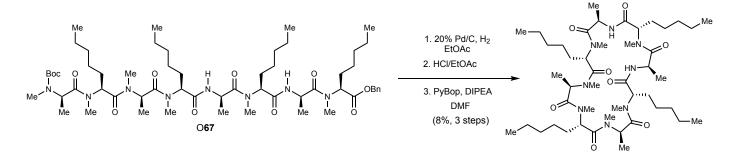
the depsipeptide (74.1 mg, 66.6 μ mol), dissolved in EtOAc (666 μ L), and treated with 10% Pd/C (14.8 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To

the crude material was added 4M HCl/EtOAc (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DMF (13.3 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (26.3 μ L, 200 μ mol) and PyBop (36.5 mg, 69.9 μ mol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 24.7 m) afforded the 24-membered macrocycle (5.5 mg, 8.3%) as a yellow oil. [α]_D²⁴ + 4.9 (c 0.21, CHCl₃); R_f = 0.31 (40% EtOAc in hexanes); IR (film) 2927, 2855, 1747, 1653, 1457, 1196 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₈H₈₉N₈O₈ [M+H]⁺ 905.6803, found 905.6800. MPT-5-242



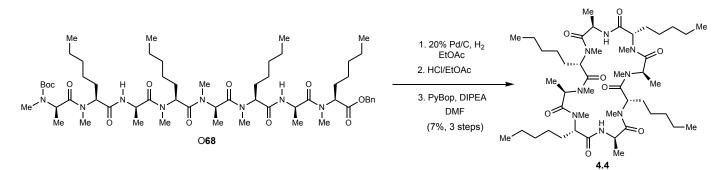
(3R,6S,9R,12S,15R,18S,21R,24S)-1,3,4,7,9,10,13,15,16,19,21-undecamethyl-6,12,18,24-tetrapentyl-

1,4,7,10,13,16,19,22-octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (4.2). A round-bottom flask was charged with the depsipeptide (116 mg, 110 μmol), dissolved in EtOAc (1.1 mL), and treated with 10% Pd/C (23.2 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCI/EtOAc (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. Dry DMF (24.0 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (43.4 µL, 330 µmol) and PyBop (60.0 mg, 115 µmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 24.1 m) afforded the 24-membered macrocycle (6.0 mg, 7.5%) as a light yellow oil. $[\alpha]_D^{24}$ – 8.1 (c 0.30, CHCl₃); R_f = 0.55 (60% EtOAcEtOAc in hexanes); IR (film) 3380, 2925, 2854, 1744, 1661, 1458, 1202, 1092 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₇H₈₇N₈O₈ [M+H]⁺ 891.6641, found 891.6640. MPT-4-224



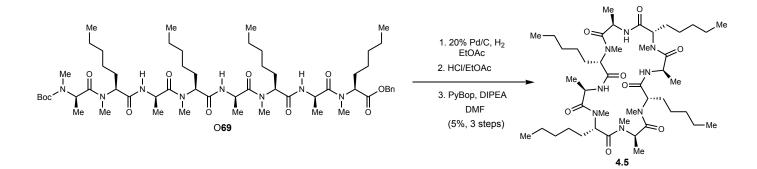
(3R,6S,9R,12S,15R,18S,21R,24S)-1,3,4,7,9,10,13,15,19,21-decamethyl-6,12,18,24-tetrapentyl-1,4,7,10,13,16,19,22-

octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (4.3). A round-bottom flask was charged with the depsipeptide (64.0 mg, 59.2 µmol), dissolved in EtOAc (592 µL), and treated with 10% Pd/C (12.8 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCI/EtOAc (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. Dry DMF (11.9 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (23.4 μL, 178 μmol) and PyBop (32.4 mg, 62.2 μmol) were added. The reaction was stirred at 0 °C for 0.5 h, and then allowed to warm to ambient temperature to stir for an additional 2.5 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd ag NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 22.9 m) afforded the 24-membered macrocycle (4.2 mg, 8.1%) as a light yellow oil. $[\alpha]_D^{24} - 3.7$ (c 0.16, CHCl₃); R_f = 0.50 (60% EtOAc in hexanes); IR (film) 3319, 2926, 2857, 1745, 1692, 1456, 1155, 1028 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₆H₈₅N₈O₈ [M+H]⁺ 877.6490, found 877.6486. MPT-4-251

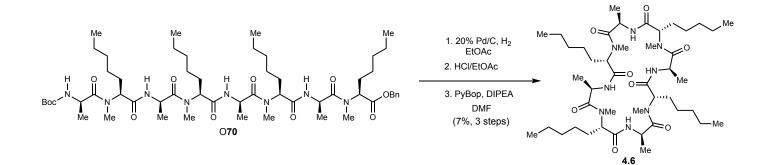


(3R,6S,9R,12S,15R,18S,21R,24S)-1,3,4,7,9,10,13,15,19,21-decamethyl-6,12,18,24-tetrapentyl-1,4,7,10,13,16,19,22octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (4.4). A round-bottom flask was charged with the depsipeptide (102 mg, 94.0 μmol), dissolved in EtOAc (940 μL), and treated with 10% Pd/C (18.8 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir

for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/EtOAc (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. Dry DMF (19.0 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (37.3 μ L, 282 μ mol) and PyBop (51.6 mg, 99.1 μ mol) were added. The reaction was stirred at 0 °C for 0.5 h, and then allowed to warm to ambient temperature to stir for an additional 2.5 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 23.2 m) afforded the 24-membered macrocycle (5.3 mg, 6.6%) as a light yellow oil. [α]²⁴_D - 3.2 (c 0.13, CHCl₃); R_f = 0.34 (60% EtOAc in hexanes); IR (film) 3287, 2922, 2860, 1744, 1649, 1555, 1456, 1210 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₆H₈₈N₉O₈ [M+NH₄]⁺ 894.6756, found 894.6753. MPT-3-103

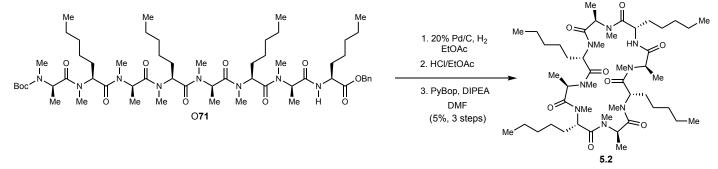


(3R,65,9R,125,15R,185,21R,245)-1,3,4,7,9,13,15,19,21-nonamethyl-6,12,18,24-tetrapentyl-1,4,7,10,13,16,19,22octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (4.5). A round-bottom flask was charged with the depsipeptide (76 mg, 71.0 µmol), dissolved in EtOAc (710 µL), and treated with 10% Pd/C (15.2 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCI/dioxane (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. Dry DMF (14.2 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (9.34 μL, 215 μmol) and PyBop (13.0 mg, 74.6 μmol) were added. The reaction was stirred at 0 °C for 1.5 h, and then allowed to warm to ambient temperature to stir for an additional 1 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd ag NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 23.0 m) afforded the 24-membered macrocycle (3.0 mg, 4.9%) as a light yellow oil. $\left[\alpha\right]_{D}^{24}$ – 7.2 (c 0.14, CHCl₃); R_f = 0.18 (60% EtOAcin hexanes); IR (film) 3340, 2934, 2861, 1747, 1666, 1456, 1391, 1084 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₅H₈₆N₉O₈ [M+NH₄]⁺ 880.6599, found 880.6603. MPT-5-254



(3R,6S,9R,12S,15R,18S,21R,24S)-1,3,7,9,13,15,19,21-octamethyl-6,12,18,24-tetrapentyl-1,4,7,10,13,16,19,22-

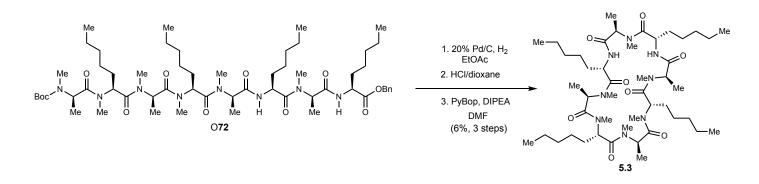
octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (4.6). A round-bottom flask was charged with the depsipeptide (136 mg, 128 µmol), dissolved in EtOAc (1.3 mL), and treated with 10% Pd/C (27.2 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCI/EtOAc (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DMF (25.7 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (50.5 µL, 384 µmol) and PyBop (69.8 mg, 134 µmol) were added. The reaction was stirred at 0 °C for 0.5 h, then allowed to warm to ambient temperature and stir for an additional 2.5 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd ag NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 23.1 m) afforded the 24-membered macrocycle (7.6 mg, 7.0%) as a light yellow oil. $[\alpha]_D^{24} - 9.9$ (c 0.23, CHCl₃); R_f = 0.48 (60% EtOAc in hexanes); IR (film) 3443, 2821, 2851, 1741, 1650, 1260, 1019 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₄H₈₁N₈O₈ [M+H]⁺ 849.6177, found 849.6176. MPT-4-081



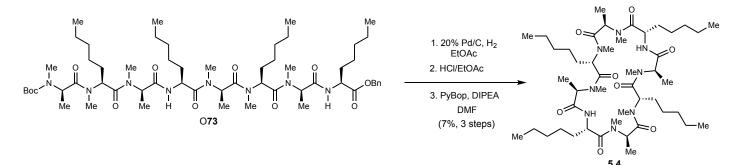
(35,6R,95,12R,155,18R,215,24R)-1,4,6,7,10,12,13,16,18,19,24-undecamethyl-3,9,15,21-tetrapentyl-

1,4,7,10,13,16,19,22-octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (5.2). A round-bottom flask was charged with the depsipeptide (95.9 mg, 86.9 μmol), dissolved in EtOAc (869 μL), and treated with 10% Pd/C (19.2 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/EtOAc (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction

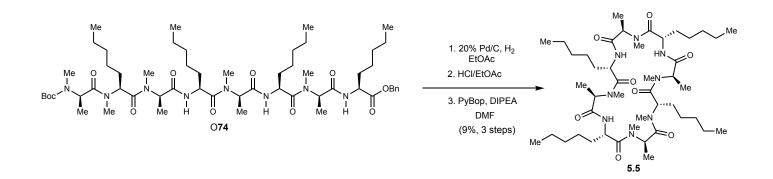
mixture was concentrated and added to a flame-dried round bottom flask. DMF (17.4 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (34.3 µL, 261 µmol) and PyBop (47.5 mg, 91.2 µmol) were added. The reaction was stirred at 0 °C for 0.5 h, and then allowed to warm to ambient temperature to stir for an additional 2.5 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 24.2 m) afforded the 24-membered macrocycle (4.1 mg, 5.3%) as a yellow oil. $[\alpha]_D^{24}$ - 5.1 (*c* 0.12, CHCl₃); R_f = 0.61 (60% EtOAc in hexanes); IR (film) 3478, 2922, 2851, 1740, 1633, 1455, 1260 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₇H₈₇N₈O₈ [M+H]⁺ 891.6647, found 891.6643. MPT-4-184



(35,6R,9S,12R,15S,18R,21S,24R)-1,4,6,7,10,12,13,18,19,24-decamethyl-3,9,15,21-tetrapentyl-1,4,7,10,13,16,19,22octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (5.3). A round-bottom flask was charged with the depsipeptide (90.2 mg, 83.2 µmol), dissolved in EtOAc (832 µL), and treated with 10% Pd/C (18.0 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/dioxane (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. DMF (16.7 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (32.8 µL, 250 µmol) and PyBop (44.9 mg, 87.4 µmol) were added. The reaction was stirred at 0 °C for 0.5 h, and then allowed to warm to ambient temperature to stir for an additional 2.5 h. The reaction mixture was poured into cold 10% ag citric acid and extracted with EtOAc. The combined organic layers were washed with satd ag NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 22.9 m) afforded the 24-membered macrocycle (4.3 mg, 5.9%) as a light yellow oil. $\left[\alpha\right]_{D}^{24}$ - 3.3 (c 0.19, CHCl₃); R_f = 0.55 (60% EtOAc in hexanes); IR (film) 3284, 2929, 2860, 1744, 1649, 1554, 1210, 1066 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₆H₈₅N₈O₈ [M+H]⁺ 877.6490, found 877.6489. MPT-5-149



(35,6R,9S,12R,15S,18R,21S,24R)-1,4,6,7,12,13,16,18,19,24-decamethyl-3,9,15,21-tetrapentyl-1,4,7,10,13,16,19,22octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (5.4). A round-bottom flask was charged with the depsipeptide (66.9 mg, 61.7 µmol), dissolved in EtOAc (617 µL), and treated with 10% Pd/C (13.4 mg). The reaction flask was evacuated with light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCI/EtOAc (1.00 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. Dry DMF (12.4 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (24.3 μL, 185 μmol) and PyBop (33.4 mg, 64.5 μmol) were added. The reaction was stirred at 0 °C for 0.5 h, then allowed to warm to ambient temperature and stir for an additional 2.5 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 23.1 m) afforded the 24-membered macrocycle (3.9 mg, 7.2%) as a light yellow oil. $\left[\alpha\right]_{D}^{24}$ - 4.2 (c 0.11, CHCl₃); R_f = 0.37 (60% EtOAc in hexanes); IR (film) 3419, 2923, 2852, 1747, 1653, 1541, 1260, 1100 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃); HRMS (EI): Exact mass calcd for C₄₆H₈₅N₈O₈ [M+H]⁺ 877.6490, found 877.6486. MPT-5-212



(35,6R,95,12R,155,18R,215,24R)-1,4,6,7,12,13,18,19,24-nonamethyl-3,9,15,21-tetrapentyl-1,4,7,10,13,16,19,22octaazacyclotetracosan-2,5,8,11,14,17,20,23-octaone (5.5). A round-bottom flask was charged with the depsipeptide (98 mg, 91.5 μmol), dissolved in EtOAc (915 μL), and treated with 10% Pd/C (19.6 mg). The reaction flask was evacuated with

light vacuum. Hydrogen (balloon) was added, and the flask was cycled once more. The reaction was allowed to stir for 1 h. After purging the flask with argon, the crude reaction mixture was filtered through Celite. To the crude material was added 4M HCl/EtOAc (1.0 mL) and the reaction mixture was allowed to stir for 30 min. The reaction mixture was concentrated and added to a flame-dried round bottom flask. Dry DMF (21.4 mL) was added and the reaction was cooled to 0 °C. Once at 0 °C, DIPEA (36.1 µL, 275 µmol) and PyBop (50.0 mg, 96.1 µmol) were added. The reaction was stirred at 0 °C for 0.5 h, and then allowed to warm to ambient temperature to stir for an additional 2.5 h. The reaction mixture was poured into cold 10% aq citric acid and extracted with EtOAc. The combined organic layers were washed with satd aq NaHCO₃, brine, and then dried and concentrated. Preparative HPLC purification (5 – 95% aqueous acetonitrile, 210 nm, flow rate: 18 mL/min, R_t = 23.9 m) afforded the 24-membered macrocycle (6.1 mg, 9.0%) as a light yellow oil. $[\alpha]_D^{24} - 2.1$ (*c* 0.12, CHCl₃); R_f = 0.43 (60% EtOAc in hexanes); IR (film) 3292, 2929, 2843, 1654, 1637, 1560, 1458 cm⁻¹; ¹H NMR (600 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹H NMR spectrum; ¹³C NMR (150 MHz, CDCl₃) This compound exists in multiple conformations, causing significant peak overlap. Refer to image of the ¹³C NMR spectrum; HRMS (EI): Exact mass calcd for C₄₅H₈₃N₈O₈ [M+H]⁺ 863.6328, found 863.6325. MPT-4-227

