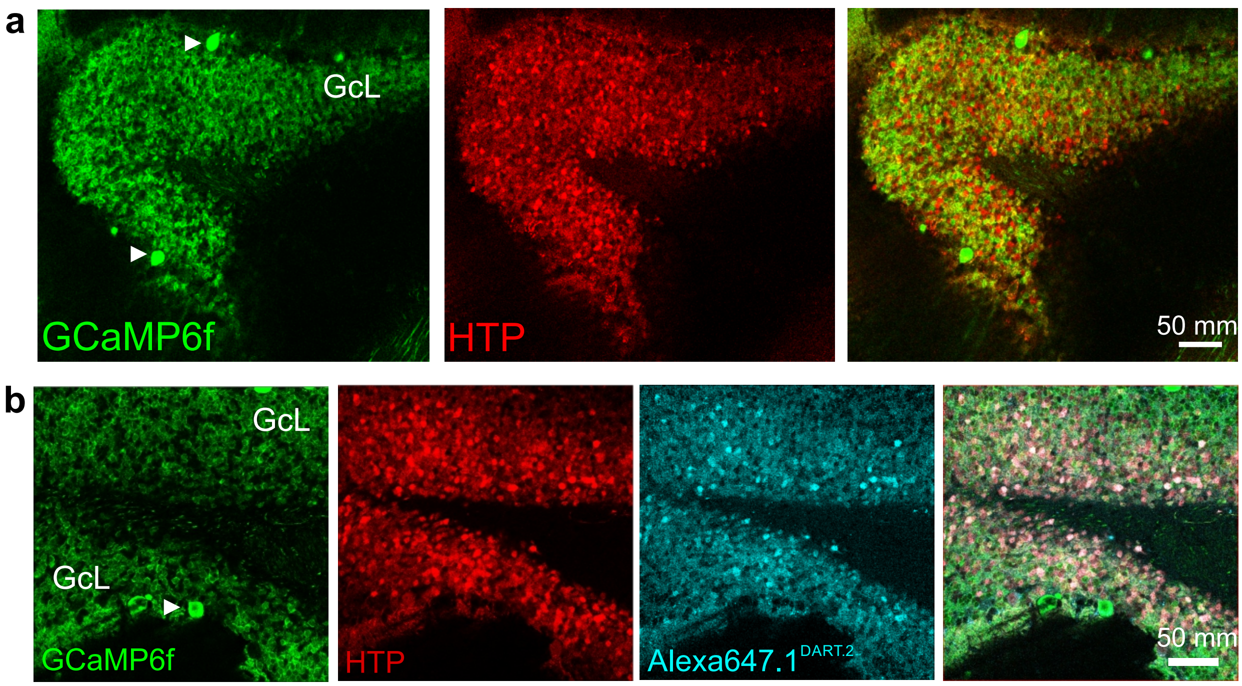
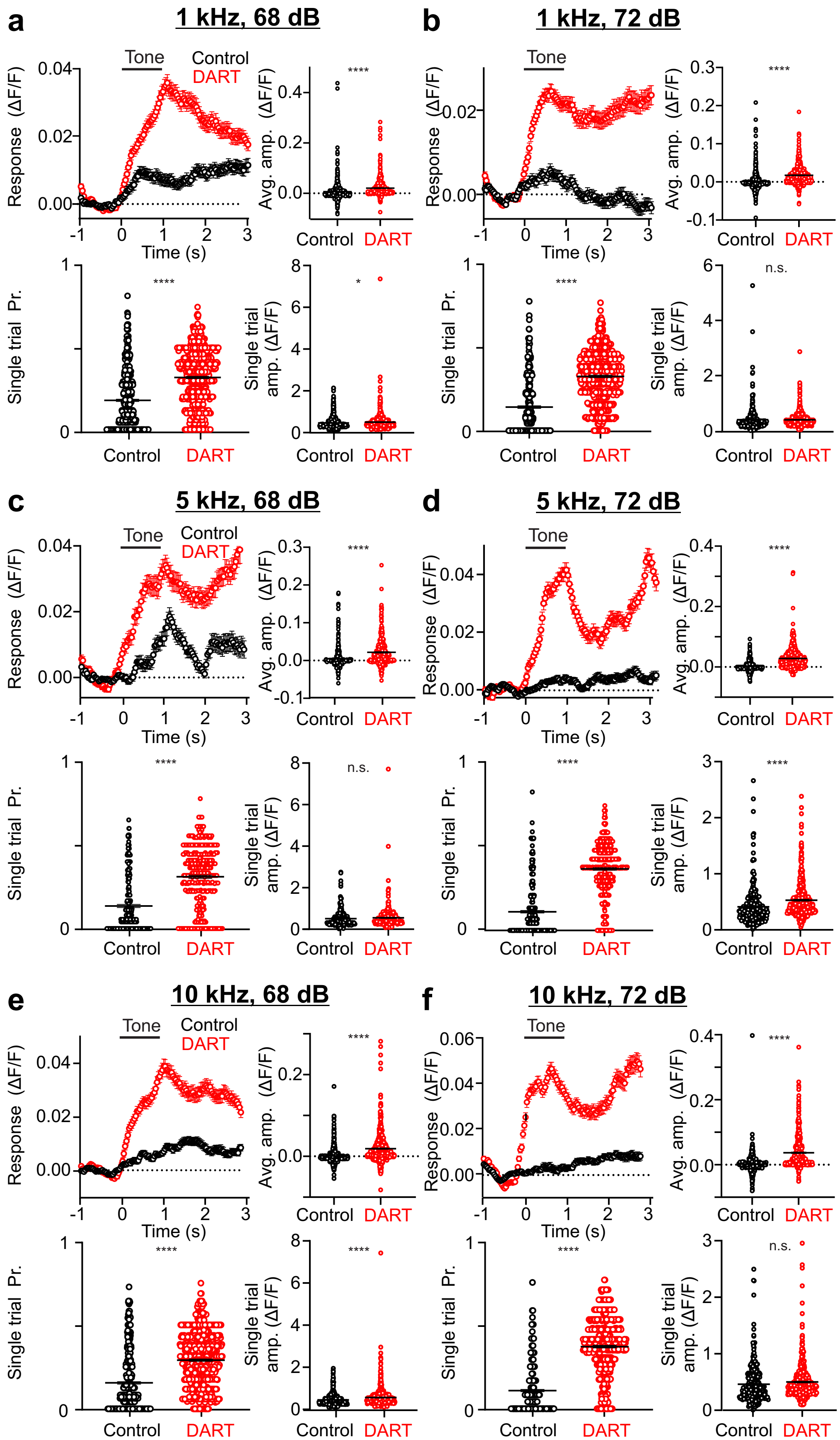


Supplementary Figure 1

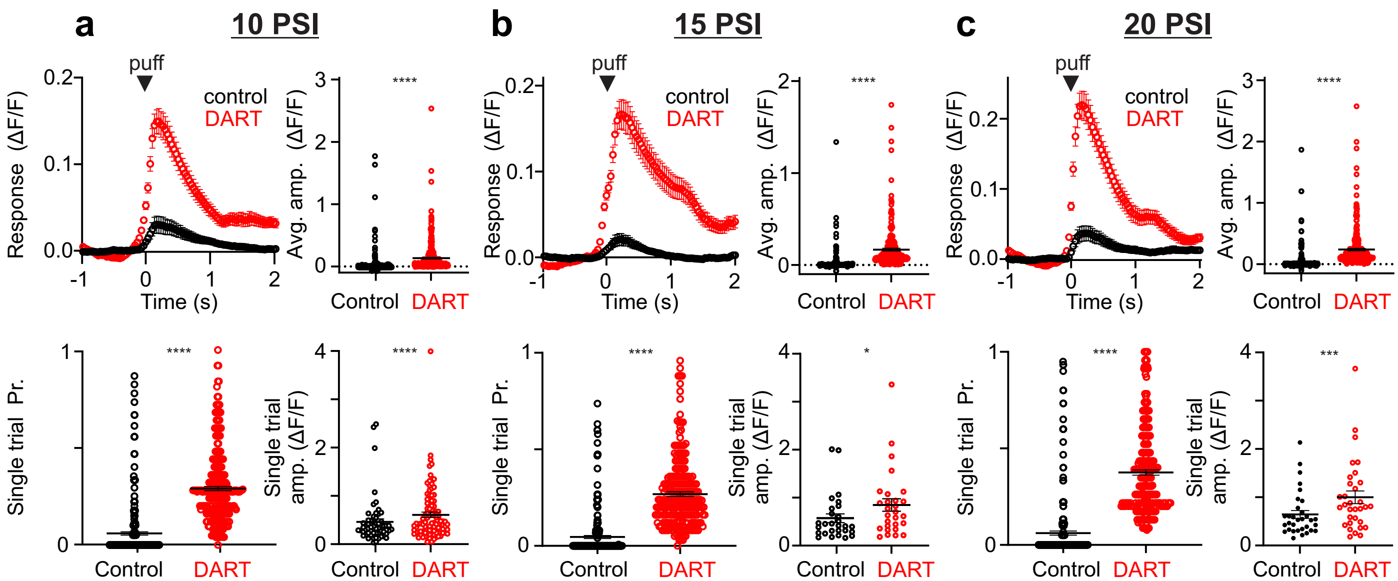
a. Example confocal images from a representative mouse expressing GCaMP6f (left) and HTP (middle) and their intersection (right) in the granule cell layer (GcL). Arrows note sparse labeling of Purkinje cells, which were excluded from analysis. **b.** Same as **a**, for a different representative mouse infused with Alexa647.1$^{DART.2}$ (cyan, middle right) and intersection of all three labels (right). Histology (GCaMP6f and HTP labeling) was collected for all experimental animals. Alexa647.1$^{DART.2}$ was also infused for all eyelid conditioning experiments, and a subset of imaging experiments.



