Supporting Information

Anionic Olefin Metathesis Catalysts Enable Modification of Unprotected Biomolecules in Water

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S1. Experimental.

S1.1. General Procedures. All reactions were carried out in an N₂-filled glovebox unless otherwise noted. HPLC-grade hexanes, THF, and CH₂Cl₂ were dried and degassed with a Glass Contour solvent purification system and stored under N₂ over 4 Å molecular sieves for at least 24 h prior to use. MeOH was distilled from CaH₂ under N₂ and stored as above. Metathesis catalysts **HI**¹ and **HI-I**₂,² CAAC salts³ **C1**^{Me}•HBF₄, and **C1**^{Ph}•HBF₄, 2,2-diallylpropane-1,3-diol **2**,⁴ 2,2-diallylmalonic acid **4**,⁵ diallyl ammonium chloride **5**,⁶ and olefin **6**⁷ were prepared by literature methods. CD₃OD (Cambridge Isotopes, 99.5%), 'BuOH (Sigma, 99.5%), MilliQ H₂O, and D₂O (Cambridge Isotopes, 99.5%) were freeze-pump-thaw degassed (4×) and stored under N₂ in the glovebox. Dimethyl sulfone (Me₂SO₂, 98%; internal standard for NMR analysis), diethyl diallylmalonate **3** (TCI, 98%), potassium trispyrazolyl borate (KTp; quenching agent;⁸ Sigma, 98%), **C1**^{Cy}•HBF₄ and **AM** (the last two kindly provided as a gift by Apeiron Synthesis) were used as received. LiHMDS (Sigma, 97%) was recrystallized from hexanes and stored under N₂ in the glovebox at -35 °C. The purity of all catalysts was confirmed by ¹H NMR analysis prior to use. For accuracy in metathesis experiments, solid catalysts were weighed outside the glovebox using a microanalytical balance.

NMR spectra were recorded on Bruker Avance 300, 400, and 600 MHz NMR spectrometers at 25 ± 0.5 °C. Chemical shifts (ppm) are referenced to the residual proton of the deuterated solvent for ¹H NMR spectra (CD₃OD: 3.31 ppm; CDCl₃: 7.26 ppm; DMSO-*d*₆: 2.50 ppm), for ¹³C {¹H} NMR to the carbon atom of the deuterated solvent (CD₃OD: 49.00 ppm; CDCl₃: 77.16 ppm; DMSO-*d*₆: 39.52 ppm), for ¹⁹F NMR spectra to external fluorobenzene at –164.9 ppm, and for ¹¹B NMR spectra to external BF₃•Et₂O at 0.00 ppm. Quantitative NMR experiments were used to quantify catalysis, using a standard delay time (D1) of 30 seconds. UV-vis spectra were measured with a Mettler-Toledo Easy UV spectrophotometer, pH with a Mettler-Toledo FiveEasy glass pH electrode (3.0 M KCl reference pH probe, calibrated using a set of 3 standardized buffer solutions: pH 4.00, 7.00, 11.00). Electrospray (ESI) mass spectra were acquired with a Micromass Q-TOF I Mass Spectrometer (Waters) on 30 µg/mL MeCN or MeOH solutions prepared under N₂, via injection of a 1 mL volume at 50 µL/min and nebulization with N₂ (70 psi) at 200 °C, using capillary and cone voltages of 3.5 and 40 kV, respectively, and a source temperature of 100 °C.



Chart 1. (a) CAAC iminium salts employed as precursors to sulfonated CAACS. (b) Catalysts employed.

S1.2. Synthesis of Ligands and Catalysts.

S1.2.1. Synthesis of C1s^{Me}•**HBF**₄. In a well-ventilated fumehood, a 50 mL round-bottom flask was charged with 18% fuming sulfuric acid (4 mL) and concentrated sulfuric acid (1 mL) in an ice bath. The mixture was stirred for 5 min, after which white solid C1^{Me}•HBF₄ (1.00 g, 2.90 mmol) was added in small portions over 20 min, turning into a red solution after the first 2 min. The red solution was allowed to warm to RT, stirred for 10 min, then slowly poured over ice in a 500 mL round-bottom flask bedded in an ice-bath. The



C1s^{Me}•HBF₄

resulting white suspension was neutralized with saturated NaOH to pH 7. The water was evaporated under vacuum, and the white residue was taken up in dry methanol and filtered to remove Na salts. Evaporation of the filtrate afforded a white solid, which was dried in vacuo for 2 days. Yield of $C1s^{Me} \cdot HBF_4$: 1.01 g, 2.25 mmol (78%).

¹H NMR (600 MHz, D₂O): δ 9.40 (s, 1H, *CH*N), 8.11 (d, ³*J*_{HH} = 8.4 Hz, 1H, NAr), 7.58 (d, ³*J*_{HH} = 8.4 Hz, 1H, NAr), 3.57 (m, 1H, *CH*HMe; diastereotopic), 2.73 (m, 1H, *CH*HMe; diastereotopic), 2.53 (s, 2H, *CH*₂; overlaps with *CH*HMe), 2.50 (m, 3H, *CH*HMe; diastereotopic, overlaps with CH₂), 1.66 (s, 3H, *CH*₃), 1.65 (s, 3H, *CH*₃), 1.58 (s, 3H, *CH*₃), 1.54 (s, 3H, *CH*₃), 1.24 (t, ³*J*_{HH} = 7.8 Hz, 3H, CH₂CH₃), 1.09 (t, ³*J*_{HH} = 7.1 Hz, 3H, CH₂CH₃). For fully-assigned ¹H NMR spectrum, see Figure S1. ¹³C{¹H} NMR (150 MHz, D₂O): δ 192.0 143.9, 140.6, 138.7, 132.0, 130.4, 128.0, 85.9, 48.9 47.9, 47.8, 27.8, 27.0, 25.6, 24.9, 23.1, 15.4, 14.3. ¹⁹F{¹H} NMR (150 MHz, D₂O): δ – 150.6 (s).