Characterization NMR Spectra

Figure S1. 1 H NMR (600 MHz, CDCl₃) of 14

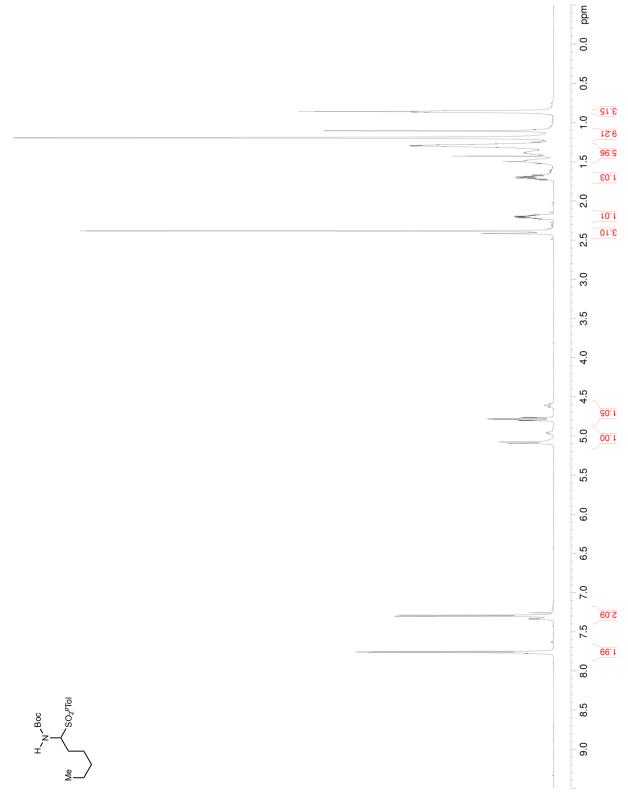


Figure S2. 13 C NMR (150 MHz, CDCl₃) of 14

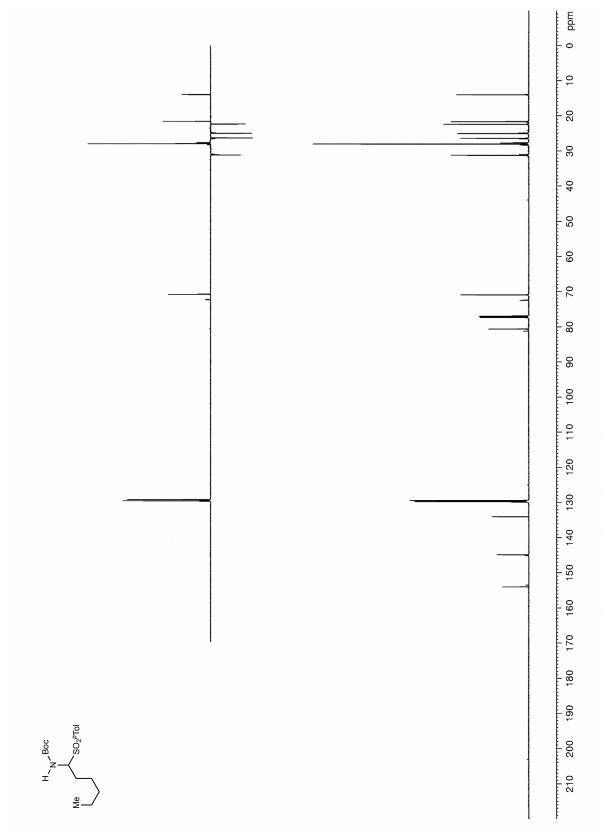


Figure S3. 1 H NMR (600 MHz, CDCl₃) of **16**

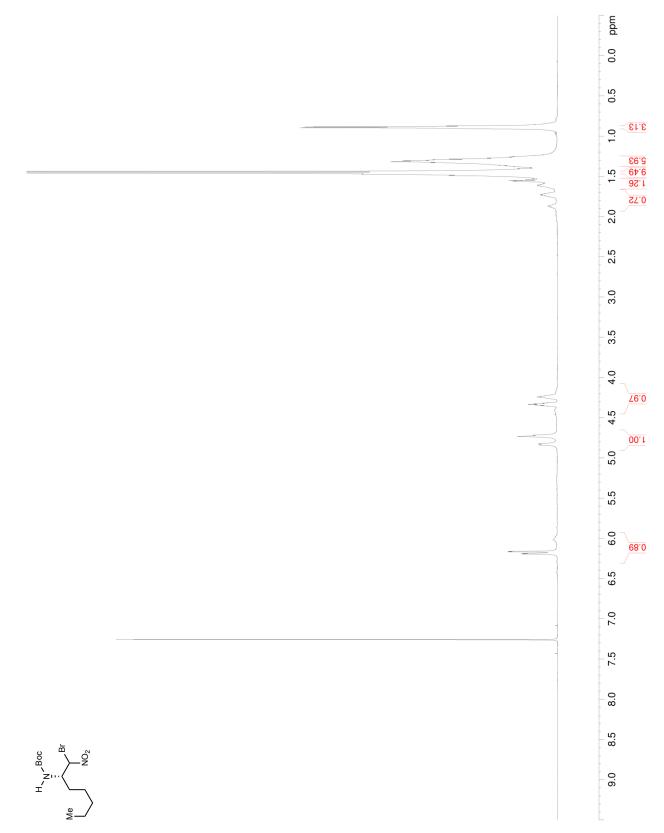


Figure S4. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 16

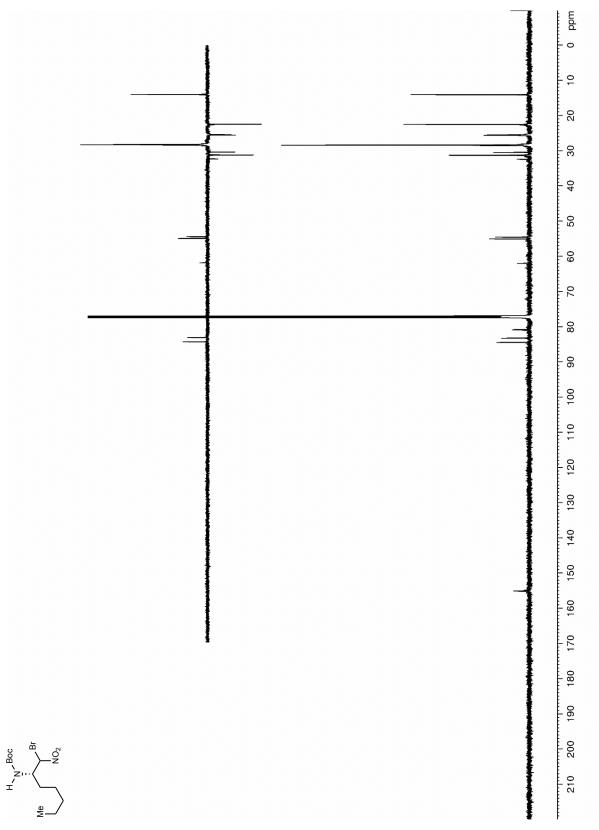


Figure S5. ¹H NMR (600 MHz, CDCl₃) of **19**

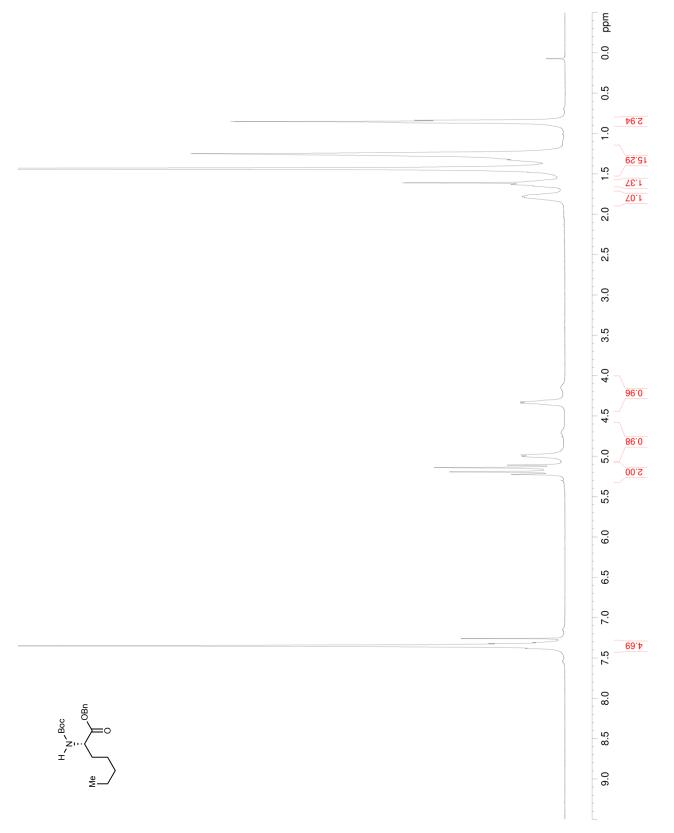


Figure S6. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 19

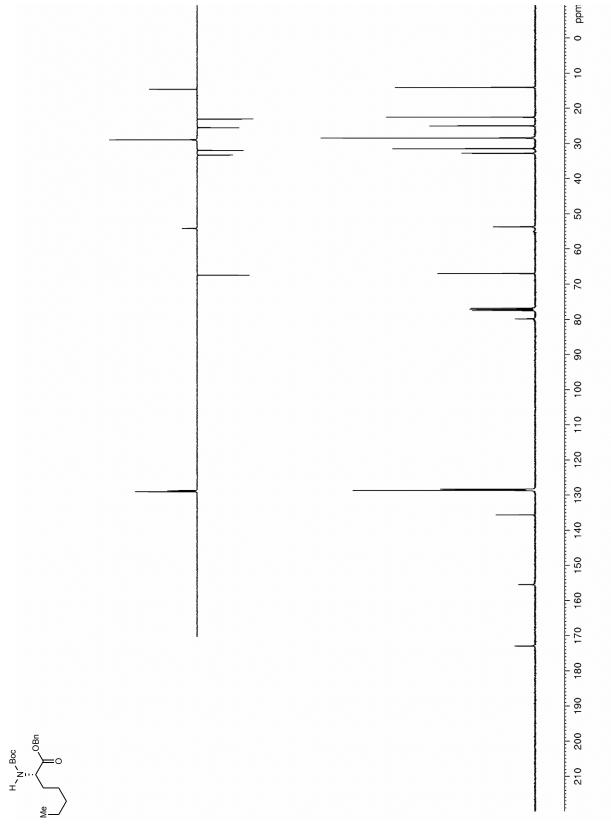


Figure S7. ¹H NMR (600 MHz, CDCl₃) of **21**

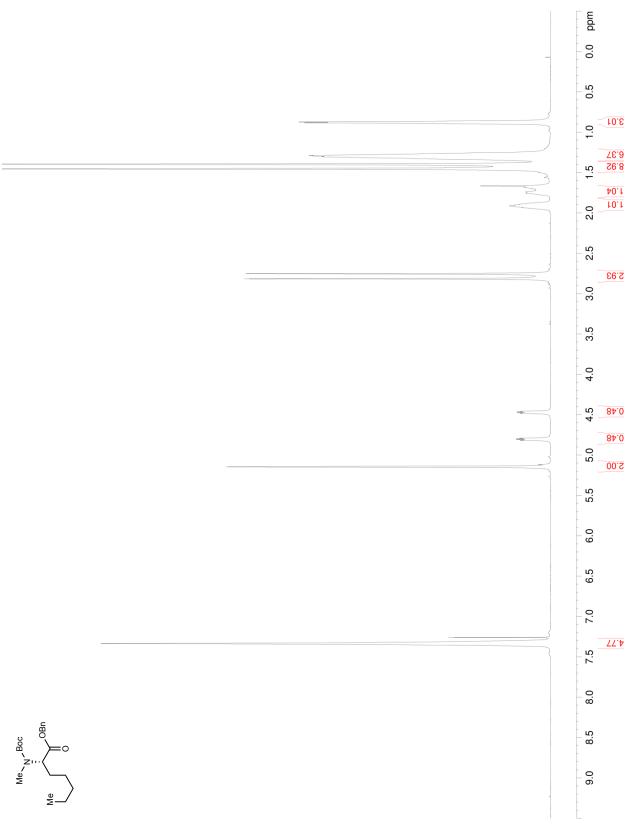


Figure S8. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 21

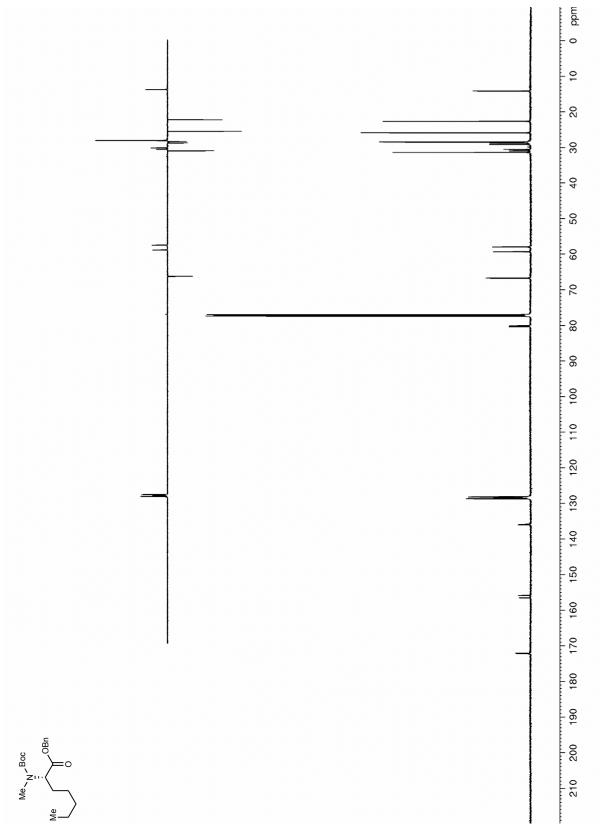


Figure S9. ¹H NMR (600 MHz, CDCl₃) of 23

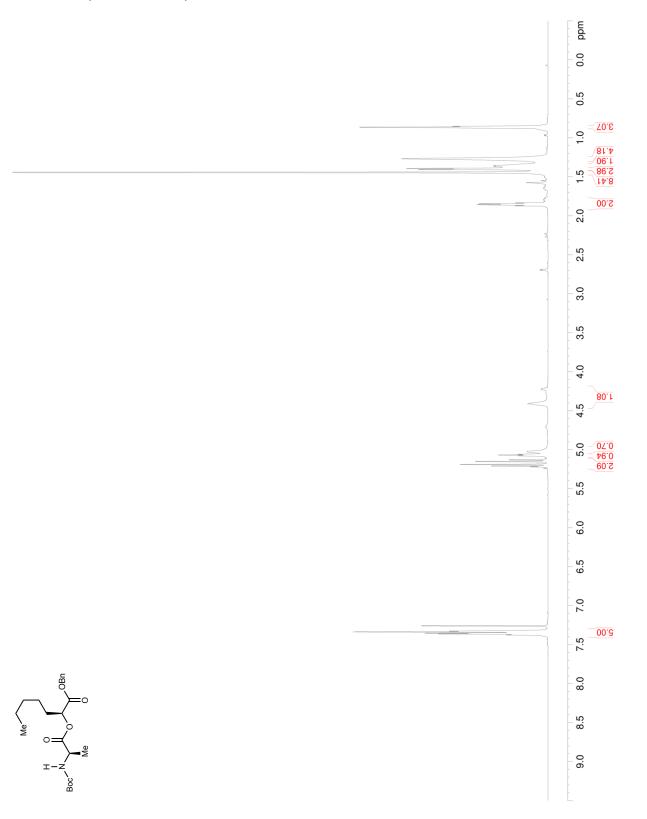
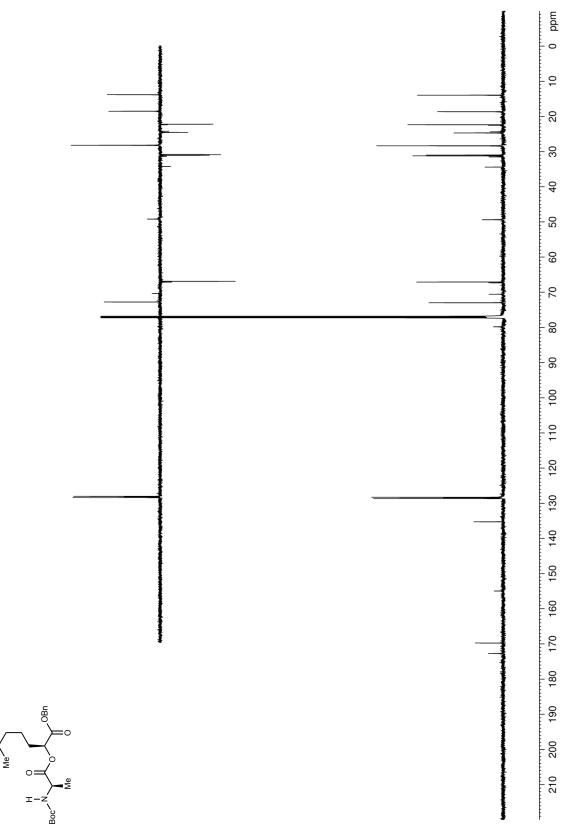
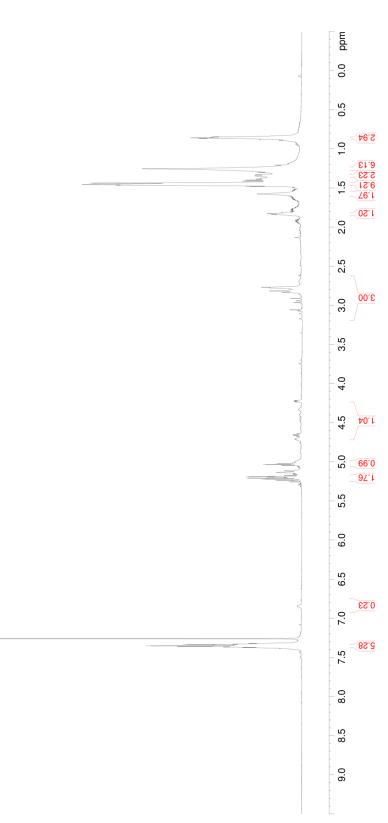


Figure S10. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 23





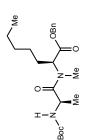
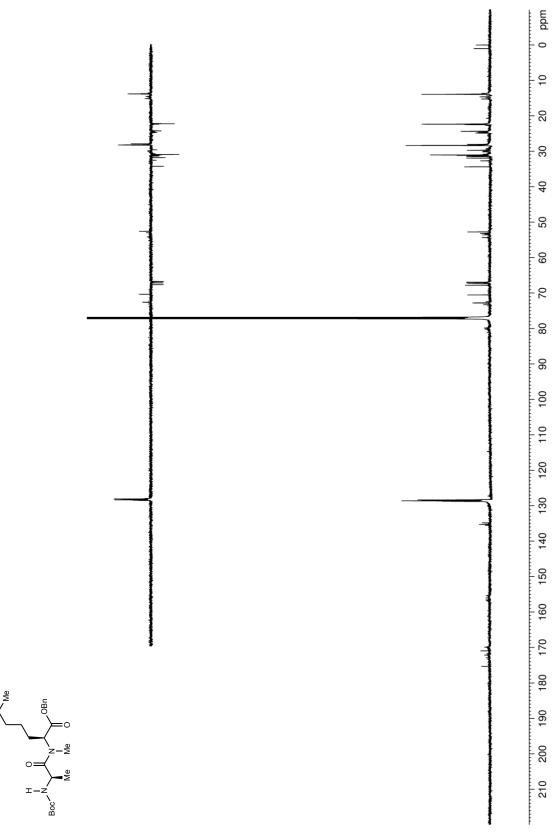


Figure S12. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 29



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Figure S13. ¹H NMR (600 MHz, CDCl₃) of 32



Me

Figure S14. ¹³C NMR (150 MHz, CDCl₃) of 32

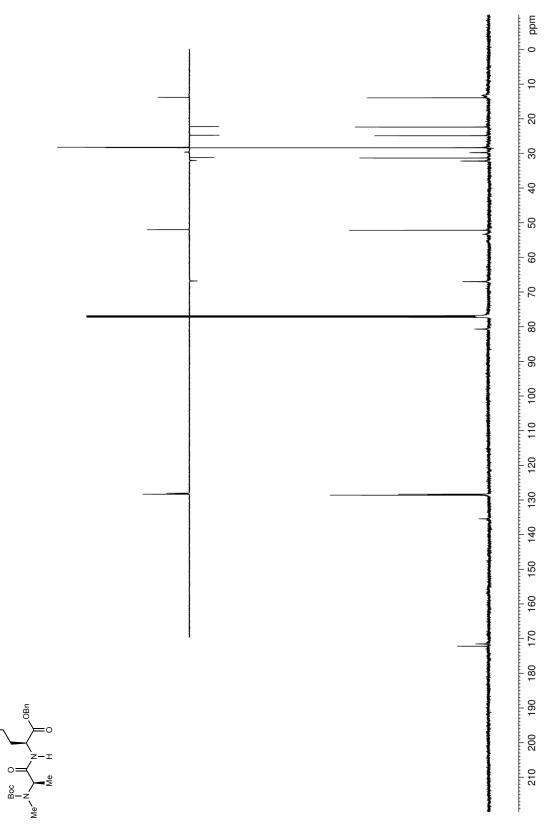


Figure S15. 1 H NMR (600 MHz, CDCl₃) of 35

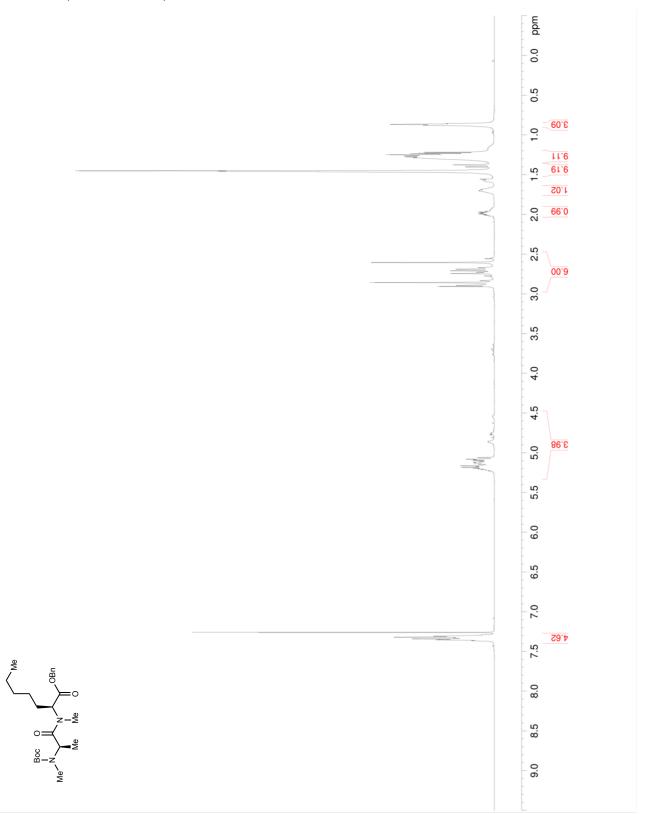


Figure S16. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 35

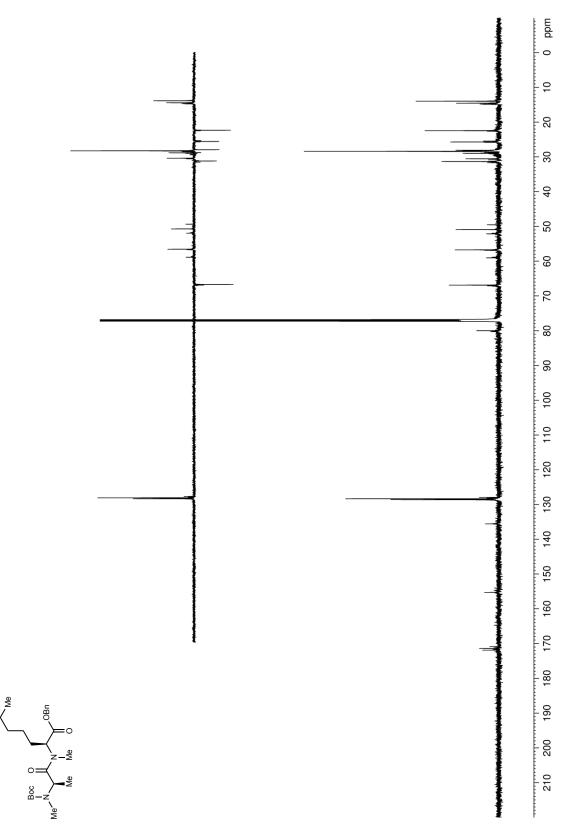
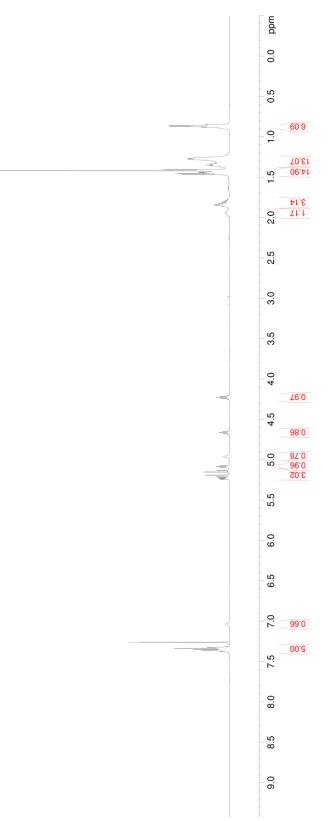


Figure S17. ¹H NMR (600 MHz, CDCl₃) of T38



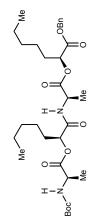
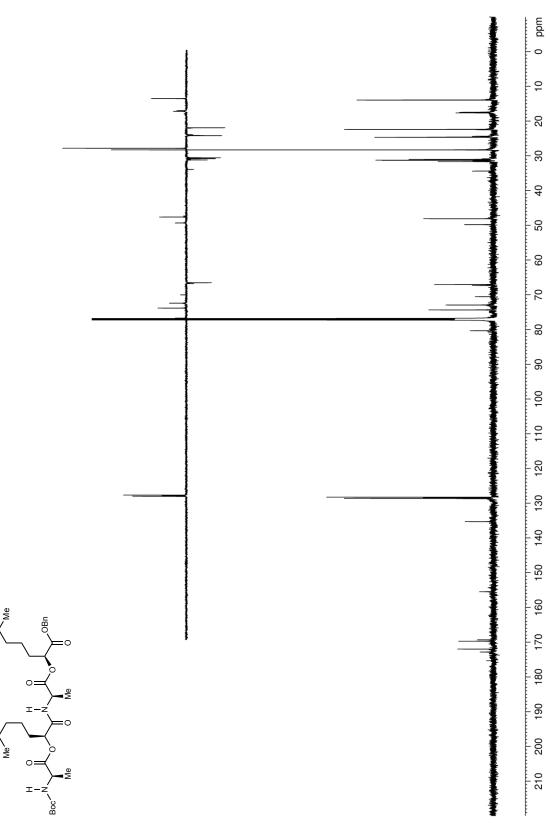


Figure S18. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of T38



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Figure S19. ¹H NMR (600 MHz, CDCl₃) of T40

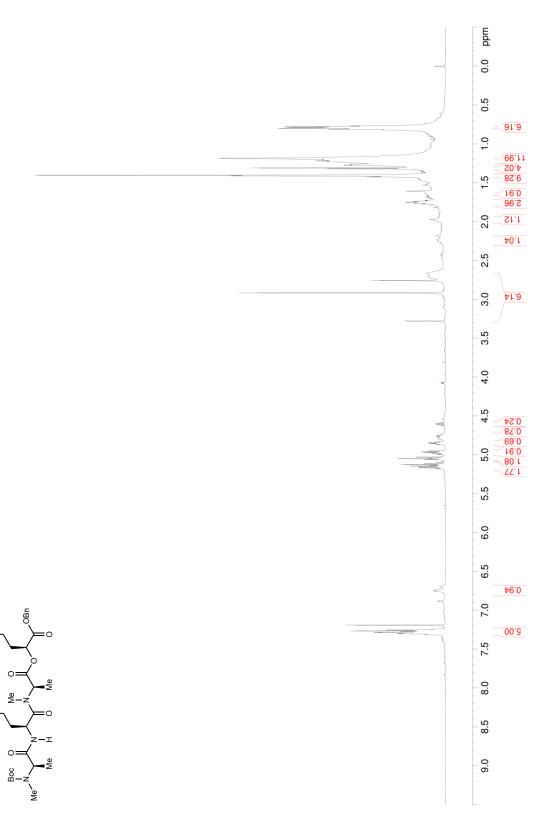
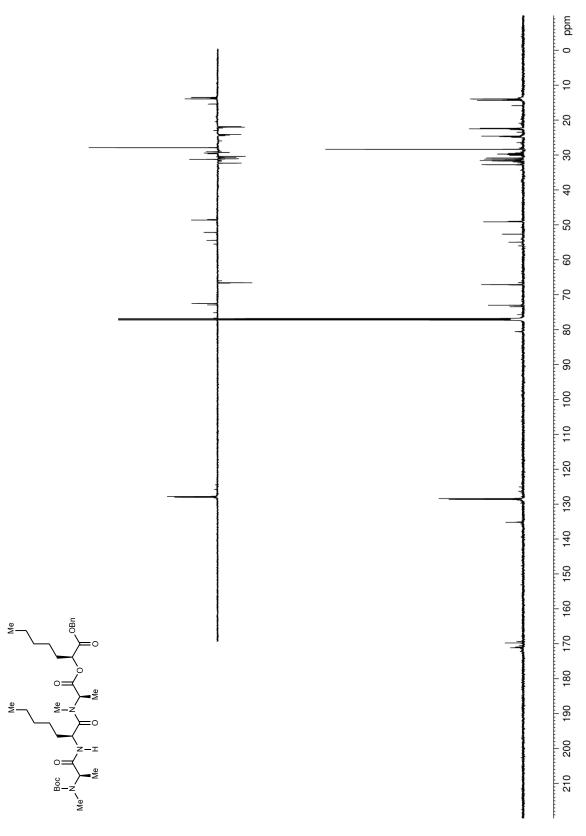


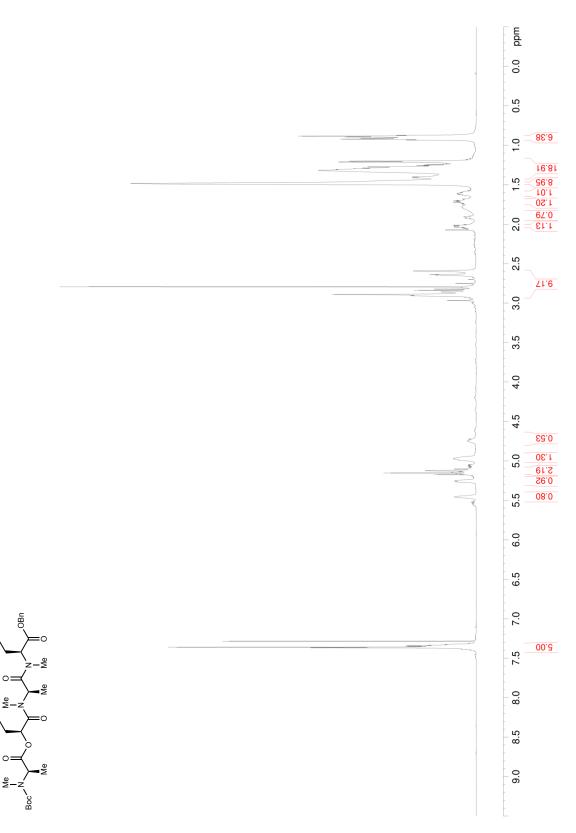
Figure S20. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of T40



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Figure S21. ¹H NMR (600 MHz, CDCl₃) of T41



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Figure S22. 13 C NMR (150 MHz, CDCl₃) of T41

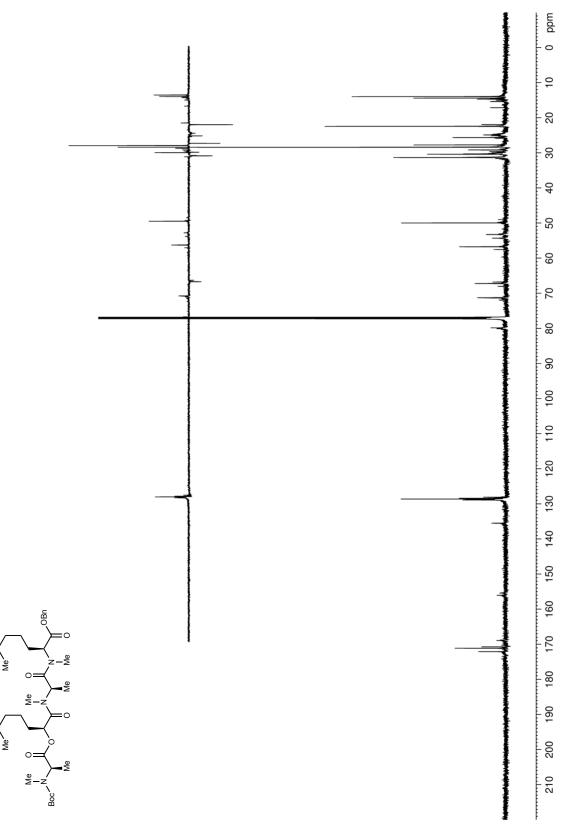
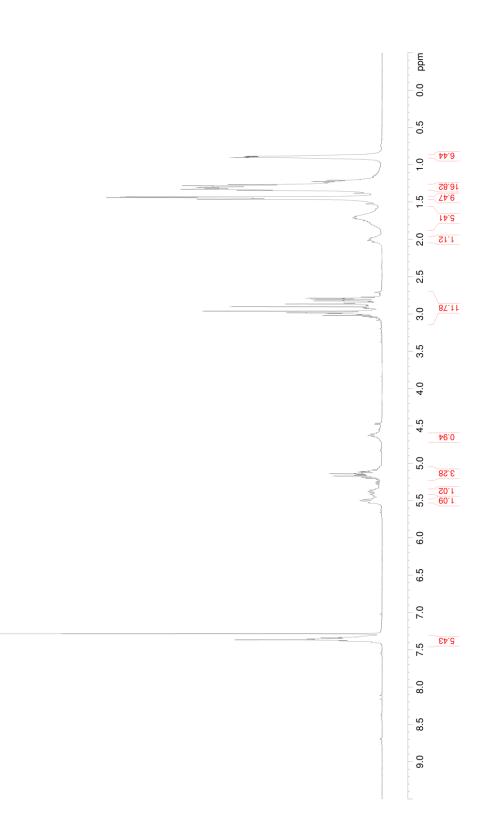


Figure S23. ¹H NMR (600 MHz, CDCl₃) of T42



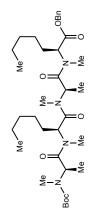


Figure S24. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of T42

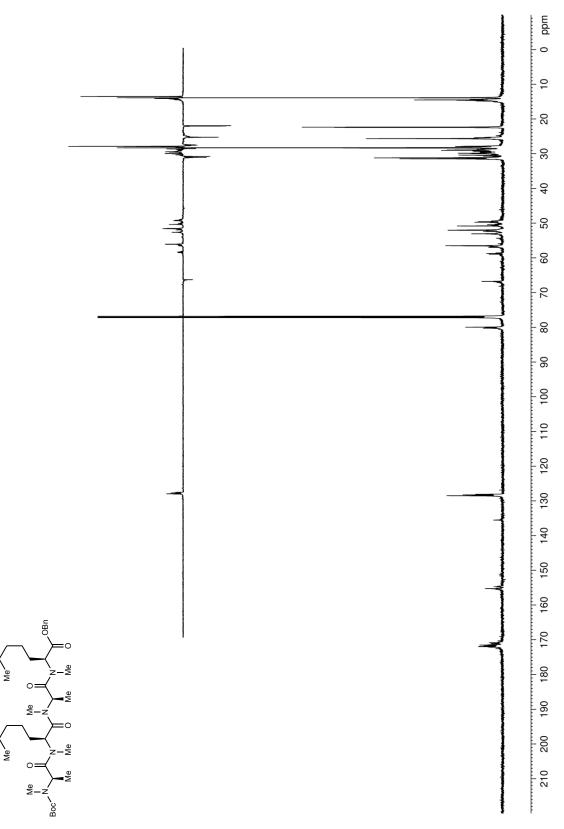
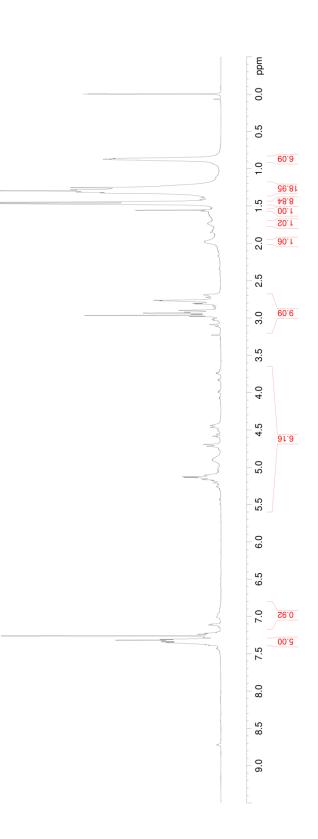


