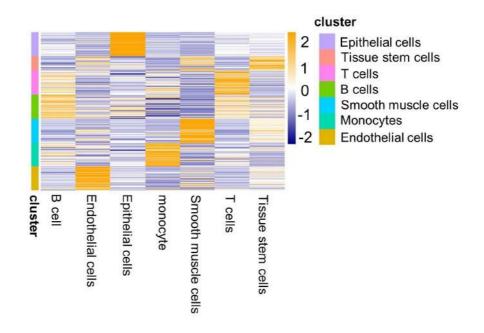
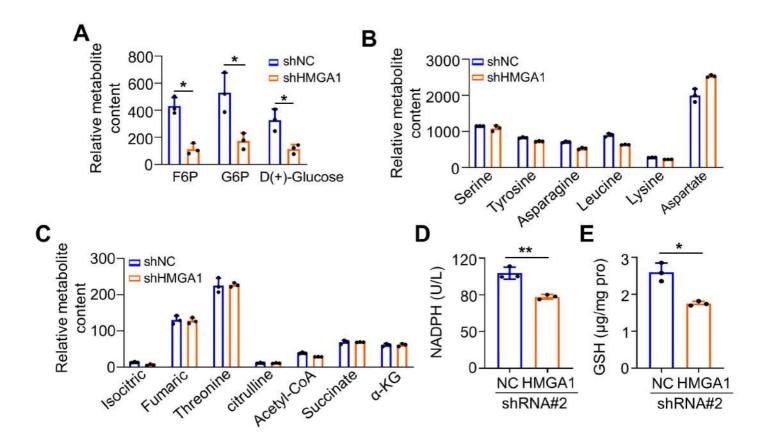
Supplementary Table S1

Gene/plasmids	Primers	Primer sequences
HMGA1-human	forward primer	CGAGAAAAGGACGGCACTGA
	reverse primer	TTGTGGTGGTTTTCCGGGTC
TKT-human	forward primer	GACAACCTTGTGGCCATTCT
	reverse primer	TCTGCTCAGCCATGTTTTTG
GAPDH-human	forward primer	AACGGATTTGGTCGTATTGG
	reverse primer	TTGATTTTGGAGGGATCTCG
HMGA1-shRNA		GAAGTGCCAACACCTAAGAGA
TKT-pcDNA 3.1	forward primer	GTGGTGGAATTCATGGAGAGCTACCACA
		AGCCTGACCAG
	reverse primer	TCTAGACTCGAGCTAGGCCTTGGTGATG
		AGGCCCCTCACA
HA-Flag-TKT	forward primer	GGACCGGTTCTAGAGCCACCATGGAGAG
		CTACCACAAGCC
	reverse primer	TCTGGAACATCGTATGGGTAGGCCTTGGT
		GATGAGGCCCC
HA-Flag-HMGA1	forward primer	GGTTCTAGAGCCACCATGAGTGAGTCGA
		GCTCGAA
	reverse primer	AGTCCTCGGAGGAGGAGCAGTACCCATA
		CGATGTTCCAGA
HA-Flag-Sp1	forward primer	GGACCGGTTCTAGAGCCACCATGAGCGA
		CCAAGATCACTC