ESI-MS (MeOH): Calc'd for C₁₈H₂₇NO₃SH⁺ ([M–BF₄]⁺), *m/z* 338.1790. Found: *m/z* 338.1740.

S1.2.2. Synthesis of C1_s^{Cy}•HBF₄. As above, with C1^{Cy}•HBF₄ (1.01 g, 2.62 SO3- Na+ mmol). Yield of white crystalline C1s^{Cy}•HBF₄: 1.04 g, 2.13 mmol (82%). Et ¹H NMR (600 MHz, D₂O): δ 9.49 (s, 1H, CHN⁺), 8.13 (d, ³J_{HH} = 8.2 Hz, 1H, + BF₄-NAr), 7.59 (d, ${}^{3}J_{HH} = 8.2$ Hz, 1H, NAr), 3.57 (m, 1H, CHHMe; diastereotopic), 2.73 (m, 1H, CHHMe; diastereotopic), 2.61(m, 1H, CHHMe; diastereotopic), 2.55 (d, ${}^{3}J_{HH} = 7.7$ Hz, 2H, CH₂; overlaps with CHHMe), 2.52 (m, 1H, CHHMe; C1s^{Cy}·HBF₄ diastereotopic, overlaps with CH₂), 2.11–1.48 (m, 10H, Cy), 1.60 (s, 3H, CH₃), 1.54 (s, 3H, CH₃), 1.24 (t, ${}^{3}J_{HH} = 7.2$ Hz, 3H, CH₂CH₃), 1.09 (t, ${}^{3}J_{HH} = 7.2$ Hz, 3H, CH₂CH₃). For fully-assigned ¹H NMR spectrum, see Figure S2. ¹³C{¹H} NMR (150 MHz, D₂O): δ 191.4, 143.9, 140.6, 138.7, 132.2, 130.4, 128.0, 85.1, 53.1, 48.9, 45.3, 34.5, 33.6, 28.3, 27.4, 25.0, 24.3, 23.1, 21.2(5), 25.1(8) 15.4, 14.3. ${}^{19}F{}^{1}H{}$ NMR (150 MHz, D₂O): δ –150.5 (s). ESI-MS (MeOH): Calc'd for C₂₁H₃₁NO₃SNa⁺ ([M–BF₄]⁺), *m/z* 400.1917. Found: *m/z* 400.1919.

S1.2.3. Attempted synthesis of monosulfonated $C1_{s}^{Ph}$ -HBF₄. Carrying out this reaction as for $C1_{s}^{Me}$ -HBF₄, with $C1^{Ph}$ -HBF₄ (1.00 g, 2.46 mmol) resulted in formation of a 1:1 mixture of two polysulfonated products, and was therefore not pursued further. A ¹H NMR spectrum showing the product mixture is provided in Figure S3.



S1.2.4. Failed synthesis of $RuCl_2(H_2IMes-SO_3Na)(=CHAr)$ (HII-SO₃⁻ Na⁺) via direct sulfonation. In a well-ventilated fumehood, a 50 mL Schlenk flask connected to the Schlenk line was charged with 18% fuming sulfuric acid (1.00 mL) and concentrated sulfuric acid (0.25 mL) in an ice bath. The mixture was stirred for 5 min, after which green solid HII (50 mg, 0.080 mmol)

was added all at once, turning immediately into a black solution. The solution was allowed to warm to RT and was slowly poured into another Schlenk flask under N_2 bedded in an ice-bath. The resulting dark-brown suspension was neutralized with saturated NaOH to pH 5. The water was evaporated under vacuum, and the brown and white residue was taken up in dry methanol inside of a glovebox and filtered to remove Na salts. Evaporation of the filtrate afforded a brown solid. No product signals were observed by ¹H NMR analysis: Figure S4.

S1.2.5. Synthesis of $RuI_2(C1_S^{Me})(=CHAr)$, $HC1_S^{Me}-I_2$. A white suspension of $C1_S^{Me}$ ·HBF₄ (800 mg, 1.78 mmol, 2 equiv) and LiHMDS (297 mg, 1.78 mmol, 2.0 equiv) in 10 mL THF was transferred to a thermostatted oil bath set at 60 °C, and stirred for 10 min. The solution turned yellow within 5 min, and a homogeneous solution formed within 10 min. (In comparison, a heterogeneous mixture was present even after 4 h at RT). Dropwise addition to a green solution of HI-I₂ (700 mg, 0.893 mmol) in THF (10 mL) caused



immediate formation of a green-yellow suspension. The reaction was stirred at 60 °C, with periodic removal of aliquots for ³¹P NMR analysis (THF). Once no signal for **HI-I₂** remained (2 h), the solvent was evaporated under reduced pressure to give a dark green oil. Chromatography on silica gel in air (99:1 CH₂Cl₂:MeOH), isolation of the green band, and evaporation of solvent gave a green solid, which was washed with benzene (3×2 mL) to remove a yellow impurity and dried. Yield of green **HC1s^{Me}-I₂**: 502 mg, 0.463 mmol (52%).

¹H NMR (300 MHz, CD₃OD): δ 15.64 (s, 1H, [Ru]=*CH*), 8.26 (d, ³*J*_{HH} = 9 Hz, 1H, NAr), 7.65 (t, ³*J*_{HH} = 8 Hz, 1H, Ar *CH*), 7.53 (d, ³*J*_{HH} = 9 Hz, 1H, NAr), 7.16 (d, ³*J*_{HH} = 8 Hz, 1H, Ar *CH*), 7.00 (m, 1H, Ar *CH*), 6.86 (m, 1H, Ar *CH*), 5.35 (sept, ³*J*_{HH} = 5 Hz, 1H, *CH*Me₂), 3.31 (m, *CH*HMe + CAAC backbone *CH*₂; overlaps with solvent peak), 3.07 (m, 2H, *CH*HMe; diastereotopic), 2.61 (m, 1H, *CH*HMe; diastereotopic), 2.20 (s, 3H, *CH*₃), 1.90 (d, ³*J*_{HH} = 5 Hz, 3H, ^{*i*}Pr *CH*₃), 1.32 (s, 3H, *CH*₃), 1.16 (s, 3H, *CH*₃), 1.06 (t, ³*J*_{HH} = 8 Hz, 3H, *CH*₂*CH*₃), 0.90 (t, ³*J*_{HH} = 8 Hz, 3H, *CH*₂*CH*₃). For fully-assigned ¹H NMR spectrum, see Figure S5.