Figure S25. ¹H NMR (600 MHz, CDCl₃) of T43



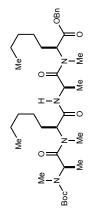
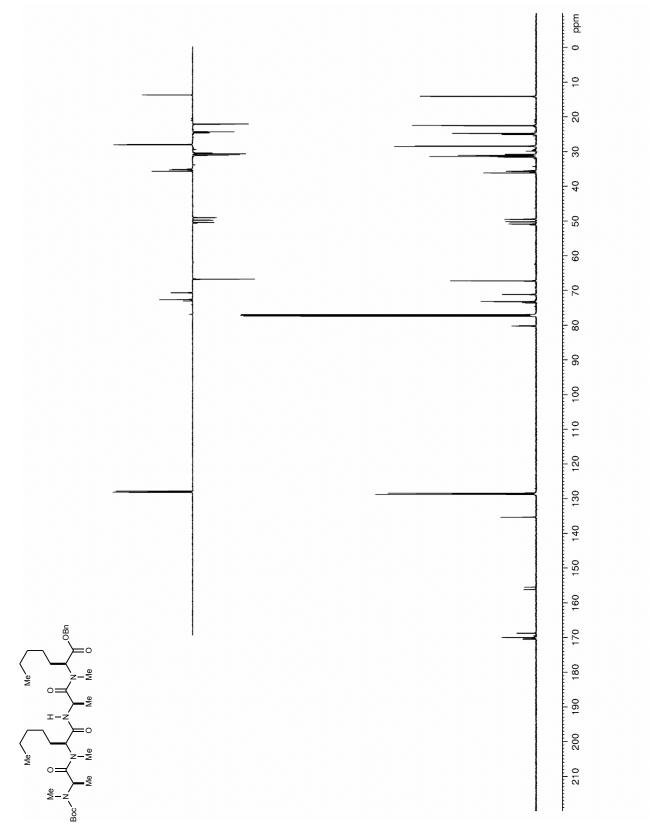


Figure S26. ¹³C NMR (150 MHz, CDCl₃) of T43



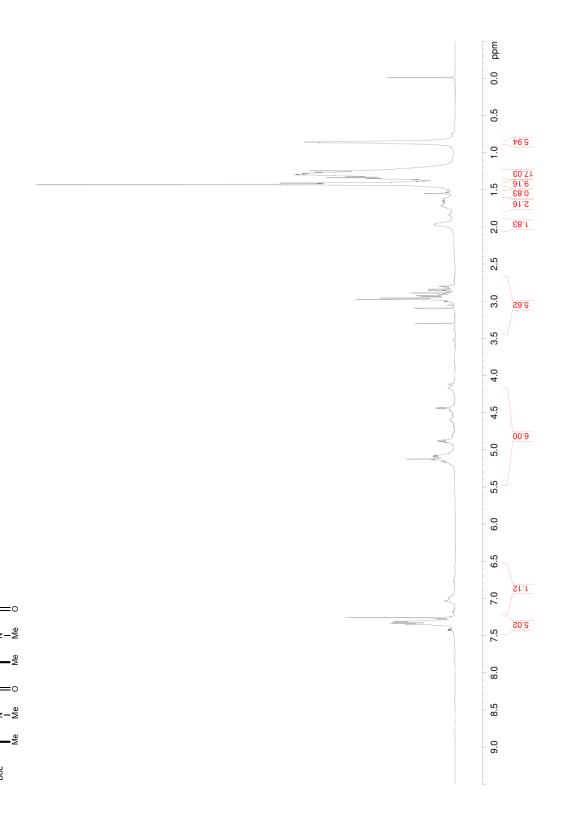
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Supporting Information

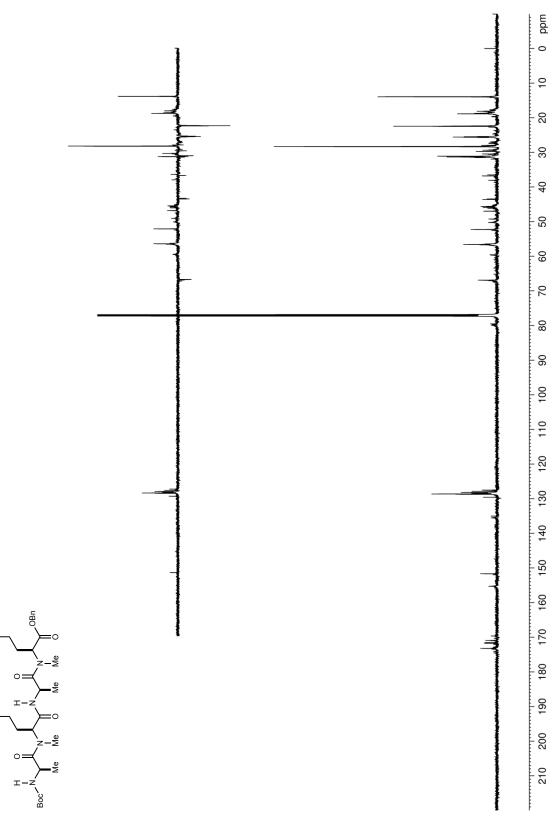
Figure S27. ¹H NMR (600 MHz, CDCl₃) of T44

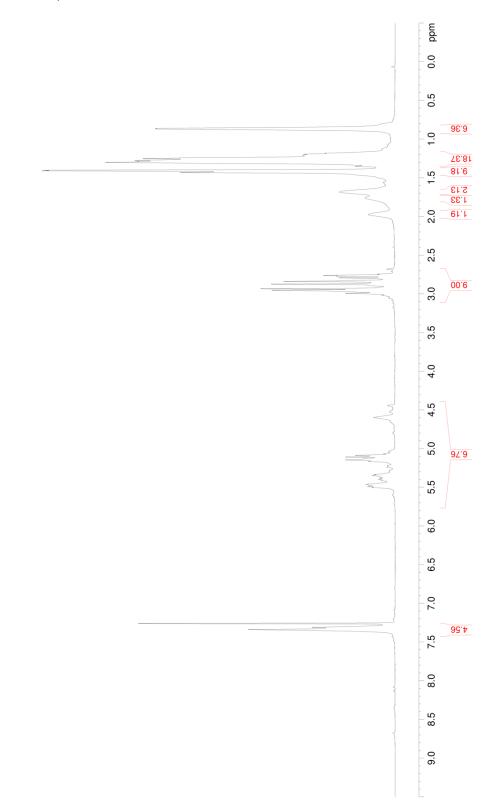


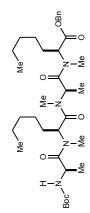
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Figure S28. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of T44



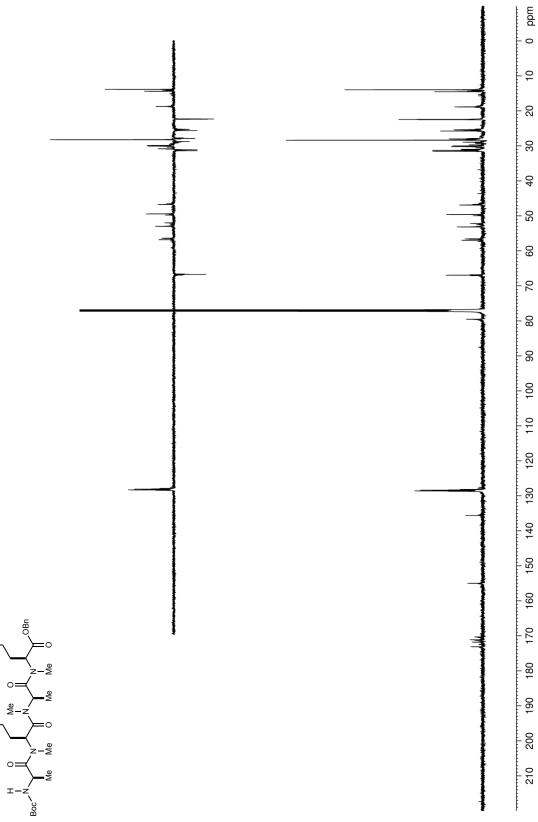




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Figure S30. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of T45



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Figure S31. ¹H NMR (600 MHz, CDCl₃) of T46

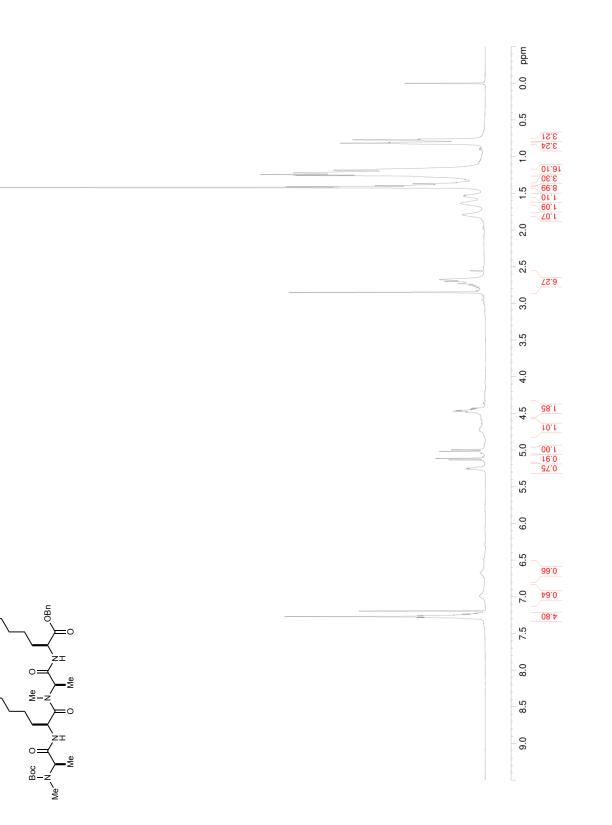
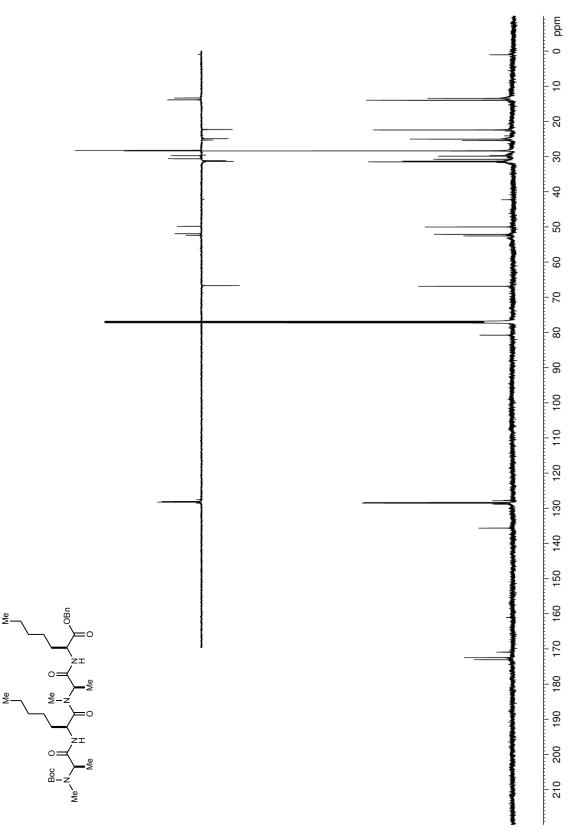


Figure S32. $^{\rm 13}{\rm C}$ NMR (150 MHz, CDCl₃) of T46



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Figure S33. ¹H NMR (600 MHz, CDCl₃) of T47

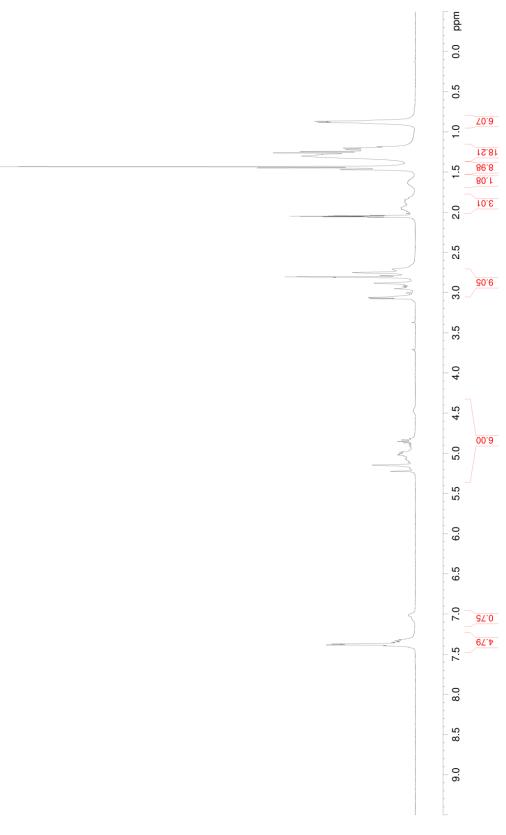


Figure S34. $^{\rm 13}{\rm C}$ NMR (150 MHz, CDCl₃) of T47

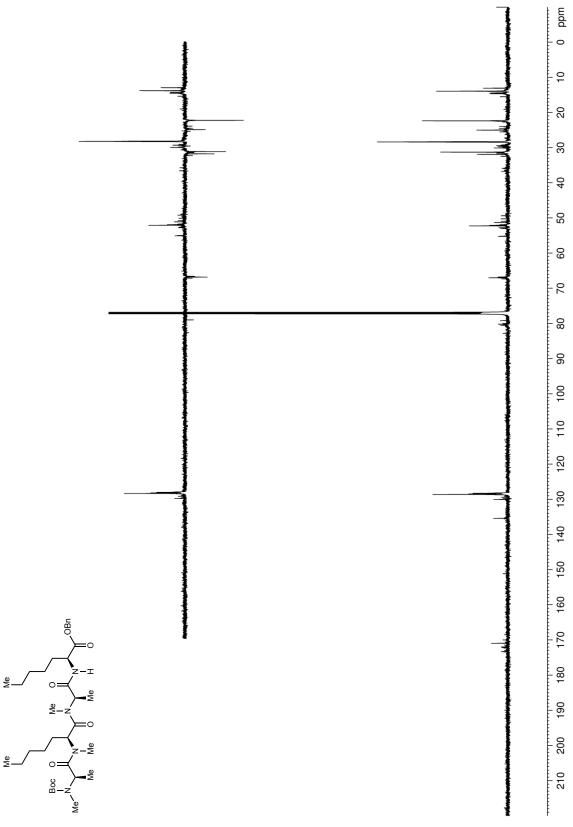


Figure S35. ¹H NMR (600 MHz, CDCl₃) of O49

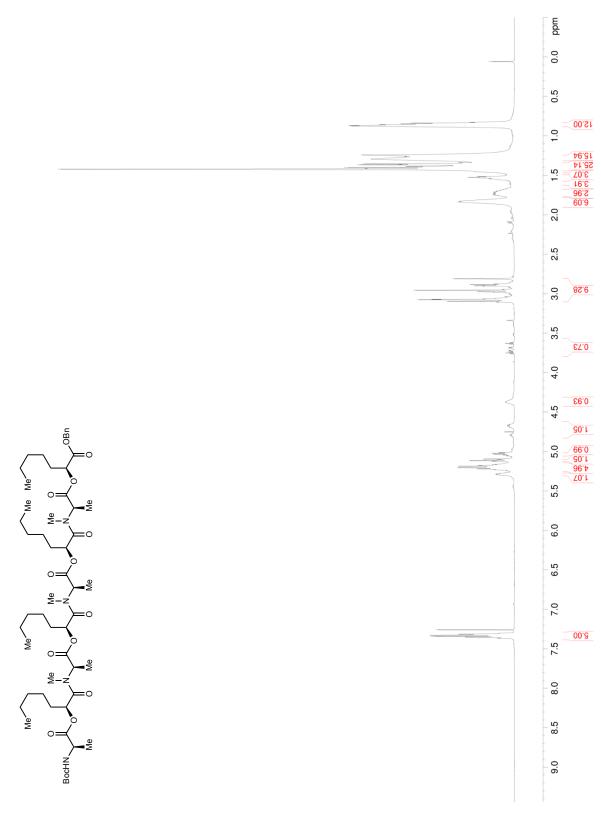
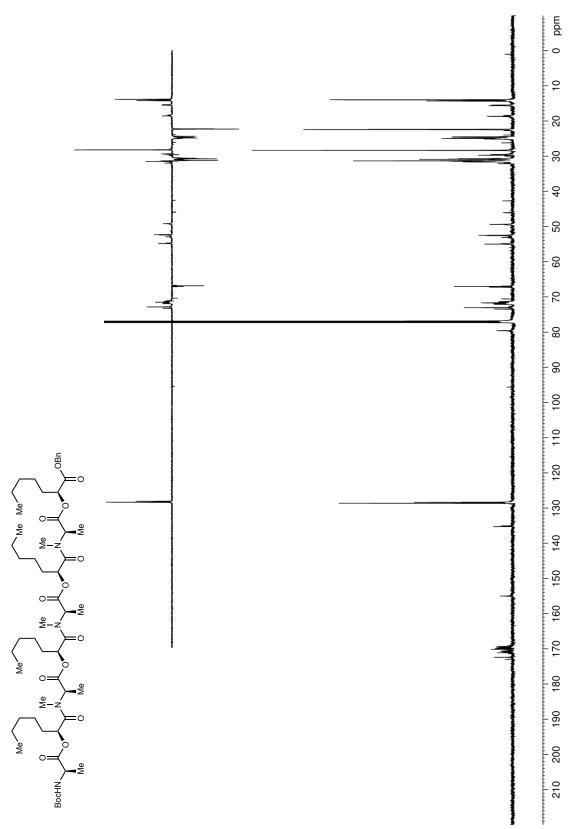
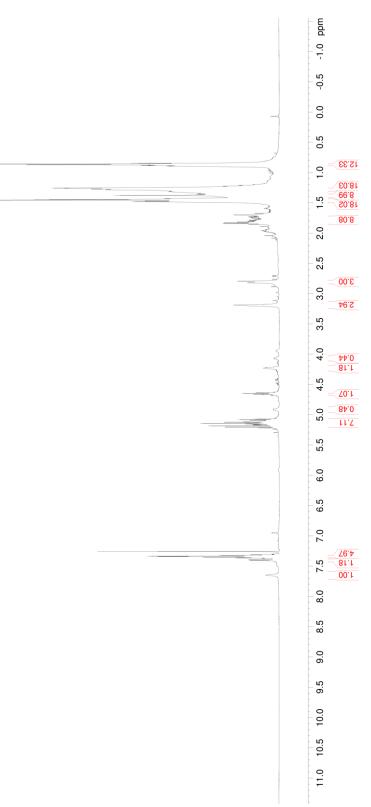
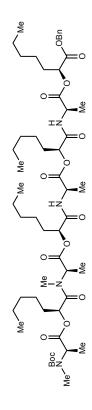


Figure S36. $^{\rm 13}{\rm C}$ NMR (150 MHz, CDCl₃) of O49

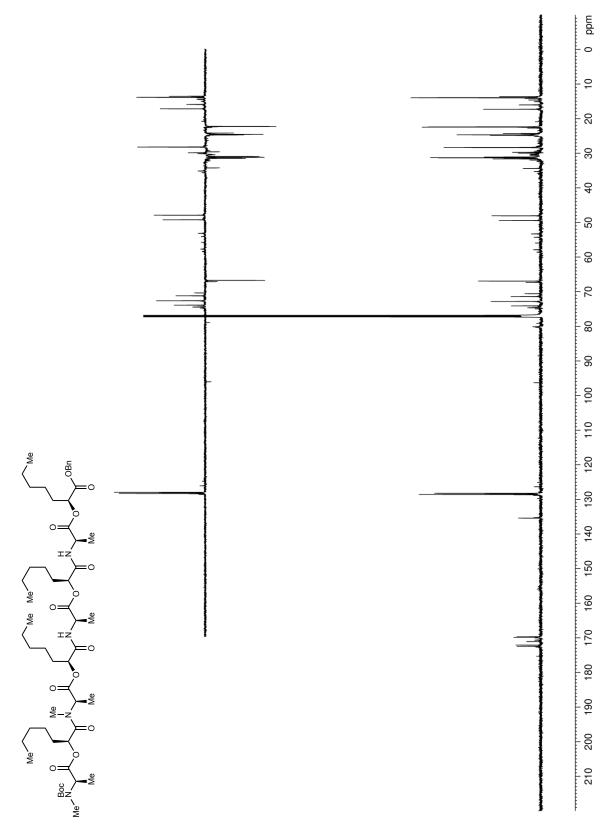






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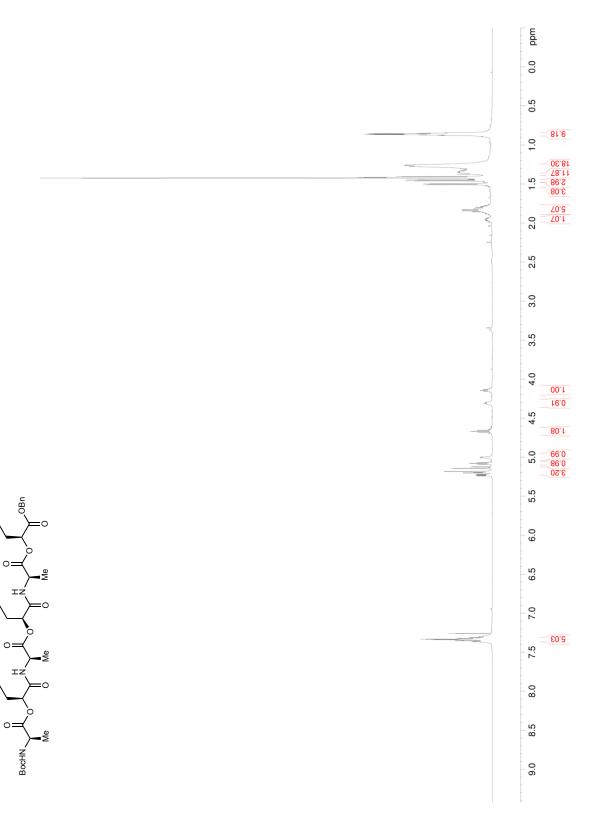
Figure S38. $^{\rm 13}\text{C}$ NMR (150 MHz, CDCl_3) of O50



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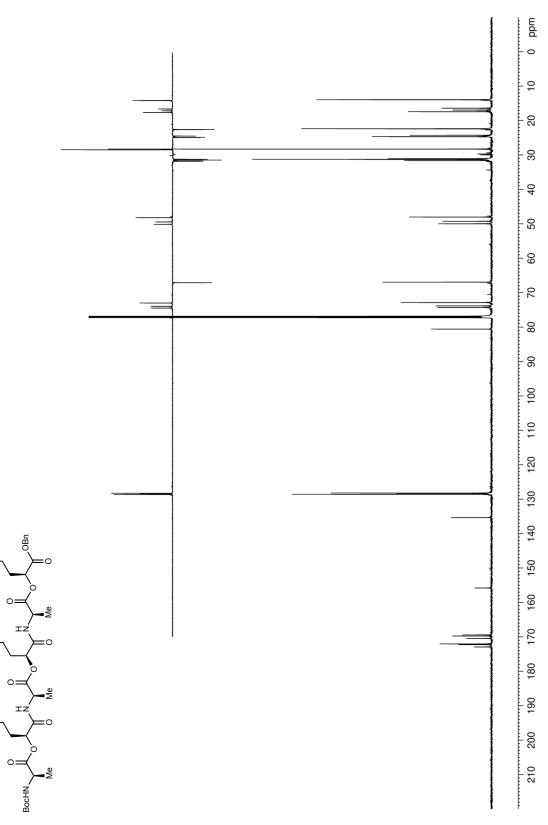
Figure S39. ¹H NMR (600 MHz, CDCl₃) of H51

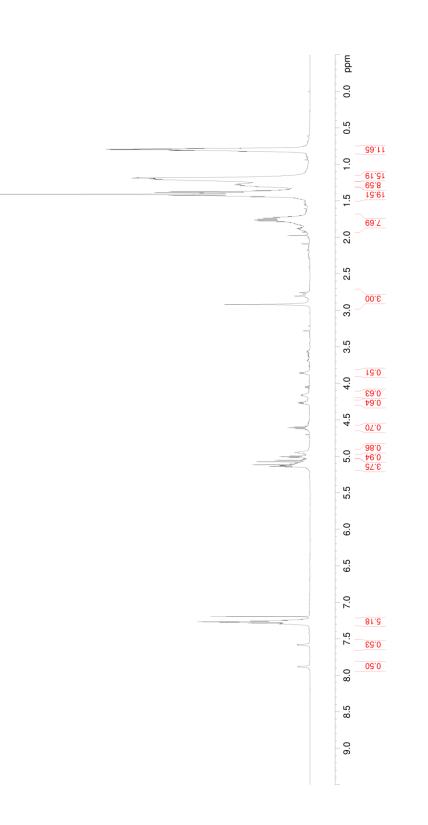


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Figure S40. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of H51





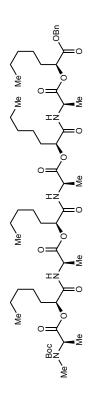


Figure S42. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of O52

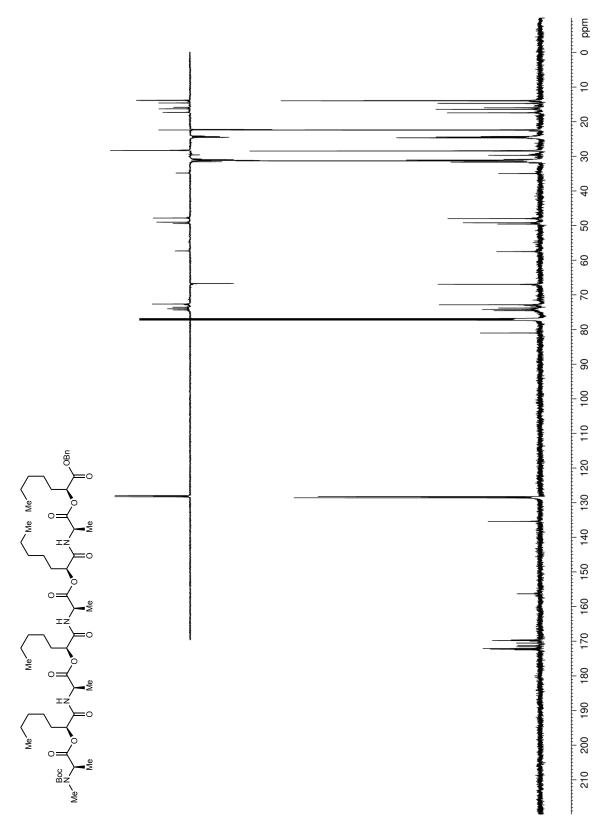
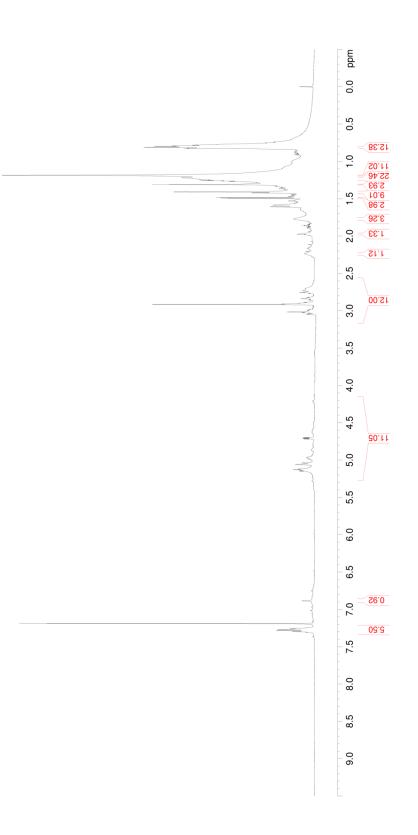


