

An Improved SNAP-ADAR Tool Enables Efficient RNA Base Editing to Interfere with Post-translational Protein Modification

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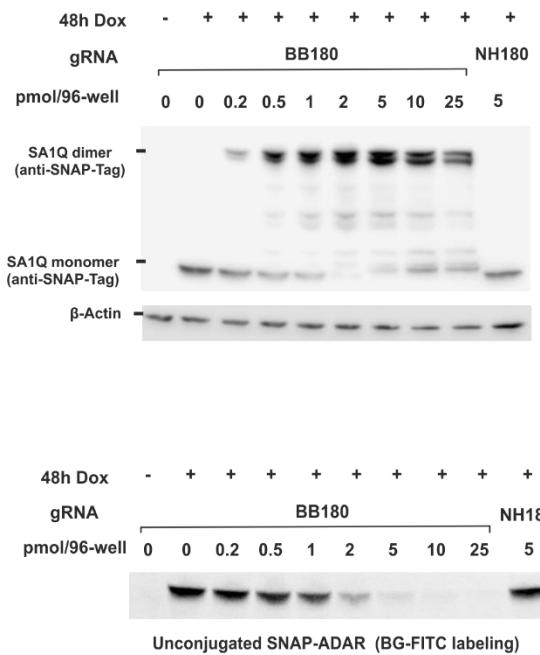
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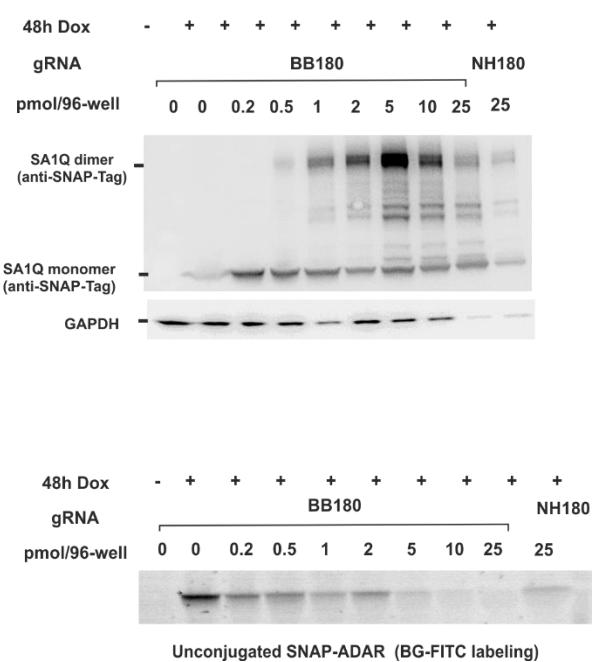
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Supplementary Figures

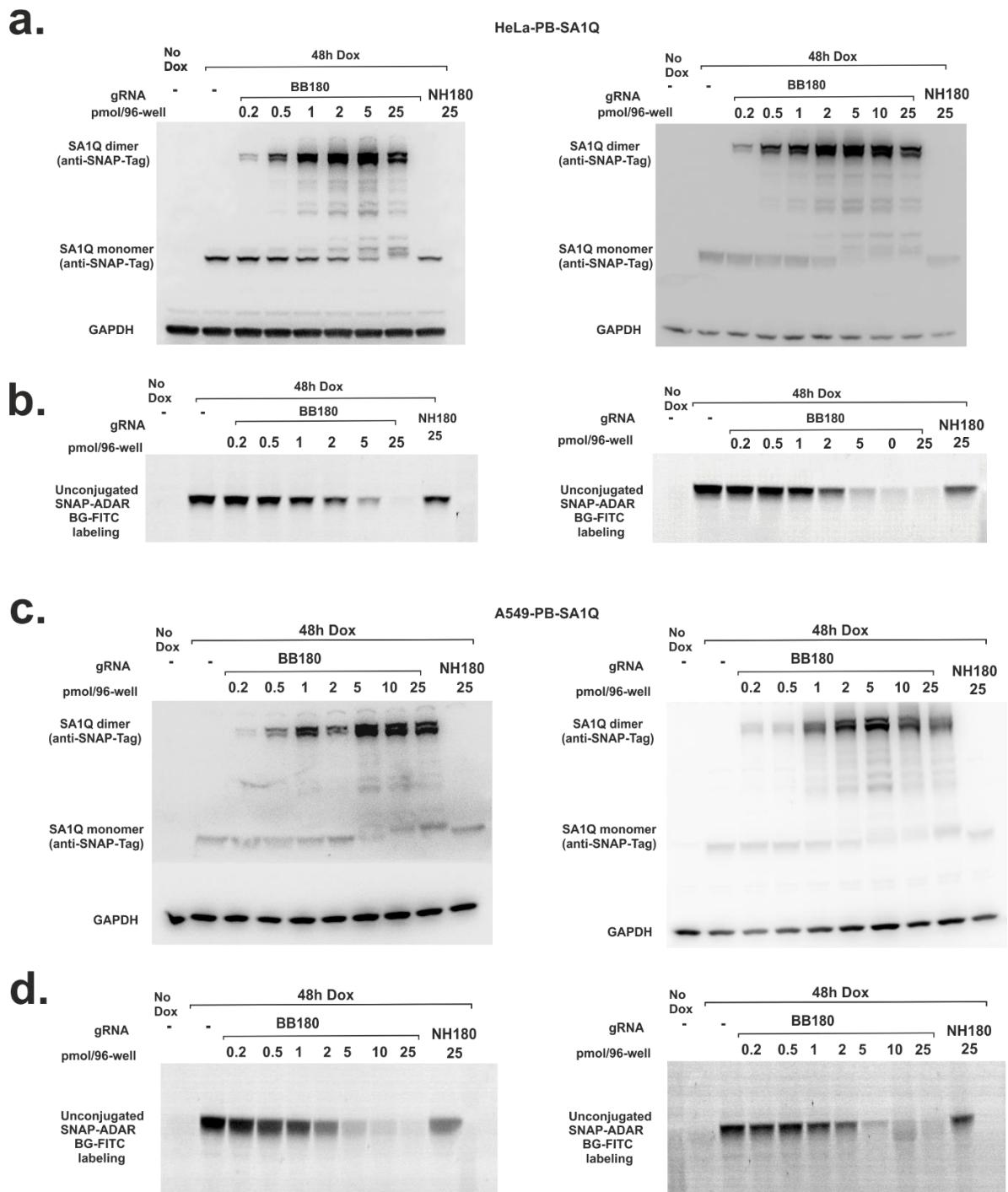
a.