Supplementary Figure 2

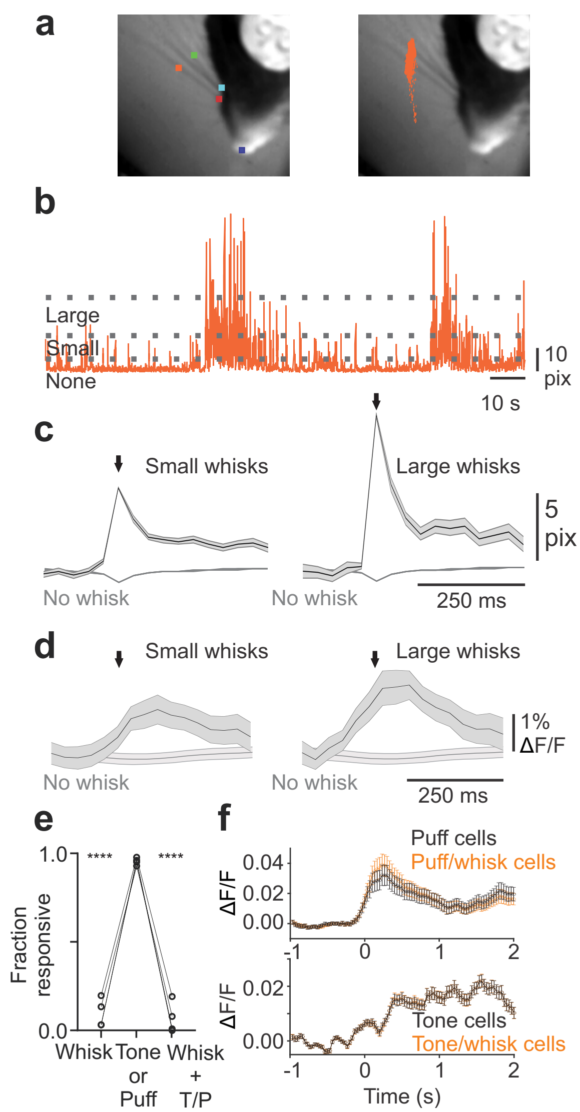
Supplementary Figure 2

a. Top left, mean time course of responses during 1 kHz, 68 dB tone presentation before (black) and after (red) DART infusion. Error is SEM across cells. Top right, mean response amplitudes for individual cells before (black) and after (red) DART infusion. Black lines are mean ± SEM across cells. Bottom left, same as top right, for response probability. Bottom right, same as top right, for mean responses on all trials with significant responses (n = 689 cells, n = 458 for single values; 6 mice). **b.** Same as **a**, for 1 kHz, 72 dB tones (n = 541 cells, n = 296 for single trial values; 5 mice). **c.** Same as **a**, for 5 kHz, 68 dB tones (n = 381 cells; n = 184 for single trial values; 3 mice). **d.** Same as **a**, for 5 kHz, 72 dB tones (n = 493 cells, n = 251 for single trial values; 3 mice). **e.** Same as **a**, for 10 kHz, 68 dB tones (n = 638 cells, n = 409 for single trial values; 6 mice). **f.** Same as **a**, for 10 kHz, 72 dB tones (n = 566 cells; n = 292 for single trial values; 3 mice). *$p<0.05$, ****$p<0.0001$, paired t-test (**a-f**).



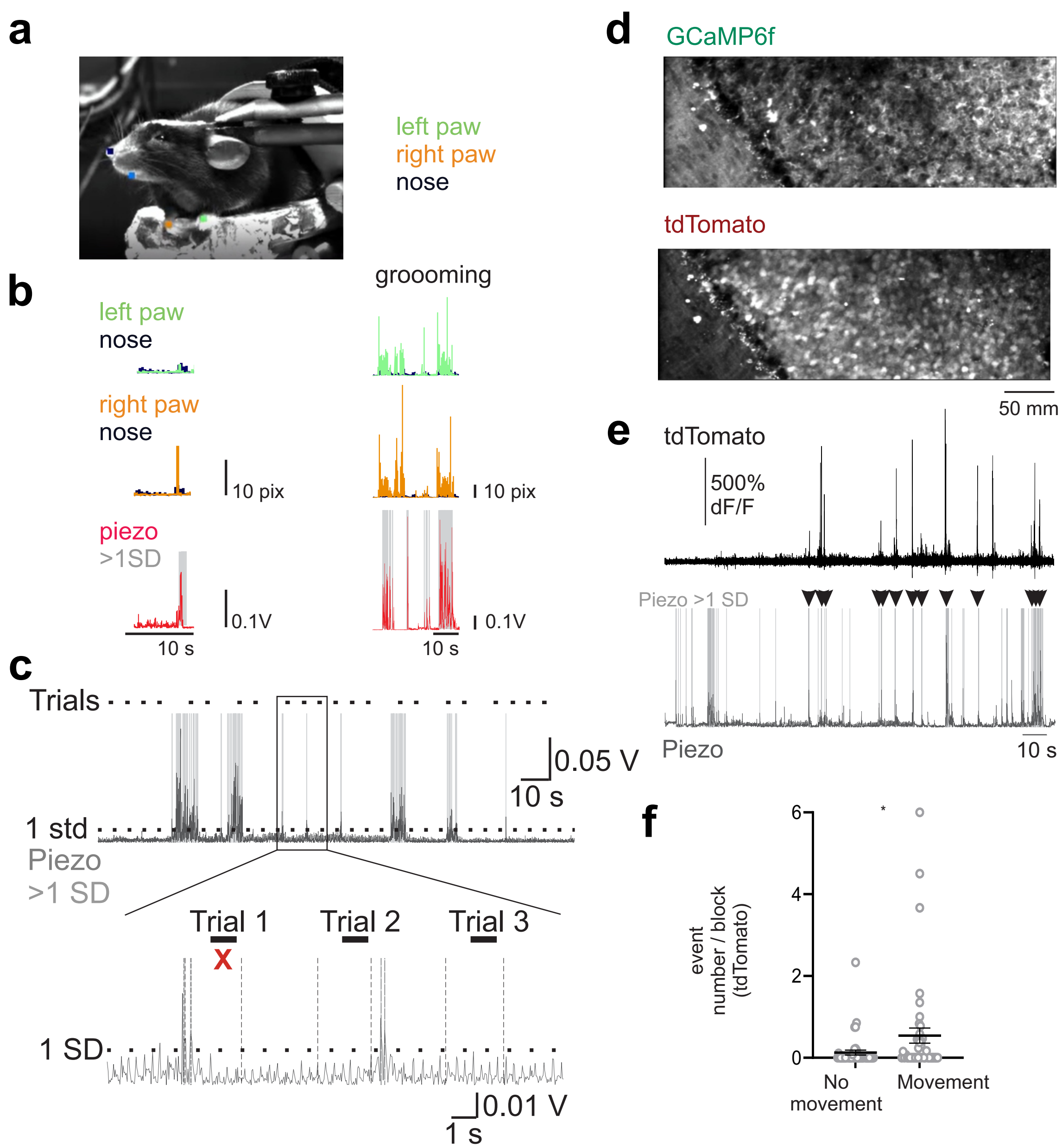
Supplementary Figure 3

a. Top left, mean time course of responses to a 10 PSI air puff before (black) and after (red) DART infusion. Error is SEM across cells. Top right, mean response amplitudes for individual cells before (black) and after (red) DART infusion. Black lines are mean ± SEM across cells. Bottom left, same as top right for response probability. Bottom right, same as top right for mean responses on all trials with significant responses (n = 283 cells; n = 58 for single trial values; 6 mice). **b.** Same as **a**, for 20 PSI air puff (n = 234 cells, n=28 for single trial values; 3 mice). **c.** Same as **a**, for 30 PSI air puff (n = 261 cells, n = 32 for single trial values; 3 mice). ****p*<0.001, *****p*<0.0001, paired t-test (**a-c**).