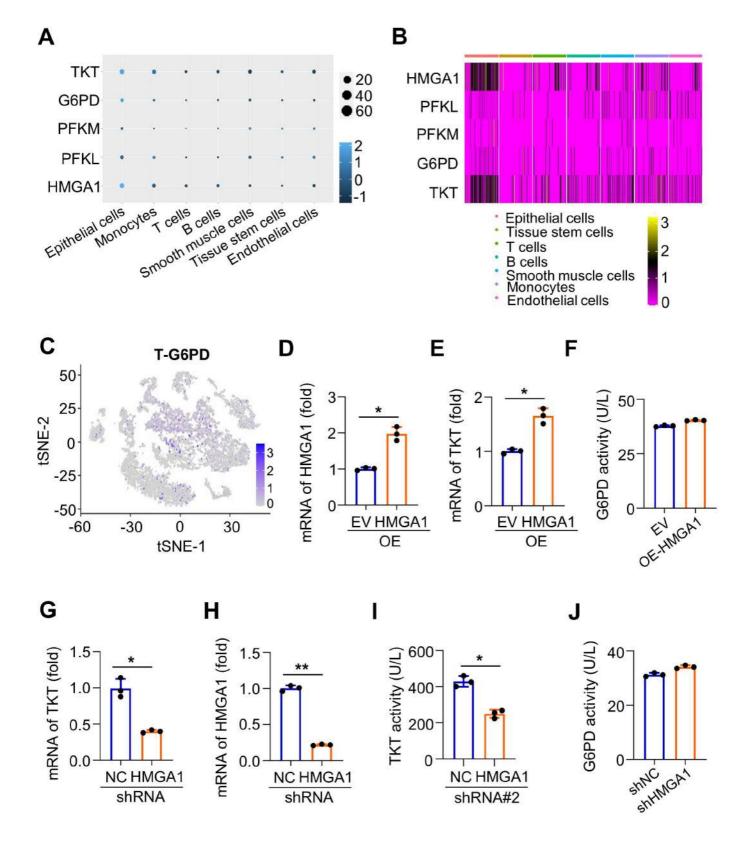
	reverse primer	TCTGGAACATCGTATGGGTAGAAGCCATT
		GCCACTGATAT
siRNA-TKT	forward primer	CCGGCAAAUACUUCGACAATTA
	reverse primer	UUGUCGAAGUAUUUGCCGGTT
TKT promoter-P1	forward primer	GGTACCGAGCTCTTTTTCCATTACTCCTA
		GGTCCCCAGA
	reverse primer	ACTTAGATCGCAGATTACAAATTATTAGG
		GCCTAGAAACA
	forward primer	GGTACCGAGCTCTTCAATAATCTCCCTAA
TKT promoter-P2		AAGTCAGTAG
TKT promoter-12	reverse primer	ACTTAGATCGCAGATTCGAAAGCCTCTCA
		GTCTCCCTCCT
	forward primer	GGTACCGAGCTCTTGACAAGTCCACGGA
TKT promoter-P3		GGACTGTGTGG
	reverse primer	ACTTAGATCGCAGAT
		CGCGAGCCCATCCCCCGCGCCACCA
ChIP-R1	forward primer	ACTGTGTGGAGGAGGGAGAC
	reverse primer	AGGCGCTGGGCCGCTGCGGA
ChIP-R2	forward primer	GGAGACTGAGAGGCTTTCGA
	reverse primer	GGGCCGCCTCCTGCCCGCTC
ChIP-R3	forward primer	CCCCAGGCGGGGCGGGCTG
	reverse primer	AGGCCGGGCGCGGGGG



Supplementary Fig. 1 The tSNE map of the single cell analysis in ESCCs and the scape was colored by cell subtypes and shown in Fig. 1A. The heatmap showed the expression levels of different cell subtype signatures in scRNA-seq data from the GEO database (GSE188900 dataset).

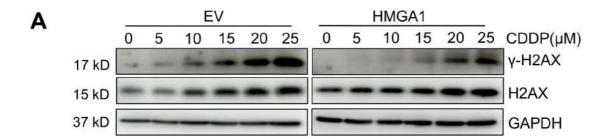


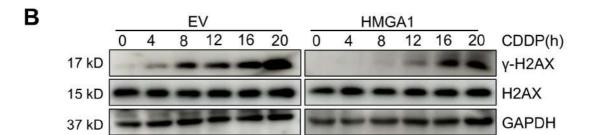
Supplementary Fig. 2 Levels of glycolytic and amino acid metabolites in the targeted metabolomics analysis of HMGA1 intact and deficient KYSE30 cells. A. Relative levels of glycolytic metabolites in KYSE30-ctrl and KYSE30-shHMGA1 cells (n = 3). B, C. Relative levels of amino acids and tricarboxylic acid cycle intermediate metabolite in KYSE30-ctrl and KYSE30-shHMGA1 cells (n = 3). D, E. Relative levels of NADPH and GSH in KYSE30 cells with HMGA1 knockdown. Data are presented as the means \pm S.D., and significant differences are indicated as * P < 0.05, ** P < 0.01.

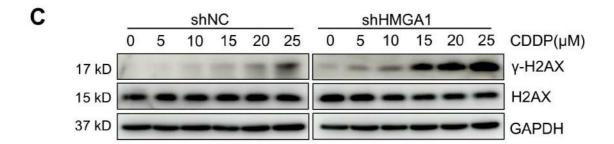


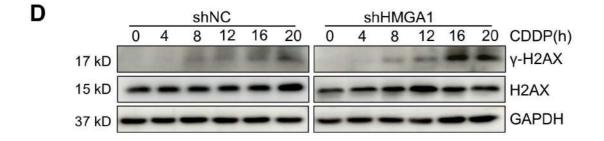
Supplementary Fig. 3 Metabolic enzymes in HMGA1-manipulated ESCC cells. A.