Figure S43. ¹H NMR (600 MHz, CDCl₃) of O54



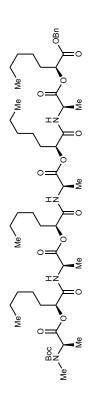
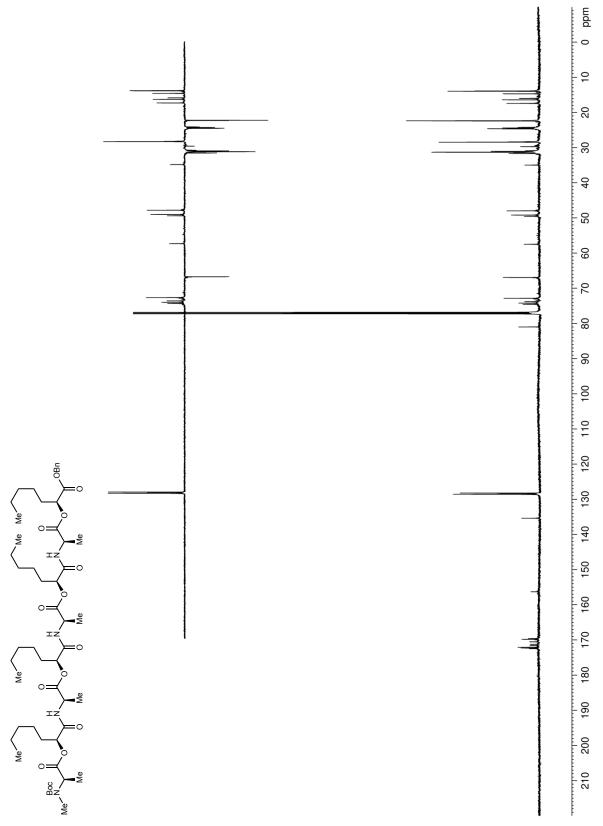
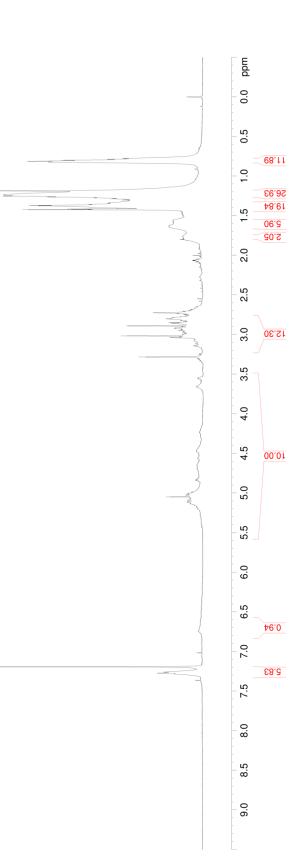


Figure S44. $^{\rm 13}{\rm C}$ NMR (150 MHz, CDCl₃) of O54





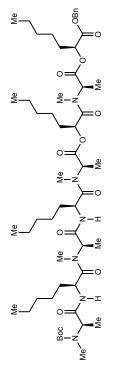


Figure S46. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 055

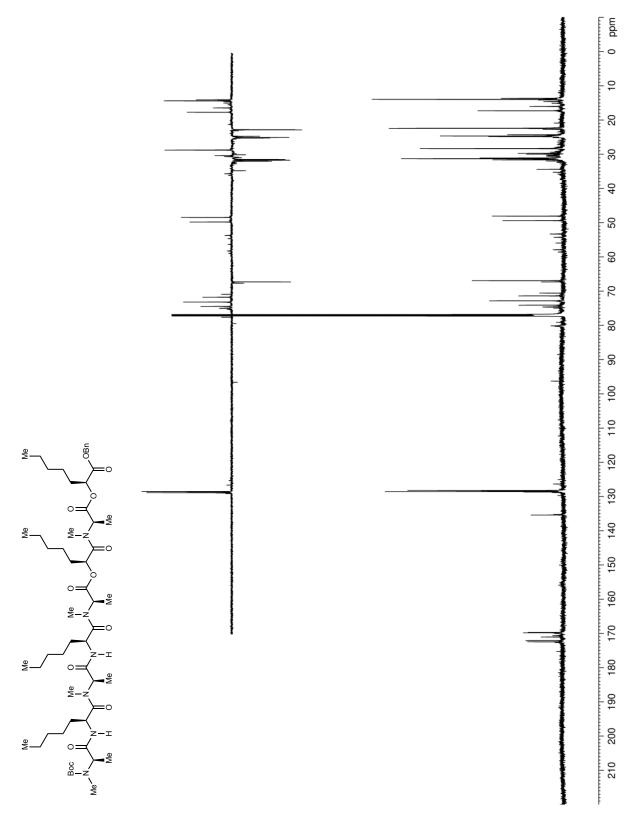


Figure S47. ¹H NMR (600 MHz, CDCl₃) of O56

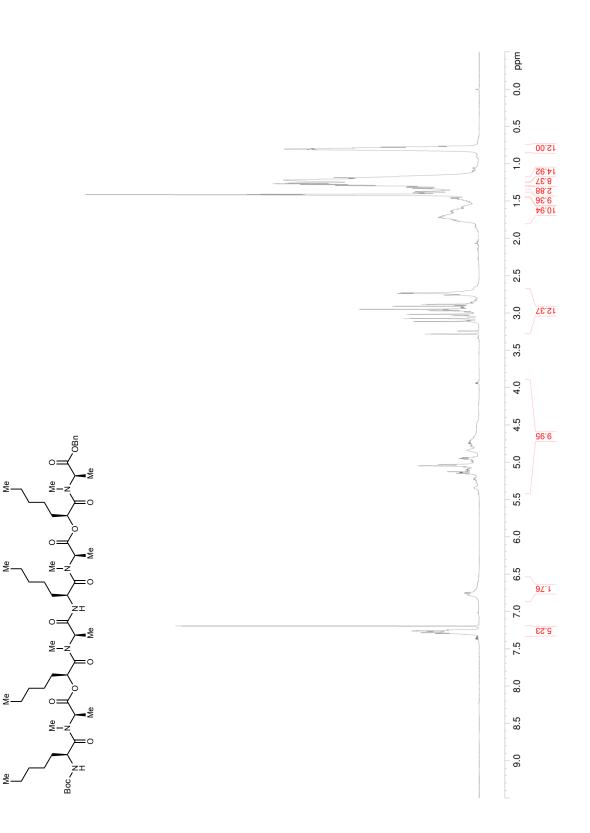


Figure S48. $^{\rm 13}\text{C}$ NMR (150 MHz, CDCl₃) of O56

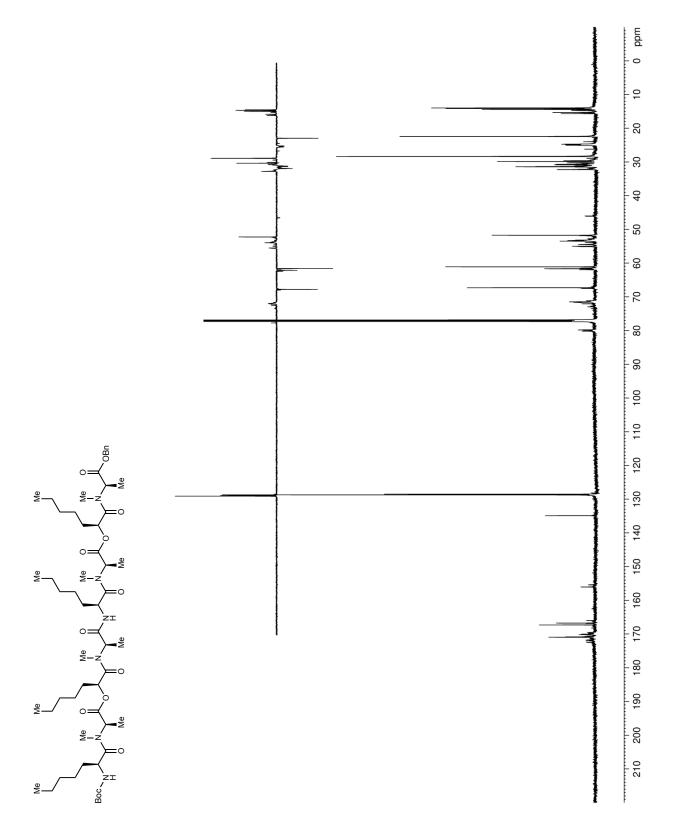
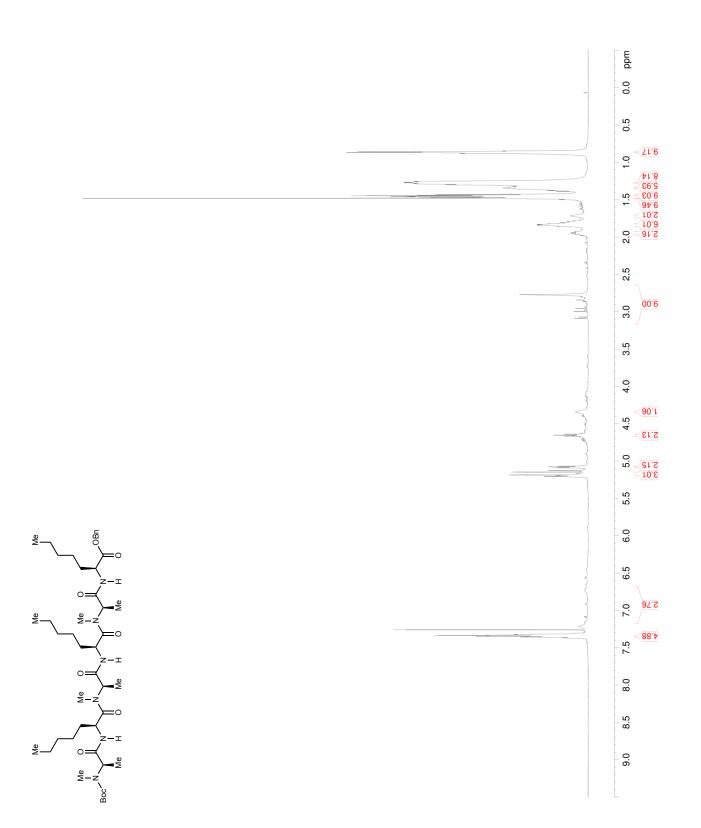


Figure S49. ¹H NMR (600 MHz, CDCl₃) of H57



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Figure S50. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of H57

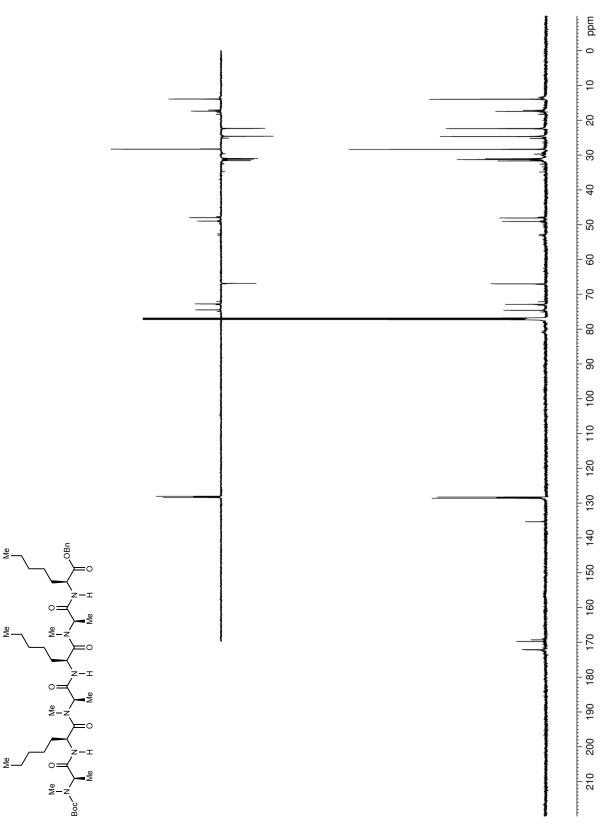


Figure S51. ¹H NMR (600 MHz, CDCl₃) of O58 (Boc deprotected)

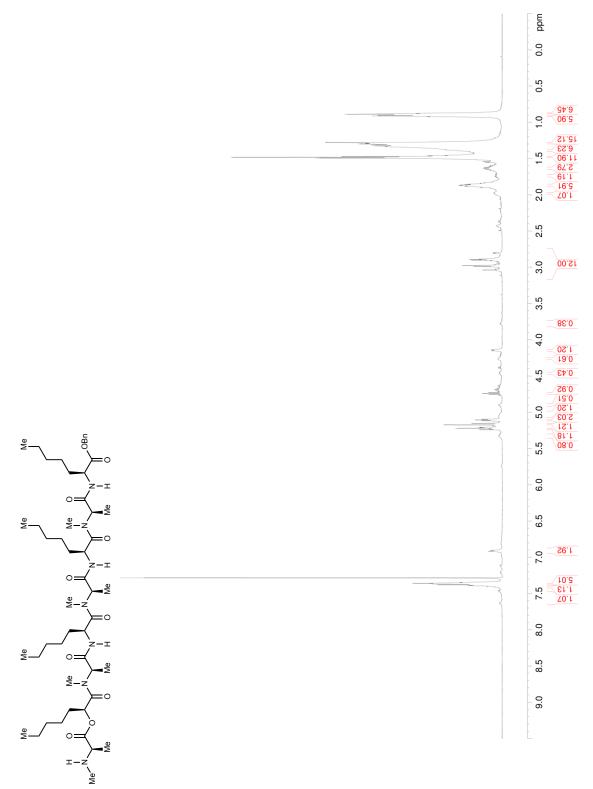


Figure S52. ¹³C NMR (150 MHz, CDCl₃) of O58 (Boc deprotected)

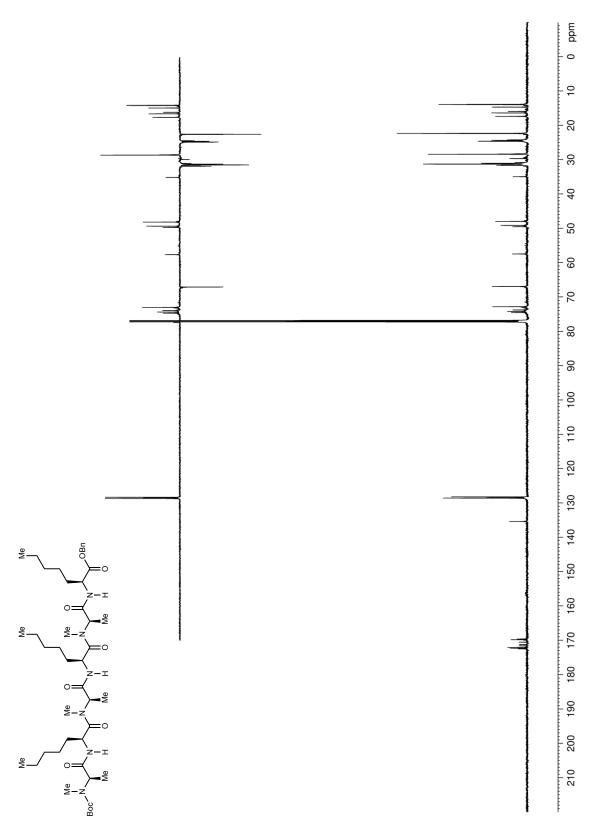


Figure S53. ¹H NMR (600 MHz, CDCl₃) of O59

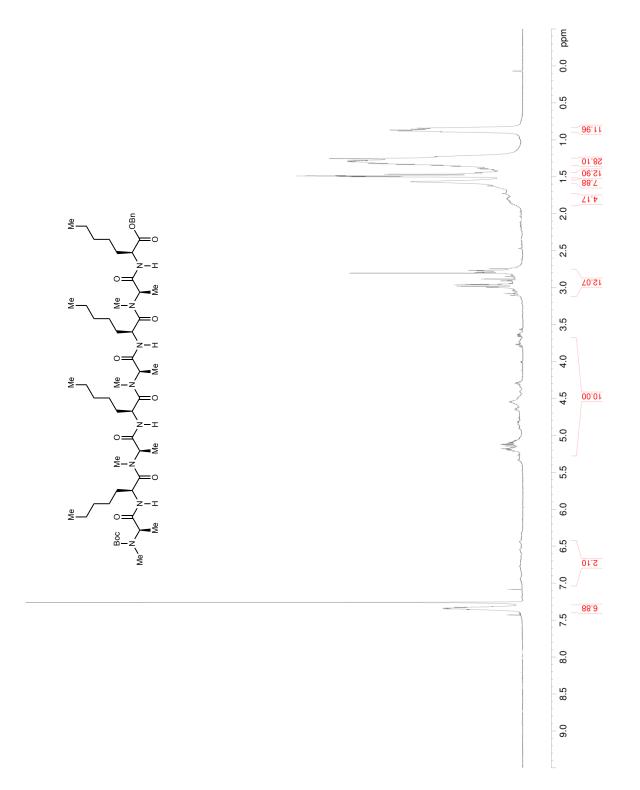


Figure S54. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of O59

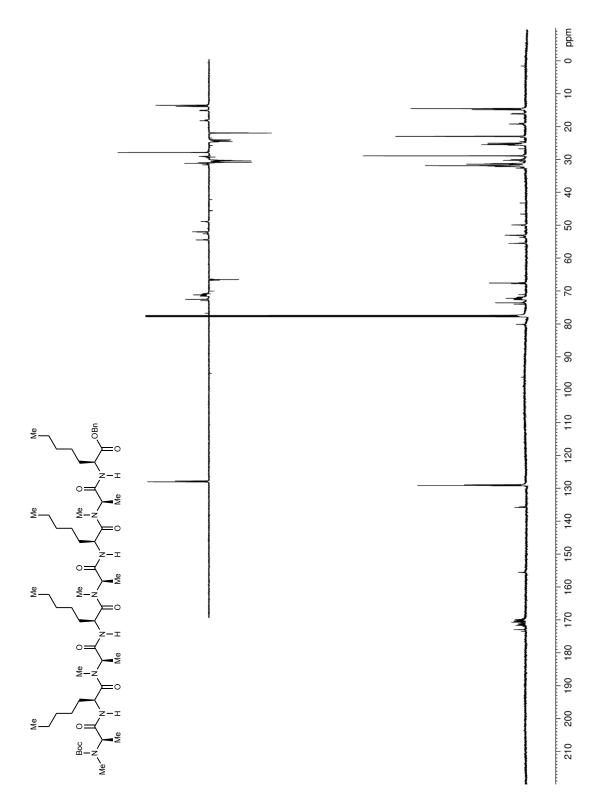
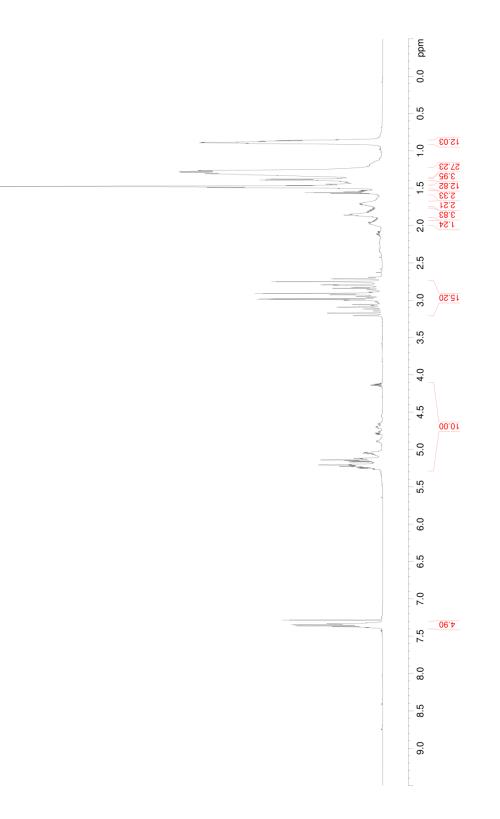


Figure S55. ¹H NMR (600 MHz, CDCl₃) of O60



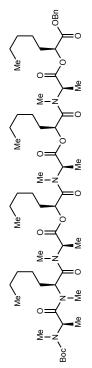


Figure S56. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of O60

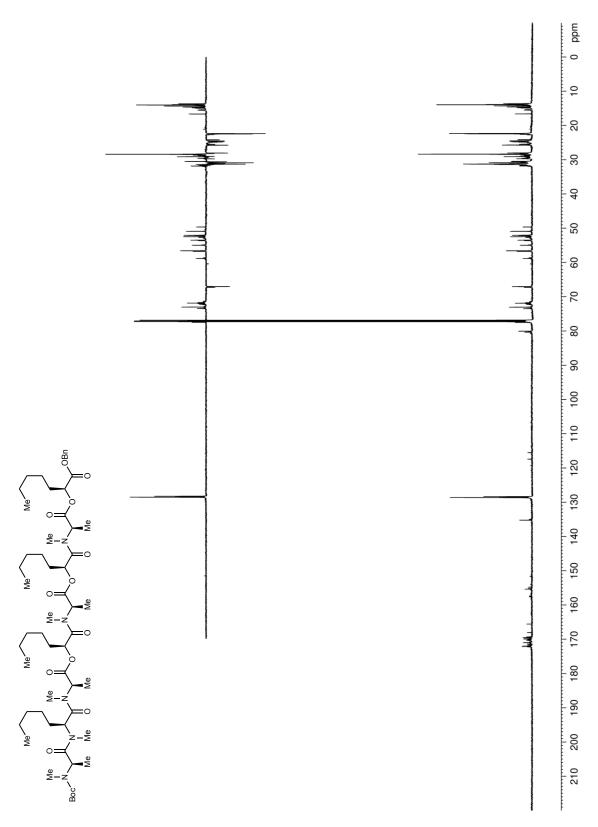


Figure S57. ¹H NMR (600 MHz, CDCl₃) of O61

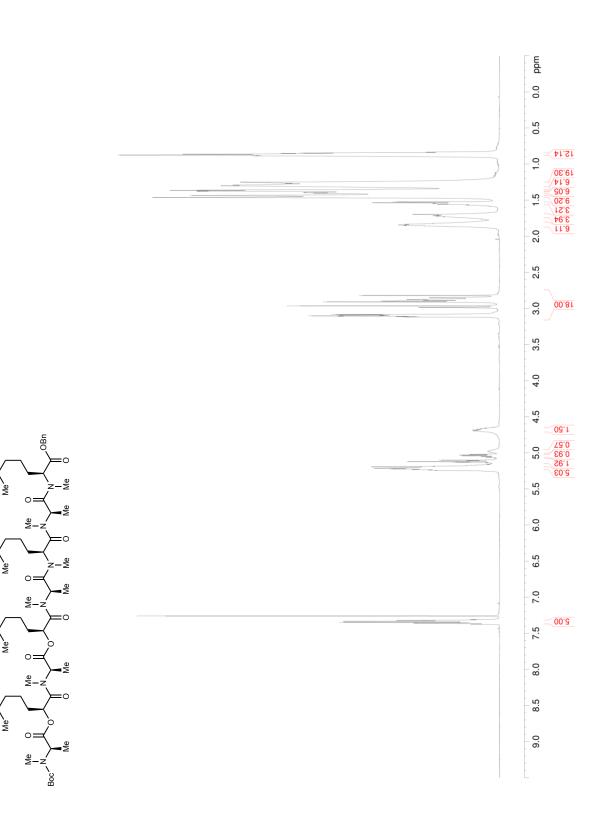


Figure S58. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 061

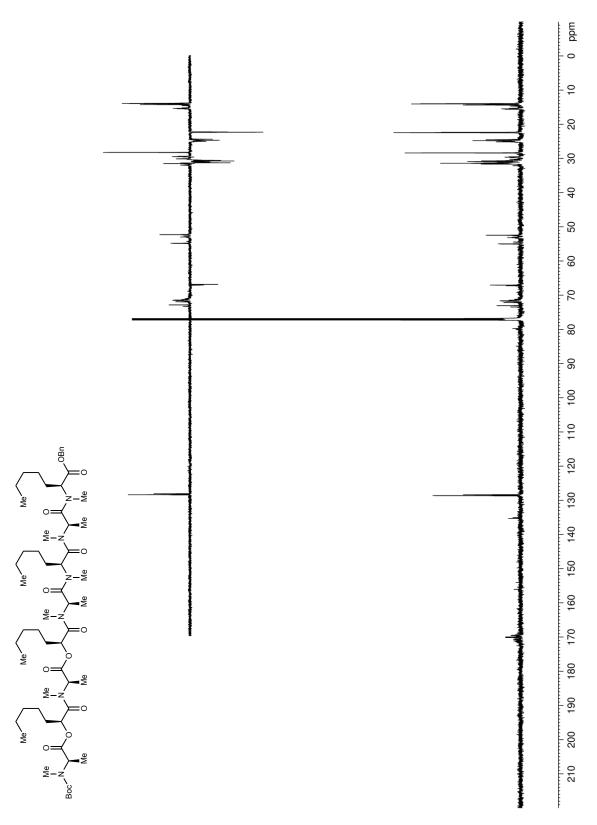
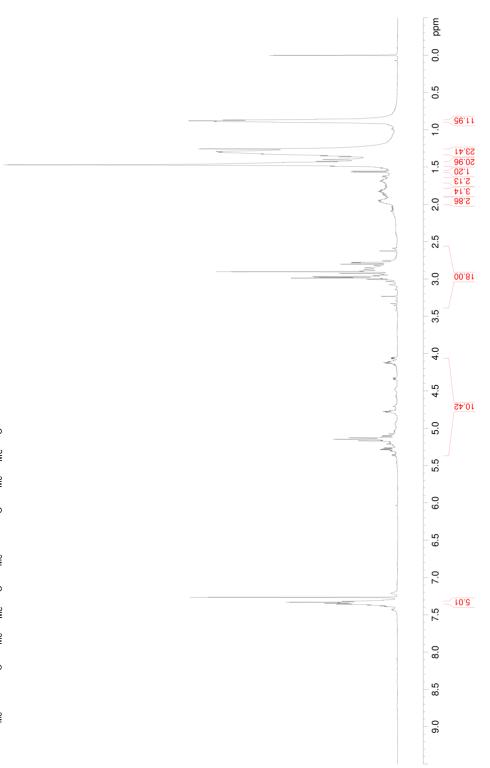