¹³C{¹H} NMR (125 MHz, CD₃OD): δ 290.3 ([Ru]=*C*H; not observed: detected by ¹H-¹³C HSQC), 271.3 (CAAC *C*:), 153.4, 145.8, 143.7, 143.2, 142.7, 138.5, 131.0, 129.7, 125.7, 123.8, 121.4, 113.7, 79.6, 75.6, 55.0, 50.6, 48.2, 32.0, 31.9, 28.4, 27.4, 26.1, 24.4, 21.8, 17.0, 13.8.

ESI-MS (MeCN): Calc'd for C₂₈H₃₈I₂NO₄SRu⁻ ([M–Na]⁻), *m/z* 839.9660. Found: *m/z* 839.9662.

S1.2.6. Synthesis of $\text{RuI}_2(\text{C1}_8^{\text{Cy}})$ (=CHAr), HC1_8^{Cy} -I₂. As for HC1_8^{Me} -I₂, using C1_8^{Cy} •HBF₄ (311 mg, 0.638 mmol, 2 equiv), LiHMDS (106 mg, 0.638 mmol, 2.0 equiv) and HI-I₂ (250 mg, 0.310 mmol). Yield of green HC1_8^{Cy} -I₂: 271 mg, 0.294 mmol (93%).

¹H NMR (300 MHz, CD₃OD): δ 15.82 (s, 1H, [Ru]=*CH*), 8.26 (d, ³*J*_{HH} = 8 Hz, 1H, NAr), 7.65 (m, 1H, Ar *CH*), 7.51 (d, ³*J*_{HH} = 8 Hz, 1H, NAr), 7.16 (d, ³*J*_{HH} = 9 Hz, 1H, Ar *CH*), 7.00 (m, 1H, Ar *CH*), 6.86 (m, 1H, Ar *CH*), 5.32 (sept,



= 9 Hz, 1H, Ar CH), 7.00 (m, 1H, Ar CH), 6.86 (m, 1H, Ar CH), 5.32 (sept, $HC1_{s}^{Cy}-I_{2}^{3}J_{HH} = 6$ Hz, 1H, CHMe₂), 3.35 (s, 2H, CAAC backbone CH₂), 3.13 (m, 3H, 3 diastereotopic CHHMe protons overlap), 2.55 (m, 2H, Cy + CHHMe; diastereotopic), 2.76–1.42 (m, 9H, Cy), 1.88 (br s, 6H, 'Pr CH₃), 1.31 (s, 4H, CH₃ + Cy), 1.14 (s, 3H, CH₃), 1.05 (t, ${}^{3}J_{HH} = 8$ Hz, 3H, CH₂CH₃), 0.98 (t, ${}^{3}J_{HH} = 8$ Hz, 3H, CH₂CH₃), 0.89 (m, 1H, Cy). For fully-assigned ¹H NMR spectrum, see Figure S6.

¹³C{¹H} NMR (150 MHz, CD₃OD): δ 293.0 ([Ru]=*C*H), 273.3 (CAAC *C*:), 154.8, 147.2, 145.1, 144.8, 144.2, 140.0, 132.5, 131.0, 127.0, 125.3, 122.7, 115.1, 80.9, 76.9, 63.0, 54.8, 44.5, 39.8, 39.0, 30.6, 29.3, 27.4, 26.7, 25.9, 24.5, 24.4, 23.3, 18.4, 15.2.

ESI-MS (MeCN): Calc'd for C₃₁H₄₂I₂NO₄SRu⁻ ([M–Na]^{-β}), *m/z* 879.9973. Found: *m/z* 879.9983.

S1.2.7. Failed synthesis of RuCl₂(C1s^{Me})(=CHAr) (HC1s^{Me}) via attemped ligand exchange with HI. As for the successful synthesis of HC1s^{Me}-I₂ above, but using C1s^{Me}•HBF₄ (134 mg, 0.28 mmol, 2 equiv), LiHMDS (48 mg, 0.28 mmol, 2.0 equiv) and HI (100 mg, 0.166 mmol). Yield of isolated impure green-yellow HC1s^{Me}: 8 mg, 0.01 mmol (7%). For ¹H NMR spectrum of this material, see Figure S7. NMR analysis of the crude reaction mixture (Figure S8) indicated the presence of benzyl derivative 1a and trifluoroborane adduct C1s^{Me}-BF₃ (1b).



Key signals for zwitterionic **1a**, observed in situ: ¹H-¹³C HMBC (CD₃OD): 3.81 (s, CH₂). Correlations: 131.4 (aromatic *C*H), 140.1 (4° aromatic *C*), 95.3 (4° aliphatic *C*), 154.5 ppm (4° iminium *C*). ESI-MS (MeOH): Calc'd for C₂₈H₃₉NO₄SNa⁺ ([M+Na]⁺), *m/z* 508.2492. Found: *m/z* 508.2471.



Key signals for C1s^{Me}-BF₃ (1b), observed in situ: see Table S1 and Figure S9.

ESI-MS (MeOH): Calc'd for C₁₈H₂₃BF₃O₃NS⁻ ([M–Na]⁻), *m/z* 404.1684. Found: *m/z* 404.1670.