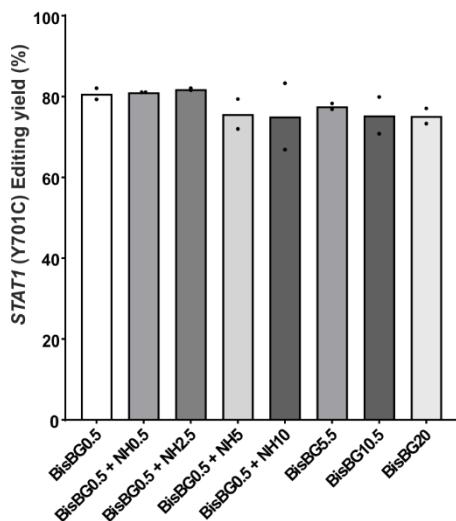
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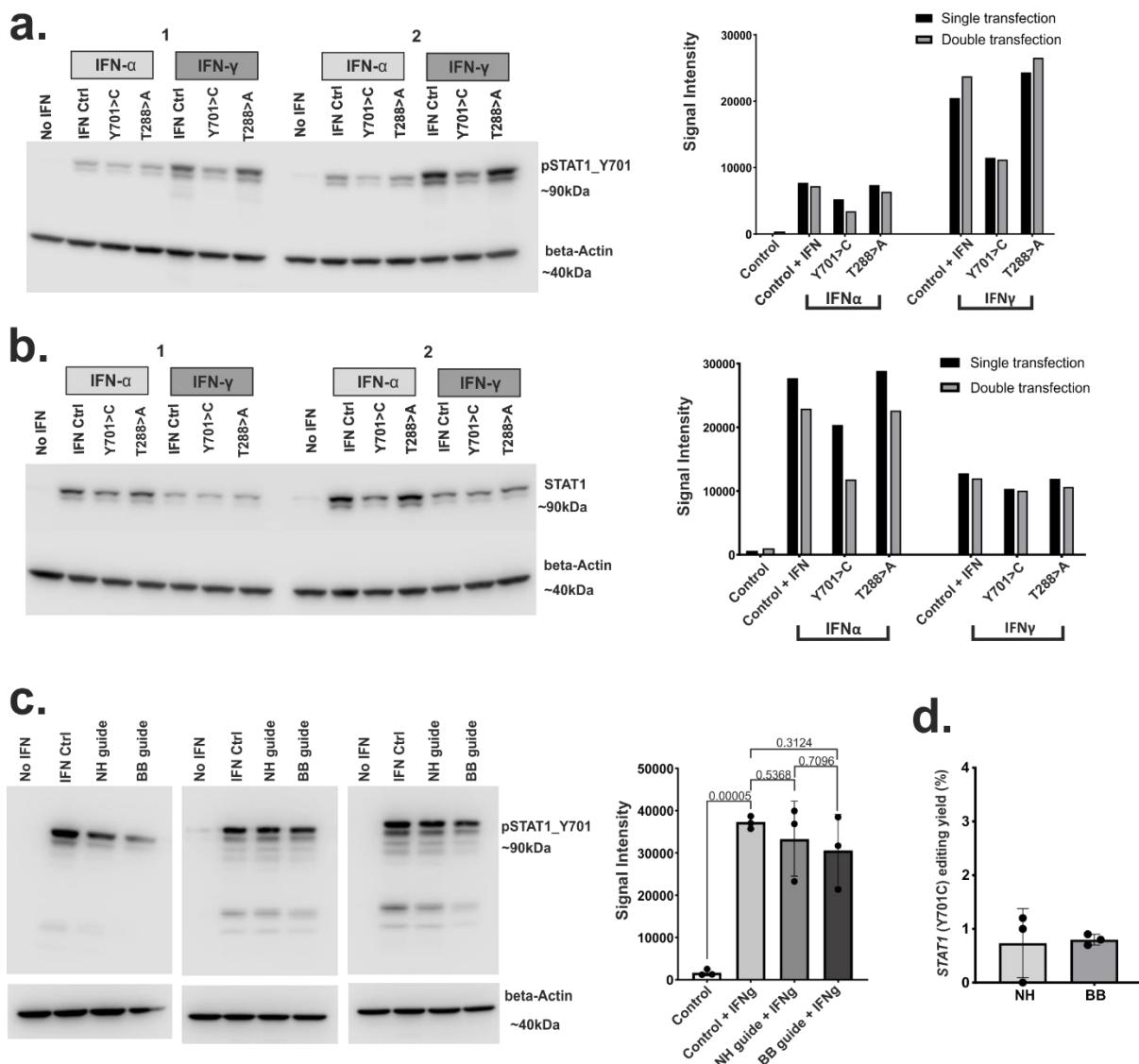
Supplementary Figure 1 Blots showing guideRNA dose-dependent dimerization of the effector SA1Q in Flp-In T-Rex 293 – SA1Q cells induced with 10ng/ml Doxycycline for 48 hrs. Cells were transfected with indicated amounts of BB180 (BisBG-25 nt + LNA gRNA against STAT1 Y701 site) or NH180 (gRNA lacking the BisBG moiety for SA attachment). **A)** and **B)** are biological replicates (N=2) of Fig.1d, with Western blot (blot on top) and BG-FITC -protein/gRNA conjugation assay blot (blot in the bottom) respectively, showing gRNA dose-dependent of SA1Q dimerization in cells induced for 48hrs with Doxycycline. GAPDH/ β -Actin served as loading controls. All guide RNA amounts denote pmol/96-well with 150 μ l total volume. Western blotting and BG-FITC staining were done as described in the methods section. Full blots are available in the Source Data file.



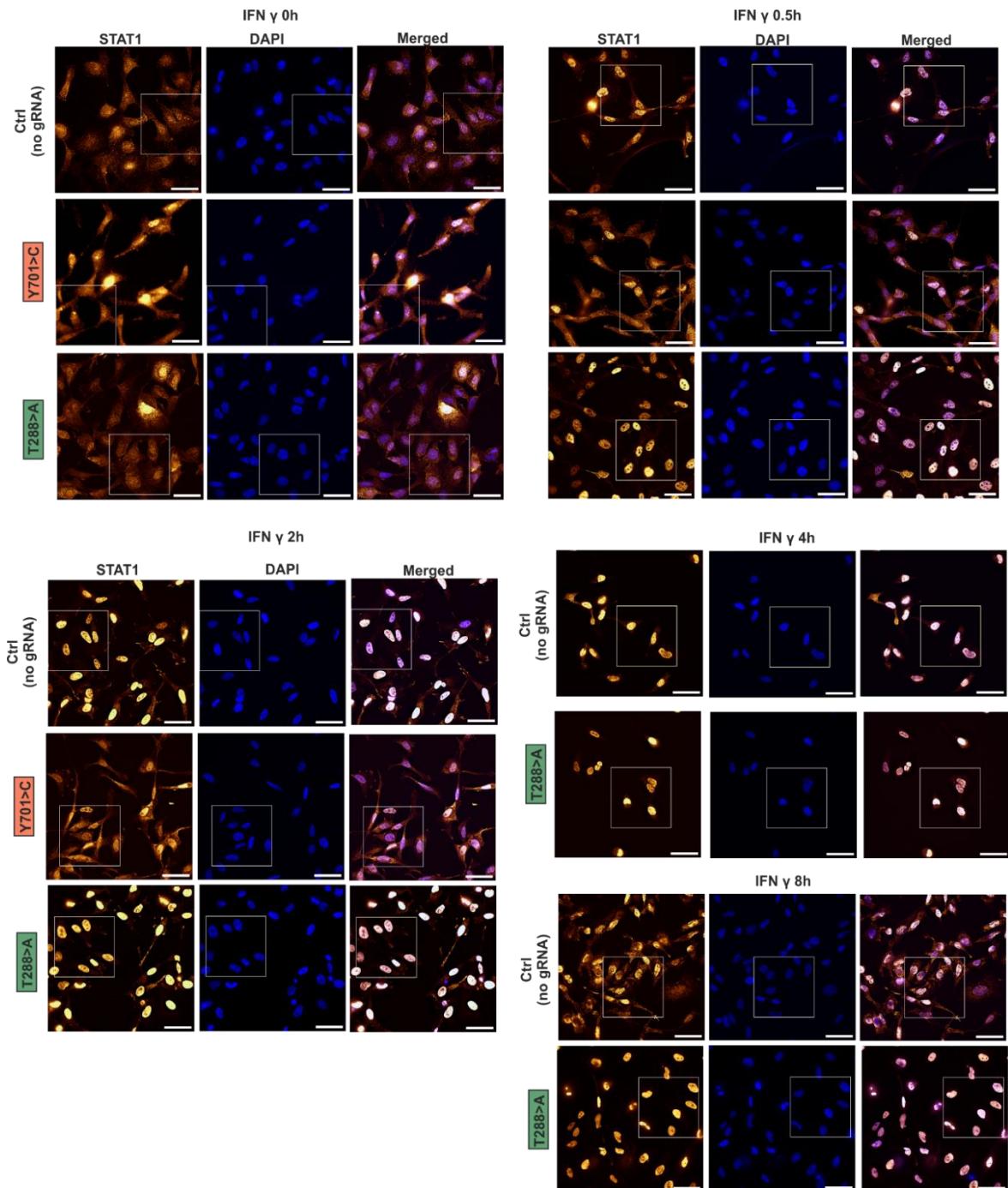
Supplementary Figure 2. Blots showing biological replicates (N=2) of SNAP-ADAR protein expression and its conjugation with gRNA studied in Hela-PB-SA1Q and A549-PB-SA1Q cells using Western blot and BG-FITC -protein/gRNA conjugation assay. Cells were transfected with different amounts of BisBG-25 nt + LNA gRNA against STAT1 Y701 site (BB180) in 96-well plate set up with 1 μ g/ml doxycycline induction for 48h. NH180 denotes gRNA lacking the BisBG moiety for SA attachment. **A)** and **B)** show Western blots and BG-FITC blots (replicates) respectively for Hela-PB-SA1Q cells. **C)** and **D)** show Western blots and BG-FITC blots (replicates) respectively for A549-PB-SA1Q cells. GAPDH served as loading control. All guide RNA amounts denote pmol/96-well with 150 μ l total volume. Western blotting and BG-FITC staining were done as described in the methods section. Full blots are available in the Source Data file.