Figure S59. ¹H NMR (600 MHz, CDCl₃) of O62



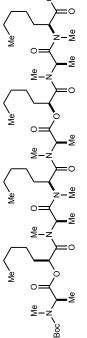


Figure S60. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of O62

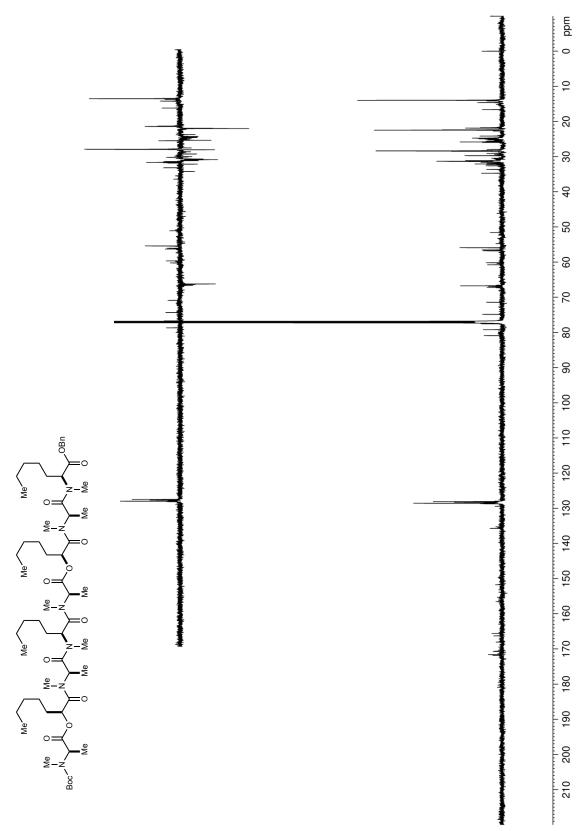
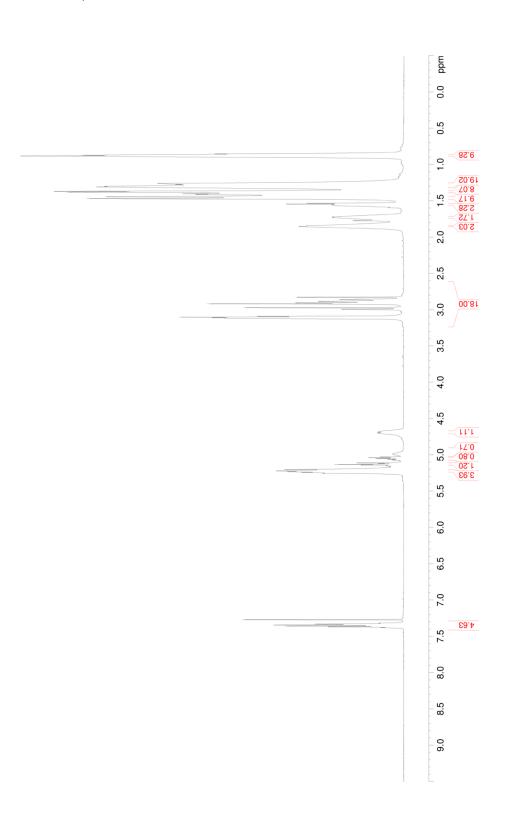
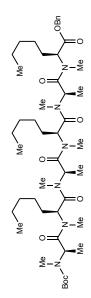


Figure S61. ¹H NMR (600 MHz, CDCl₃) of H63





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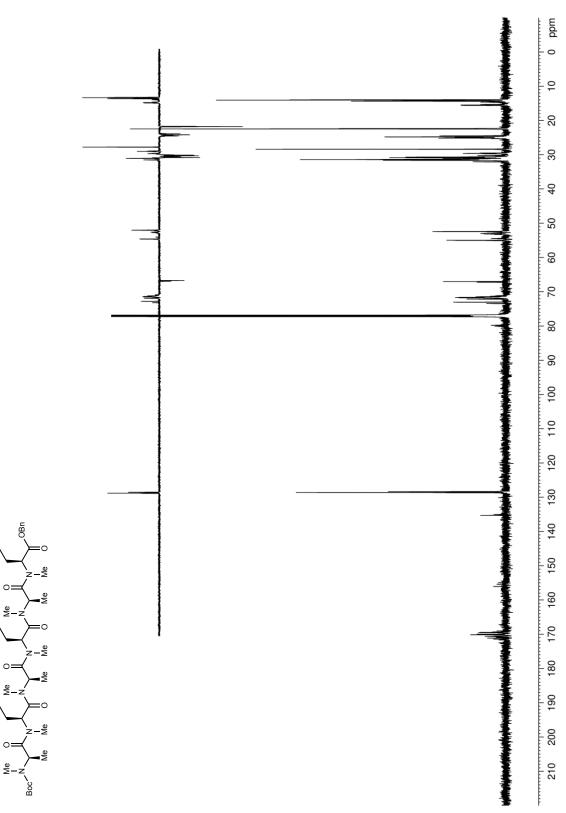


Figure S63. ¹H NMR (600 MHz, CDCl₃) of O64

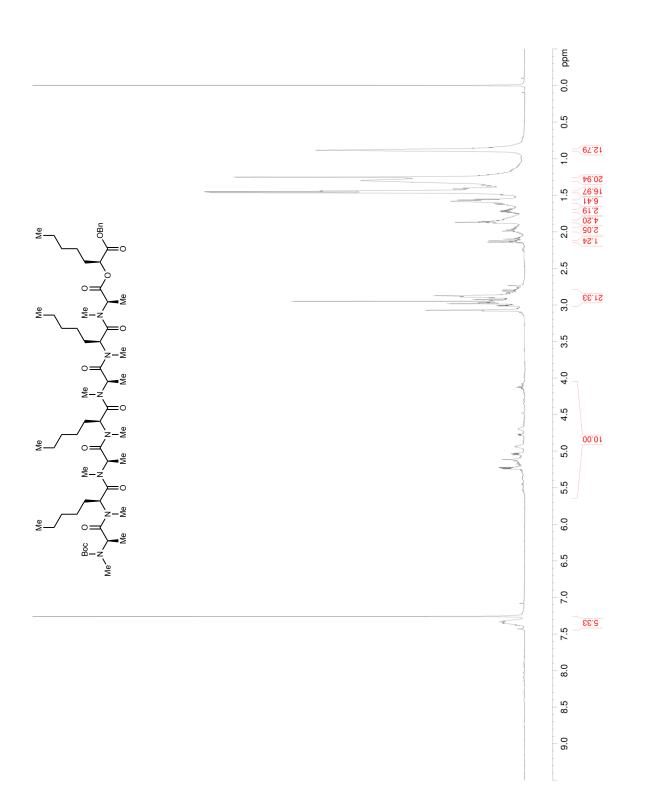


Figure S64. $^{\rm 13}\text{C}$ NMR (150 MHz, CDCl₃) of O64

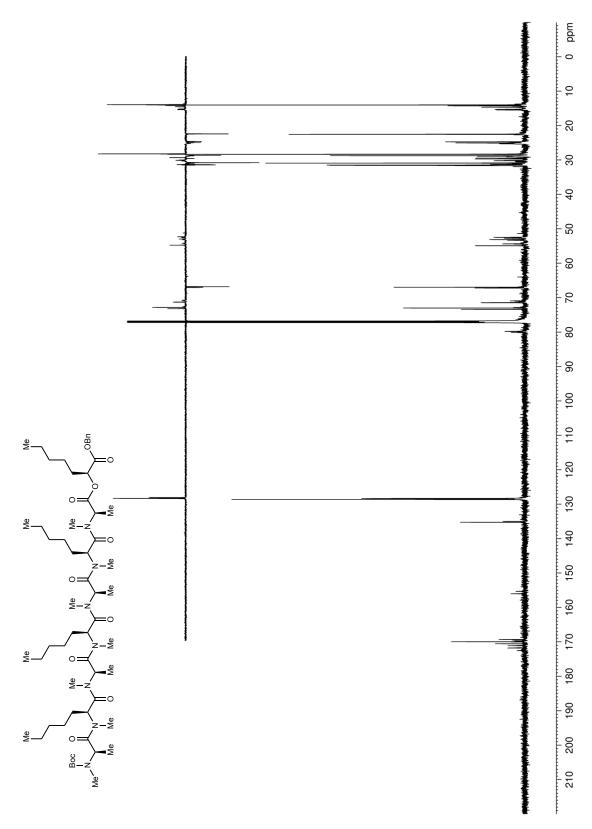


Figure S65. ¹H NMR (600 MHz, CDCl₃) of O65

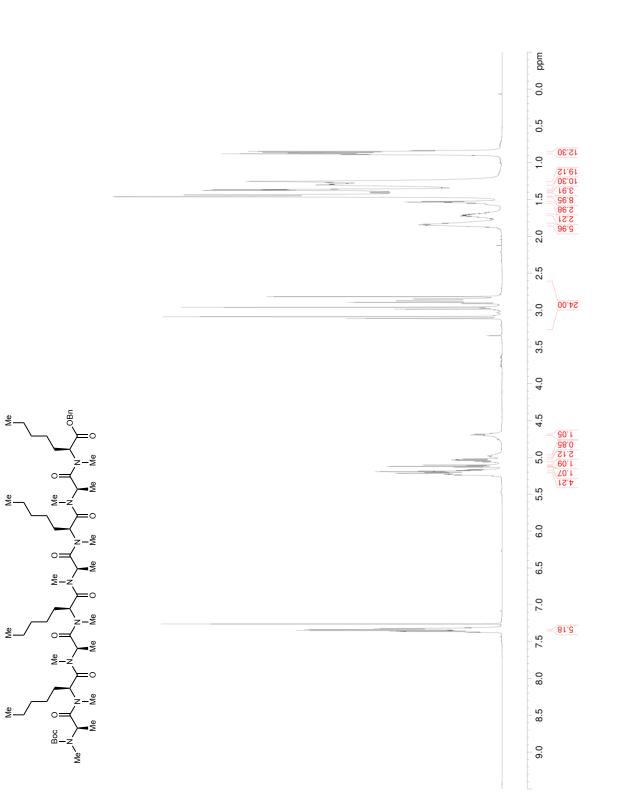


Figure S66. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of O65

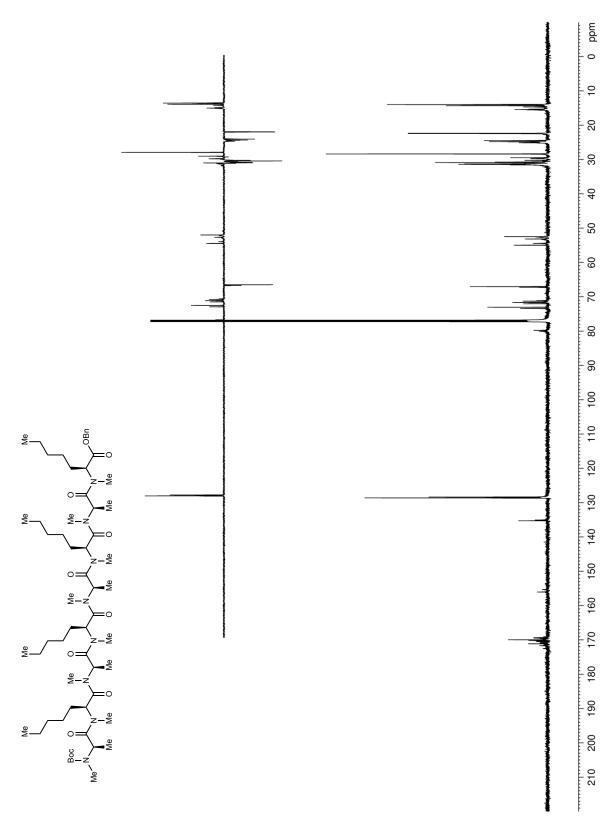
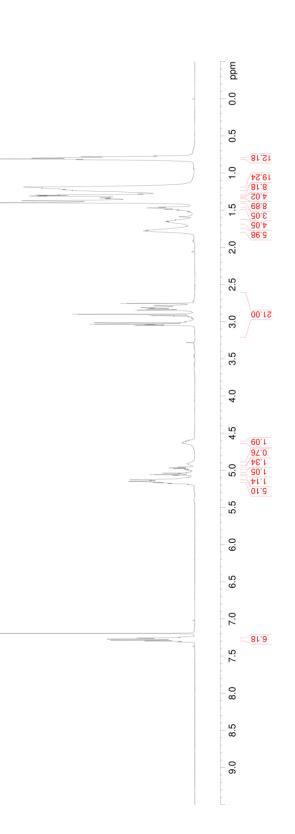


Figure S67. ¹H NMR (600 MHz, CDCl₃) of O66



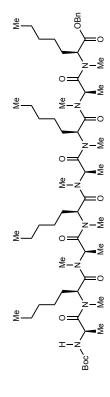
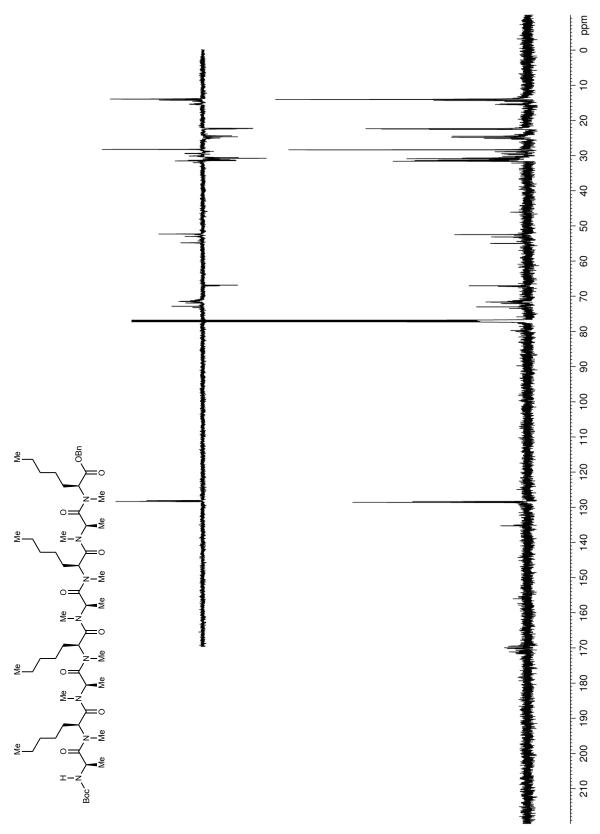
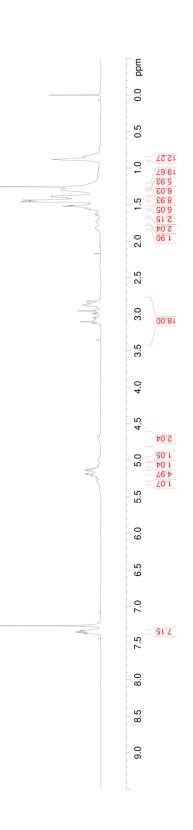


Figure S68. $^{\rm 13}\text{C}$ NMR (150 MHz, CDCl₃) of O66





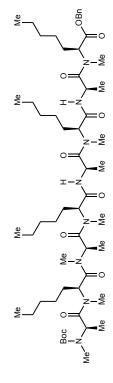


Figure S70. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 067

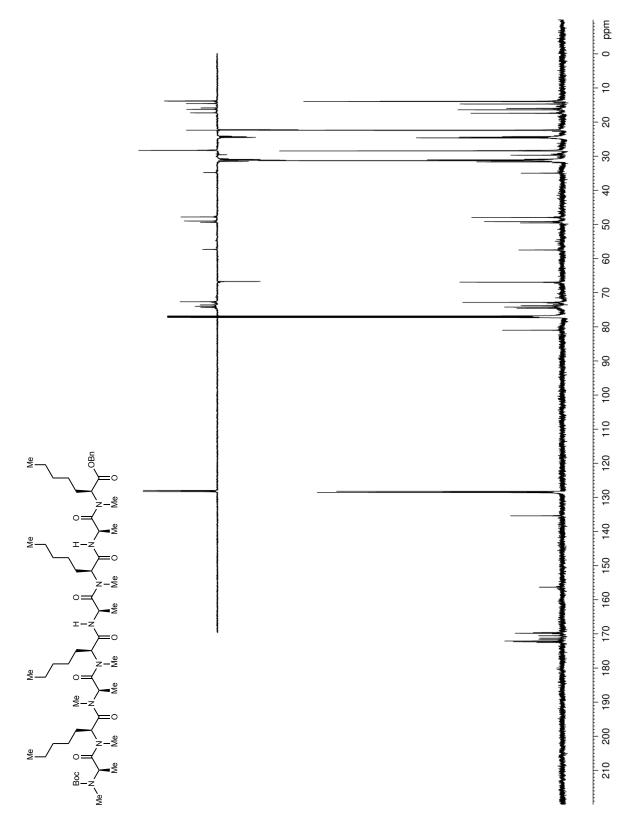
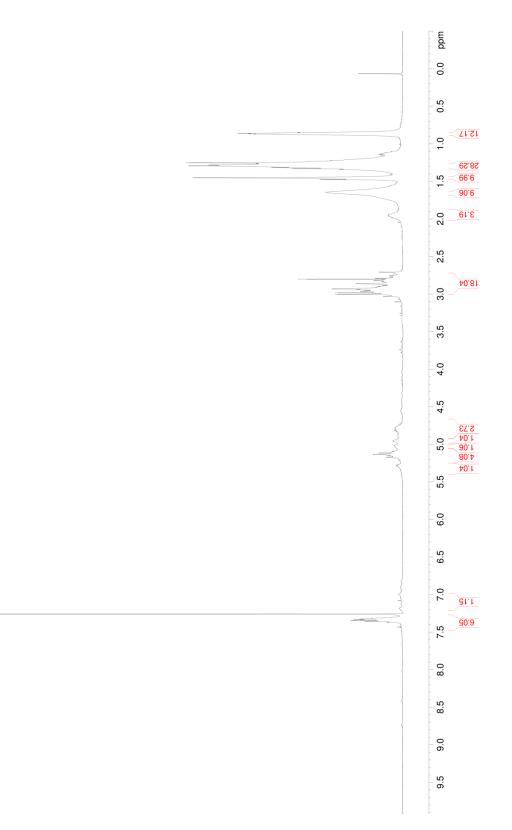


Figure S71. ¹H NMR (600 MHz, CDCl₃) of O68



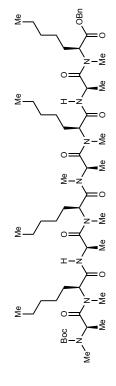


Figure S72. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of O68

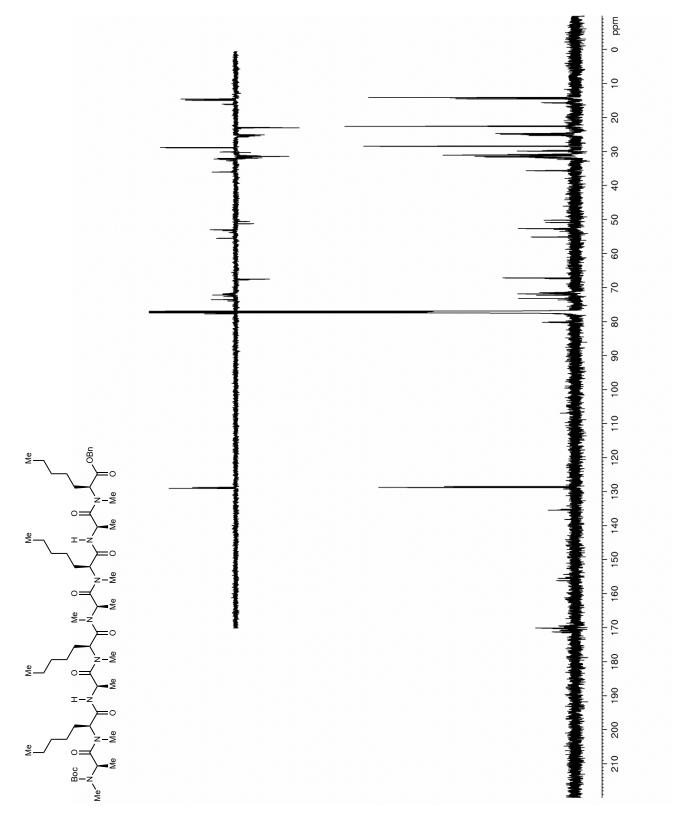
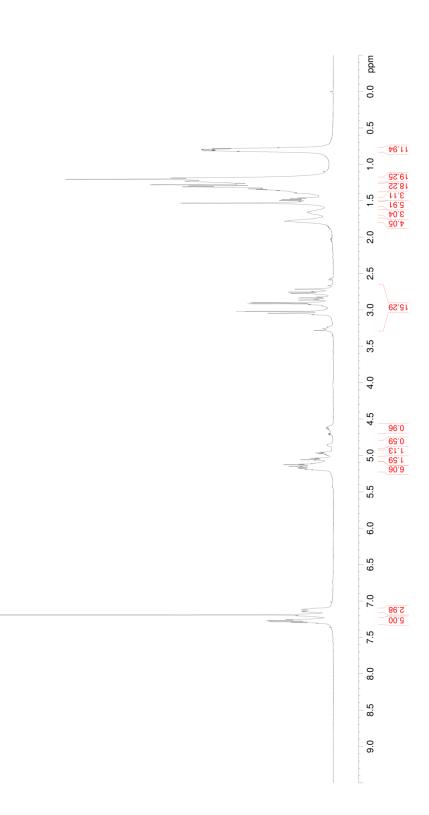


Figure S73. ¹H NMR (600 MHz, CDCl₃) of O69



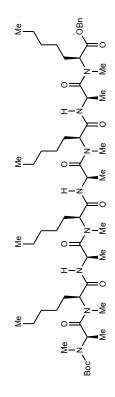
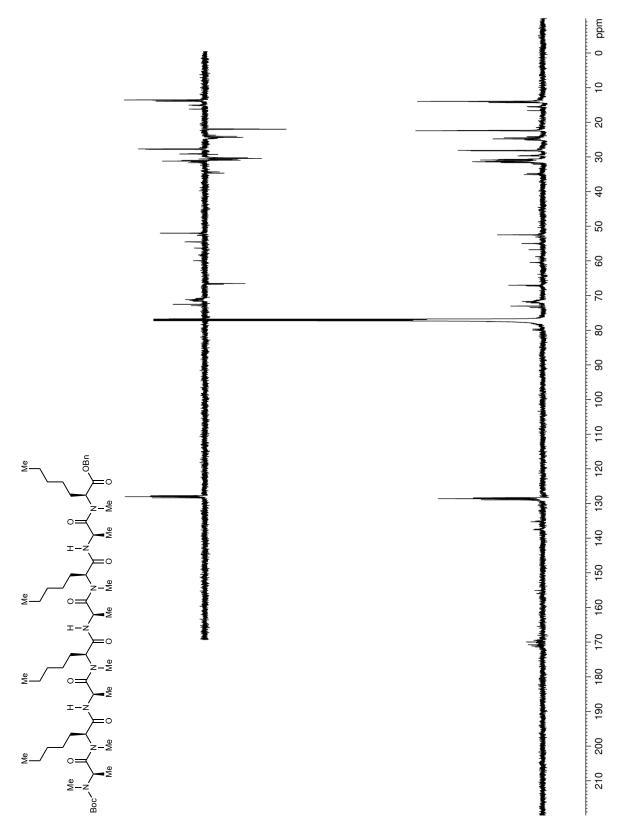