Compound	Solvent	¹⁹ F (282 MHz)	¹¹ B{ ¹ H} (96 MHz)
$SO_{3}^{-} Na^{+}$ $Ft = BF_{3}$ $Ft = BF_{3}$ $C1_{s}^{Me}-BF_{3}$ $(1b)$	CD ₃ OD	-139.5 ppm (q, ${}^{1}J_{\text{F-B}}$ = 39 Hz)	$-0.52 \text{ ppm } (q, {}^{1}J_{B-F} = 41 \text{ Hz})$ (overlaps with BF ₄ ⁻)
C1 ^{Me} -BF ₂	C ₆ D ₆	-139.7 (q, ${}^{1}J_{\text{F-B}}$ = 36 Hz)	0.06 (q, ${}^{1}J_{B-F} = 36 \text{ Hz}$) (overlaps with BF ₄ ⁻)

Table S1. Key NMR signals for 1b (C1s^{Me}-BF₃): comparison with known⁹ values for C1^{Me}-BF₃.

S1.2.8. Failed synthesis of RuCl₂(C1_S^{Cy})(=CHAr) (HC1_S^{Cy}) via reaction with HI. The reaction was carried out as for HC1_S^{Me}-I₂, but using C1_S^{Cy}•HBF₄ (179 mg, 0.366 mmol, 2 equiv) and LiHMDS (61 mg, 0.366 mmol, 2.0 equiv) and adding the mixture to HI (110 mg, 0.183 mmol). Yield of impure brown-yellow solid containing HC1_S^{Cy}: 13 mg, 0.017 mmol (13%). For ¹H NMR spectrum, see Figure S10.



S1.2.9. Synthesis of RuCl₂(C1_S^{Me})(=CHAr), HC1_S^{Me}. Solid AgCl (82 mg, 0.57 mmol, 5 equiv) was added to a yellow-green solution of HC1_S^{Me}-I₂ (100 mg, 0.115 mmol) in MeOH. ¹H NMR

analysis after stirring at RT for 24 h revealed 10% of the mixed-halide species $HC1s^{Me}$ -I (Table S2), which disappeared over a further 24 h. The green suspension was filtered through Celite to remove Ag salts. The product was washed through with CH₂Cl₂ (3 × 5 mL), the combined filtrate was concentrated to a minimum volume, and hexanes was added. The precipitate was filtered off, washed with cold hexanes (3×1 mL), and dried under vacuum. Yield of green HC1s^{Me}: 76 mg, 0.11 mmol (92%).



¹H NMR (300 MHz, CD₃OD): δ 16.99 (s, 1H, [Ru]=*CH*), 8.31 (d, ³*J*_{HH} = 8 Hz, 1H, NAr), 7.61 (dd, ³*J*_{HH} = 8 Hz, ⁴*J*_{HH} = 2.1 Hz, 1H, Ar *CH*), 7.52 (d, ³*J*_{HH} = 8 Hz, 1H, NAr), 7.16 (d, ³*J*_{HH} = 8 Hz, 1H, Ar *CH*), 6.93 (m, 2H, Ar *CH*), 5.25 (sept, ³*J*_{HH} = 5 Hz, 1H, *CH*Me₂), 3.45 (m, 1H, *CH*HMe; diastereotopic), 3.31 (detected by ¹H-¹³C HMBC, CAAC backbone *CH*₂, overlaps with residual CHD₂OD), 2.86 (m, 1H, *CH*HMe; diastereotopic), 2.69 (m, 1H, *CH*HMe; diastereotopic), 2.47 (m, 1H, *CH*HMe; diastereotopic), 2.19 (s, 3H, *CH*₃), 1.97 (s, 3H, *CH*₃), 1.74 (d, ³*J*_{HH} = 5 Hz, 6H, ⁴Pr *CH*₃), 1.36 (s, 3H, *CH*₃), 1.13 (s, 3H, *CH*₃), 0.94 (t, ³*J*_{HH} = 7 Hz, 3H, *CH*₂*CH*₃), 0.82 (t, ³*J*_{HH} = 7 Hz, 3H, *CH*₂*CH*₃). For fully-assigned ¹H NMR spectrum, see Figure S11.

¹³C{¹H} NMR (125 MHz, CD₃OD): δ 295.2 ([Ru]=*C*H; not observed: detected by ¹H-¹³C HSQC) 268.3 (CAAC *C*:), 154.3, 147.1, 145.0, 144.6, 143.8, 140.0, 132.1, 130.9, 127.2, 124.6, 123.1, 114.5, 80.8, 76.3, 57.6, 52.5, 30.1, 29.8, 29.2, 28.4, 26.7, 25.2, 22.4, 22.4, 16.1, 14.7

ESI-MS (MeCN): Calc'd for C₂₈H₃₈Cl₂NO₄SRu⁻ ([M–Na]⁻), *m/z* 656.0944. Found: *m/z* 656.0889.

Complex	δ (ppm)	Proportion		
		At 24 h	At 48 h	
HC1s ^{Me}	16.99	90%	100%	
HC1s ^{Me} -I	16.37, 16.33 (rotamers)	10%	0%	
HC1s ^{Me} -I ₂	15.63	0%	0%	

Table S2. Chemical shifts and product distribution in halide exchange with HC1_s^{Me}.

S1.2.10. Attempted synthesis of $\operatorname{RuCl}_2(\operatorname{C1}_S^{\operatorname{Me}})(=\operatorname{CHAr})$, $\operatorname{HC1}_S^{\operatorname{Me}}$ via salt exchange with NaCl. To a yellow-green solution of $\operatorname{HC1}_S^{\operatorname{Me}}$ -I₂ (50 mg, 0.058 mmol) in MeOH (5 mL) was added NaCl (678 mg, 11.6 mmol, 200 equiv) an let stir at RT. After 24 h, ¹H NMR analysis (see Figure S12) showed 6% starting material and 31% of the mixed-halide species $\operatorname{HC1}_S^{\operatorname{Me}}$ -I. The suspension was stirred for an additional 24 h, after which 20% $\operatorname{HC1}_S^{\operatorname{Me}}$ -I and 3% $\operatorname{HC1}_S^{\operatorname{Me}}$ -I₂ remained. The suspension was filtered off and subjected to a second round of NaCl treatment for 48 h. ¹H NMR analysis showed 7% $\operatorname{HC1}_S^{\operatorname{Me}}$ -I remaining.