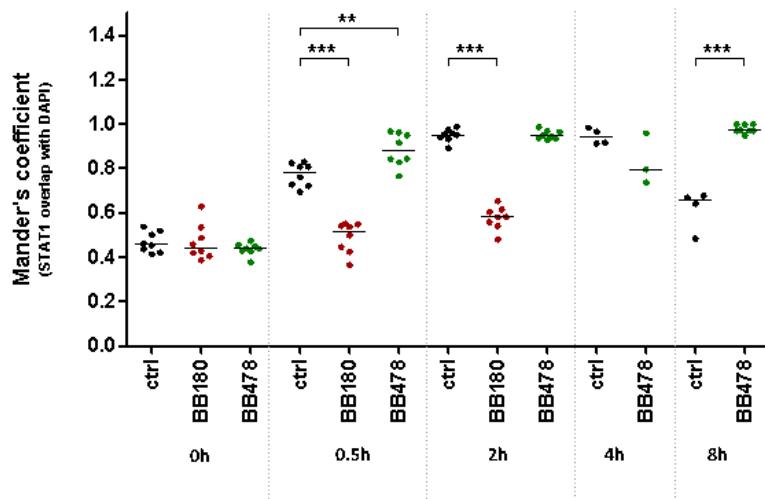
Supplementary Figure 3. Competitive editing experiment of editing-competent BisBG guide RNAs with editing-incompetent amino guide RNA of the same sequence and chemical modification. In Flp-In T-REx 293 SA1Q cells, the editing-competent BisBG 180 guide (0.5 pmol per 96 well) and an increasing amount (0.5-10 pmol per 96-well) of the same editing-incompetent NH180 control guide RNA against the transcript STAT1 (Y701>C site) were co-transfected. Even a 20x excess of amino control guide RNA (NH180) did not reduce the editing yield with 0.5 pmol BisBG guide RNA, indicating that the guide RNAs act on the target RNA rather in a hit-and-run fashion than staying at the target for a long time. Furthermore, editing yield for increasing BisBG guide RNA concentrations (up to 20 pmol/96 well) are shown. The data is presented as N=2 biological replicates, as indicated by individual data points. Source data are provided in Source Data file.



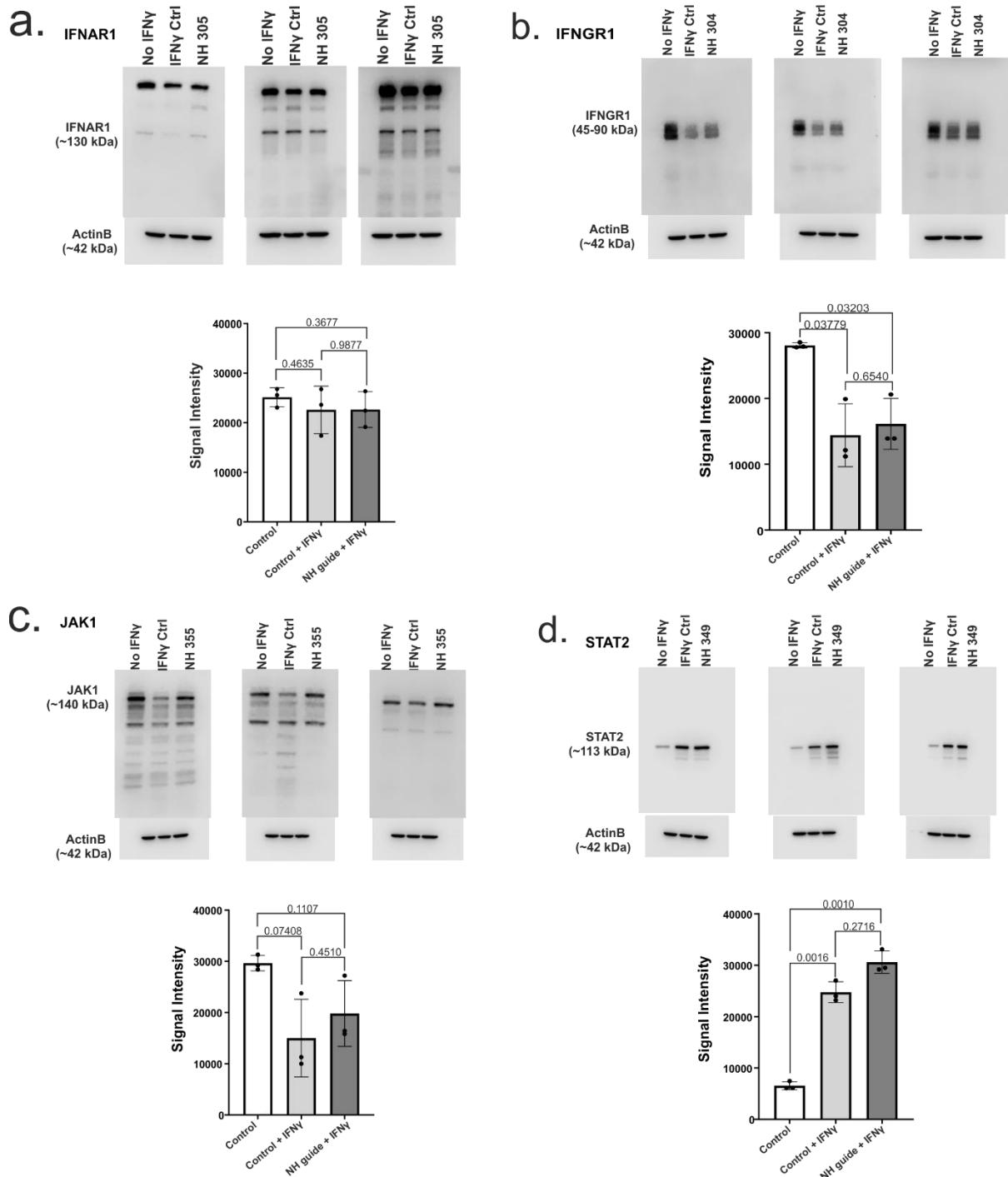
Supplementary Figure 4. Analysis of STAT1 activation (tyrosine 701 phosphorylation) by Western Blot in response to up (T288>A) and down regulating (Y701>C) PTMi. **A)** Shows phospho-STAT1 and **B)** shows total STAT1 in response to various PTMi settings and induction by interferon- α and γ , as indicated. Experiment **1 in A)** and **B)**: Dox (1 μ g/ml, 72h total), single transfection (T1) in 24 well plate with 2×10^5 cells and 8 pmol guide per well, 2000 U/ml IFN α or γ induction 24h prior to lysis. Experiment **2 in A)** and **B)**: Dox (1 μ g/ml, 120h total), double transfection (T2) with 48h in between, in 24 well plate with 2×10^5 cells and 8 pmol guide per well, 2000 U/ml IFN- α or - γ induction 24h prior to lysis. **C)** Effect of an editing-incompetent guide RNA that can (BB482 = BisBG 482) or cannot (NH482) recruit the SNAP-ADAR effector on target protein expression (pSTAT1) by Western Blot. The bar graph includes a statistical analysis of the band intensities, with P values indicated. Applied was a 2-winged t-test, N=3; P>0.05 is interpreted as non-significant. **D)** The editing-incompetent guide RNA BisBG 482 was unable to induce the Y701>C edit in STAT1. Source data are provided as a Source Data file.



Supplementary Figure 5. Immunofluorescence imaging of total STAT1 cellular localization upon different gRNA treatment vs control (no gRNA), at various time points post IFN- γ induction. Insets show zoomed-in areas shown in Fig. 5d. For detailed protocol and list of antibodies, see the Methods section and Supplementary Table 1, respectively. Images were taken with a Nikon Eclipse Ti2-E inverted fluorescent microscope, equipped with a photometrics® Prime 95B camera and a lumencor® Aurall light engine. All pictures were recorded using a 60x oil objective (numerical aperture 1.4) and Olympus IMMOIL-F30CC immersion oil. The excitation wavelengths and corresponding filter sets used to record each channel are specified in Supplementary Table 2. The same acquisition settings were chosen for each channel for all images. The images were 3D deconvoluted, and further processed in FIJI ImageJ¹. Amber LUT, used to display STAT1 protein, was obtained from ImageJ Wiki – NucMed LUT list². For more details on LUT, see Source Data.