Figure S74. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 069



Supporting Information

Figure S75. ¹H NMR (600 MHz, CDCl₃) of O70

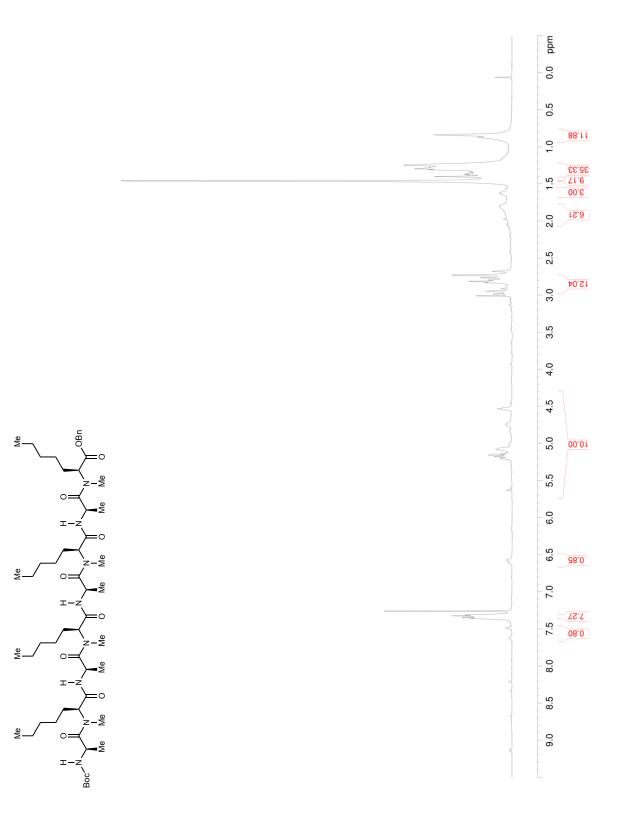


Figure S76. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of O70

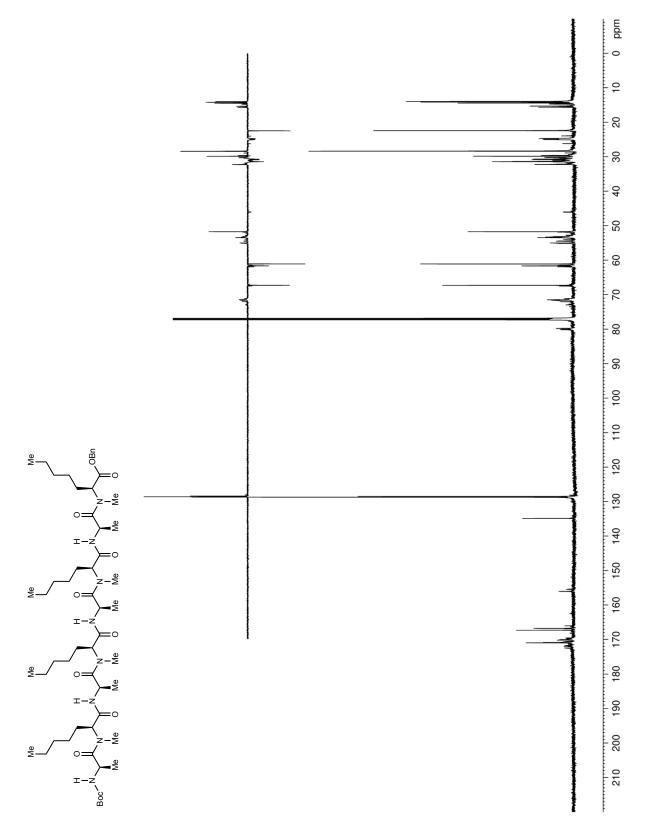


Figure S77. ¹H NMR (600 MHz, (CDCl₃) of **O71**

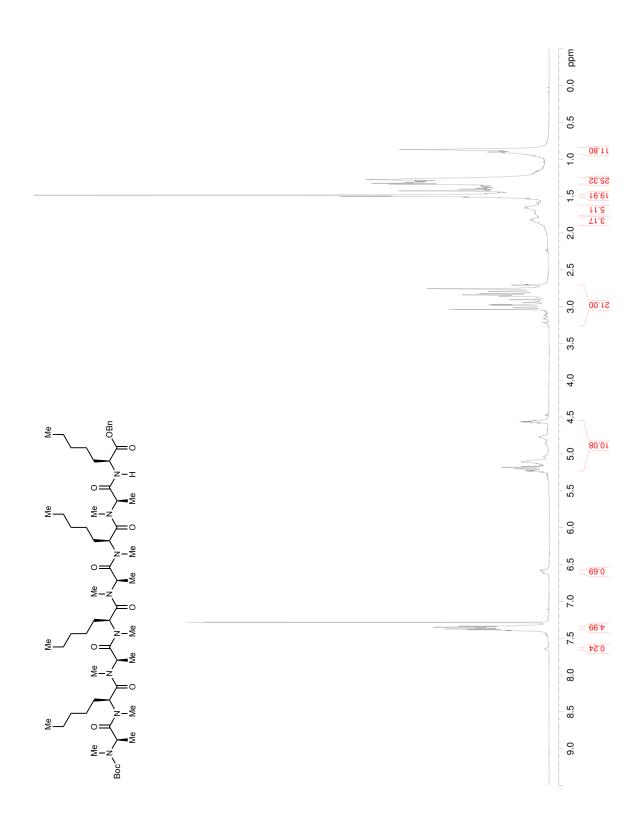


Figure S78. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of O71

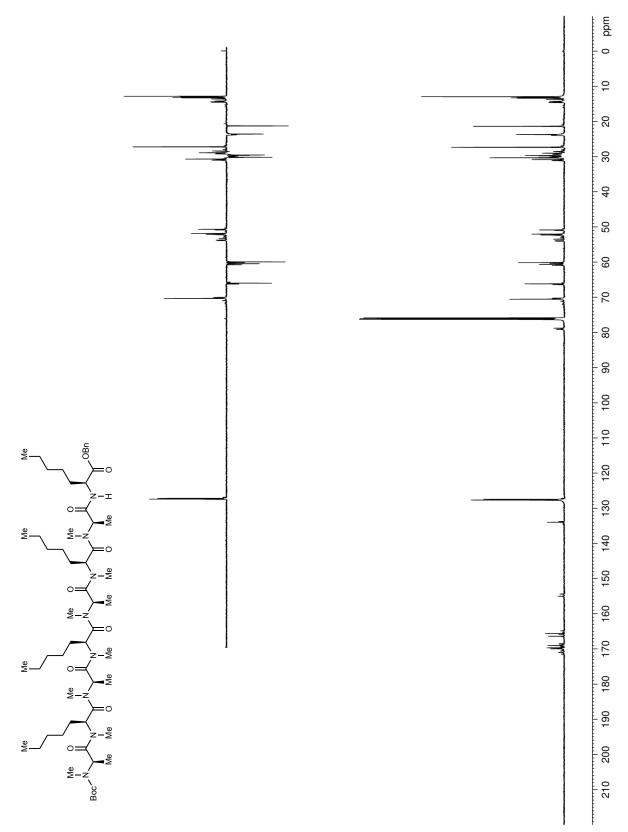


Figure S79. ¹H NMR (600 MHz, CD₃COCD₃) of O72

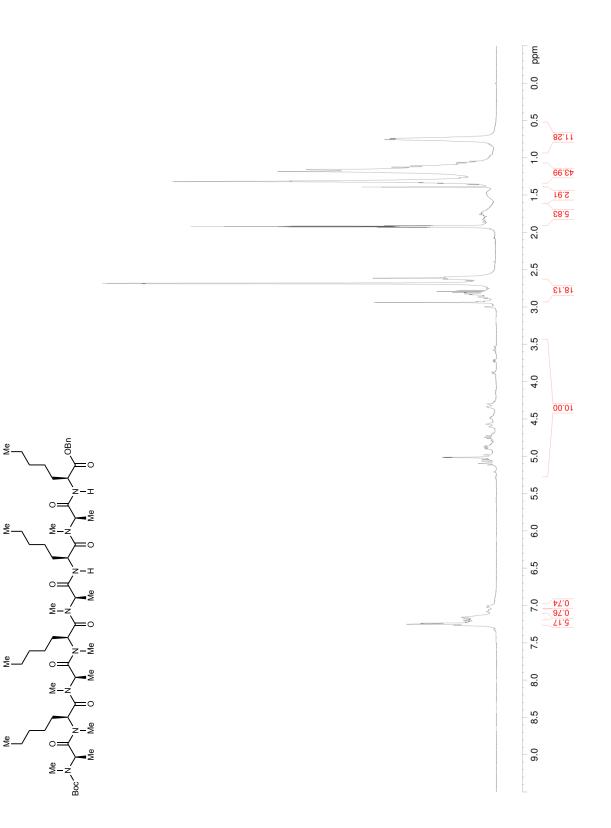


Figure S80. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 072

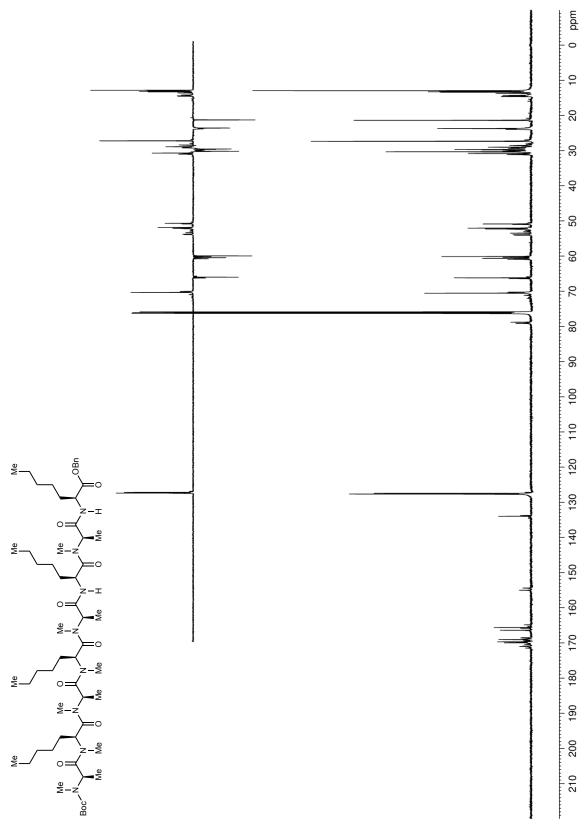


Figure S81. ¹H NMR (600 MHz, CDCl₃) of **O73**

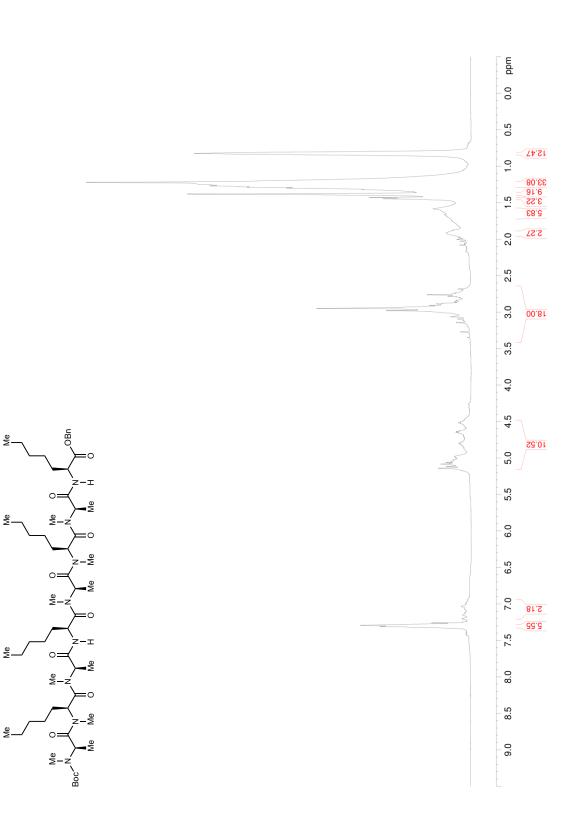


Figure S82. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 073

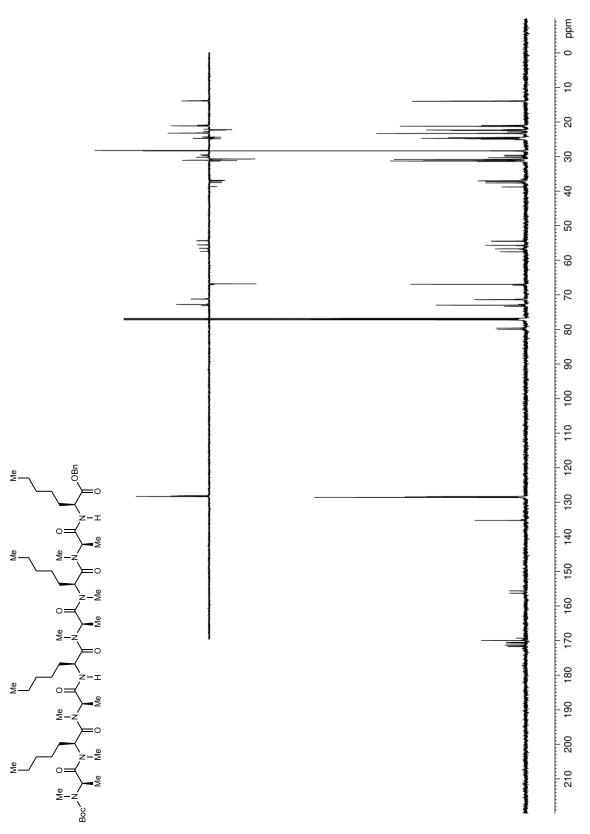
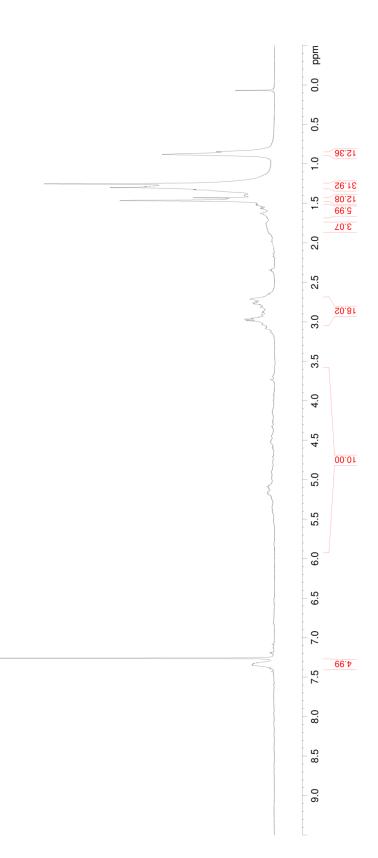


Figure S83. ¹H NMR (600 MHz, CDCl₃) of O74



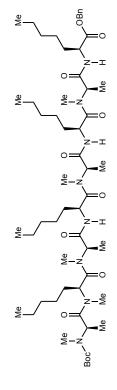


Figure S84. $^{\rm 13}\text{C}$ NMR (150 MHz, CDCl₃) of 074

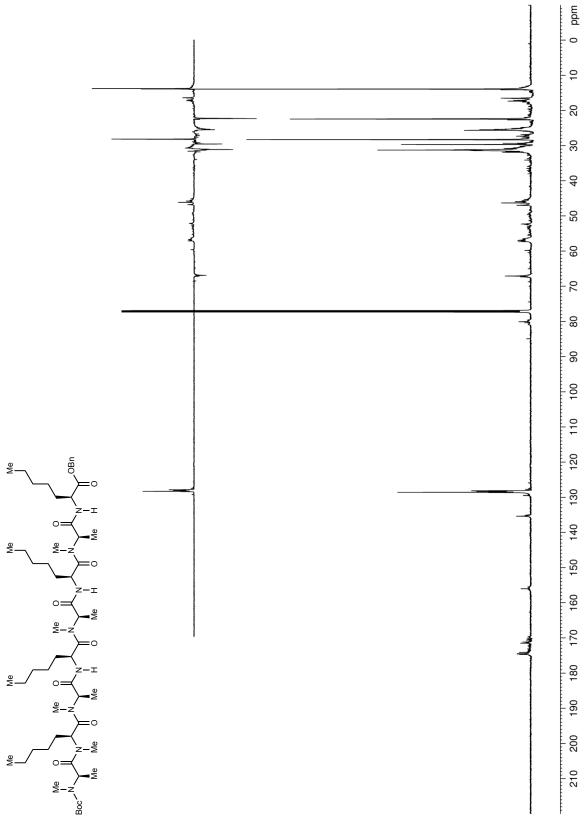
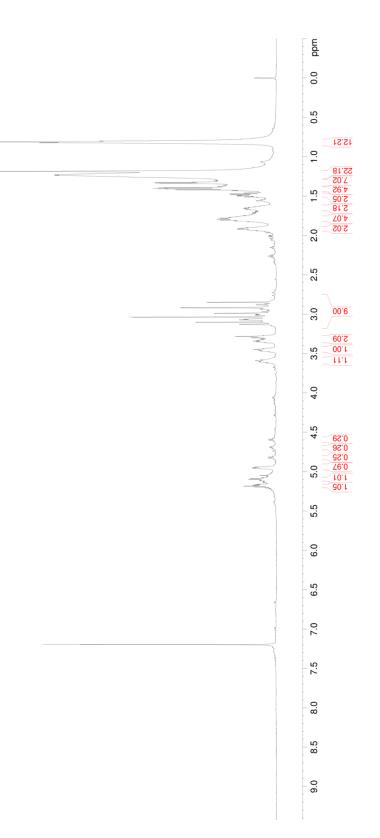
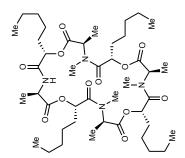


Figure S85. ¹H NMR (600 MHz, CDCl₃) of 1.1





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Figure S86. $^{\rm 13}{\rm C}$ NMR (150 MHz, CDCl₃) of 1.1

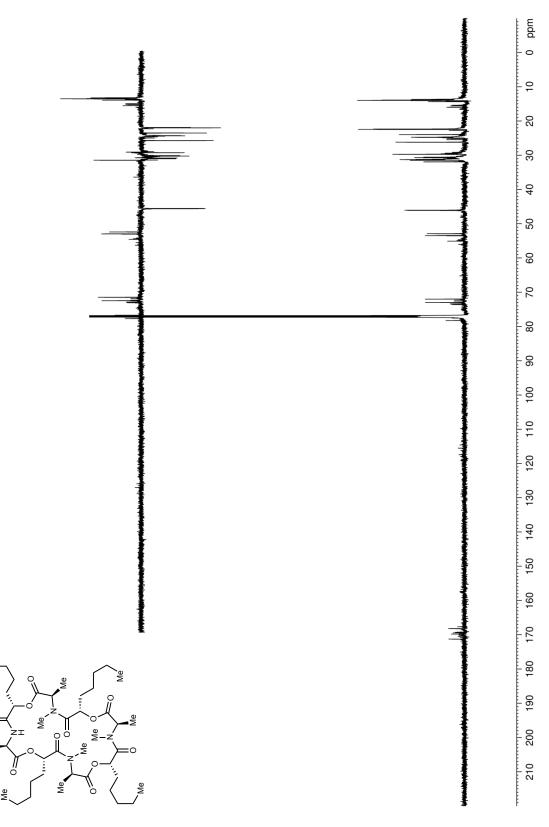


Figure S87. 1 H NMR (600 MHz, CDCl₃) of **1.2**

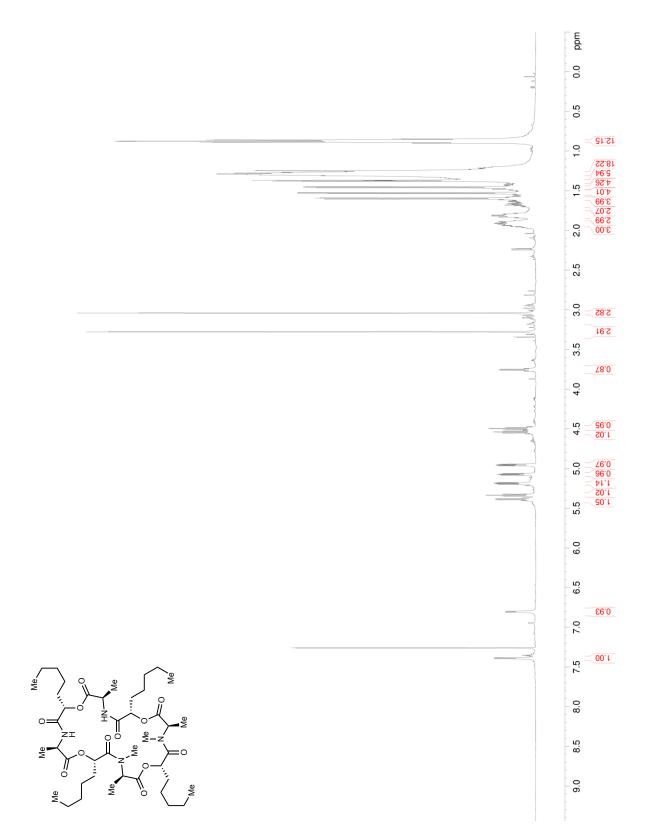


Figure S88. $^{\rm 13}{\rm C}$ NMR (150 MHz, CDCl₃) of 1.2

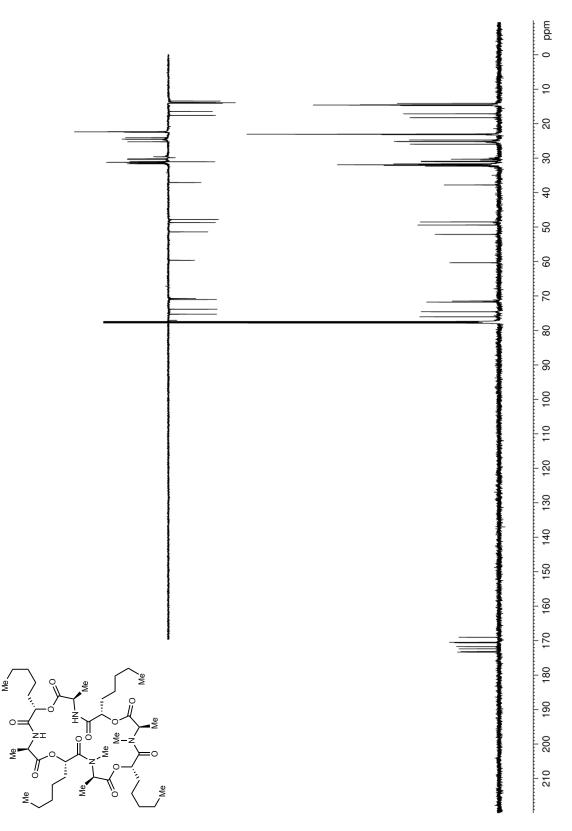
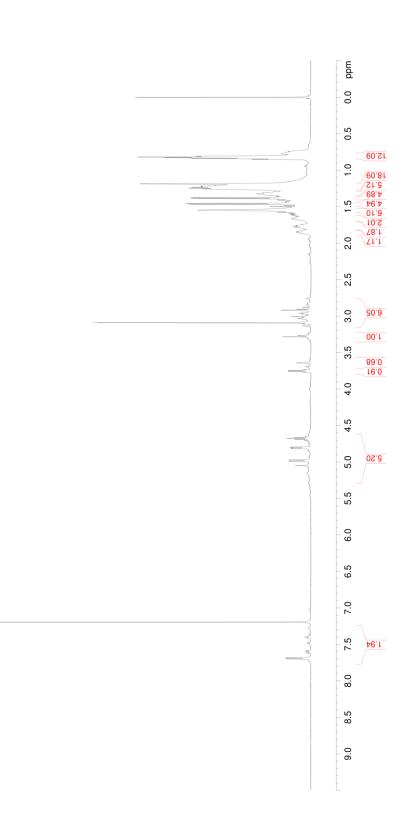


Figure S89. ¹H NMR (600 MHz, CDCl₃) of 1.3



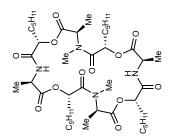


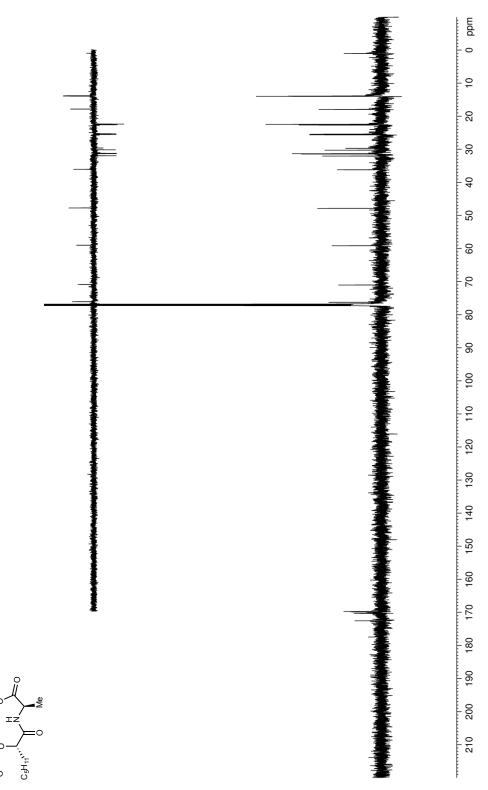
Figure S90. ¹³C NMR (150 MHz, CDCl₃) of **1.3**

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C5H11

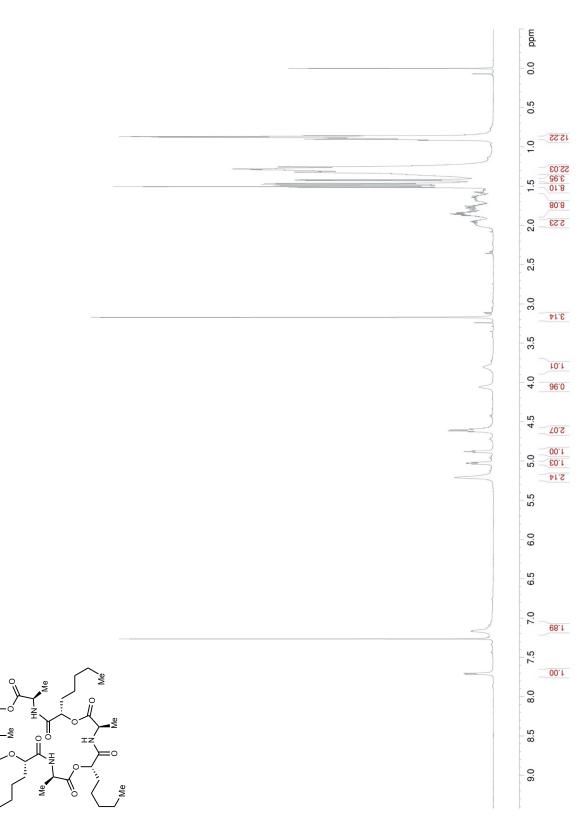
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Figure S92. 13 C NMR (150 MHz, CDCl₃) of 1.4

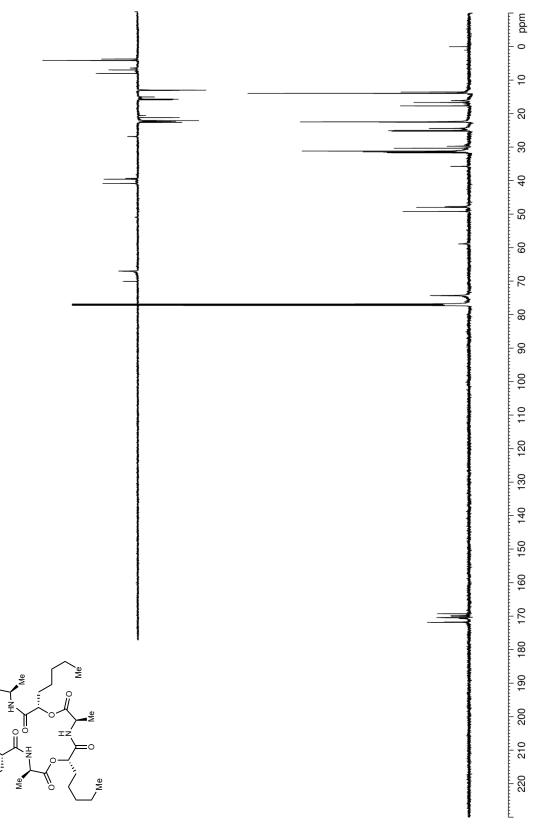


Figure S93. ¹H NMR (600 MHz, CD₃COCD₃) of 2.1

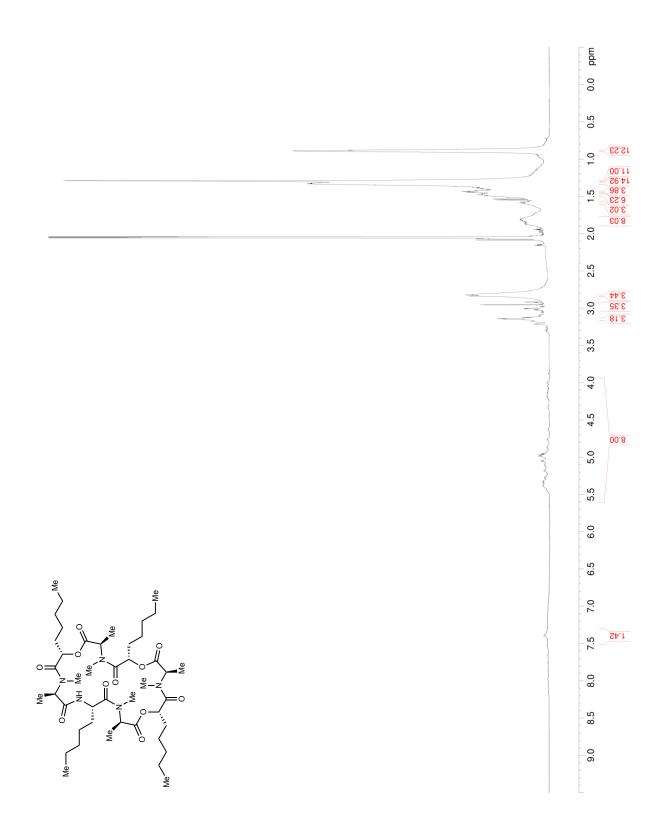


Figure S94. ¹³C NMR (150 MHz, CDCl₃) of 2.1

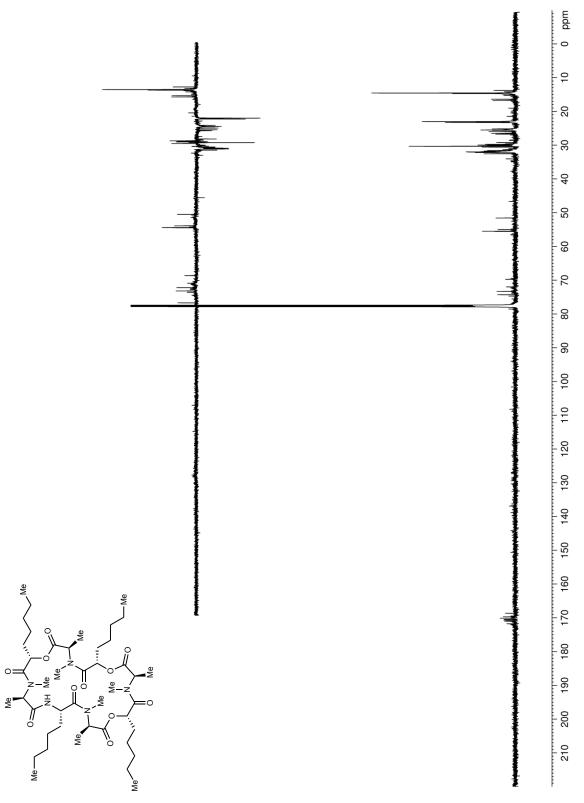
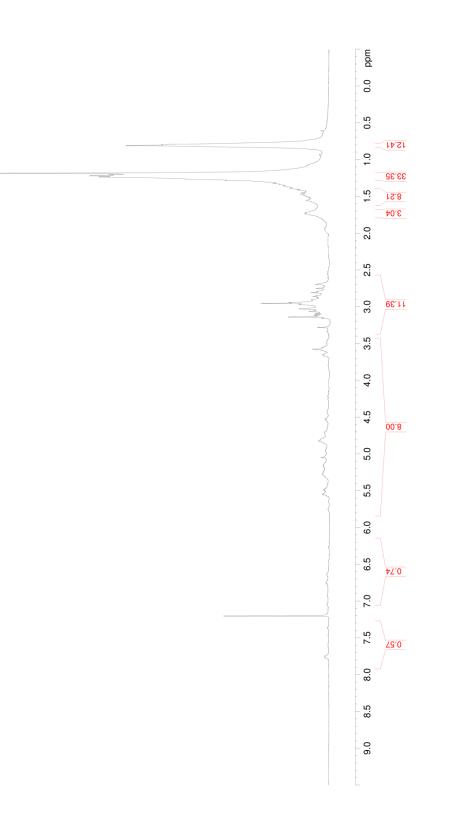
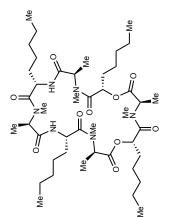