The dot plot map showed the expression of HMGA1 and different metabolic enzyme genes in a variety of cell subtypes in ESCCs from scRNA-seq data in the GEO database (GSE188900 dataset). **B**. The tSNE map showed expression levels of HMGA1 and various metabolic enzyme genes in different cell subtypes in ESCCs. **C**. The tSNE map showed expression of G6PD in ESCCs from scRNA-seq data in the GEO database (GSE188900 dataset). **D**, **E**. Levels of HMGA1 (**D**) and TKT (**E**) mRNA were determined by RT-qPCR in HMGA1-overexpressed TE13 cells. Samples were normalized to GAPDH mRNA. **F**. The enzymatic activity of G6PD was measured in HMGA1-overexpressed TE13 cells. **G**, **H**. Levels of HMGA1 (**G**) and TKT (**H**) mRNA were determined by RT-qPCR in HMGA1-knocked down KYSE30 cells. Samples were normalized to GAPDH mRNA. **I**. The enzymatic activity of TKT was measured in HMGA1-knocked down KYSE30 cells. **J**. The enzymatic activity of G6PD was measured in HMGA1-knocked down KYSE30 cells. Data are presented as the means \pm S.D., and significant differences are indicated as * P < 0.05, ** P < 0.01. n = 3.

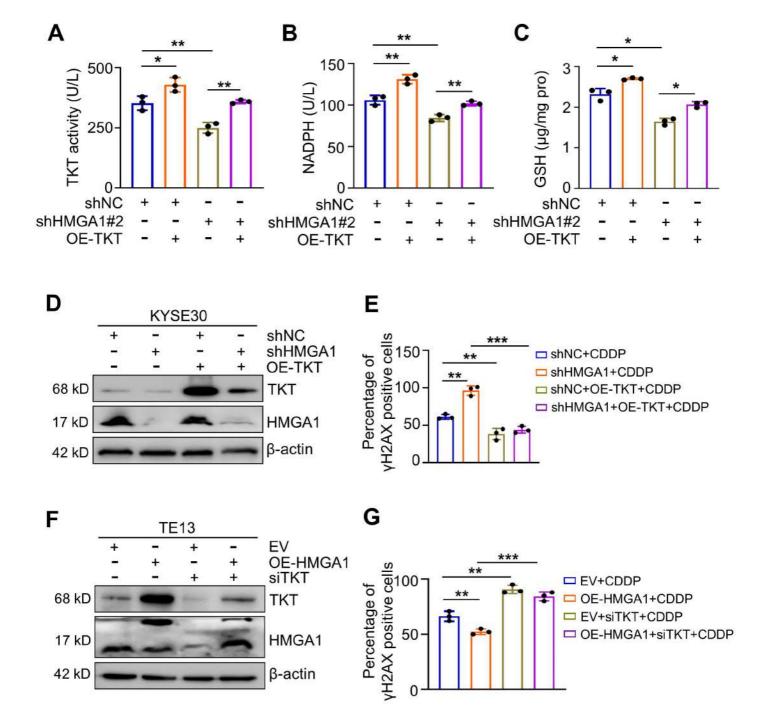








Supplementary Fig. 4 Alterations of γ -H2AX in HMGA1-manipulated KYSE30 and TE13 cells. A. The TE13 cells with or without HMGA overexpression were treated with different concentrations of CDDP for 16 h and the cell lysates were analyzed by western blotting (WB). B. The TE13 cells with or without HMGA overexpression were treated with CDDP (10 μ M) for different time, and the cell lysates were analyzed by WB. C. The KYSE30 cells with or without HMGA knockdown were treated with different concentrations of CDDP for 16 h and the cell lysates were analyzed by WB. D. The KYSE30 cells with or without HMGA knockdown were treated with CDDP (10 μ M) for different time, and the cell lysates were analyzed by WB.



Supplementary Fig. 5 TKT mediates HMGA1-upregulated PPP. A - C. Control and HMGA1-knocked down KYSE30 cells were transfected with Flag-tagged TKT. TKT enzyme activity (A), NADPH (B), and GSH (C) were measured. D. Control and HMGA1-knocked down KYSE30 cells were transfected with pcDNA3.1/TKT. Western blotting was used for determining the expression of HMGA1 and TKT. E. Immunofluorescence was performed for the detection of γ-H2AX in cells treated with 10 μM CDDP for 16 h. The percentage of γ-H2AX-positive ESCC cells was calculated. Only cells with more than 5 y-H2AX lesions are considered positive. The percentage of y-H2AX-positive cells is quantified based on DNA repair activity. Count at least 100 cells per sample per experiment. F. Control and HMGA1-overexpressed TE13 cells were transfected with TKT siRNA. Western blotting was used for determining the expression of HMGA1 and TKT. G. Immunofluorescence was performed for the detection of y-H2AX in cells treated with TKT siRNA and 10 µM CDDP. The experiment was performed as described in (E). Data are presented as the means ± S.D., and significant differences are indicated as * P < 0.05, ** P < 0.01, and *** P < 0.001. n = 3.