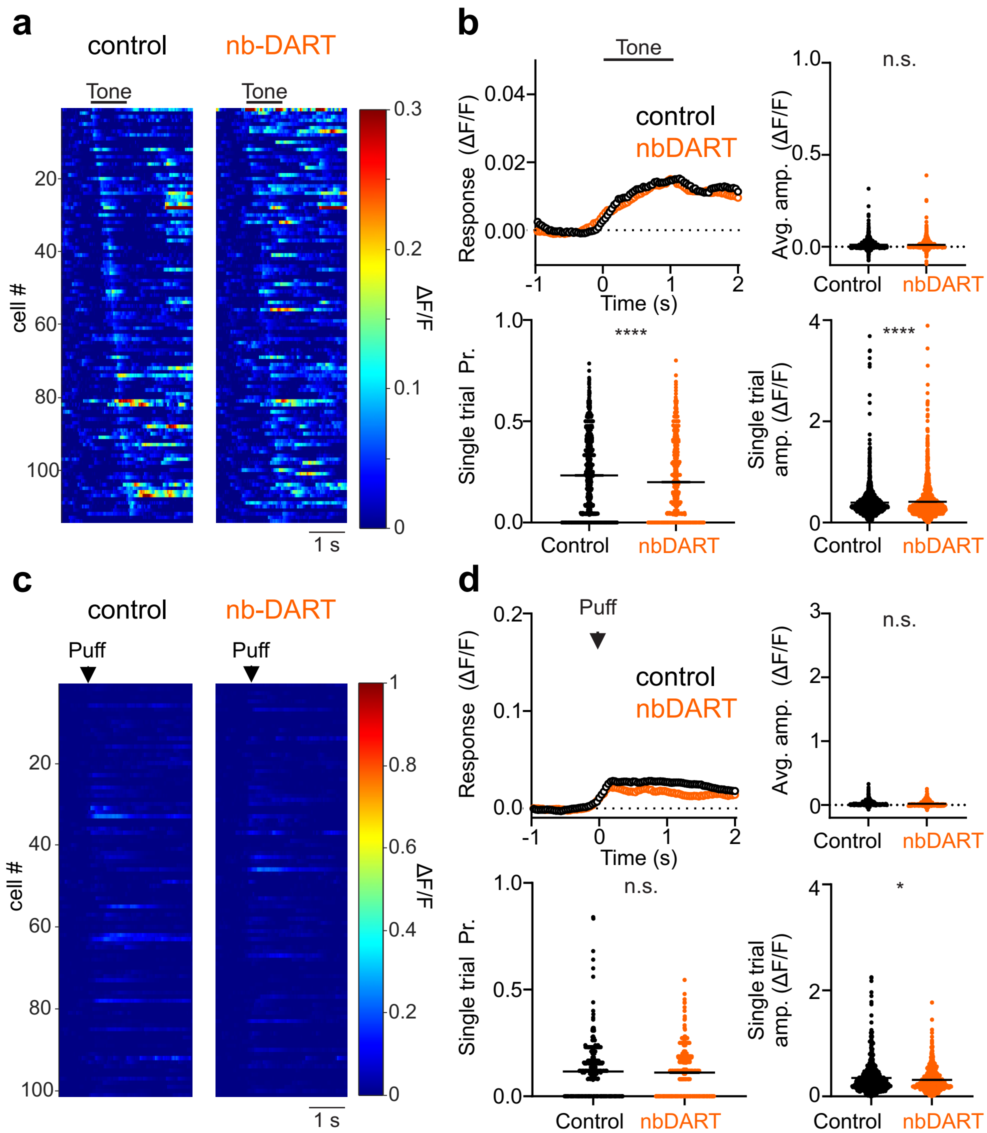
Supplementary Figure 4

a. Left, example image from a representative mouse with features labeled for automated identification of two whisker root (light blue and red) and midpoints (orange and green), as well as a stationary feature (snout, dark blue) used to estimate tracking jitter. Right, image from **a** with x and y coordinates for anterior whisker midpoint for all frames overlaid. **b.** Time course of displacement (in pixels) of whisker midpoint labeled in **a**. Dashed lines indicate thresholds used to segregate whisks according to magnitude. **c.** Left, mean ± SEM of low amplitude whisks occurring between trials and in the absence of body movement during an example experiment (n = 816 whisks). Mean ± SEM of frames without whisker movement plotted behind. Right, same for high amplitude whisks measured during the same experiment (n=287 whisks). **d.** Left, mean ± SEM fluorescence of cells modulated during small whisks and fluorescence of the same cells during frames without movement (n = 16 cells). Right, same for independently identified cells modulated during large whisks (n = 16 cells). Cells had the same identity. **e.** Fraction of the total responsive cells from 4 mice with significant activity during whisking, a sensory stimulus (auditory or somatosensory), or both; RM ANOVA: p = 0.0013, Tukey's multiple comparison's test: stim v. whisk: p = 0.0025, stim v. w+s: p = 0.0036, whisk v. w+s: p = 0.5782). **f.** Top, Time course of mean responses to a somatosensory stimulus for all cells significantly responsive to the stimulus (orange; n=366 cells) and cells significantly responsive to the stimulus not modulated during whisking (black; n = 306 cells; paired t-test of mean during stimulus window: control = 0.0343 ± 0.0066, DART = 0.0278 ± 0.0065, p = 0.4864). Error is SEM. Bottom, same as top for cells significantly responsive to an auditory stimulus (orange: n = 432 cells; black: n = 424; paired t-test of mean during stimulus window: control = 0.0123 ± 0.0015, DART = 0.0122 ± 0.0014, p = 0.9686).



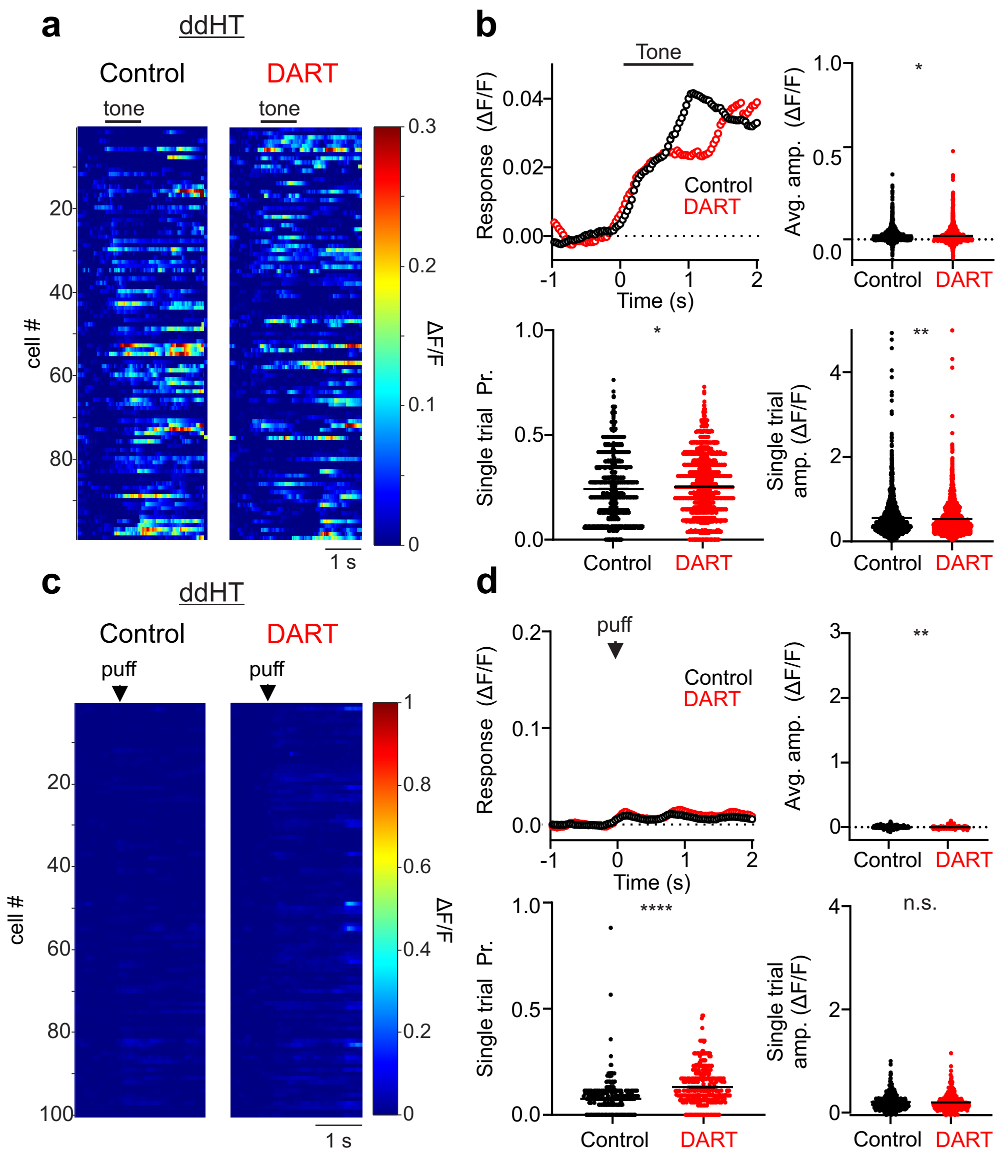
Supplementary Figure 5

a. Example image from a representative mouse following automated identification of left (green) and right (orange) paws, nose (dark blue) and mouth (light blue). **b.** Time course of displacement (in pixels) of left paw and nose (top), right paw and nose (middle) during left paw lift (left) and grooming (right). Bottom, simultaneous motion measurement using a piezoelectric sensor under the mouse's torso. Shaded gray regions reflect motions greater than 1 standard deviation that were discarded. **c.** Example raw piezo sensor trace at low (top) and high (bottom) temporal resolution. Dashes at top of each trace indicate times of auditory or somatosensory stimulus presentation; dashed line indicates 1 standard deviation threshold. Shaded regions indicate epochs that exceed this threshold; red x indicates trial that was discarded. **d.** Example field of view with GCaMP6f (top) and tdTomato (bottom) expression. **e.** Time course of changes in fluorescence in the tdTomato channel (top) after x-y motion correction and fluctuations in the piezo sensor. Note that changes in tdTomato fluorescence correspond with periods of animal movement (arrowheads), likely corresponding to uncorrected effects of z-motion. **f.** Quantification of the mean number of tdTomato events detected during blocks in which the mouse was stationary or moving ($n = 44$ blocks; $p = 0.00177$, paired t-test). Error is SEM across blocks.



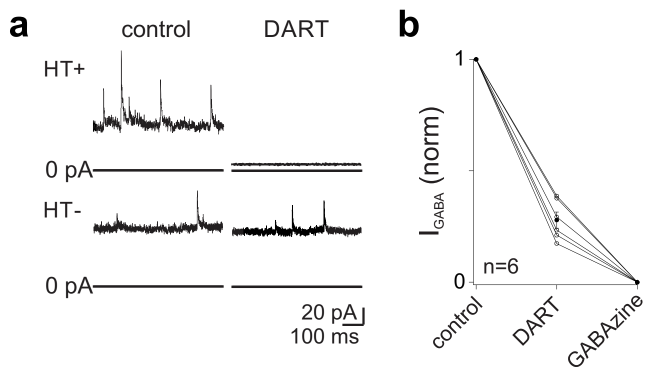
Supplementary Figure 6

a. Heatmap of average responses to auditory stimuli in control conditions (left) and after infusion of **gabazine.1**[nbDART] (nbDART, right) in an example mouse with HTP expression. **b.** Top left, mean time course of responses during tone presentation before (black) and after (orange) nbDART infusion. Error is SEM across cells. Top right, mean response amplitudes for individual cells before (black) and after (orange) nbDART infusion. Black lines are mean ± SEM across cells. Bottom left, same as top right, for response probability. Bottom right, same as top right, for mean responses on all trials with significant responses (n = 2044 cells, n = 1114 for single trial values; 3 mice). **c-d.** Same as **a-b**, for responses to somatosensory air puffs (n = 770 cells, n = 259 for single trial values; 3 mice). **p*<0.05, *****p*<0.0001, paired t-test (**b,d**).