S1.2.11. Synthesis of $RuCl_2(C1s^{Cy})$ (=CHAr), $HC1s^{Cy}$. As for $HC1s^{Me}$, using $HC1s^{Cy}$ -I₂ (100 mg, 0.111 mmol), AgCl (79 mg, 0.55 mmol, 5.0 equiv) in MeOH (5 mL). Yield of green $HC1s^{Cy}$: 72 mg, 0.10 mmol (90%). Table S3 shows chemical shifts and yields of relevant species.

¹H NMR (300 MHz, CD₃OD): δ 17.12 (s, 1H, [Ru]=*CH*), 8.32 (d, ³*J*_{HH} = 8 Hz, 1H, NAr), 7.62 (dd, ³*J*_{HH} = 8 Hz, ⁴*J*_{HH} = 2 Hz, 1H, Ar *CH*), 7.53 (d, ³*J*_{HH} = 8 Hz, 1H, NAr), 7.15 (d, ³*J*_{HH} = 8 Hz, 1H, Ar *CH*), 6.96 (m, 2H, Ar *CH*), 5.24



(sept, ${}^{3}J_{\text{HH}} = 6$ Hz, 1H, CHMe₂), 3.44, (m, 1H, CHHMe; diastereotopic), 3.31 (detected by ${}^{1}\text{H}{}^{-13}\text{C}$ HMBC, CAAC backbone CH₂, overlaps with CHD₂OD), 2.86 (m, 1H, CHHMe; diastereotopic),

2.69 (m, 1H, C*H*HMe; diastereotopic), 2.47 (m, 1H, C*H*HMe; diastereotopic), 2.22–1.24 (m, 9H, Cy), 1.76 (d, ${}^{3}J_{HH} = 6$ Hz, 6H, , i Pr CH₃; overlaps with Cy), 1.35 (s, 3H, CH₃; overlaps with Cy), 1.15 (s, 3H, CH₃), 0.95 (t, ${}^{3}J_{HH} = 7$ Hz, 3H, CH₂CH₃), 0.82 (t, ${}^{3}J_{HH} = 7$ Hz, 3H, CH₂CH₃). For fully-assigned ¹H NMR spectrum, see Figure S13.

¹³C{¹H} NMR (150 MHz, CD₃OD): δ 297.6 ([Ru]=*C*H), 268.1 (CAAC carbene), 154.3, 147.1, 144.9, 144.7, 143.9, 139.9, 132.2, 130.9, 127.2, 124.7, 123.1, 114.6, 80.8, 76.2, 63.8, 54.8, 49.0, 38.2, 34.4, 30.4, 28.9, 26.8, 26.7, 25.2, 24.3, 23.7, 22.5, 16.1, 14.7.

ESI-MS (MeCN): Calc'd for C₃₁H₄₂Cl₂NO₄SRu⁻ ([M–Na]⁻), *m/z* 695.1265. Found: *m/z* 695.1270.

Complex	δ (ppm)	Proportion		
		At 24 h	At 48 h	
HC1s ^{Cy}	17.12	87%	100%	
HC1s ^{Cy} -I	16.52, 16.47 (rotamers)	13%	0%	
HC1s ^{Cy} -I ₂	15.83	0	0%	

Table S3. Chemical shifts and speciation in halide exchange with $HC1_S^{Cy}$.

S1.2.12. Determining the solubility of the sulfonated catalysts in various solvents. To a 4 mL vial charged with 20 mg solid catalyst, solvent was added via gas-tight syringe in 100 μ L increments. The vial was shaken after every portion of solvent was added. The volume was recorded when homogeneity was achieved. Table S4 shows the solubility data.

Catalyst	Water	MeOH	THF	CH ₂ Cl ₂	Hexanes
HC1s ^{Me} -I2	5	>100	40	>100	—
HC1s ^{Cy} -I ₂	2	>100	50	>100	_
HC1s ^{Me}	17	>100	33	>100	_
HC1s ^{Cy}	9	>100	40	>100	_

Table S4. Solubility of the sulfonated catalysts in mg/mL.

S1.3. Synthesis of Novel Uridine Substrate 7 and Metathesis Dimer 7'

S1.3.1. Synthesis of hex-1-enyl-tagged uridine 7. A 25 mL high-pressure vessel was charged with uridine (2.00 g, 8.19 mmol), hex-5-en-1-yl methanesulfonate (1.60 g, 9.01 mmol, 1.1 equiv), K_2CO_3 (2.30 g, 16.4 mmol, 2.0 equiv), and DMF (10 mL) in air, capped and stirred at 80 °C in an oil bath for 24 h. The salts were then filtered off, and the product washed through with EtOAc (2×30 mL). After adding water (2×20 mL), the aqueous layer was extracted with EtOAc (2×30 mL). The combined organic layer was then washed



with water (20 mL), dried (NaSO₄) and dried under vacuum at 50 °C for a day. The resulting white solid was purified by column chromatography (silica gel, MeOH/EtOAc 5:95) and recrystallized from boiling EtOAc, with hexanes as counter-solvent. Yield of white 7: 1.51 g, 4.50 mmol (55%).

¹H NMR (600 MHz, CDCl₃): 7.63 (d, ${}^{3}J_{HH} = 8.2$ Hz, 1H, =CHN), 5.78 (d, ${}^{3}J_{HH} = 8.0$ Hz,1H, CH=CH₂, overlaps with CHC=O), 5.76 (d, ${}^{3}J_{HH} = 7.6$ Hz, 1H, CHC=O, overlaps with CH=CH₂), 5.67 (d, ${}^{3}J_{HH} = 4.5$ Hz, 1H, CHN), 5.03–4.91 (m, 2H, =CH₂), 4.38 (m, 1 H, CHOH_C), 4.35 (m, 1H, CHOH_B), 4.21 (m, 1H, OCHCH₂), 4.06 (d, ${}^{3}J_{HH} = 3.0$ Hz, 1H, OH_C), 3.97 (m, 1H, CHHOH_A), 3.91 (dd, ${}^{3}J_{HH} = 7.8$, Hz, ${}^{4}J_{HH} = 3.2$ Hz, 2H, NCH₂), 3.90 (m, 1H, CHHOH_A), 3.22 (d, ${}^{3}J_{HH} = 4.0$ Hz, 1H, OH_B), 2.62 (br s, 1H, OH_A), 2.08 (m, 2H, CH₂CH=), 1.62 (q, ${}^{3}J_{HH} = 8.1$ Hz, 2H, NCH₂CH₂), 1.42 (q, ${}^{3}J_{HH} = 7.6$ Hz, 2H, CH₂CH₂CH=). For fully-assigned ¹H NMR spectrum, see Figure S14.