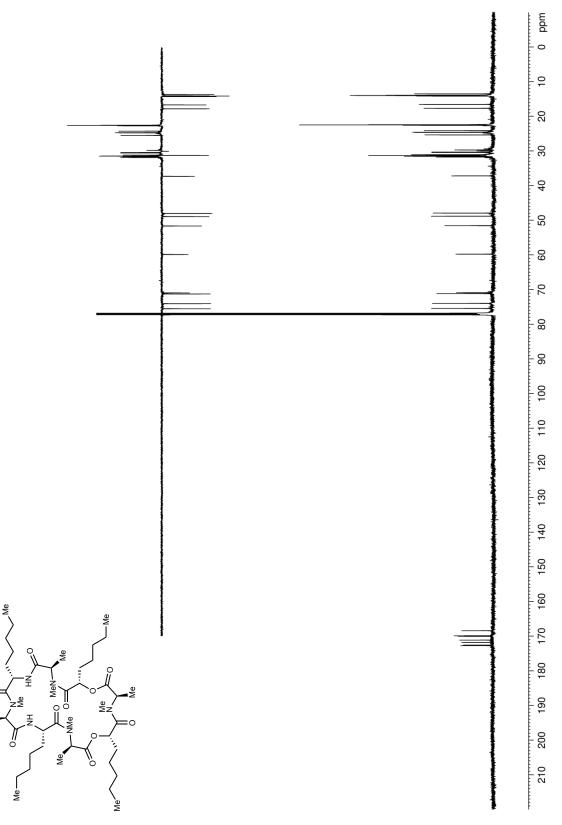
Figure S95. ¹H NMR (600 MHz, CDCl₃) of 2.2





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Figure S96. $^{\rm 13}{\rm C}$ NMR (150 MHz, CDCl₃) of 2.2



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Figure S97. ¹H NMR (600 MHz, CDCl₃) of 2.3

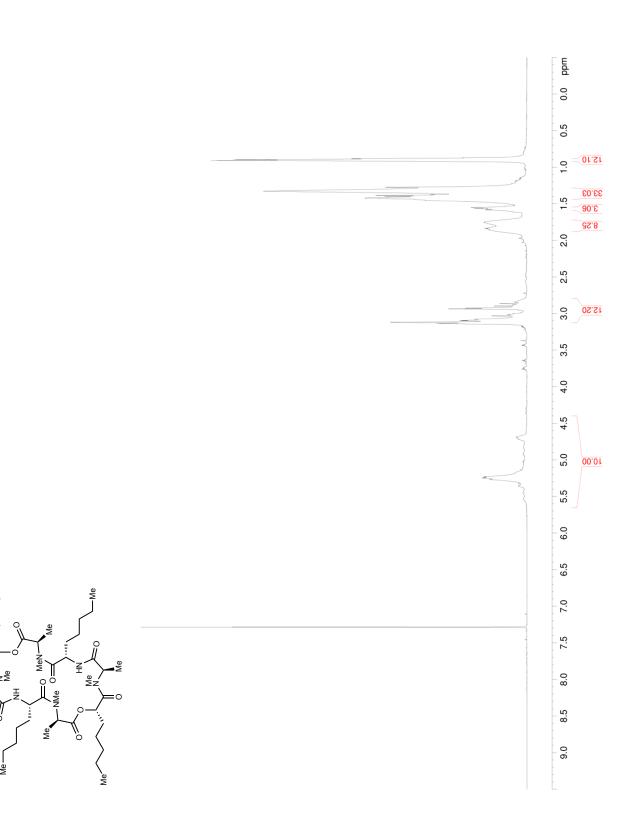


Figure S98. ¹³C NMR (150 MHz, CDCl₃) of 2.3

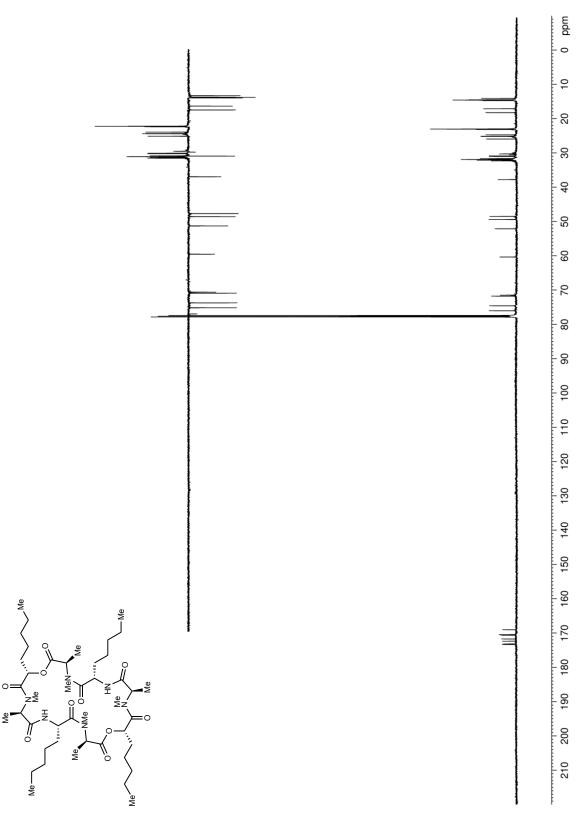
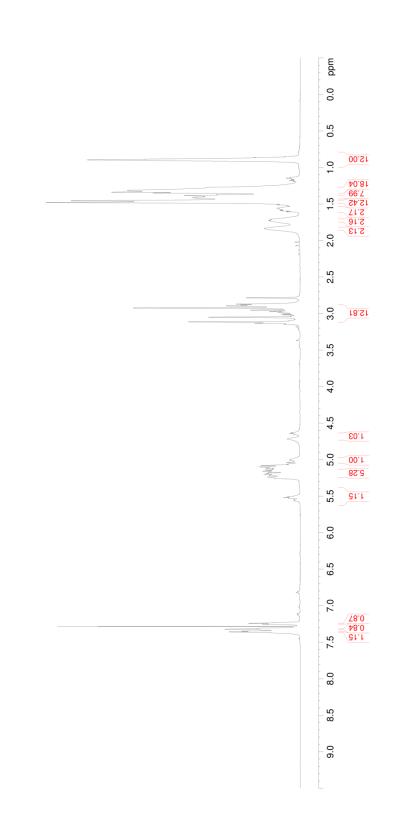


Figure S99. ¹H NMR (600 MHz, CDCl₃) of 2.4



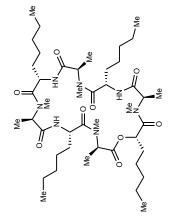


Figure S100. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 2.4

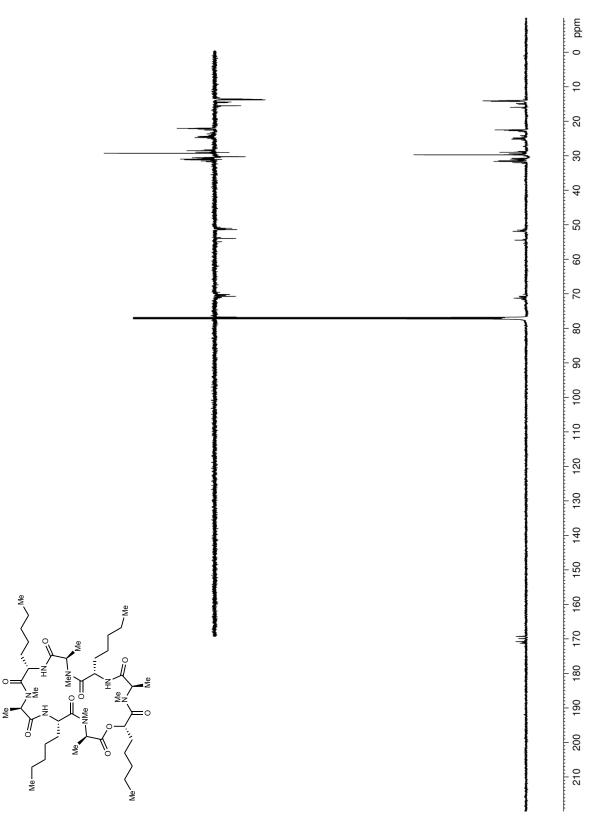
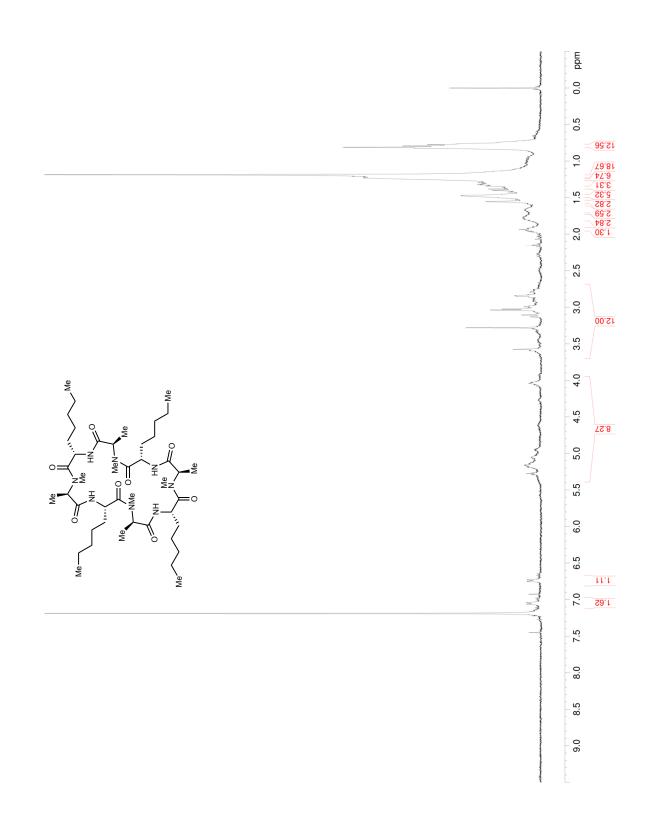


Figure S101. 1 H NMR (600 MHz, CDCl₃) of 2.5



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Figure S102. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 2.5

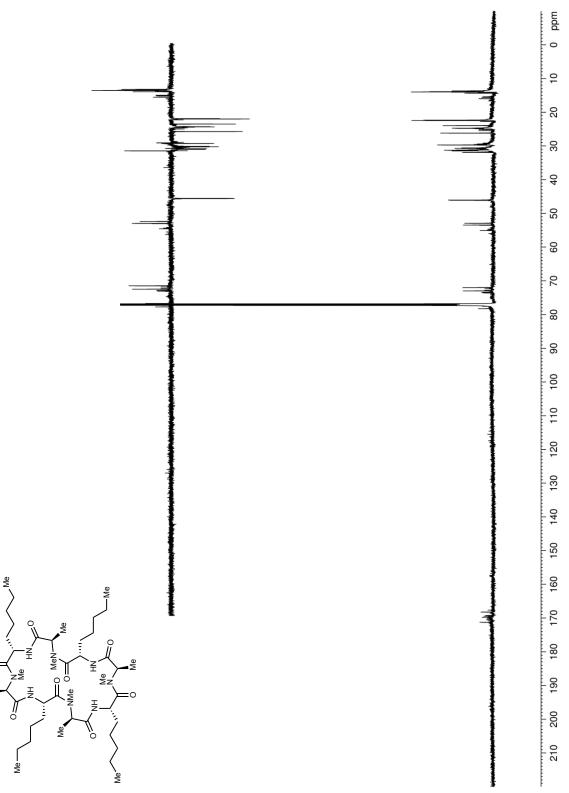
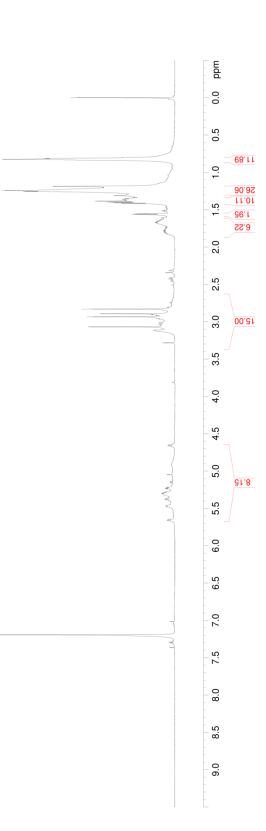


Figure S103. 1 H NMR (600 MHz, CDCl₃) of 3.1



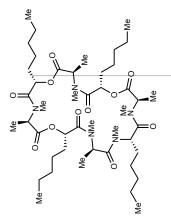
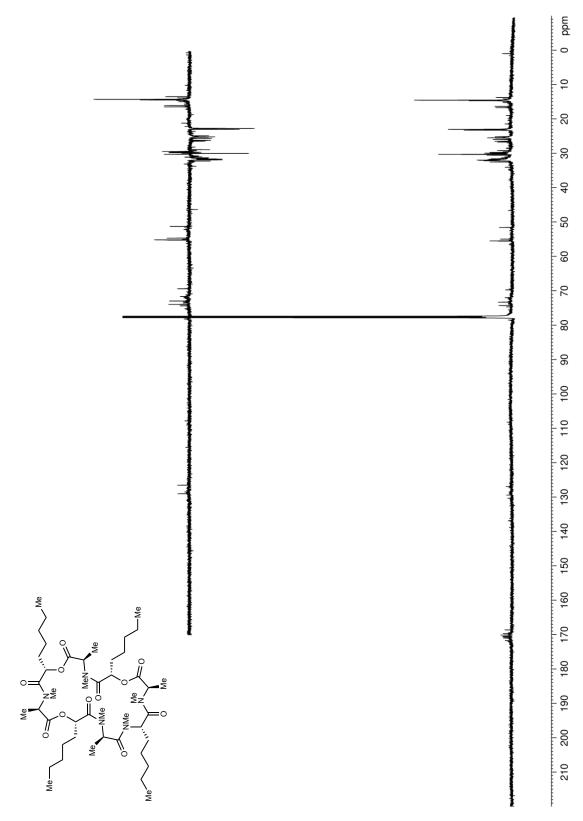


Figure S104. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 3.1



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Figure S105. 1 H NMR (600 MHz, CDCl₃) of 3.2

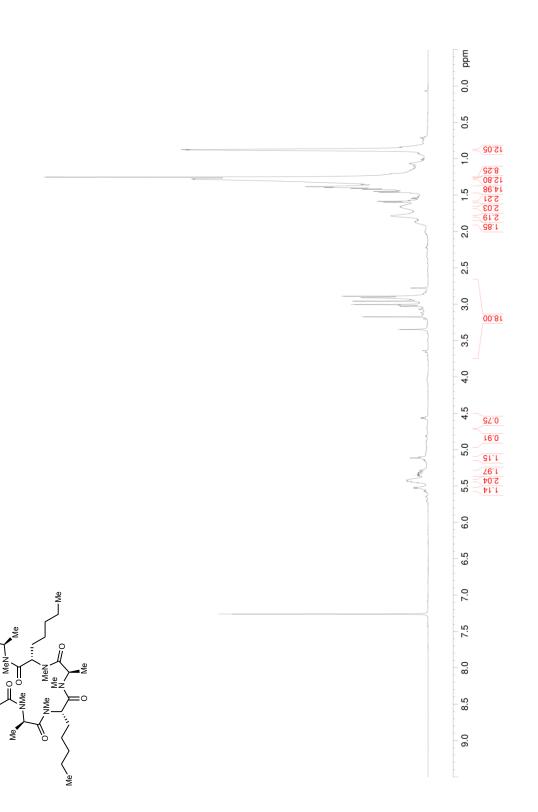


Figure S106. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 3.2

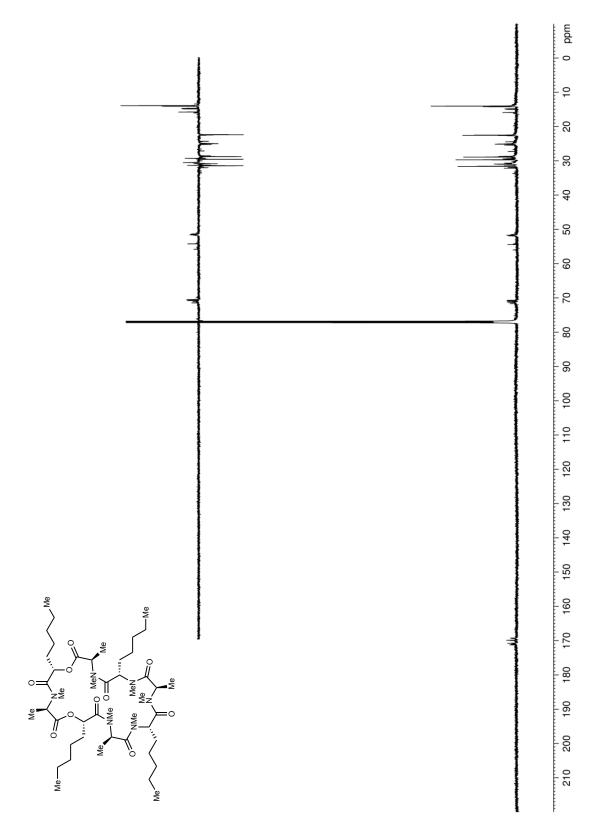


Figure S107. 1 H NMR (600 MHz, CDCl₃) of 3.3

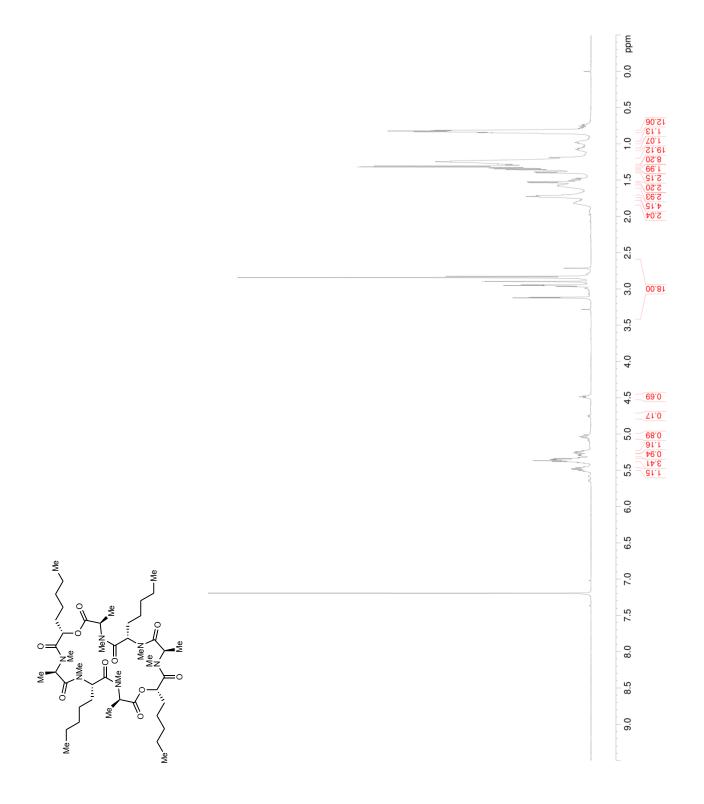
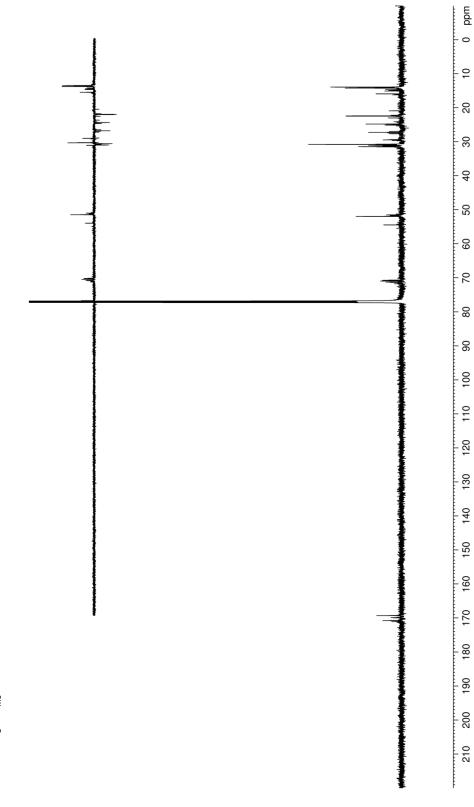
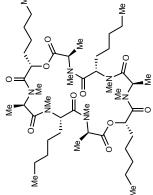


Figure S108. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 3.3





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Figure S109. 1 H NMR (600 MHz, CDCl₃) of 3.4

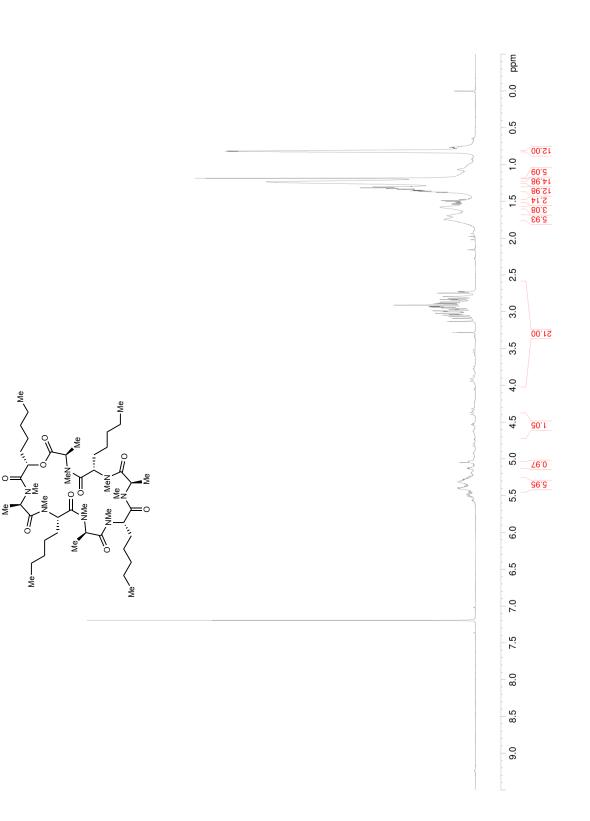
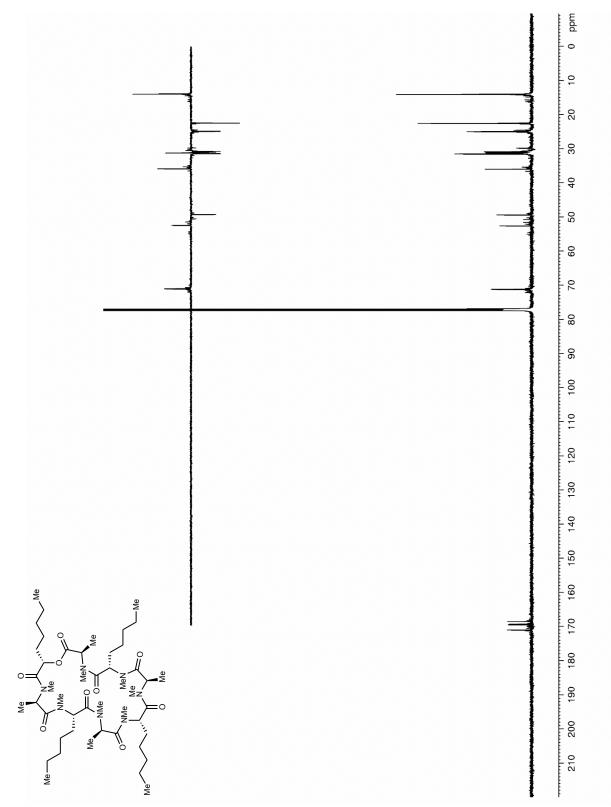
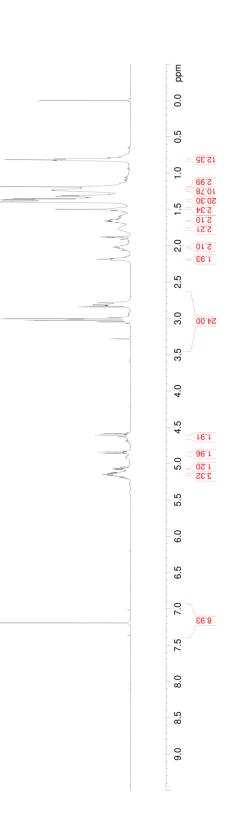