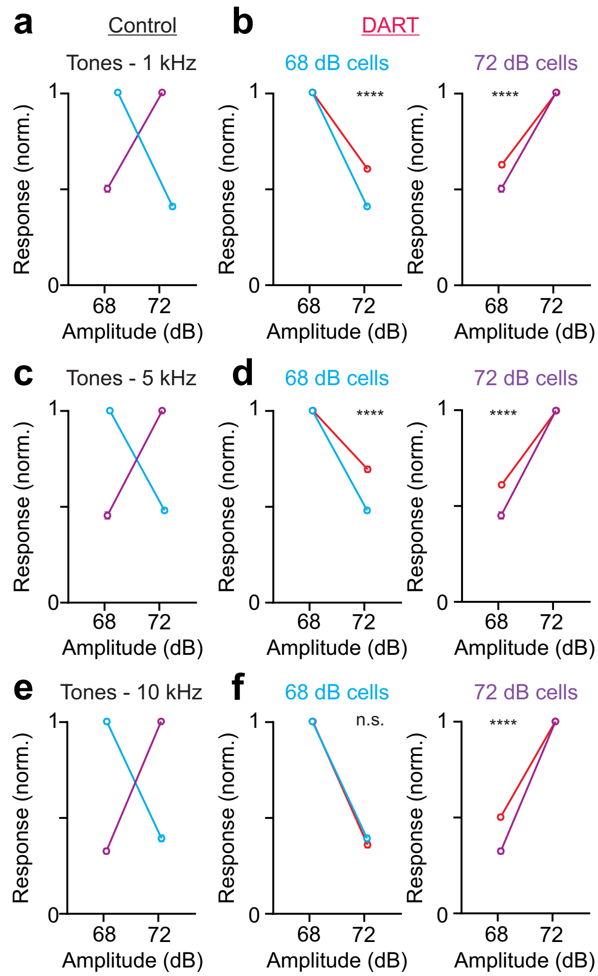
Supplementary Figure 7

a. Heatmap of average responses to auditory stimuli in control conditions (left) and after infusion of DART (right) in an example mouse with ddHTP expression. **b.** Top left, mean time course of responses during tone presentation before (black) and after (red) DART infusion. Error is SEM across cells. Top right, mean response amplitudes for individual cells before (black) and after (red) DART infusion. Black lines are mean ± SEM across cells. Bottom left, same as top right, for response probability. Bottom right, same as top right, for mean responses on all trials with significant responses (n = 1853, n = 1723 for single trial values; 2 mice). **c-d.** Same as **a-b**, for responses to somatosensory air puffs (n = 443 cells, n = 266 for single trial values; 2 mice). **p*<0.05, ***p*<0.01, *****p*<0.0001, paired t-test (**b,d**).



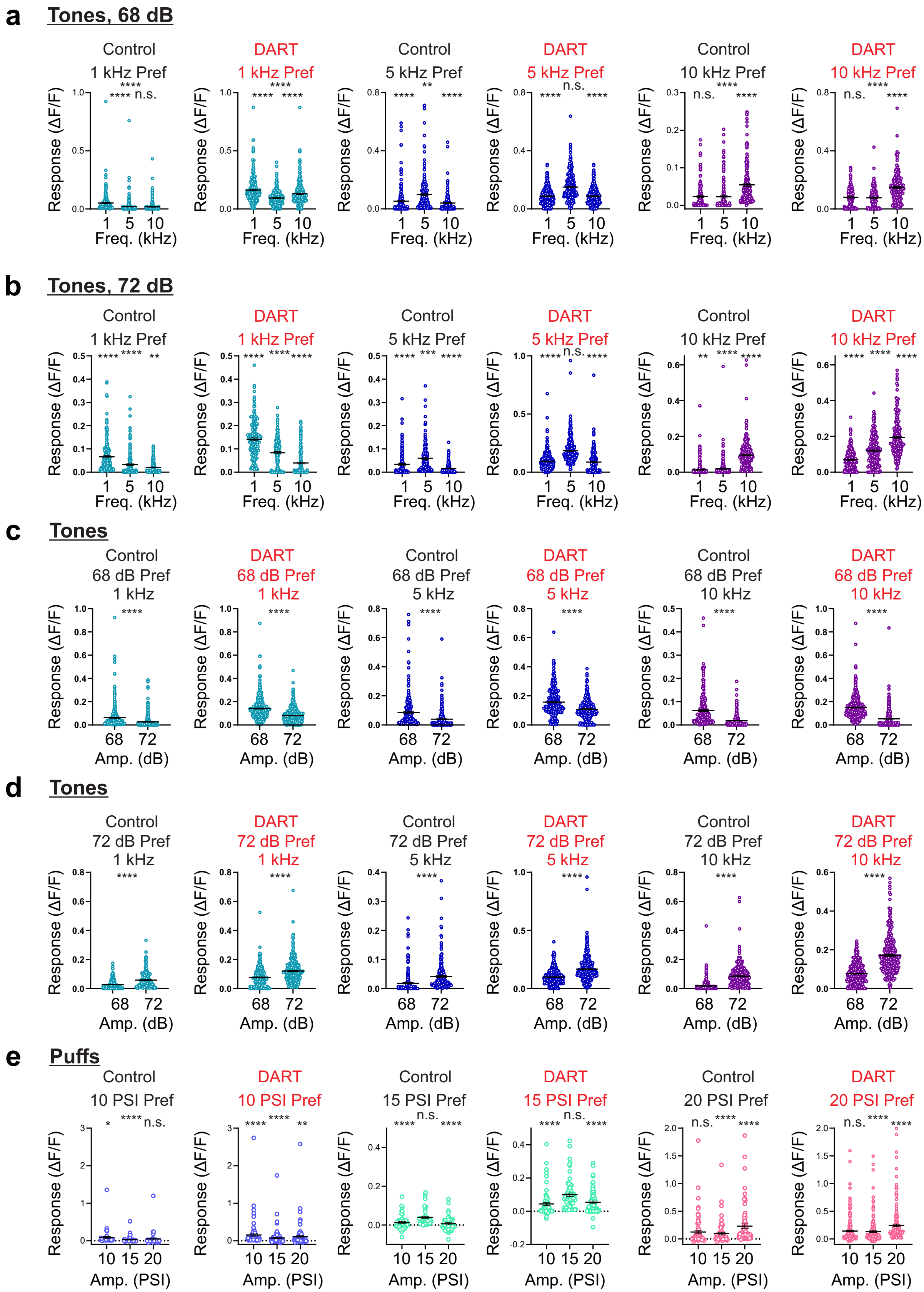
Supplementary Figure 8

a. Top, Example voltage-clamp recording at 0 mV from a granule cell expressing HTP. Wash-in of 100 nM gabazine.1$^{DART.2}$ (10x lower than *in vivo* infusions) blocks GABAergic inhibition, both spontaneous IPSCs and the tonic component of GABAergic inhibition. Bottom, the same concentration of DART does not block inhibition in a granule cell that does not express HTP. **b.** Across HTP expressing cells (n=6 cells), 100 nM gabazine.1$^{DART.2}$ blocked the total GABAergic current (sIPSCs + tonic current) by 73 $\pm$ 4%. Error is SEM. All procedures for *in vitro* voltage-clamp measurements from granule cells were performed as described previously[44].



Supplementary Figure 9

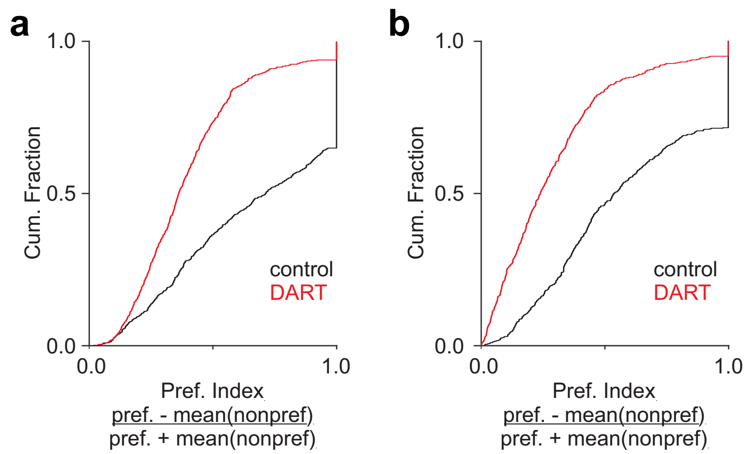
a. Normalized responses for cells that prefer 68 dB (cyan; n = 342) or 72 dB (purple; n = 207) in response to 1 kHz tones. Error is SEM across cells, n = 3 mice. **b.** Normalized responses for cells that prefer 68 dB (left) or 72 dB (right) in control (cyan/purple) and after DART infusion (red; 68 dB: n = 390 cells, 72 dB: n = 323 cells). Data is the same as in **a** for control conditions. **c-d.** Same as **a-b**, for 5 kHz tones: control, 68 dB: n = 293, 72 dB: n = 213; DART, 68 dB: n = 220, 72 dB: n = 469 **e-f.** Same as **a-b**, for 10 kHz tones: control, 68 dB: n = 241, 72 dB: n = 367; DART, 68 dB: n = 381, 72 dB: n = 344. ****p<0.0001, paired t-test (**b,d,f**).



