Bioinformatic, enzymatic and structural characterization of Trichuris suis hexosaminidase HEX-2

Zuzanna Dutkiewicz^{1,†}, Annabelle Varrot², Karen J. Breese³, Keith A. Stubbs^{3,4}, Lena Nuschy¹, Isabella Adduci⁵, Katharina Paschinger¹ and Iain B. H. Wilson^{1,*}

¹ Institut für Biochemie, Department für Chemie, Universität für Bodenkultur, Muthgasse 18, 1190

Wien, Austria

² Univ. Grenoble Alpes, CNRS, CERMAV, 38000 Grenoble, France

³ School of Molecular Sciences, University of Western Australia, Crawley, WA 6009, Australia

⁴ ARC Training Centre for Next-Gen Technologies in Biomedical Analysis, School of Molecular

Sciences, University of Western Australia, Crawley, WA 6009, Australia

⁵ Institut für Parasitologie, Department für Pathobiologie, Veterinärmedizinische Universität Wien,

Veterinärplatz 1, A-1210 Wien, Austria

+ Current address: Institut für Mikrobiologie, Universität Innsbruck, A-6020 Innsbruck, Austria

* Correspondence to: iain.wilson@boku.ac.at

Supporting Information

Supplementary Figures S1-S6



Supplementary Figure S1A: Full phylogenetic tree of nematode subfamily 1 GH20 hexosaminidases – these data underly **Figure 2** in the main text. *D. melanogaster* FDL (fused lobes hexosaminidase) was used as an out group. Bootstrap values of more than 70 are shown.



Supplementary Figure S1B: Full phylogenetic tree of all nematode GH20 hexosaminidases – these data underly **Figure 2** in the main text. *D. melanogaster* FDL was used as an out group. Bootstrap values of more than 70 are shown.

A) Full theoretical sequence:

1 MTGKKCAFVQ HKYSKLRQDY VLQDDNEVAT GRSVADSNVR RYFTVMKVYR WRGKPAFAFI TVLTVILLII GYHTITSRHG 81 DTVIHEGVFQ RGAAMRKRTV YGQVDEKSSP TSTSTDLTMK TTTTYSQNLK EPPPSNVFIP KRRIVHLDLK GAAPKPQHFR 161 AFFEYFVRIG ATGILIEWED MFPYEGRLSD LRNGDAYSAD DVRMILSTAD QLRLEVIPLV QTIGHLEWLL KTHKFYSFRE 241 NPRNPQSVCV SNAEAVDLVL HLVDQVMAFH KDYGQFV**HIG ADE**VYQYGEC SRCVARMNKE NLRREDLLLR HIV<u>N</u>SKHVK 321 TKYGKNVLMW HDMIANIDAS LAEKYDLKNL VEPVLWNYAE DLEAFLPMGI WETFSAMVPY MWGSSAFKGA DSPTRYHSNV 401 KHYLENHISW IKQMSTASEK FREFRGLIFT GWQRYDHFAV LCEFLPIGIP SLTVNMLTIR NGRFDASVND QAISIMQCVT 481 GSDVKGDLYG CRFPGSDIYH HVQLLHEKKG EIEKLLLQQS VQGWLSNIAI DY<u>NMS</u>SPWYM NLIVPDLMTY KNQMIELSLN 561 IRQAMLEMFY ENAVDEFLFT YVDPVINHLQ RLLDRATAIQ RRDEFPVRPF PIKRTIDTTR

B) Long secreted form:

HEGVFQ RGAAMRKRTV YGQVDEKSSP TSTSTDLTMK TTTTYSQNLK EPPPSNVFIP KRRIVHLDLK GAAPKPQHFR
161 AFFEYFVRIG ATGILIEWED MFPYEGRLSD LRNGDAYSAD DVRMILSTAD QLRLEVIPLV QTIGHLEWLL KTHKFYSFRE
241 NPRNPQSVCV SNAEAVDLVL HLVDQVMAFH KDYGQFVHIG ADEVYQYGEC SRCVARMNKE NLRREDLLLR HIV<u>NVS</u>KHVK
321 TKYGKNVLMW HDMIANIDAS LAEKYDLKNL VEPVLWNYAE DLEAFLPMGI WETFSAMVPY MWGSSAFKGA DSPTRYHSNV
401 KHYLENHISW IKQMSTASEK FREFRGLIFT GWQRYDHFAV LCEFLPIGIP SLTVNMLTIR NGRFDASVND QAISIMQCVT
481 GSDVKGDLYG CRFPGSDIYH HVQLLHEKKG EIEKLLLQQS VQGWLSNIAI DY<u>NMS</u>SPWYM NLIVPDLMTY KNQMIELSLN
561 IRQAMLEMFY ENAVDEFLFT YVDPVINHLQ RLLDRATAIQ RRDEFPVRPF PIKRTIDTTR AVDHHHHH