Supplementary Figure 6. Colocalization analysis. Mander's coefficient for the overlap of total STAT1 protein signal with nuclear DAPI signal was determined and plotted for each condition using the JACoP plug-in³ in FIJI ImageJ¹. An unbiased sectioning of biological replicate images (N = 2 for most, N = 1 for some) into quadrants was done. Significance for N= 3,4 or 8 sections was determined by an unpaired, two-tailed t-test, P ≤ 0.05 = *, P ≤ 0.01 = **, P ≤ 0.001 = ***, P>0.05 = nonsignificant. Individual data points are given, the median is denoted by a black line. Source data are provided as a Source Data file.



Supplementary Figure 7. Analysis of the effect of guide RNA transfection with or without IFN- γ induction on the protein expression of four different PTMi targets by Western Blot. The bar graphs show the statistical analysis of the band intensities, with P values indicated. Applied was a 2-winged t-test, N=3; P>0.05 is interpreted as non-significant. **A)** For the IFNAR1 target. **B)** For the IFNGR1 target. **C)** For the JAK1 target. **D)** For the STAT2 target. Dox (1 μ g/ml, 120h total), double transfection (T2) with 48 h in between, in 24 well plate with 2×10^5 cells and 8 pmol guide per well, 2000 u/ml IFN- γ induction 24 h prior to cell lysis. Full Western blots and Source data are provided as a Source Data file.

References

1. Schneider, C. A., Rasband, W. S. & Eliceiri, K. W. NIH Image to ImageJ: 25 years of image analysis. *Nat Methods* **9**, 671–675 (2012).
2. https://imagej.net/ij/download/luts/NucMed_Image_LUTs/.
3. BOLTE, S. & CORDELIÈRES, F. P. A guided tour into subcellular colocalization analysis in light microscopy. *J Microsc* **224**, 213–232 (2006).

Supplementary Tables

Supplementary Table 1 All antibodies were diluted in 5% dry milk in 1x TBST buffer.

Antibody	Cat.no.	Manufacturer	Dilution
Mouse α-Beta-Actin	A5441	Sigma	1: 5000
Rabbit α-SNAP-tag	P9310S	New England Biolabs	1: 1000
α-SNAP Tag Polyclonal Antibody	CAB4255	Thermo-Fisher Scientific	1: 1000
Mouse-α-GAPDH Loading Control Monoclonal Antibody (GA1R)	MA5-15738	Thermo-Fisher Scientific	1: 1000
Mouse-α-GAPDH Monoclonal Antibody (6C5)	AM4300	Thermo-Fisher Scientific	1: 1000
Rabbit-α-Gapdh (D16H11) XP	5174T/5174S	Cell Signalling	1: 2000
pSTAT1(Y701)	9167	Cell Signalling	1: 1000
STAT1 mouse	9176	Cell Signalling	1: 1000
STAT1 (42H3) Rabbit	9175	Cell Signalling	1: 1000
Goat α-Rabbit Alexa Fluor 594	8889S	Cell Signalling	1: 1000
Goat α-Rabbit HRP	111-035-003	Jackson Immuno Research Laboratories	1: 10000
Goat α-Mouse HRP	115-035-003	Jackson Immuno Research Laboratories	1: 10000
IFNAR1 rabbit	MA5-32006	Thermo-Fisher Scientific	1: 1000
JAK1 mouse	50996	Cell Signaling	1: 1000
STAT1 Rabbit	9172	Cell Signaling	1: 1000
IFNGR1 rabbit	34808	Cell Signaling	1: 1000
STAT2 rabbit	72604	Cell Signaling	1: 1000

Supplementary Table 2 Excitation wavelengths and corresponding filter sets used with Nikon Eclipse Ti2-Emicroscope.

Fluorophore	Excitation	Filter
eGFP/FITC	475 nM	Excitation: Chroma 89402x Beamsplitter: Chroma 89402bs Emission: Chroma 89402m
Atto594	575 nM	Excitation: Semrock 585/29 BrightLine HC Beamsplitter: Chroma T610LPXR Emission: Semrock 650/60 BrightLine HC
NucBlue (DAPI)	390 nM	Excitation: Chroma 89402x Beamsplitter: Chroma 89402bs Emission: Chroma 89402m

Gene and protein sequences for SNAP-ADAR versions used:

Gene & protein sequence of the SA1Q as expressed from engineered Flp-In-TREx 293 genome. E>Q site is highlighted in cyan.

	10	20	30	40	50	60
1	ATGGACAAAGACTGCGAAATGAAGCGCACCAACCCTGGATAGCCCTCTGGGCAAGCTGGAA					
1	M D K D C E M K R T T L D S P L G K L E					
	70	80	90	100	110	120
61	CTGTCTGGGTGCGAACAGGGCCTGCACCGTATCATCTTCCTGGGCAAAGGAACATCTGCC					
21	L S G C E Q G L H R I I F L G K G T S A					
	130	140	150	160	170	180
121	GCCGACGCCGTGGAAGTGCCTGCCAGCCGCCGTGCTGGCGGACCAGAGCCACTGATG					
41	A D A V E V P A P A A V L G G P E P L M					
	190	200	210	220	230	240
181	CAGGCCACCGCCTGGCTCAACGCCTACTTTACCAGCCTGAGGCCATCGAGGAGTTCCCT					
61	Q A T A W L N A Y F H Q P E A I E E F P					
	250	260	270	280	290	300
241	GTGCCAGCCCTGCACCACCCAGTGTTCAGCAGGAGAGCTTACCCGCCAGGTGCTGTGG					
81	V P A L H H P V F Q Q E S F T R Q V L W					
	310	320	330	340	350	360
301	AAACTGCTGAAAGTGGTGAAGTTCGGAGAGGTACAGCTACAGCCACCTGGCCGCCCTG					
101	K L L K V V K F G E V I S Y S H L A A L					
	370	380	390	400	410	420
361	GCCGGCAATCCGCCGCCACCGCCCGCGTGAAACCGCCCTGAGCGGAAATCCGTGCC					
121	A G N P A A T A A V K T A L S G N P V P					
	430	440	450	460	470	480
421	ATTCTGATCCCCTGCCACCGGGTGGTGCAGGGCGACCTGGACGTGGGGGCTACGAGGGC					
141	I L I P C H R V V Q G D L D V G G Y E G					
	490	500	510	520	530	540
481	GGGCTGCCGTGAAAGAGTGGCTGCTGGCCCACGAGGGCCACAGACTGGCAAGCCTGGG					
161	G L A V K E W L L A H E G H R L G K P G					
	550	560	570	580	590	600
541	CTGGGTCTGCAGGCAGGGCGGCCAGGGTCTGGCGGGCAGTAAGGCAGAACGCATG					
181	L G P A G G G A P G S G G S K A E R M					
	610	620	630	640	650	660
601	GGTTTCACAGAGGTAACCCCAGTGACAGGGGCCAGTCTCAGAAGAACTATGCTCCTC					
201	G F T E V T P V T G A S L R R T M L L L					
	670	680	690	700	710	720
661	TCAAGGTCCCCAGAACAGCACAGCCAAAGACACTCCCTCTCACTGGCAGCACCTCCATGAC					
221	S R S P E A Q P K T L P L T G S T F H D					
	730	740	750	760	770	780
721	CAGATAGCCATGCTGAGCCACCGGTGCTTCAACACTCTGACTAACAGCTTCCAGCCCTCC					
241	Q I A M L S H R C F N T L T N S F Q P S					