Supplementary Figure 10

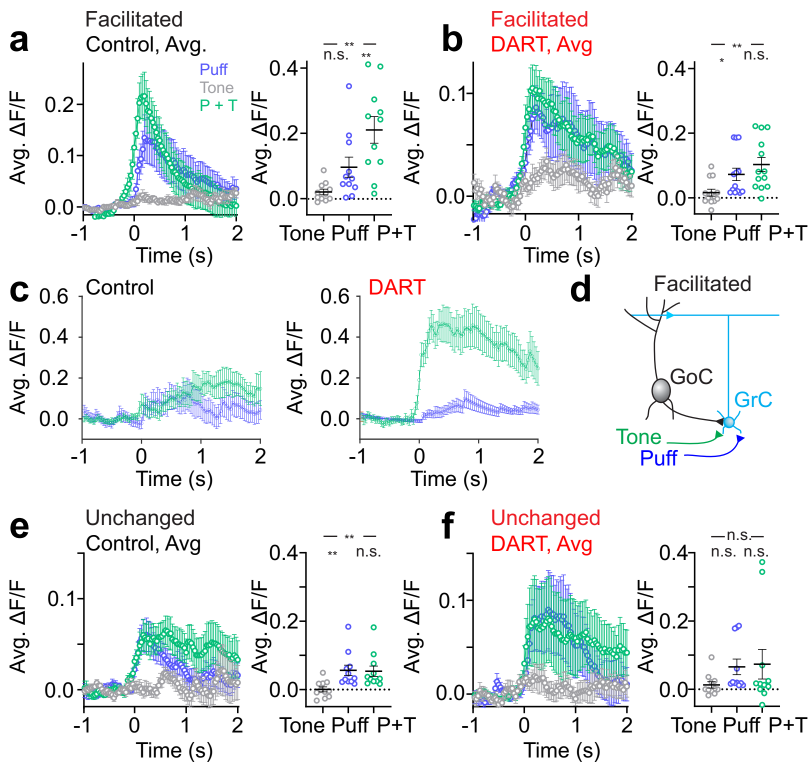
Supplementary Figure 10

a. Left, maximum responses (ΔF/F) for cells significantly responsive to any 68 dB tones that prefer 1 kHz in control (n = 227 cells) and DART (n = 250 cells) conditions. n = 3 mice for tone preferences. Middle, same as left for cells that prefer 5 kHz: control: n = 211 cells; DART: n = 297 cells. Right, same as left for cells that prefer 10 kHz: control: n = 166 cells; DART: n = 169 cells. **b.** Same as a for cells significantly responsive to any 72 dB tones: 1 kHz, control: n = 184 cells, DART: n = 170 cells; 5 kHz, control: n = 141 cells, DART: n = 331 cells; 10 kHz, control: n = 313 cells, DART: n = 224). **c.** Left, maximum responses (ΔF/F) for cells significantly responsive to 1 kHz at any amplitude that prefer 68 dB in control (n = 342 cells) and DART (n = 207 cells) conditions. Middle, same as left for cells responsive to 5 kHz that prefer 68 dB in control (n = 293 cells) and DART (n = 220 cells) conditions. Right, same as left for cells responsive to 10 kHz that prefer 68 dB in control (n = 241 cells) and DART (n = 381 cells) conditions. **d.** Same as c for cells that prefer 72 dB: 1 kHz, control: n=207 cells, DART: n = 323 cells; 5 kHz, control: n = 213 cells, DART: n = 469 cells; 10 kHz, control: n = 367 cells, DART: n = 344 cells. **e.** Left, maximum responses (ΔF/F) for somatosensory stimulus responsive cells that prefer 10 PSI in control (n = 45 cells) and DART (n = 100 cells) conditions. n = 3 mice for puff preferences. Middle, same as left for cells that prefer 15 PSI: control: n = 47 cells, DART: n = 69 cells. Right, same as left for cells that prefer 20 PSI: control: n = 74 cells, DART: n = 202 cells. Error bars are SEM. *p<0.05, **p<0.01, ****p<0.0001, RM ANOVA (**a,b,e**), paired t-test (**c**).



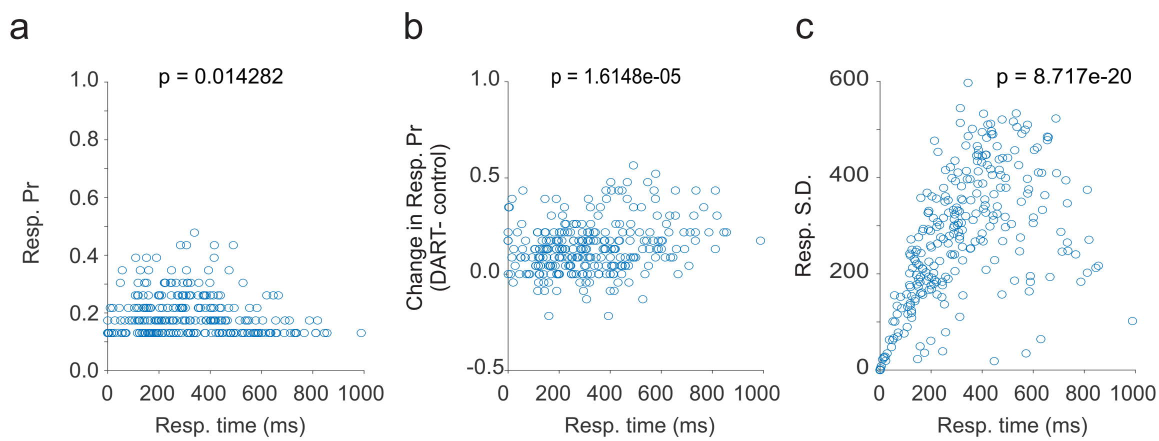
Supplementary Figure 11

a. Cumulative fraction of granule cells responding to 72 dB tones show a larger fraction with a preference index of 1 (perfect selectivity) in control conditions as compared to DART (n = 636). **b.** Same as **a.**, for granule cells responsive to 68 dB tones (n = 592).



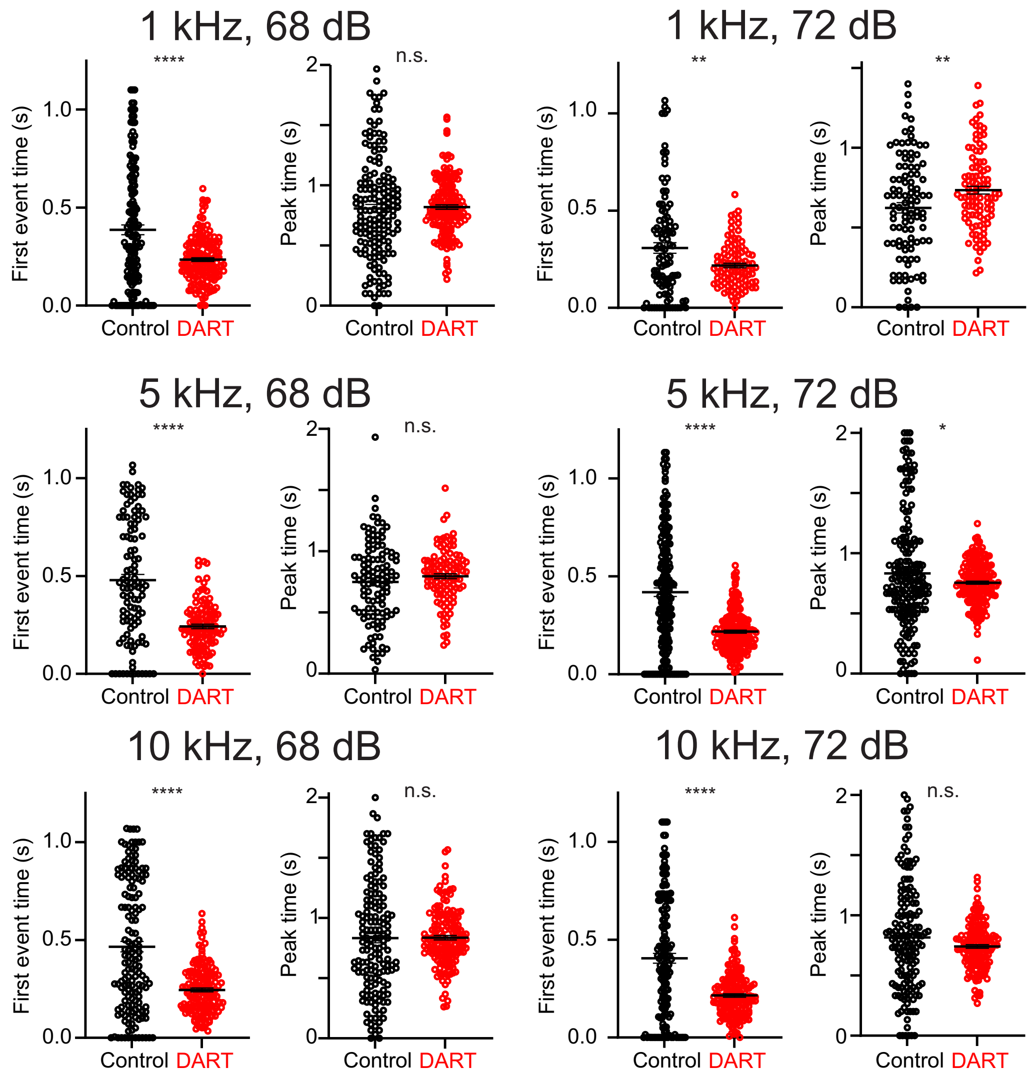
Supplementary Figure 12

a. Left, average time course of responses to uni- (tone: gray; puff: blue) and multisensory (tone + puff: green) stimuli for facilitated cells (subset matched to DART, n = 13 cells). Error is SEM across cells, n = 3 mice. Right, amplitude of responses to tone, puff and tone + puff for all facilitated cells. **b.** Same as **a**, after DART infusion. **c.** Example cell that was modestly facilitated by the tone+puff (green) as compared to the puff alone (blue) in control conditions (left, puff n = 17 trials, puff+tone n = 24 trials), and facilitation was greatly enhanced after DART infusion (right, puff n=16 trials, puff+tone n=24 trials). Error bars are SEM across trials. **d.** Proposed cerebellar circuit mediating facilitation. **e-f.** Same as **a-b**, for unchanged cells (n = 11 cells). Note that DART does not change the integration for these cells. *p<0.05, **p<0.01, RM ANOVA with Tukey's multiple comparisons (**a,b,e,f**).



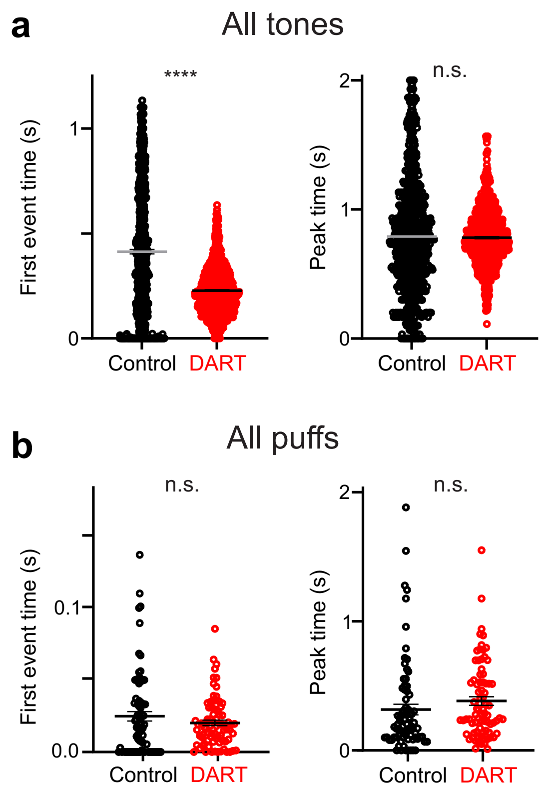
Supplementary Figure 13

a. Scatter of response time and probability for cells with 2 or more significant trials (n = 275 cells) This relationship has a significant negative slope from a linear regression (using the function lmfit in Matlab). **b.** Same as **a**, for change in response probability after DART infusion (n = 275 cells). This relationship has a significant positive slope. **c.** Same as **a**, for response standard deviation (S.D). This relationship has a significant positive slope.