¹³C{¹H} NMR (150 MHz, CDCl₃): δ 162.7, 151.9, 138.8, 138.5, 114.9, 102.0, 93.7, 85.9, 75.3, 71.0, 62.2, 41.3, 33.5, 27.1, 26.3.

ESI-MS (MeOH): Calc'd for C₁₅H₂₂N₂O₆Na⁺ ([M+Na]⁺), *m/z* 349.1478. Found: *m/z* 349.1381.

S1.3.2. Synthesis and characterization of uridine dimer 7'. To a colourless solution of 7 (100 mg, 0.306 mmol, 100 equiv) in CH_2Cl_2 (5 mL) was added solid green $nGC1^{Ph}$ (10 mg, 0.15 mmol, 5 mol%). The resulting red suspension was stirred at RT for 24 h. After cooling the suspension to RT, the solvent was decanted, and the pink residue was washed with CH_2Cl_2 (3×2 mL) to yield a white solid (a mixture of product with 6% starting material) which was purified by chromatography on silica gel (MeOH/EtOAc 5:95). Yield of white 7': 35 mg, 0.11 mmol (37%).



¹H NMR (600 MHz, DMSO-*d*₆): 7.95 (d, ${}^{3}J_{HH} = 8.1$ Hz, 2H, =C*H*N), 5.80 (d, ${}^{3}J_{HH} = 5.0$ Hz, 2H, C*H*N), 5.75 (d, ${}^{3}J_{HH} = 8.1$ Hz, 2H, C*H*C=O), 5.38 (m, 1.2H, *E*-C*H*=), 5.33 (m, 0.4H, *Z*-C*H*=), 4.02 (m, 2H, C*H*OH_C), 3.97 (m, 2H, OC*H*CH₂), 3.85 (m, 2H, C*H*OH_B), 3.77 (m, 4H, NC*H*₂), 3.64 (m, 2H, C*H*HOH_A), 3.56 (m, 2H CH*H*OH_A), 1.98 (m, 4H, C*H*₂CH=), 1.50 (m, 4H, NCH₂C*H*₂), 1.29 (m, 4H, C*H*₂CH=). For fully-assigned ¹H NMR spectrum, see Figure S15.

¹³C{¹H} NMR (150 MHz, DMSO-*d*₆): δ 161.9, 150.7, 139.1, 130.0, 129.6, 100.9, 88.8, 84.8, 73.7, 69.6, 60.6, 31.7, 26.6, 26.4.

ESI-MS (MeOH): Calc'd for C₂₈H₄₀N₄O₁₂Na⁺ ([M+Na]⁺), *m/z* 647.2540. Found: *m/z* 647.2559.

S1.4. Catalytic Performance of Sulfonated Catalysts.

S1.4.1. Representative RCM reaction with diene 2. Diene 2 (16 mg, 0.10 mmol), NaCl (117 mg, 0.4 mmol, 20 equiv), and dimethyl sulfone, Me₂SO₂ (9 mg, 0.10 mmol, 1 equiv; internal standard) were dissolved in 0.92 mL D₂O; final concentration 100 mM 2. A 50 μ L aliquot was removed for NMR analysis to establish the initial ratio of 2:Me₂SO₂. To the stirred solution was added 0.1 mol% HC1s^{Me} (14 μ L of a stock solution of 10.1 mg HC1s^{Me} in 2.00 mL D₂O). Aliquots were removed periodically, quenched with KTp in THF (10 mg/mL; 10 equiv vs starting Ru) and analyzed (NMR). Table S5 shows conversions of 2 and yields of 2' (from the 2H olefinic signal for 2' at 5.73 ppm; Figure S16, assigned by analogy to the reported signal in CDCl₃ at 5.65 ppm).¹⁰

HO OH Ru HO OH 2 $D_2O, 2 M NaCl$ $2'$						
Catalyst	mol%	T (°C)	Atmosphere	Conversion (%)	Yield 2' (%)	TON
HC1s ^{Me} -I ₂	0.05	70	N_2	3	3	60
HC1s ^{Cy}	0.05	70	N_2	2	2	40
HC1s ^{Me}	0.05	70	N_2	32	32	640
HC1s ^{Me}	0.05	70	Air	23	23	460
AM	0.05	70	N_2	22	22	420
HC1s ^{Me}	0.5	70	N_2	100	100	200
HC1s ^{Me}	0.5	70	Air	95	95	190
HC1s ^{Me}	2^b	70	N_2	85	80	40
AM	2^b	70	N_2	100	30	15
HC1s ^{Me}	0.05	RT	N_2	91	9	180

Table S5. Yields, conversions, and TONs in RCM of diol 2 by water-soluble catalysts.

^aNumerical data for Fig. 3 in main text. Agreement in replicate run averages ±2%. ^bAt 4 h, 0 NaCl.

S1.4.2. RCM of 2 in 'BuOH:D₂O. As above, in 0.92 mL 'BuOH:D₂O (1:1). 'BuOH was removed under vacuum prior to NMR analysis. Yields and conversions appear in Table S6.