	790	800	810	820	830	840
781	TTGCTCGGCCGCAAGATTCTGGCCGCCATTATGAAAAAAGACTCTGAGGACATGGGT					
261	L L G R K I L A A I I M K K D S E D M G					
	850	860	870	880	890	900
841	GTCGTCGTCAGCTGGAACAGGGAATCGCTGTGTAAAAGGAGATTCTCTCAGCCTAAA					
281	V V V S L G T G N R C V K G D S L S L K					
	910	920	930	940	950	960
901	GGAGAAACTGTCAATGACTGCCATGCAGAAATAATCTCCGGAGAGGCTTCATCAGGTT					
301	G E T V N D C H A E I I S R R G F I R F					
	970	980	990	1000	1010	1020
961	CTCTACAGTGAGTTAATGAAATAACAACCCCAGACTGCGAAGGGATAGTATATTGAACCT					
321	L Y S E L M K Y N S Q T A K D S I F E P					
	1030	1040	1050	1060	1070	1080
1021	GCTAAGGGAGGAGAAAAGCTCCAAATAAAAAGACTGTGTCAATTCCATCTGTATATCAGC					
341	A K G G E K L Q I K K T V S F H L Y I S					
	1090	1100	1110	1120	1130	1140
1081	ACTGCTCCGTGTGGAGATGGCGCCCTTTGACAAGTCCTGCAGCGACC GTGCTATGGAA					
361	T A P C G D G A L F D K S C S D R A M E					
	1150	1160	1170	1180	1190	1200
1141	AGCACAGAATCCGCCACTACCCCTGTCTCGAGAATCCCAAACAAGGAAAGCTCCGCACC					
381	S T E S R H Y P V F E N P K Q G K L R T					
	1210	1220	1230	1240	1250	1260
1201	AAGGTGGAGAACCGGA CAAGGCACAATCCCTGTGGAATCCAGTGACATTGTGCCTACGTGG					
401	K V E N G Q G T I P V E S S D I V P T W					
	1270	1280	1290	1300	1310	1320
1261	GATGGCATT CGCTCGGGAGAGACTCCGTACCATGTCCTGTAGTGACAAAAT CCTACGC					
421	D G I R L G E R L R T M S C S D K I L R					
	1330	1340	1350	1360	1370	1380
1321	TGGAACGTGCTGGGCCTGCAAGGGGCACTGTTGACCCACTTCCTGCAGCCCATTATCTC					
441	W N V L G L Q G A L L T H F L Q P I Y L					
	1390	1400	1410	1420	1430	1440
1381	AAATCTGTACATTGGGTTACCTTCAGCCAAGGGCATCTGACCCGTGCTATTGCTGT					
461	K S V T L G Y L F S Q G H L T R A I C C					
	1450	1460	1470	1480	1490	1500
1441	CGTGTGACAAGAGATGGAGTCATTTGAGGATGGACTACGACATCCCTTATTGTCAAC					
481	R V T R D G S A F E D G L R H P F I V N					
	1510	1520	1530	1540	1550	1560
1501	CACCCCAAGGTTGGCAGAGTCAGCATATATGATTCCAAAAGGCAATCCGGGAAGACTAAG					
501	H P K V G R V S I Y D S K R Q S G K T K					
	1570	1580	1590	1600	1610	1620
1561	GAGACAAGCGTCAACTGGTGTCTGGCTATGACCTGGAGATCCTGGACGGTACC					
521	E T S V N W C L A D G Y D L E I L D G T					
	1630	1640	1650	1660	1670	1680
1621	AGAGGCACTGTGGATGGGCCACGGAATGAATTGTCGGGGTCTCCAAAAGAACATTTT					
541	R G T V D G P R N E L S R V S K K N I F					
	1690	1700	1710	1720	1730	1740

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561 L L F K K L C S F R Y R R D L L R L S Y

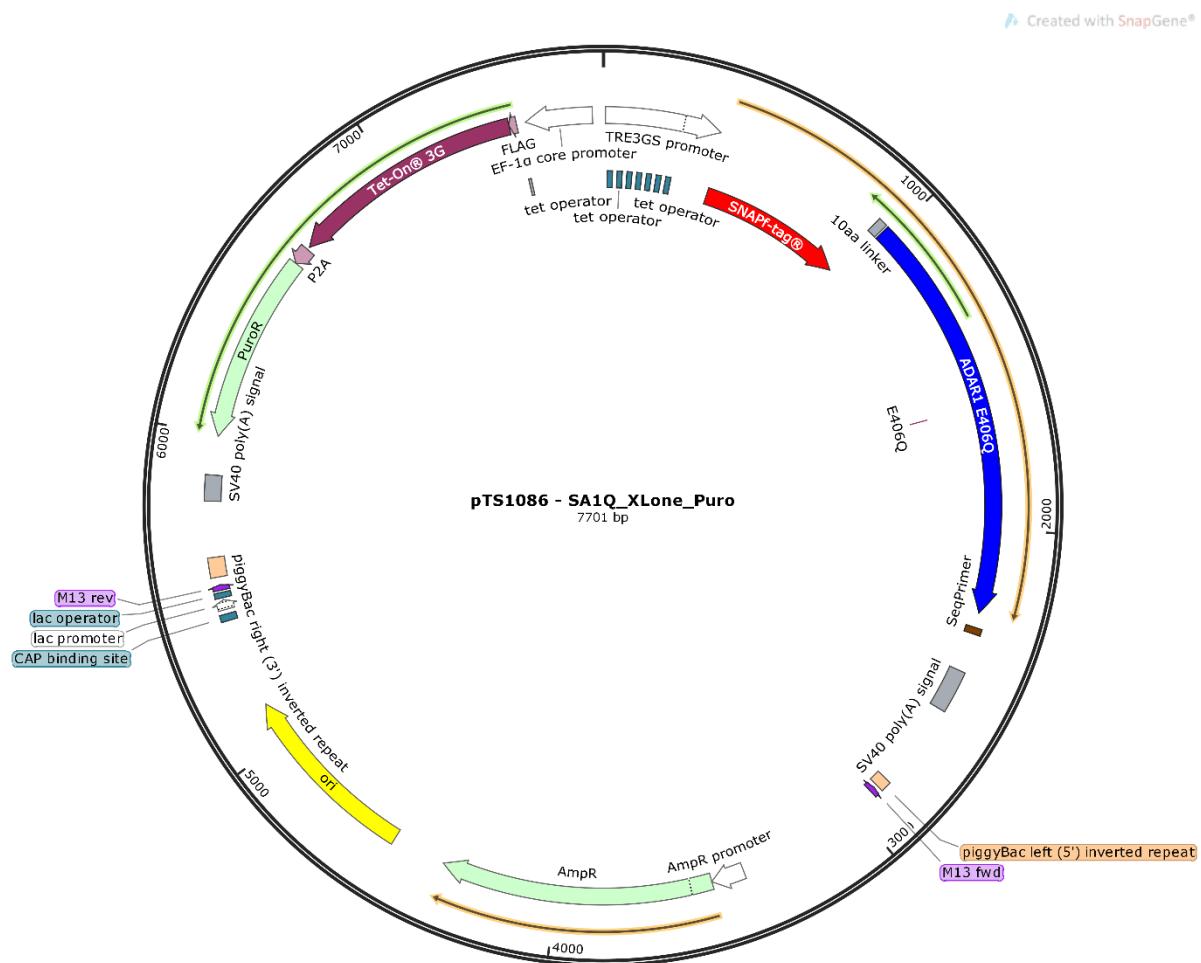
1741 1750 1760 1770 1780 1790 1800
581 GGTGAGGCCAAGAAAGCTGCCCGTGACTACGAGACGGCCAAGAACTACTTCAAAAAGGC
G E A K K A A R D Y E T A K N Y F K K G

1801 1810 1820 1830 1840 1850 1860
601 CTGAAGGATATGGGCTATGGGAACTGGATTAGCAAACCCCAGGAGGAAAAGAACTTTAT
L K D M G Y G N W I S K P Q E E K N F Y

1861 1870
621 CTCTGCCAGTA
L C P V

Plasmid-maps & sequences:

Plasmid-map and sequence of PiggyBac plasmid SA1Q_XLone_Puro (pTS1086): The cyan highlighted site denotes E>Q mutation in hyperactive SNAP ADAR (SA1Q). The PiggyBac plasmid SA1_XLone_Puro (pTS1074) differs from this plasmid at this site with a GAA codon (Glu) instead of CAA codon(Gln).