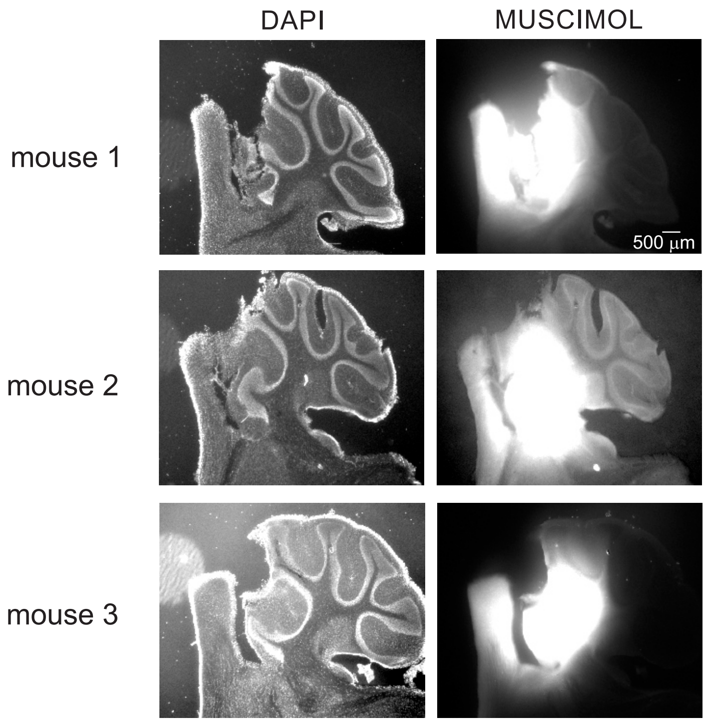
Supplementary Figure 14

Mean first (left) and peak (right) event times of significant trials before (black) and after (red) DART infusion for all individual cells with significant trial responses in both conditions. Overall mean ± SEM across cells overlaid. 1 kHz, 68 dB: n = 166 cells; 1 kHz, 72 dB: n = 101 cells; 5 kHz, 68 dB: n = 110 cells; 5 kHz, 72 dB: n = 188 cells; 10 kHz, 68 dB: n = 154 cells; 10 kHz, 72 dB: n = 157 cells; 3 mice. **p*<0.05, ***p*<0.01, *****p*<0.0001, paired t-test.



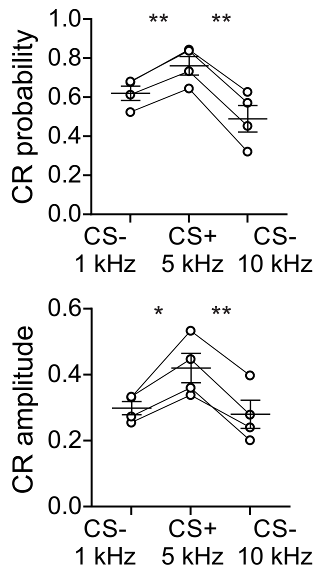
Supplementary Figure 15

a. Mean first (left) and peak (right) event times of significant trials before (black) and after (red) DART infusion for all individual cells with significant trial responses to auditory stimuli in both conditions. Overall mean ± SEM across cells overlaid (n = 876 cells, 3 mice). **b.** Same as **a**, for responses to air puffs (n = 104 cells, 3 mice). ****p<0.0001, paired t-test (**a-b**).



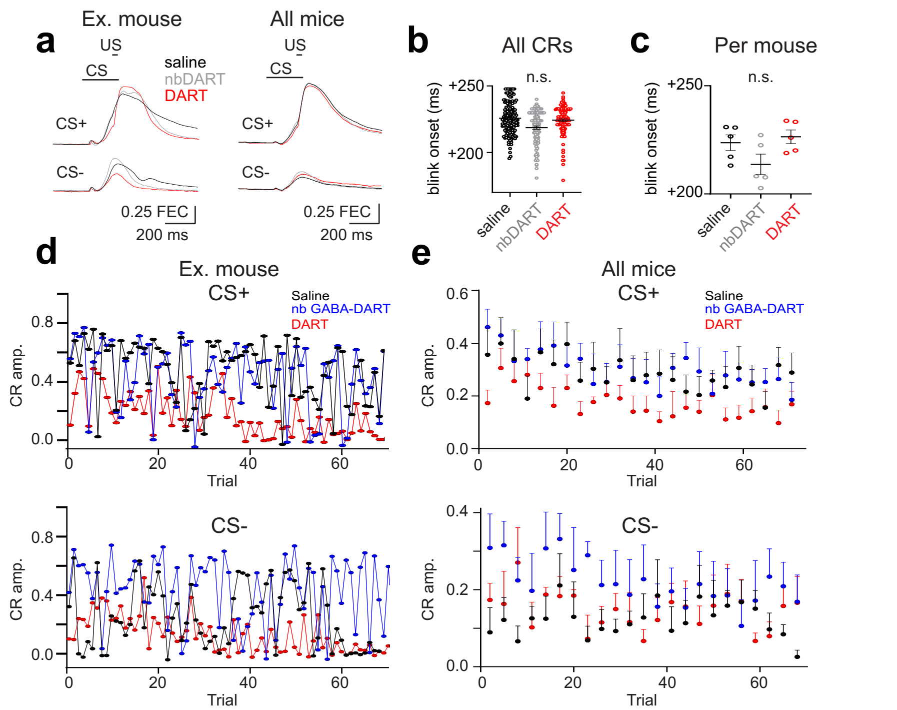
Supplementary Figure 16

Example widefield images of DAPI (left) and muscimol (right) fluorescence from three mice. Muscimol injection was targeted at the floor of the primary fissure, spreading outward to include the anterior interpositus.



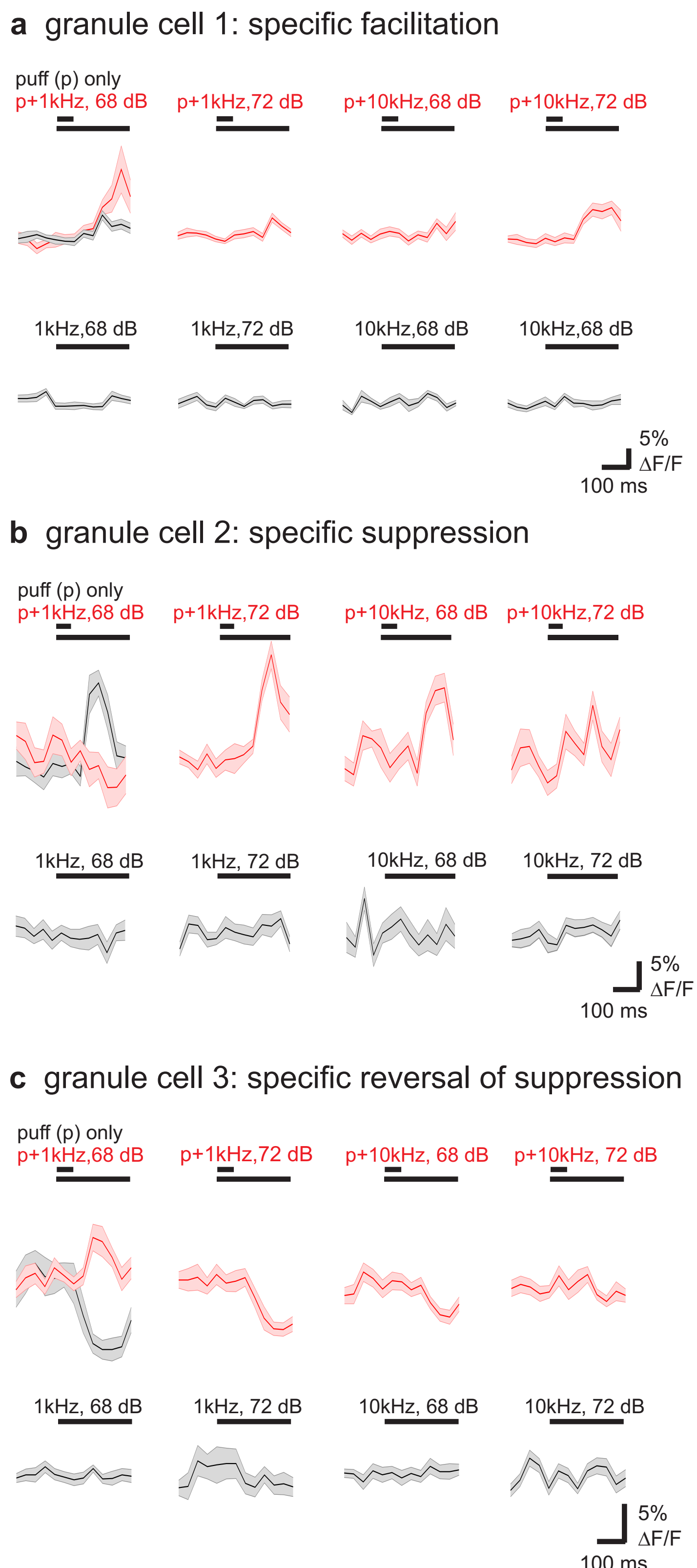
Supplementary Figure 17

Individual mean CR probability (top) and amplitude (bottom) during CS+ trials and CS- trials with either a 1 kHz or 10 kHz tone. Overall mean ± SEM across mice overlaid (n = 4 mice). **p*<0.05, ***p*<0.01, RM ANOVA with Dunnett's multiple comparisons.