Figure S110. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 3.4





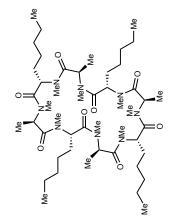


Figure S112. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 3.5

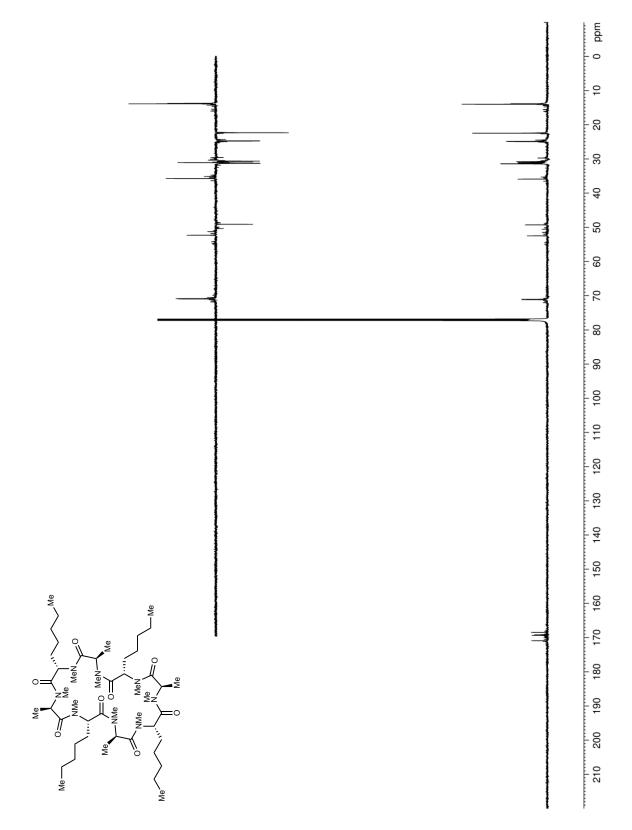
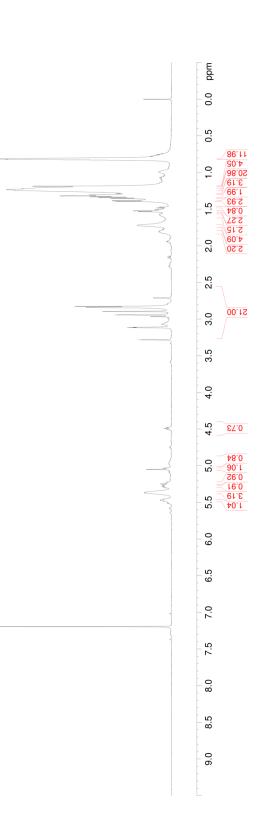


Figure S113. 1 H NMR (600 MHz, CDCl₃) of 4.2



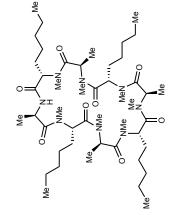


Figure S114. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 4.2

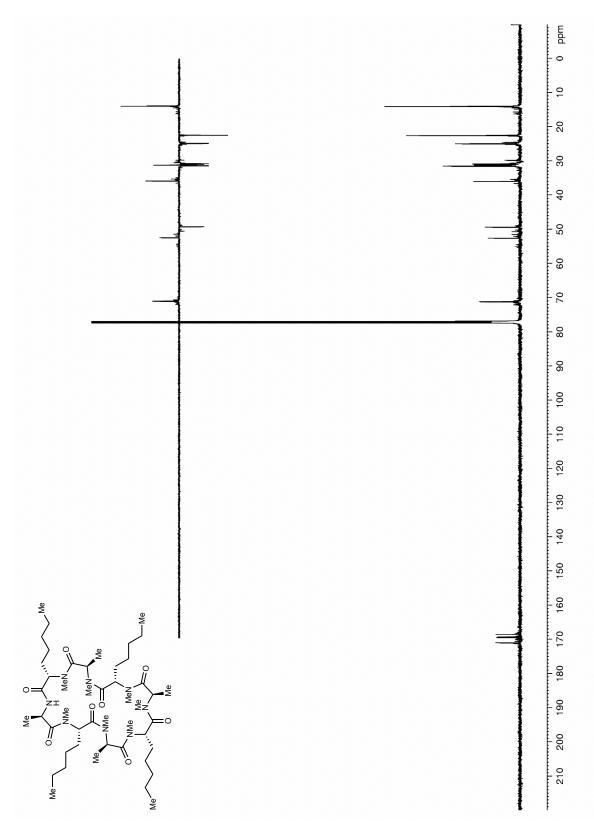


Figure S115. 1 H NMR (600 MHz, CDCl₃) of 4.3

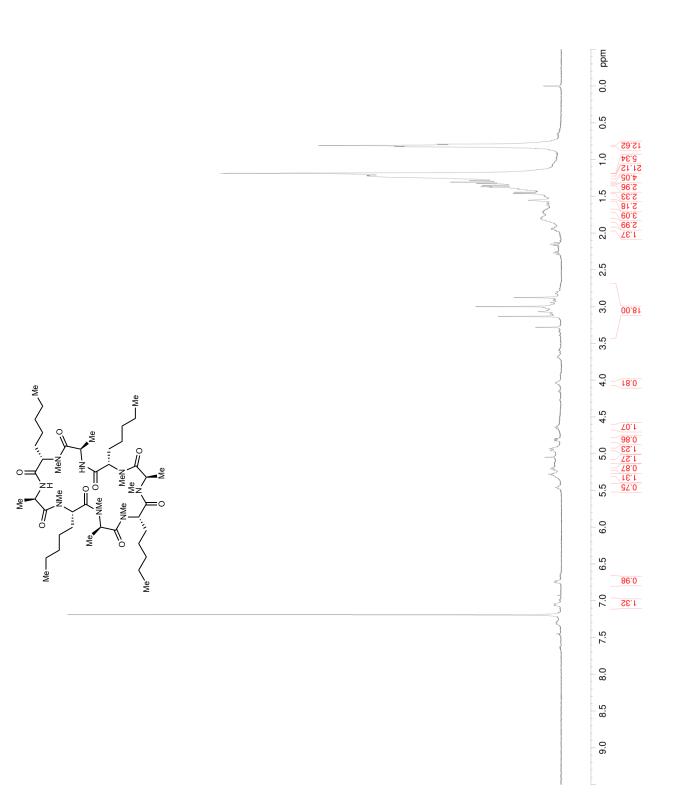


Figure S116. $^{\rm 13}C$ NMR (150 MHz, CDCl₃) of 4.3

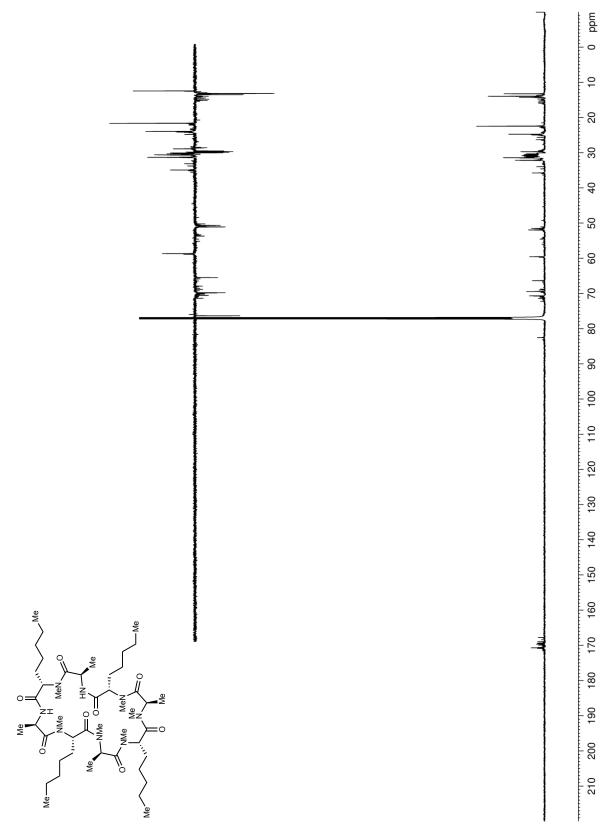


Figure S117. 1 H NMR (600 MHz, CDCl₃) of 4.4

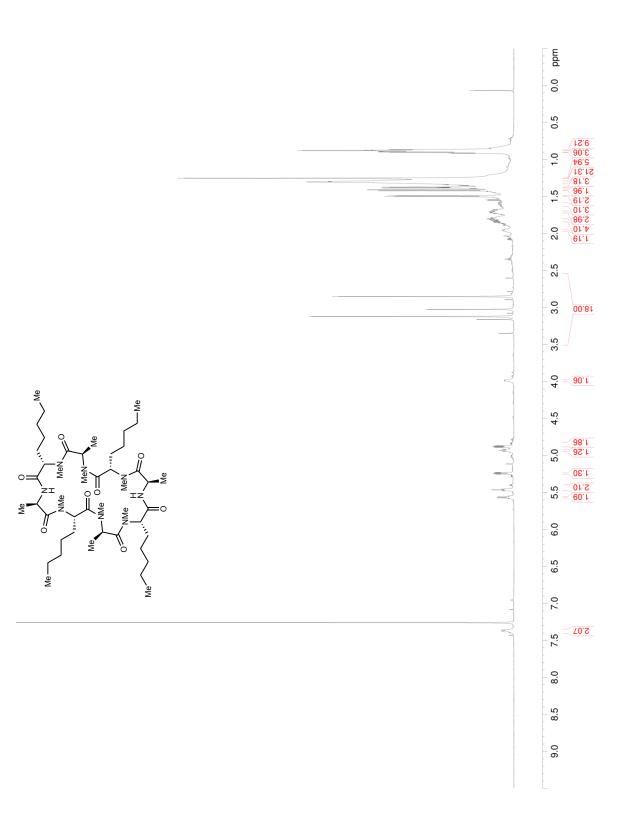


Figure S118. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 4.4

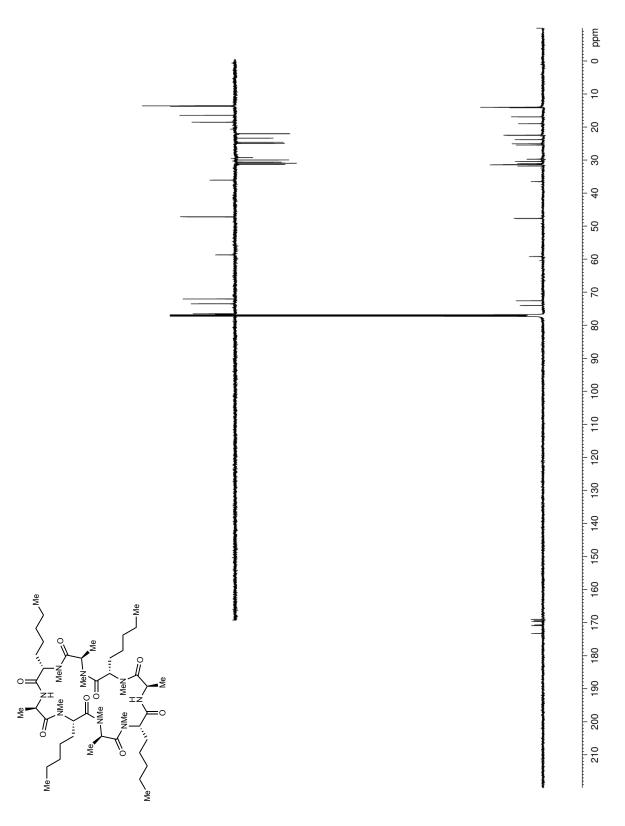


Figure S119. 1 H NMR (600 MHz, CDCl₃) of 4.5

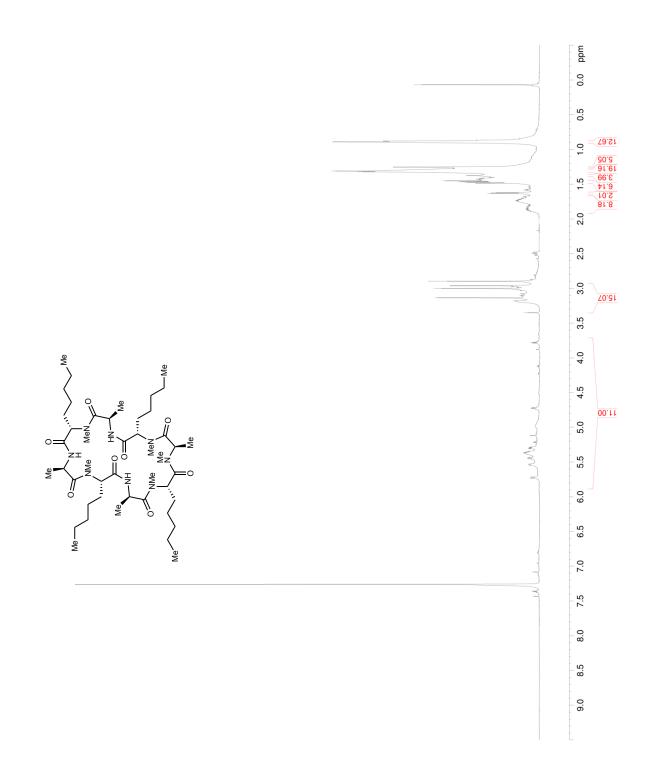


Figure S120. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 4.5

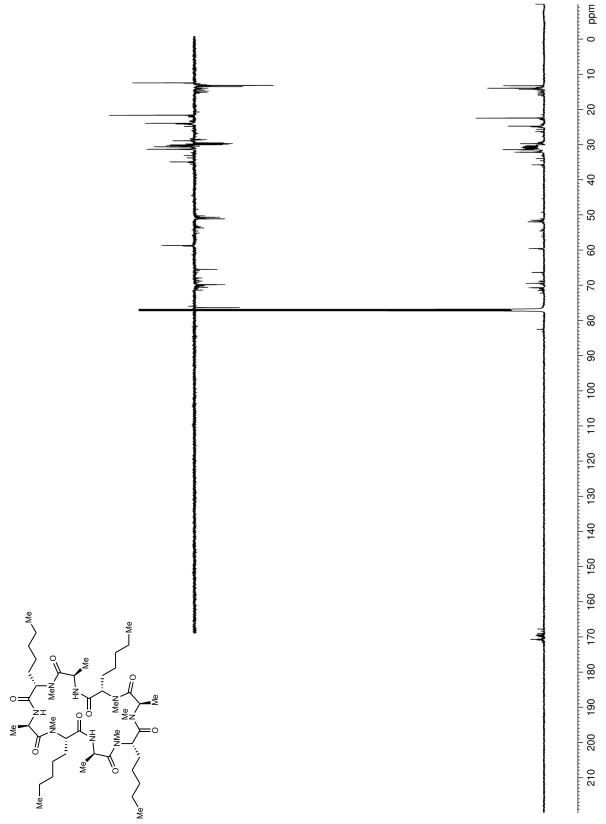


Figure S121. 1 H NMR (600 MHz, CDCl₃) of 4.6

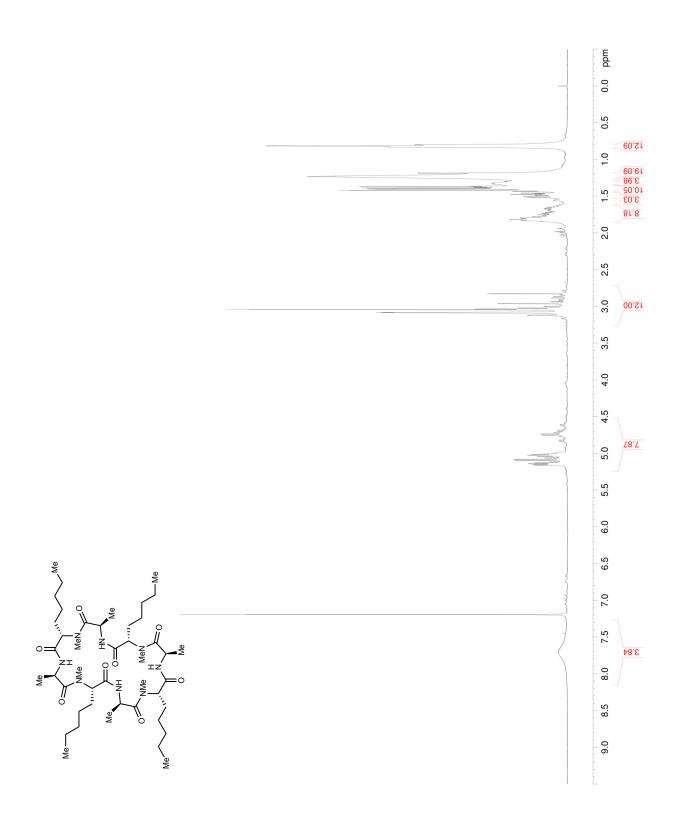
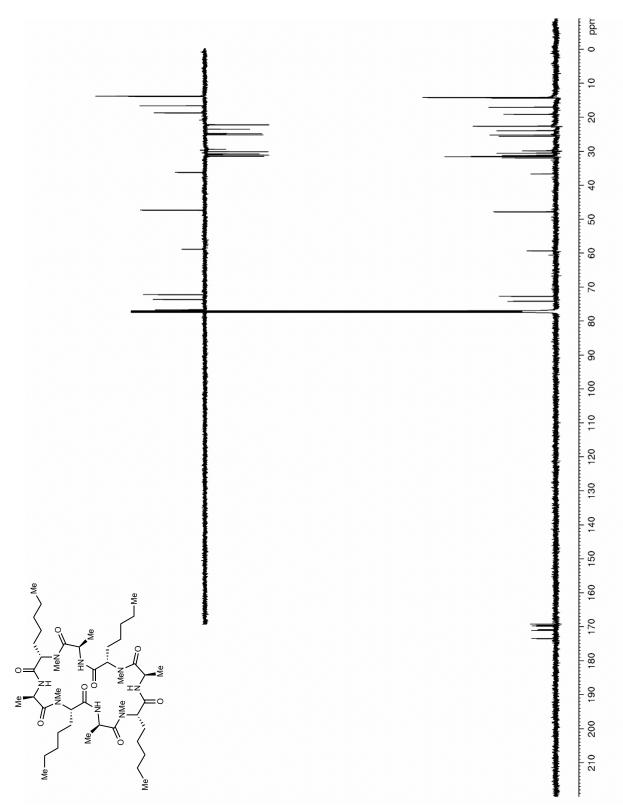
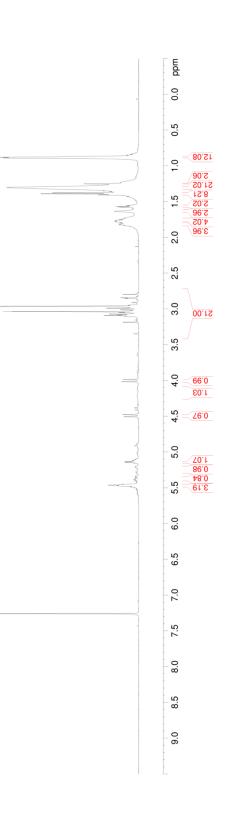


Figure S122. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 4.6





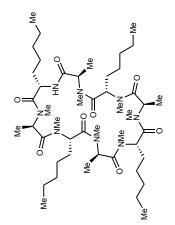


Figure S124. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 5.2

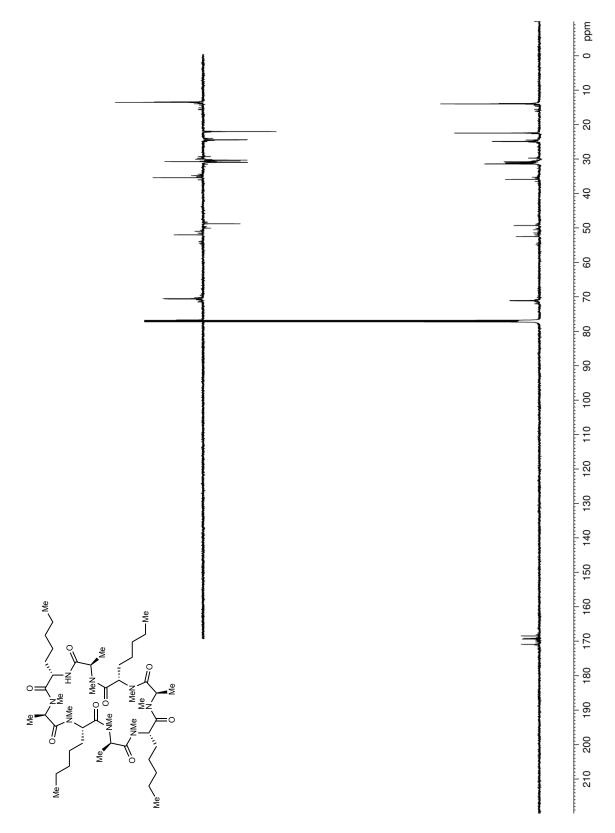
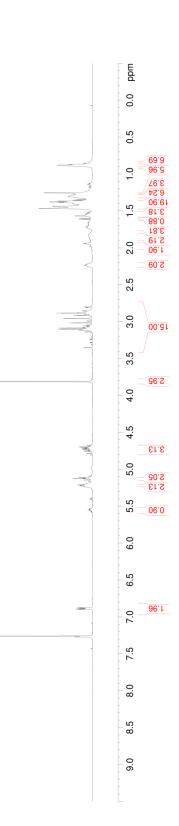
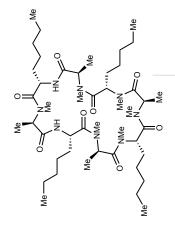


Figure S125. 1 H NMR (600 MHz, CDCl₃) of 5.3



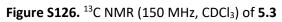




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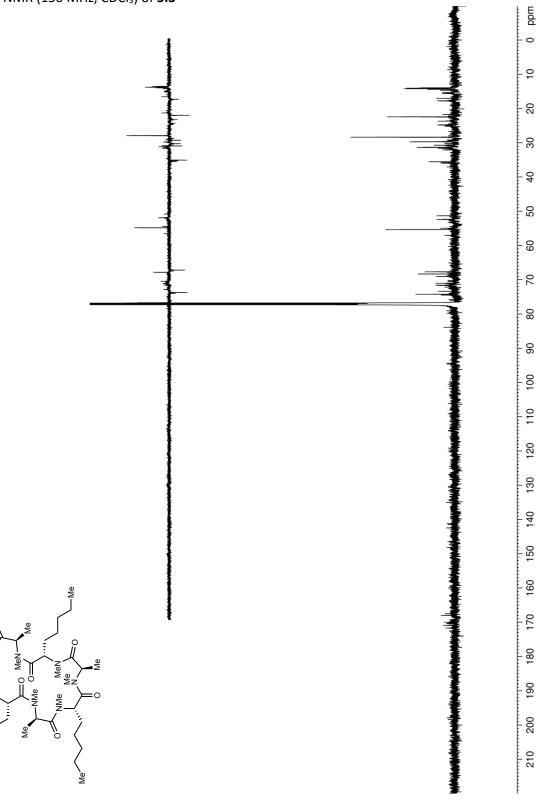
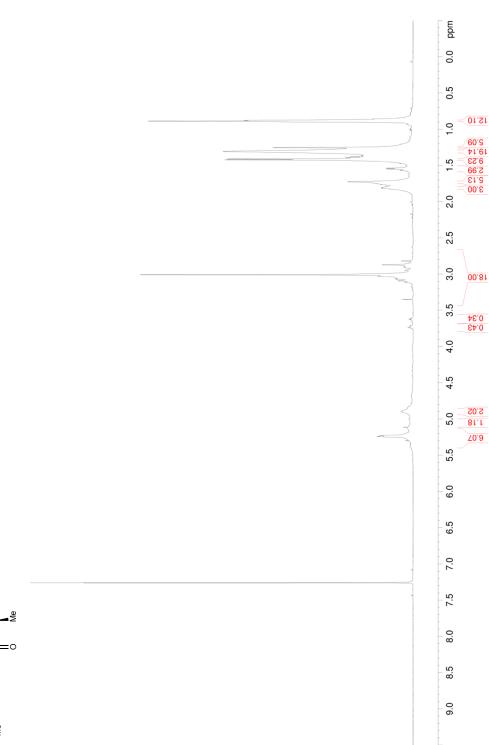
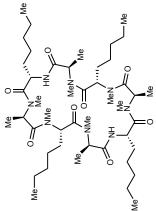


Figure S127. 1 H NMR (600 MHz, CDCl₃) of 5.4





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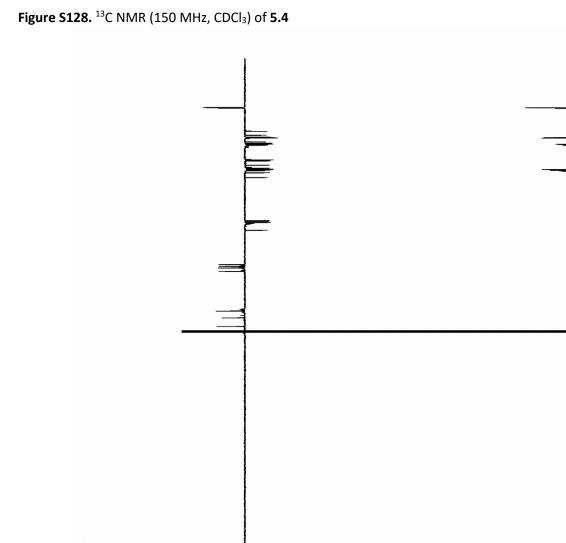
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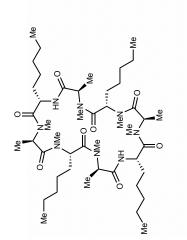
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Figure S129. 1 H NMR (600 MHz, CDCl₃) of 5.5

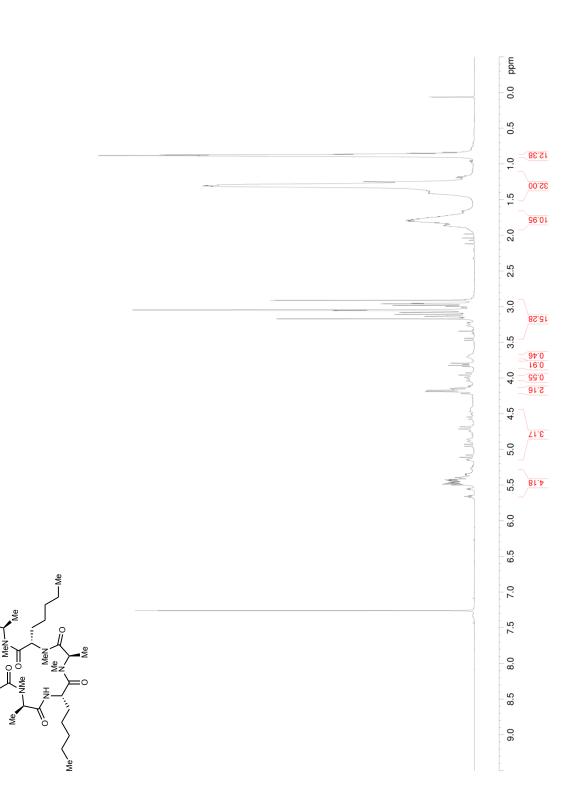


Figure S130. $^{\rm 13}C$ NMR (150 MHz, CDCl_3) of 5.5