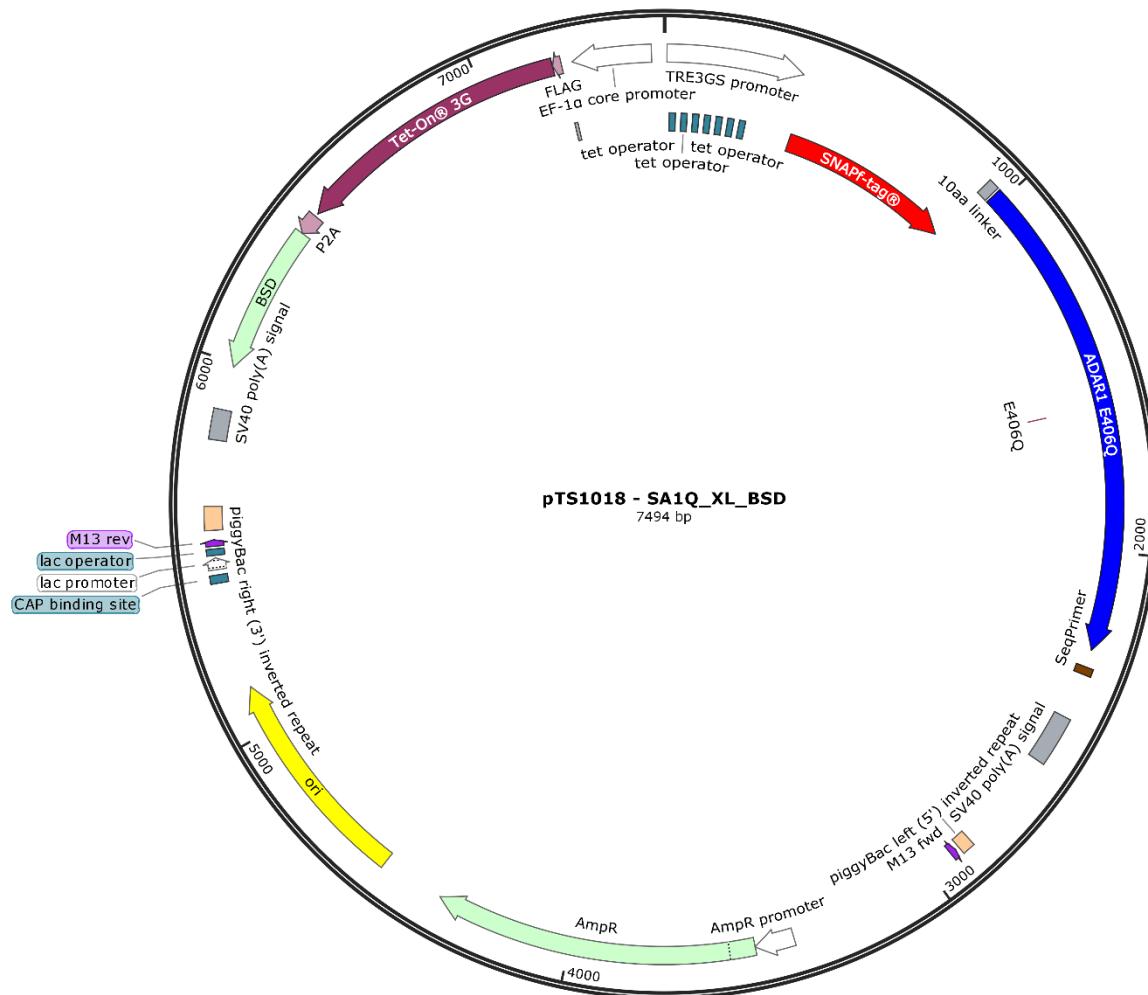
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121	TTACTCCCTA	TCAGTGATAG	AGAACGTATG	ACCAGTTAC	TCCCTATCAG	TGATAGAGAA
181	CGTATCTACA	GTTTACTCCC	TATCAGTGAT	AGAGAACGTA	TATCCAGTTT	ACTCCCTATC
241	AGTGATAGAG	AACGTATAAG	CTTGCTTAT	GTAAACCAGG	GCCGCTATAA	AAGAGTGCTG
301	ATTTTTGAG	TAAAACCAA	TTCCACAAACA	CTTTGTCTT	ATACCAACTT	TCCGTACAC
361	TTCCCTACCC	CGTAAAGGTA	CCGGGGCGC	CACCATGGAC	AAAGACTGCG	AAATGAAGCG
421	CACCAACCTG	GATAGCCCTC	TGGGCAAGCT	GGAACTGTCT	GGGTGCGAAC	AGGGCTGCA
481	CCGTATCATC	TTCCCTGGGCA	AAGGAACATC	TGCCGCCGAC	GCCGTGGAAG	TGCCTGCCCC
541	AGCCGCCGTG	CTGGGCGGAC	CAGAGCCACT	GATGCAGGCC	ACCCGCTGGC	TCAACGCCCTA
601	CTTTCACCA	CCTGAGGCCA	TCGAGGAGTT	CCCTGTGCCA	GCCCTGCACC	ACCCAGTGT
661	CCAGCAGGAG	AGCTTACCC	GCCAGGTGCT	GTGGAAACTG	CTGAAAGTGG	TGAAGTTCGG
721	AGAGGTCATC	AGCTACAGCC	ACCTGGCCGC	CCTGGCCGGC	AATCCCAGCG	CCACCGCCGC
781	CGTAAAACC	GCCCTGAGCG	GAAATCCCCT	GCCCCATTCTG	ATCCCCCTGCC	ACCGGGTGGT
841	GCAGGGCGAC	CTGGACGTGG	GGGGCTACGA	GGGCGGGCTC	GCCGTGAAAG	AGTGGCTGCT
901	GGCCCCACGAG	GGCCACAGAC	TGGGCAAGGCC	TGGGCTGGGT	CCTGCAGGCC	GAGGGCGCGC