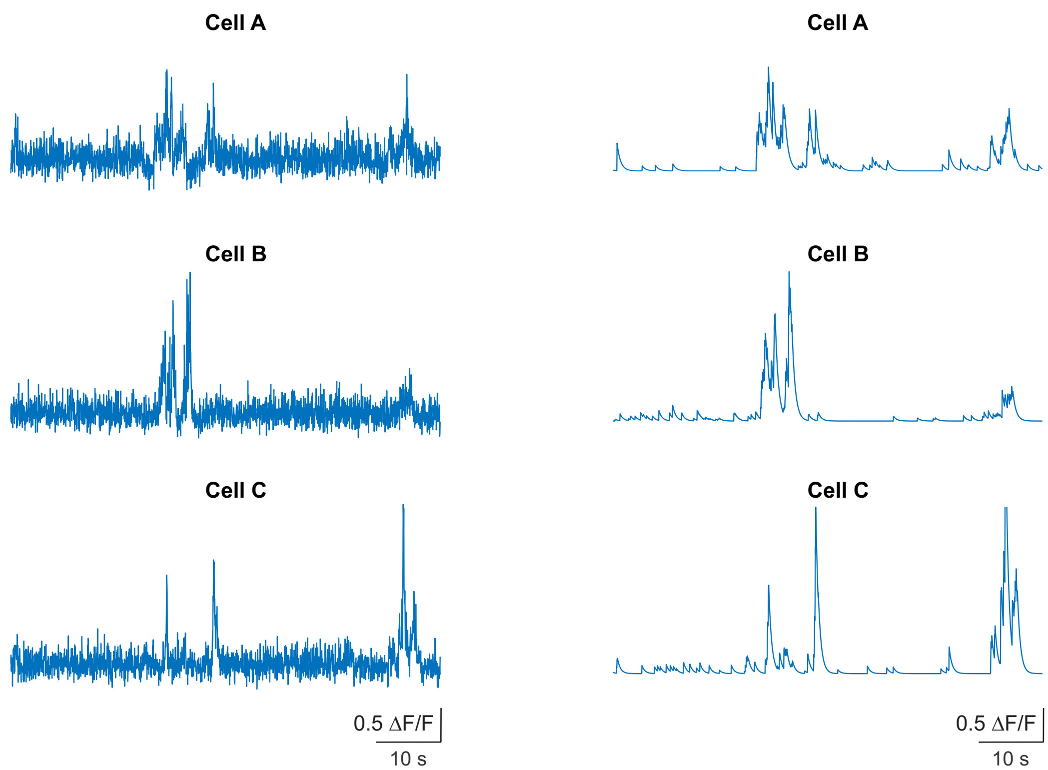
Supplementary Figure 18

a. Average time courses of fractional eyelid closure (FEC) in response to conditioned stimuli for an example mouse (left) and across mice (right) that were either paired (CS+, top) or unpaired (CS-, bottom) with an unconditioned stimulus (US) after saline (black), nbDART (gray) or DART (red) infusion **b.** Time of blink onset for all CRs in each condition for all CRs across mice (saline: n = 578 trials, nb-DART: n = 327, DART: n = 339; RM ANOVA *p*<0.0001, Tukey's multiple comparison's test: saline v. DART: p = 0.5837, saline v. nbDART: *p*<0.0001, nbDART v. DART: p = 0.0015). Overall mean ± SEM across CRs overlaid. **c.** Summary of average time of blink onset for each mouse individually across conditions. Overall mean ± SEM across mice (n = 5) overlaid (p = 0.2690, RM ANOVA). **d.** CR amplitudes across trials for an example mouse for CS+ trials (top) and CS- trials (bottom) in sessions following infusion of saline (black), nb-DART (blue), or DART (red). **e.** Same as **d**, averaged across mice (n = 5). Error bars are +/- SEM.



Supplementary Figure 19

a. Example granule cell with a sensory response that was selectively facilitated by a 1 kHz, 68 dB tone. Top row, puff alone (black), puff + tune (red). Bottom row, tone alone (black). Shaded error is SEM across trials. **b.** Example granule cell with a sensory response that was selectively suppressed by a 1 kHz, 68 dB tone. Shaded error is SEM across trials. **c.** Example granule cell with a sensory response that was selectively relieved of suppression by a 1 kHz, 68 dB tone. Shaded error is SEM across trials.



Supplementary Figure 20

Raw (left) and processed (right) time courses for three example cells. Following demixing to isolate the time courses from regions of interest corresponding to individual granule cells (left), the CaImAn analysis pipeline utilizes whitening steps to isolate calcium transients (right) as described previously[80].

Supplemental Table 1

Figure	Panel	Test	p-Value
1	E	Paired t-test	1.00E-15
	F	Paired t-test	1.00E-15
	G	Paired t-test	8.08E-11
	J	Paired t-test	1.00E-15
	K	Paired t-test	1.00E-15
	L	Paired t-test	9.16E-09
2	A, left	Paired t-test	1.83E-07
	A, right	Paired t-test	5.58E-03
	B, left	Paired t-test	1.83E-07
	B, right	Paired t-test	5.58E-03
3	B, left, 1kHz->5kHz	RM ANOVA	1.00E-15
	B, left, 5kHz->10kHz	RM ANOVA	1.16E-03
	B, left, 1kHz->10 kHz	RM ANOVA	1.00E-15
	B, right, 1kHz->5kHz	RM ANOVA	1.00E-15
	B, right, 5kHz->10kHz	RM ANOVA	1.00E-15
	B, right, 1kHz->10 kHz	RM ANOVA	1.00E-15
	C, left, 1kHz->5kHz	RM ANOVA	4.20E-14
	C, left, 5kHz->10kHz	RM ANOVA	9.30E-14
	C, left, 1kHz->10 kHz	RM ANOVA	1.75E-04
	C, right, 1kHz->5kHz	RM ANOVA	4.68E-13
	C, right, 5kHz->10kHz	RM ANOVA	4.68E-13
	C, right, 1kHz->10 kHz	RM ANOVA	5.59E-01
	D, left, 1kHz->5kHz	RM ANOVA	1.88E-03
	D, left, 5kHz->10kHz	RM ANOVA	9.42E-13
	D, left, 1kHz->10 kHz	RM ANOVA	9.42E-13
	D, right, 1kHz->5kHz	RM ANOVA	5.20E-14
	D, right, 5kHz->10kHz	RM ANOVA	4.70E-14
	D, right, 1kHz->10 kHz	RM ANOVA	4.70E-14
	F, left, 5kHz	Paired t-test	2.81E-03
	F, left, 10kHz	Paired t-test	3.13E-03
	F, middle, 1kHz	Paired t-test	3.23E-01
	F, middle, 10kHz	Paired t-test	1.20E-02

	F, right, 1kHz	Paired t-test	5.00E-15
	F, right, 5kHz	Paired t-test	1.00E-15
	H, left, 5kHz	Paired t-test	1.00E-15
	H, left, 10kHz	Paired t-test	1.00E-15
	H, middle, 1kHz	Paired t-test	1.31E-08
	H, middle, 10kHz	Paired t-test	1.58E-08
	H, right, 1kHz	Paired t-test	1.03E-01
	H, right, 5kHz	Paired t-test	1.49E-02
	J, left, 15 PSI	Paired t-test	2.40E-01
	J, left, 20 PSI	Paired t-test	1.45E-03
	J, middle, 10 PSI	Paired t-test	3.82E-02
	J, middle, 20 PSI	Paired t-test	1.33E-04
	J, right, 10 PSI	Paired t-test	8.78E-03
	J, right, 15 PSI	Paired t-test	6.28E-03
4	A	Paired t-test	8.52E-08
	B	Paired t-test	2.66E-02
	C	Paired t-test	4.93E-07
	D	Paired t-test	3.46E-01
	E, left	one-sided Wald test	4.14E-07
	E, Middle	one-sided Wald test	4.00E-03
	E, Right	one-sided Wald test	1.17E-06
5	B, Right, tone->puff	RM ANOVA	6.75E-01
	B, Right, puff+tone->puff	RM ANOVA	9.26E-09
	B, Right, puff+tone->tone	RM ANOVA	6.89E-09
	D, Right, tone->puff	RM ANOVA	9.59E-10
	D, Right, puff+tone->puff	RM ANOVA	8.95E-01
	D, Right, puff+tone->tone	RM ANOVA	9.80E-10
	E, Right, tone->puff	RM ANOVA	1.60E-01
	E, Right, puff+tone->puff	RM ANOVA	1.00E-15
	E, Right, puff+tone->tone	RM ANOVA	1.00E-15
	F, Right, tone->puff	RM ANOVA	2.91E-04
	F, Right, puff+tone->puff	RM ANOVA	3.23E-04
	F, Right, puff+tone->tone	RM ANOVA	3.81E-01

	K	Z test	2.30E-03
6	E	KS test	5.31E-08
	F	KS test	9.86E-01
	H	Paired t-test	1.00E-15
7	G, top	Paired t-test	3.60E-03
	G, bottom	Paired t-test	4.00E-04
	H, top	Paired t-test	3.19E-02
	H, bottom	Paired t-test	3.14E-02
	I, top	Paired t-test	1.89E-01
	I, bottom	Paired t-test	2.52E-01
S2	A, Top, Right	Paired t-test	5.90E-14
	A, Bottom, Left	Paired t-test	1.00E-15
	A, Bottom, Right	Paired t-test	1.99E-02
	B, Top, Right	Paired t-test	1.00E-15
	B, Bottom, Left	Paired t-test	1.00E-15
	B, Bottom, Right	Paired t-test	4.47E-01
	C, Top, Right	Paired t-test	1.53E-11
	C, Bottom, Left	Paired t-test	1.00E-15
	C, Bottom, Right	Paired t-test	2.14E-01
	D, Top, Right	Paired t-test	1.00E-15
	D, Bottom, Left	Paired t-test	1.00E-15
	D, Bottom, Right	Paired t-test	1.39E-07
	E, Top, Right	Paired t-test	1.00E-15
	E, Bottom, Left	Paired t-test	1.00E-15
	E, Bottom, Right	Paired t-test	2.48E-07
	F, Top, Right	Paired t-test	1.00E-15
	F, Bottom, Left	Paired t-test	1.00E-15
	F, Bottom, Right	Paired t-test	1.16E-01
S3	A, Top, Right	Paired t-test	1.00E-15
	A, Bottom, Left	Paired t-test	1.00E-15
	A, Bottom, Right	Paired t-test	1.17E-05