Table S6. Yields, conversions, and TONs for RCM of 2 in D₂O vs D₂O-^tBuOH.^a

НОСОН	0.050 mol% HC1 _S ^{Me}	ностон
2	2 M NaCl, 70 °C – C ₂ H ₄	2'

Solvent	[NaCl]	Conversion of 2 (%)	Yield of 2' (%)	TON
D ₂ O	0 M	99	0.5	10
1:1 D ₂ O: ^{<i>t</i>} BuOH	0	6	6	120
D_2O	2.0	32	32	640
1:1 D ₂ O: ^{<i>t</i>} BuOH	2.0	41	37	740

^{*a*}Agreement in replicate run averages $\pm 2\%$. Phase separation occurs with added NaCl.

S1.4.3. Representative procedure for metathesis dimerization (exemplified with known⁷ **6').** β-D-galactopyranoside **6** (13 mg, 0.05 mmol), NaCl (58 mg, 1.0 mmol, 20 equiv), and dimethyl sulfone (Me₂SO₂; 5 mg, 0.10 mmol, 1 equiv, internal standard) were dissolved in 0.93 mL D₂O; final concentration 50 mM **6**. A 50 µL aliquot was removed for NMR analysis to establish the starting ratio of **6** vs Me₂SO₂. To the stirred solution was added **HC1**s^{Me} (68 µL of a stock solution of 10.0 mg **HC1**s^{Me} in 2.00 mL D₂O) to give a catalyst loading of 1 mol%. Aliquots were removed periodically, quenched with KTp in THF (10 mg/mL; 10 equiv vs starting Ru) and analyzed (NMR). Yield of known dimer **6'** quantified by integration of the olefinic signal at 5.52 ppm (1H; Figure S17).⁷ Yields and TONs are given in Table 1 in the main text. Isomerization was assessed at high catalyst loading (1 mol %) to maximize its probability, as C=C migration is known to increase at higher proportions of Ru.^{7,11}

ESI-MS (MeOH). m/z = 519 (M+Na; 6'), 505 (M+Na–14; 6"; 7% vs Σ 6'+6"). The proportion of 6" is calculated based on the assumption of equal lifetimes for 6' and 6" (internal olefins differing by one methylene unit).

S1.5. Monitoring Catalyst Stability by UV-Vis Spectroscopy.

S1.5.1. Stability of HC1s^{Me} in water. To a quartz cuvette was added H₂O (1.97 mL, pH = 7 prior to catalyst addition), and a 30 μ L aliquot of a stock solution of **HC1**s^{Me} in water (4.2 mg/mL), to give a final Ru concentration of 30 μ M. The cuvette was sealed, wrapped with Parafilm and removed from the glovebox to the spectrometer. The first UV-vis spectrum was taken 5 min after preparing the catalyst stock solution. Subsequent spectra were recorded periodically up to 24 h. UV-vis spectra showing the stability of **HC1**s^{Me} vs **AM** appear in Figure 2 in the main text.

<u>With NaCl</u>: Solid NaCl (234 mg, 4.00 mmol) was added to the cuvette prior to catalyst. UV-vis spectra for $HC1_{s}^{Me}$ and AM appear in Figure S18.

S2. NMR Spectra.





(b) ${}^{13}C{}^{1}H$ NMR spectrum of C1s^{Me}•HBF₄



Figure continues next page



(c) ^{1}H - ^{1}H COSY NMR spectrum of C1s^{Me}•HBF₄





Figure S1. NMR characterization of CAAC salt $C1s^{Me}$ •HBF₄ in D₂O.(a) ¹H NMR (600 MHz). (b) ¹³C {¹H} NMR (150 MHz). (c) ¹H-¹H COSY NMR (600 MHz). (d) ¹H-¹³C HSQC NMR (600/150 MHz). Grey dashed lines indicate key 4° carbon signals (note absence of ¹*J*_{HC} correlations). (e) ¹H-¹³C HMBC NMR (600/150 MHz).

(a) ¹H NMR spectrum of $C1_{S}^{Cy}$ •HBF₄





Figure continues next page

(e) ¹H-¹³C HMBC NMR spectrum of C1s^{Cy}•HBF₄



Figure S2. NMR characterization of CAAC salt $C1_{s}^{Cy}$ •HBF₄ in D₂O. (a) ¹H NMR (600 MHz). (b) ¹³C {¹H} NMR (150 MHz). (c) ¹H-¹H COSY NMR (600 MHz). (d) ¹H-¹³C HSQC NMR (600/150 MHz). Grey dashed lines indicate key 4° carbon signals (note absence of ¹*J*_{HC} correlations). (e) ¹H-¹³C HMBC NMR (600/150 MHz).



Figure S3. ¹H NMR spectrum (300 MHz, D₂O) of C1s^{Ph}•HBF₄ after sulfonation, showing mixture of products.



Figure S4. ¹H NMR spectrum (300 MHz, D_2O) after attempted sulfonation of HII. The inset shows the loss of signals in the alkylidene region.

(a) ¹H NMR spectrum of $HC1s^{Me}$ -I₂





(c) $^{1}\text{H}\text{-}^{1}\text{H}$ COSY NMR spectrum of HC1s^{Me} -I₂

(e) ¹H-¹³C HMBC NMR spectrum of HC1s^{Me}-I₂



Figure S5. NMR characterization of synthetic intermediate $HC1s^{Me}-I_2$ in CD₃OD. (a) ¹H NMR (300 MHz; inset shows alkylidene signal. (b) ¹³C{¹H} NMR (150 MHz; alkylidene C not observed; located by ¹H-¹³C HSQC). (c) ¹H-¹H COSY NMR (600 MHz). (d) ¹H-¹³C HSQC NMR spectrum (600/150 MHz). (e) ¹H-¹³C HMBC NMR (600/150 MHz).

(a) ¹H NMR spectrum of $HC1_S^{Cy}-I_2$







Figure continues next page

(e) ¹H-¹³C HMBC NMR spectrum of HC1_s^{Cy}-I₂



Figure S6. NMR characterization of $HC1_S^{Cy}-I_2$ in CD₃OD. (a) ¹H NMR (300 MHz; inset shows alkylidene signal, (†) indicates residual CH₂Cl₂). (b) ¹³C{¹H} NMR (150 MHz; alkylidene C not observed; located by ¹H-¹³C HSQC). (c) ¹H-¹H COSY NMR (600 MHz). (d) ¹H-¹³C HSQC NMR (600/150 MHz). Grey dashed lines indicate key 4° carbon signals (note absence of ¹*J*_{HC} correlations). (e) ¹H-¹³C HMBC NMR (600/150 MHz).