961 AGGGTCTGGC GGCGGCAGTA AGGCAGAACG CATGGGTTTC ACAGAGGTAA CCCCAGTGAC
1021 AGGGGCCAGT CTCAGAAGAA CTATGCTCCT CCTCTCAAGG TCCCCAGAAG CACAGCCAA
1081 GACACTCCCT CTCACTGGCA GCACCTCCA TGACCAGATA GCCATGCTGA GCCACCGGTG
1141 CTTCAACACT CTGACTAACCA GCTTCCAGCC CTCCTGCTC GGCGCAAGA TTCTGGCCGC
1201 CATCATTATG AAAAAAGACT CTGAGGACAT GGGTGTGTC GTCAGCTTGG GAACAGGGAA
1261 TCGCTGTGTA AAAGGAGATT CTCTCAGCCT AAAAGGAGAA ACTGTCAATG ACTGCCATGC
1321 AGAAAATAATC TCCCGGAGAG GCTTCATCATG GTTCTCTAC AGTGAGTTAA TGAAATACAA
1381 CTCCCAGACT GCGAAGGATA GTATATTGAA ACCTGCTAAG GGAGGAGAAA AGCTCAAAT
1441 AAAAAAGACT GTGTCATTCC ATCTGTATAT CAGCACTGCT CCAGTGTGGAG ATGGCCCGCT
1501 CTTTGACAAG TCCTGCAGCG ACCGTGCTAT GGAAAGCACA GAATCCGCC ACTACCTGT
1561 CTTCGAGAAT CCCAAACAAG GAAAGCTCCG CACCAAGGTG GAGAACGGAC **AAGGCACAAT**
1621 CCCTGTGGAA TCCAGTGACA TTGTGCCTAC GTGGGATGGC ATTCCGGCTCG GGGAGAGACT
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1741 ACTGTTGACC CACTTCCTGC AGCCCATTAA TCTCAAATCT GTCACATTGG GTTACCTTT
1801 CAGCCAAGGG CATCTGACCC GTGCTATTG CTGTCGTGT ACAAGAGATG GGAGTGCATT
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1981 TGATGGCTAT GACCTGGAGA TCCTGGACGG TACCAAGGGC ACTGTGGATG GGCCACGGAA
2041 TGAATTGTCC CGGGTCTCCA AAAAGAACAT TTTTCTTCTA TTTAAGAACG TCTGCTCCTT
2101 CCGTTACCGC AGGGATCTAC TGAGACTCTC CTATGGTGAG GCCAAGAAAG CTGCCGTGA
2161 CTACGAGACG GCCAAGAACT ACTTCAAAAA AGGCCTGAAG GATATGGGCT ATGGGAACCTG
2221 GATTAGCAAA CCCCAGGAGG AAAAGAACCT TTATCTCTGC CCAGTATAAA TCGATTCTAG
2281 TAGACCACCT CCCCTGCGAG CTAAGCTGGA CAGCCAATGA CGGGTAAGAG AGTACATTT
2341 TTCACTAACC TAAGACAGGA GGGCCGTAG AGCTACTGCC TAATCCAAAG ACGGGTAAAAA
2401 GTGATAAAAAA TGTATCACTC CAACCTAACAGA CAGGCGCAGC TTCCGAGGGG TTTGAGATCC
2461 AGACATGATA AGATACATTG ATGAGTTGG ACAAAACAAA ACTAGAACG AGTGAACAAAAA
2521 ATGCCTTATT TGTGAAATT GTGATGCTAT TGCCTTATT GTAACCTTA TAAGCTGCAA
2581 TAAACAAGTT TGATATCTAT AACAAAGAAAA TATATATATA ATAAGTTATC ACGTAAGTAG
2641 AACATGAAAT AACAAATATAA TTATCGTATG AGTTAAATCT TAAAGTCAC GTAAAGATA
2701 ATCATGCGTC ATTTTGACTC ACGCGGTGCT TATAGTTCAA AATCAGTGAC ACTTACCGCA
2761 TTGACAAGCA CGCCTCACGG GAGCTCAAAG CGCGCACTGA GATGCTCTAA ATGCACAGCG
2821 ACGGATTTCGC GCTATTTAGA AAGAGAGAGC AATATTCAA GAATGCATGC GTCAATTAA
2881 CGCAGACTAT CTTCTAGGG TTAAGAATTG ACTGGCCGTC GTTTTACAAC GTCGTGACTION
2941 GGAAAACCTT GGGGTTACCC AACTTAATCG CCTTGCAGCA CATCCCCCTT TCGCCAGCTG
3001 GCGTAATAGC AAAGAGGGCC GCACCGATCG CCCTTCCCAA CAGTTGCGCA GCCTGAATGG
3061 CGAATGGCGC CTGATGCGGT ATTTCTCCT TACGCATCTG TGCGGTATT CACACCGCAT
3121 ATGGTGCACT CTCAGTACAA TCTGCTCTGA TGCCGCATAG TTAAGCCAGC CCCGACACCC
3181 GCCAACACCC GCTGACGCGC CCTGACGGGC TTGTCTGCTC CGGGCATCCG CTTACAGACA
3241 AGCTGTGACC GTCTCCGGGA GCTGCATGTG TCAGAGGTT TCACCGTCAT CACCGAAACG
3301 CGCGAGACGA AAGGGCCTCG TGATACGCCT ATTTTATAG GTTAATGTCA TGATAATAAT
3361 GGTTTCTTAG ACGTCAGGTG GCACTTTCG GGGAAATGTG CGCGGAACCC CTATTGTTT
3421 ATTTTCTAA ATACATTCAA ATATGTATCC GCTCATGAGA CAATAACCCT GATAAATGCT
3481 TCAATAATAT TGAAAAGGA AGAGTATGAG TATTCAACAT TTCCGTGTCG CCCTTATTCC
3541 CTTTTTGCG GCATTTGCC TTCTGTTT TGCTCACCC GAAACGCTGG TGAAAGTAAA
3601 AGATGCTGAA GATCAGTTGG GTGCACGAGT GGGTTACATC GAACTGGATC TCAACAGCGG
3661 TAAGATCCTT GAGAGTTTC GCCCGAAGA ACGTTTCCA ATGATGAGCA CTTTTAAAGT
3721 TCTGCTATGT GGCGCGGTAT TATCCCGTAT TGACGCCGGG CAAGAGCAAC TCGGTGCGCG
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3841 GGATGGCATG ACAGTAAGAG AATTATGCAG TGCTGCCATA ACCATGAGTG ATAACACTGC
3901 GGCCAACCTA CTTCTGACAA CGATCGGAGG ACCGAAGGAG CTAACCGCTT TTTGCAACAA
3961 CATGGGGGAT CATGTAACTC GCCTTGATCG TTGGGAACCG GAGCTGAATG AAGCCATACC
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Plasmid-map and sequence of PiggyBac plasmid SA1Q_XLone_BSD (pTS1018): The cyan highlighted site denotes E>Q mutation in hyperactive SNAP ADAR (SA1Q). The PiggyBac plasmid SA1_XLone_BSD (pTS990) differs from this plasmid at this site with a GAA codon (Glu) instead of CAA codon (Gln).

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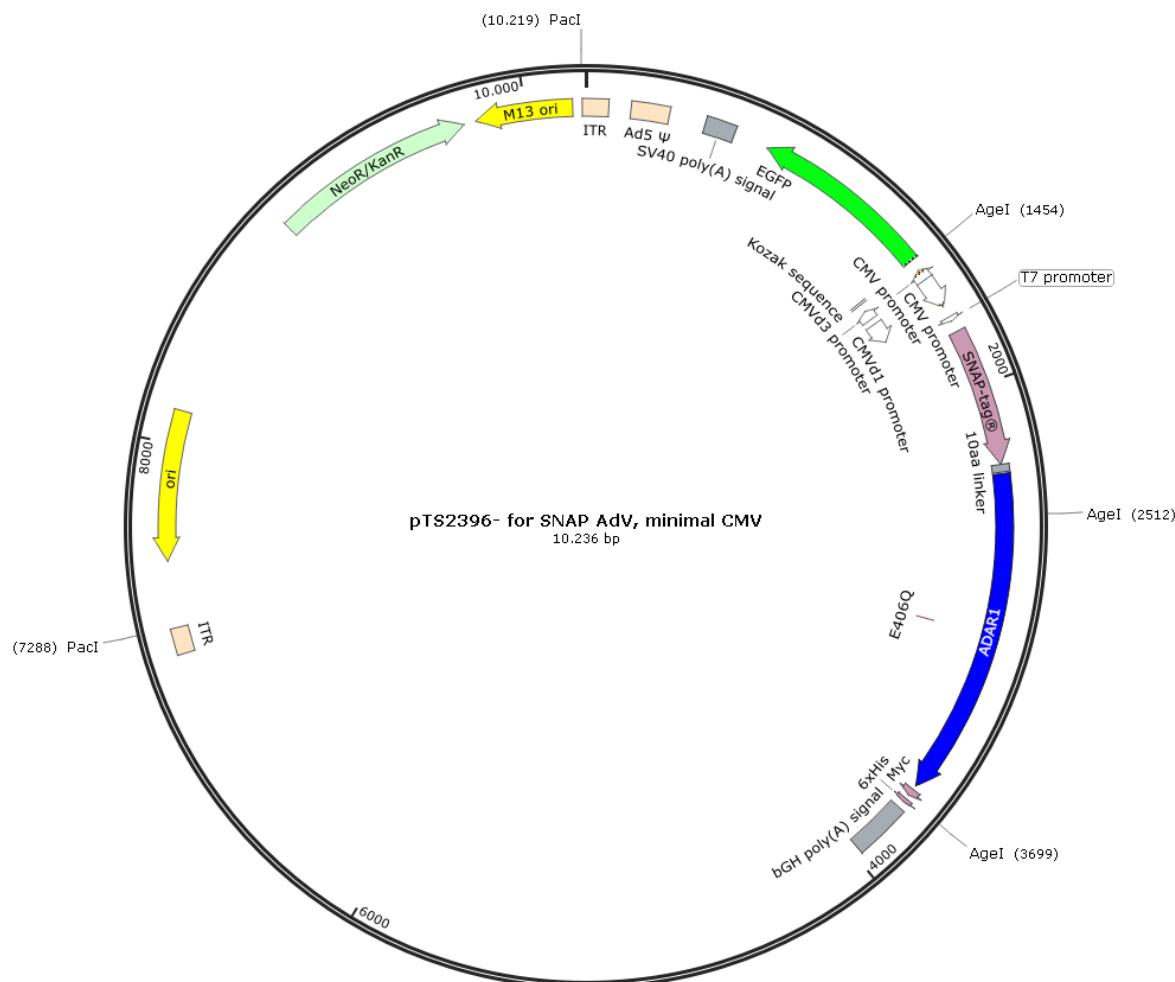
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Plasmid-map and sequence of SNAP AdV SA1Q, minimal CMV (pTS2396):