	B, Top, Right	Paired t-test	1.00E-15
	B, Bottom, Left	Paired t-test	1.00E-15
	B, Bottom, Right	Paired t-test	1.71E-02
	C, Top, Right	Paired t-test	1.00E-15
	C, Bottom, Left	Paired t-test	1.00E-15
	C, Bottom, Right	Paired t-test	4.45E-04
S4	E	RM ANOVA	1.30E-03
	E, Stim -> whisk	RM ANOVA: with Tukey's multiple comparison	2.50E-03
	E, stim -> whisk+stim	RM ANOVA: with Tukey's multiple comparison	3.60E-03
	E, whisk -> whisk+stim	RM ANOVA: with Tukey's multiple comparison	5.78E-01
	F, Top	Paired t-test	4.86E-01
	F, Bottom	Paired t-test	9.69E-01
S5	F	Paired t-test	1.77E-03
S6	B, Top, Right	Paired t-test	2.96E-01
	B, Bottom, Left	Paired t-test	8.34E-06
	B, Bottom, Right	Paired t-test	1.02E-05
	D, Top, Right	Paired t-test	6.00E-02
	D, Bottom, Left	Paired t-test	3.66E-01
	D, Bottom, Right	Paired t-test	1.10E-02
S7	B, Top, Right	Paired t-test	1.18E-02
	B, Bottom, Left	Paired t-test	4.66E-02
	B, Bottom, Right	Paired t-test	1.90E-03
	D, Top, Right	Paired t-test	7.50E-03
	D, Bottom, Left	Paired t-test	1.00E-15
	D, Bottom, Right	Paired t-test	3.25E-01
S9	B, Left	Paired t-test	1.00E-15
	B, Right	Paired t-test	1.70E-07
	D, Left	Paired t-test	1.00E-15
	D, Right	Paired t-test	2.00E-15

	E, Left	Paired t-test	1.77E-01
	E, Right	Paired t-test	1.00E-15
S10	A, Left, Control, 1kHz->5kHz	RM ANOVA	1.00E-15
	A, Left, Control, 10kHz->5kHz	RM ANOVA	4.25E-01
	A, Left, Control, 1kHz->10 kHz	RM ANOVA	1.00E-15
	A, Left, DART, 1kHz->5kHz	RM ANOVA	2.60E-14
	A, Left, DART, 10kHz->5kHz	RM ANOVA	5.30E-14
	A, Left, DART, 1kHz->10 kHz	RM ANOVA	5.90E-14
	A, Middle, Control, 1kHz->5kHz	RM ANOVA	1.00E-15
	A, Middle, Control, 10kHz->5kHz	RM ANOVA	1.00E-15
	A, Middle, Control, 1kHz->10 kHz	RM ANOVA	2.02E-03
	A, Middle, DART, 1kHz->5kHz	RM ANOVA	7.09E-13
	A, Middle, DART, 10kHz->5kHz	RM ANOVA	7.09E-13
	A, Middle, DART, 1kHz->10 kHz	RM ANOVA	6.53E-01
	A, Right, Control, 1kHz->5kHz	RM ANOVA	7.67E-01
	A, Right, Control, 10kHz->5kHz	RM ANOVA	1.80E-14
	A, Right, Control, 1kHz->10 kHz	RM ANOVA	3.10E-14
	A, Right, DART, 1kHz->5kHz	RM ANOVA	5.92E-01
	A, Right, DART, 10kHz->5kHz	RM ANOVA	3.70E-14
	A, Right, DART, 1kHz->10 kHz	RM ANOVA	3.70E-14
	B, Left, Control, 1kHz->5kHz	RM ANOVA	1.00E-15
	B, Left, Control, 10kHz->5kHz	RM ANOVA	1.16E-03
	B, Left, Control, 1kHz->10 kHz	RM ANOVA	1.00E-15
	B, Left, DART, 1kHz->5kHz	RM ANOVA	1.00E-15
	B, Left, DART, 10kHz->5kHz	RM ANOVA	1.00E-15
	B, Left, DART, 1kHz->10 kHz	RM ANOVA	1.00E-15
	B, Middle, Control, 1kHz->5kHz	RM ANOVA	4.20E-14
	B, Middle, Control, 10kHz->5kHz	RM ANOVA	9.30E-14
	B, Middle, Control, 1kHz->10 kHz	RM ANOVA	1.75E-04
	B, Middle, DART, 1kHz->5kHz	RM ANOVA	4.68E-13
	B, Middle, DART, 10kHz->5kHz	RM ANOVA	4.68E-13
	B, Middle, DART, 1kHz->10 kHz	RM ANOVA	5.59E-01
	B, Right, Control, 1kHz->5kHz	RM ANOVA	1.88E-03
	B, Right, Control, 10kHz->5kHz	RM ANOVA	9.42E-13
	B, Right, Control, 1kHz->10 kHz	RM ANOVA	9.42E-13

	B, Right, DART, 1kHz->5kHz	RM ANOVA	5.20E-14
	B, Right, DART, 10kHz->5kHz	RM ANOVA	4.70E-14
	B, Right, DART, 1kHz->10 kHz	RM ANOVA	4.70E-14
	C, Left, Control	Paired t-test	1.00E-15
	C, Left, DART	Paired t-test	1.00E-15
	C, Middle, Control	Paired t-test	1.00E-15
	C, Middle, DART	Paired t-test	1.00E-15
	C, Right, Control	Paired t-test	1.00E-15
	C, Right, DART	Paired t-test	1.00E-15
	D, Left, Control	Paired t-test	1.00E-15
	D, Left, DART	Paired t-test	1.00E-15
	D, Middle, Control	Paired t-test	1.00E-15
	D, Middle, DART	Paired t-test	1.00E-15
	D, Right, Control	Paired t-test	1.00E-15
	D, Right, DART	Paired t-test	1.00E-15
	E, Left, Control, 10PSI->15PSI	RM ANOVA	1.58E-02
	E, Left, Control, 10PSI->20PSI	RM ANOVA	7.64E-10
	E, Left, Control, 15PSI->20PSI	RM ANOVA	4.65E-01
	E, Left, DART, 10PSI->15PSI	RM ANOVA	1.84E-09
	E, Left, DART, 10PSI->20PSI	RM ANOVA	2.23E-10
	E, Left, DART, 15PSI->20PSI	RM ANOVA	4.82E-03
	E, Middle, Control, 10PSI->15PSI	RM ANOVA	4.46E-08
	E, Middle, Control, 10PSI->20PSI	RM ANOVA	4.16E-01
	E, Middle, Control, 15PSI->20PSI	RM ANOVA	1.57E-05
	E, Middle, DART, 10PSI->15PSI	RM ANOVA	1.00E-15
	E, Middle, DART, 10PSI->20PSI	RM ANOVA	1.82E-01
	E, Middle, DART, 15PSI->20PSI	RM ANOVA	1.00E-15
	E, Right, Control, 10PSI->15PSI	RM ANOVA	6.37E-02
	E, Right, Control, 10PSI->20PSI	RM ANOVA	4.35E-09
	E, Right, Control, 15PSI->20PSI	RM ANOVA	3.77E-07
	E, Right, DART, 10PSI->15PSI	RM ANOVA	2.58E-01
	E, Right, DART, 10PSI->20PSI	RM ANOVA	4.80E-14
	E, Right, DART, 15PSI->20PSI	RM ANOVA	5.20E-14
S12	A, Right, tone->puff	RM ANOVA	3.11E-02
	A, Right, tone->puff+tone	RM ANOVA	4.17E-08

	A, Right, puff->puff+tone	RM ANOVA	6.33E-04
	B, Right, tone->puff	RM ANOVA	1.57E-02
	B, Right, tone->puff+tone	RM ANOVA	3.49E-03
	B, Right, puff->puff+tone	RM ANOVA	3.03E-01
	E, Right, tone->puff	RM ANOVA	1.14E-08
	E, Right, tone->puff+tone	RM ANOVA	1.69E-08
	E, Right, puff->puff+tone	RM ANOVA	9.77E-02
	F, Right, tone->puff	RM ANOVA	1.37E-01
	F, Right, tone->puff+tone	RM ANOVA	3.67E-01
	F, Right, puff->puff+tone	RM ANOVA	9.83E-01
S13	A	Linear regression	1.42E-02
	B	Linear regression	1.61E-05
	C	Linear regression	1.00E-15
S14	1kHz, 68dB, left	Paired t-test	3.19E-08
	1kHz, 68dB, right	Paired t-test	7.74E-01
	1kHz, 72dB, left	Paired t-test	2.40E-03
	1kHz, 72dB, right	Paired t-test	7.00E-03
	5kHz, 68dB, left	Paired t-test	1.00E-15
	5kHz, 68dB, right	Paired t-test	2.26E-01
	5kHz, 72dB, left	Paired t-test	6.14E-17
	5kHz, 72dB, right	Paired t-test	4.26E-02
	10kHz, 68dB, left	Paired t-test	9.87E-14
	10kHz, 68dB, right	Paired t-test	9.21E-01
	10kHz, 72dB, left	Paired t-test	3.79E-12
	10kHz, 72dB, right	Paired t-test	5.89E-02
S15	A, left	Paired t-test	7.38E-15
	A, right	Paired t-test	6.04E-01
	B, left	Paired t-test	2.31E-01
	B, right	Paired t-test	1.48E-01
S17	Top, 5kHz->1kHz	RM ANOVA	2.30E-03
	Top, 5kHz->10kHz	RM ANOVA	1.90E-03
	Bottom, 5kHz->1kHz	RM ANOVA	3.59E-02

	Bottom, 5kHz->10kHz	RM ANOVA	4.90E-03
S18	B, saline -> DART	RM ANOVA	5.84E-01
	B, saline -> nbDART	RM ANOVA	1.11E-05
	B, DART -> nbDART	RM ANOVA	1.50E-03
	C	RM ANOVA	9.34E-02