Figure S7. ¹H NMR spectrum (300 MHz, isopropanol-d₇) of isolated crude HC1s^{Me}, prepared from HI.



(a) ¹H NMR spectrum showing the benzylidene abstraction product 1a

(c) ¹H-¹H COSY NMR spectrum showing the benzylidene abstraction product **1a**





(e) ¹H-¹³C HSQC NMR spectrum showing the benzylidene abstraction product **1a**

Figure S8. NMR spectra in CD₃OD, showing benzylidene abstraction product **1a** formed in the reaction of $HC1_{s}^{Me}$ with HI. (a) ¹H NMR (600 MHz). (b) ¹H-¹H COSY NMR (600 MHz). (c) ¹H-¹³C HSQC NMR (600/150 MHz). Grey dashed lines indicate key 4° carbon signals (note absence of ¹J_{HC} correlations). (d) ¹H-¹³C HMBC NMR (600/150 MHz).

(a) ¹⁹F NMR showing the borylation product **1b**.





Figure S9. NMR spectra for the crude reaction mixture from the synthesis of $HC1_S^{Me}$ via HI, showing signals for borane adduct $C1_S^{Me}$ -BF₃ (1b). (a) ¹⁹F NMR (282 MHz, CD₃OD). (b) ¹¹B{¹H} NMR (96 MHz, CD₃OD). Note overlap with BF₄⁻ singlet in (b).



Figure S10. ¹H NMR spectrum (300 MHz, CD₃OD) of isolated impure $HC1s^{Cy}$ prepared by ligand exchange with HI. For comparison, see spectra for material prepared via ligand exchange with HI-I₂ in Figure S13.

(a) ¹H NMR spectrum of $HC1s^{Me}$



(c) $^{1}\text{H}^{-1}\text{H}$ COSY NMR spectrum of $\text{HC1}_{\text{S}}^{\text{Me}}$



(e) ¹H-¹³C HMBC NMR spectrum of HC1s^{Me}



Figure S11. NMR characterization of $HC1s^{Me}$ in CD₃OD. (a) ¹H NMR (300 MHz, CD₃OD); inset shows alkylidene signal. (b) ¹³C{¹H} NMR (150 MHz). Alkylidene carbon signal not observed. (c) ¹H-¹H COSY NMR (600 MHz). (d) ¹H-¹³C HSQC NMR (600/150 MHz) showing correlation for alkylidene carbon. (e) ¹H-¹³C HMBC NMR (600/150 MHz).



Figure S12. ¹H NMR (300 MHz, CD₃OD) spectrum for the reaction of $HC1_S^{Me}-I_2 + 200$ equiv NaCl, after 24 h.

(a) ¹H NMR spectrum of $HC1s^{Cy}$



Figure continues next page

(c) $^{1}\text{H}^{-1}\text{H}$ COSY NMR spectrum of $\text{HC1}_{\text{S}}^{\text{Cy}}$



(d) ^{1}H - ^{13}C HSQC NMR spectrum of HC1s^{Cy}



Figure continues next page

(e) ¹H-¹³C HMBC NMR spectrum of HC1s^{Cy}



Figure S13. NMR characterization of $HC1_S^{Cy}$ in CD₃OD. (a) ¹H NMR (300 MHz; inset shows alkylidene signal. (b) ¹³C{¹H} NMR (150 MHz). (c) ¹H-¹H COSY NMR (600 MHz). (d) ¹H-¹³C HSQC NMR spectrum (600/150 MHz) (e) ¹H-¹³C HMBC NMR (600/150 MHz).

(a) ¹H NMR spectrum of novel uridine substrate 7





(c) $^{1}\text{H-}^{1}\text{H}$ COSY NMR spectrum of novel uridine substrate 7

Figure continues next page



(e) ¹H-¹³C HMBC NMR spectrum of novel uridine substrate 7

Figure S14. NMR characterization of uridine-tagged substrate 7 in CDCl₃. (a) ¹H NMR (600 MHz). (b) ${}^{13}C{}^{1}H{}$ NMR (150 MHz). (c) ${}^{1}H{}^{-1}H$ COSY NMR (600 MHz). (d) ${}^{1}H{}^{-13}C$ HSQC NMR spectrum (600/150 MHz). (e) ${}^{1}H{}^{-13}C$ HMBC NMR (600/150 MHz).



(a) ¹H NMR spectrum of novel uridine-tagged dimer 7'





(c) ¹H-¹H COSY NMR spectrum of uridine-tagged dimer 7'

(d) ¹H-¹³C HSQC NMR spectrum of uridine-tagged dimer 7'



Figure continues next page



Figure S15. NMR characterization of novel uridine dimer 7', prepared via self-metathesis, in DMSO-*d*₆. (a) ¹H NMR (600 MHz). (b) ¹³C{¹H} NMR (150 MHz). (c) ¹H-¹H COSY NMR (600 MHz). (d) ¹H-¹³C HSQC NMR (600/150 MHz). (e) ¹H-¹³C HMBC NMR (600/150 MHz).



Figure S16. Quantifying RCM of **2** by $HC1_S^{Me}$ (0.05 mol%). Representative ¹H NMR spectrum (400 MHz, D₂O) at 32% conversion. Internal standard (IS) = dimethylsulfone, Me₂SO₂.



Figure S17. Quantifying dimerization of **6** by $HC1_{s}^{Me}$ (1 mol%). Representative ¹H NMR spectrum (300 MHz, D₂O) at 100% conversion. Internal standard (IS) = dimethylsulfone, Me₂SO₂.



Figure S18. UV-vis spectra in $H_2O + 2$ M NaCl, showing λ max for dichlororuthenium complexes. (a) AM. (b) HC1s^{Me}.

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