Alias: pTS2396

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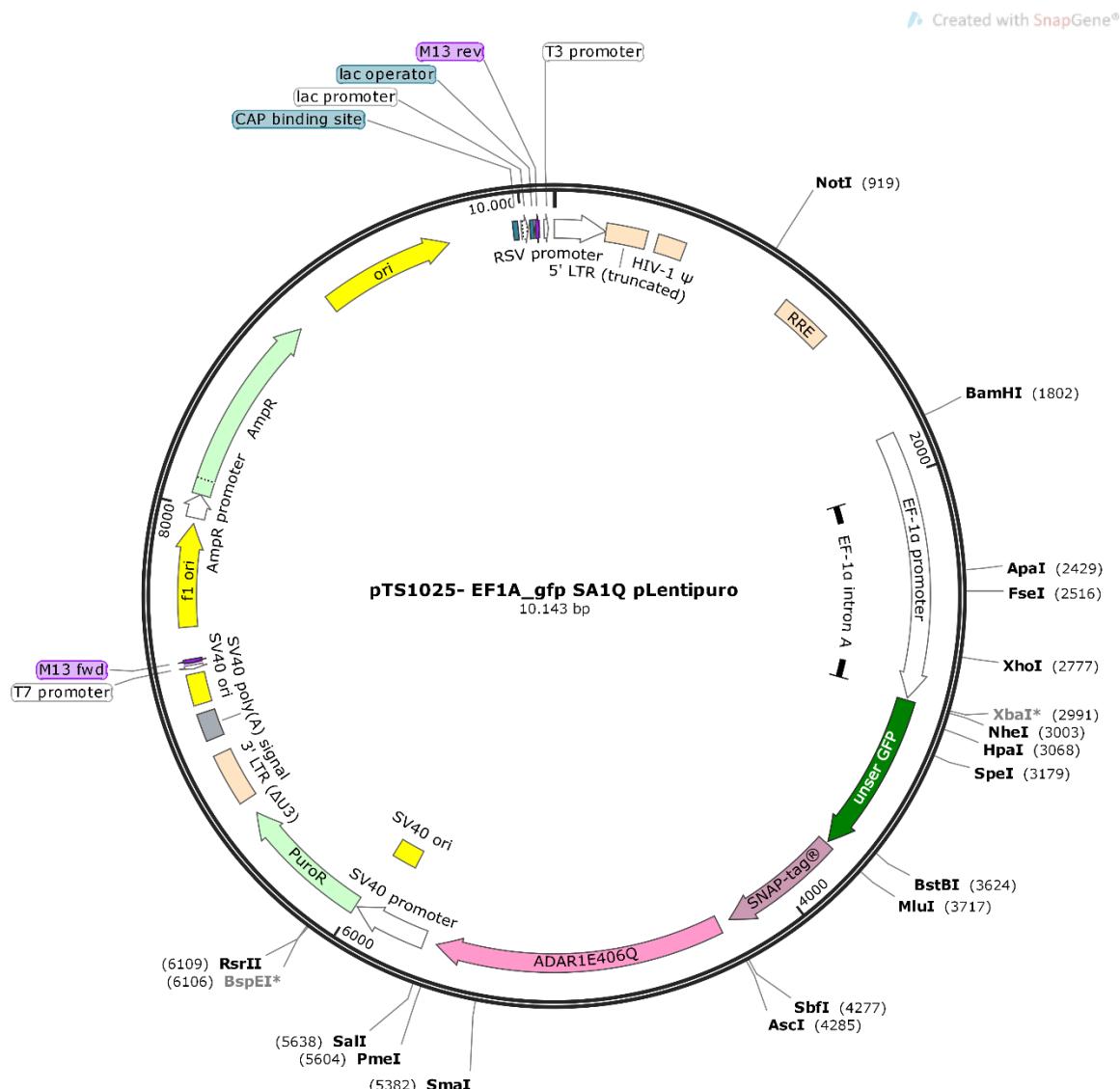
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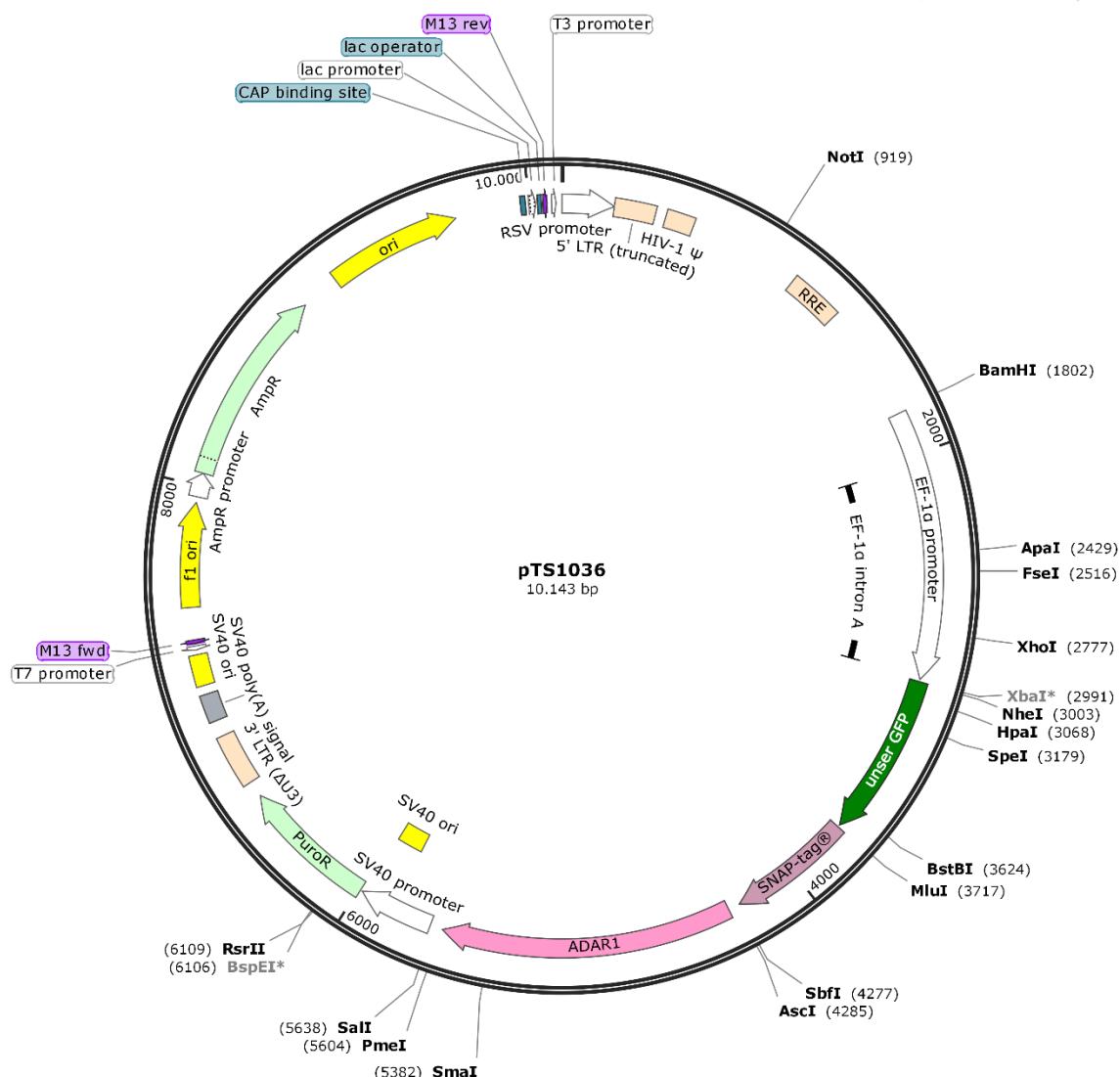
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Plasmid-map and sequence of EF1A_GFP_SA1_pLentiPuro (pTS1036):

Created with SnapGene®



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