C) Alignment with C. elegans HEX-2

Query	124	TYSQNLKEPPPSNVFIPKRRIVHLDLKGAAPKPQHFRAFFEYFVRIGATGILIEWEDMFP	183
Sbjct	96	TQEVKIERPSRDNEFY-KNVVIHFDLKGAPPKVDYFLDLLRLIAKGGATGILLEWEDMFP	154
Query	184	YEGRLSDLRNGDAYSADDVRMILSTADQLRLEVIPLVQTIGHLEWLLKTHKFYSFRENPR + G+L +N DAYS DV MILS A +L+L+VIPLVOT GHLEW+LK + +REN	243
Sbjct	155	WTGKLEQFKNTDAYSESDVDMILSEATKLKLDVIPLVQTFGHLEWILKYEEMRKYRENDA	214
Query	244	NPQSVCVSNAEAVDLVLHLVDQVMAFHKDYG-QFV HIGADE VYQYGECSRCVARMNKE PO +C+ N E V+ V ++ OV H YG F HIGADE +++G C + + N +	300
Sbjct	215	YPQVLCLGNEEGVEFVREMIRQVAKKHAKYGIPFF HIGADE AFEFGVCQESLDWIKKNGK	274
Query	301	NLRREDLLLRHIV <u>NVS</u> KHVKTKYGKNVLMWHDMIANIDASLAEKYDLKNLVEPVLWNY N R++ L L H+ +++ K + G + +L WHDM+ + D+ L + +L +++PV+W+Y	358
Sbjct	275	NGRKQLLALAHLKAIAEFAKQQTGDSTQILAWHDMLKDFDSRLIKNLELGQIIQPVVWDY	334
Query	359	AEDLEAFLPMGIWETFSAMVPYMWGSSAFKGADSPTRYHSNVKHYLENHISWIKQMSTAS +E++ L I+ + P MW SSA+KGA+ P+ S V+HY N+ +WI+	418
Sbjct	335	$\verb+SENI-ITLNDYIFSALAENFPTMWASSAYKGANYPSASTSEVRHYETNNRNWIRTKQNQE$	393
Query	419	EKFRE-FRGLIFTGWQRYDHFAVLCEFLPIGIPSLTVNMLTIRNG-RFD-ASVNDQAISI KF+ F+G+I TGWORYDH A LCE LPIG S+ + M N D +A ++	475
Sbjct	394	RKFKNGFQGIIVTGWQ RYDHLAGLCETLPIGTASMMLQMQIALNAPALDLEGTRQKAATL	453
Query	476	MQCVTGSDVKGDLYGCRFPGSDIYHHVQLLHEKKGEIEKLLLQQSVQGWLS ++C V G V + C++ G Y O L E+ K + GW +	526
Sbjct	454	LECQGFNVDGVKVVSNQCKYRGFQTYLIYQSEVPNLFARIDSELSKNHHLMGWAN	508
Query	527	NIAIDYNMSSPWYMNLIVPDLMTYKNQMIELSLNIRQAMLEMFYENAVDEFLFTYVDPVI YN+S WY ++P + O + ++R +M ++++EN +DEF++ + +	586
Sbjct	509	$RYNRKY\underline{NIS}QNWYHREMLPFVQQLVGQYDRVESDLRASMKDLYFENTIDEFIYENLGEMS$	568
Query	587	NHLQRLLDRATAIQRRDEFPVRPFPIKR 614 L L+ + + + P R FPIK+	
Sbict	569	EKLHGYLEEIORLDKLRAWPKRHFPIKK 596	

Supplementary Figure S2: (A) Sequence of the predicted subfamily 1 hexosaminidase (HEX-2) from *T. suis* (hypothetical protein M514_07548; NCBI accession KFD67184). The predicted transmembrane domain absent from all recombinant protein constructs is in red, the potential N-glycosylation sites are underlined and the conserved HIGADE active site region is indicated in bold; the actual initial methionine residue is unknown but two are N-terminal to the transmembrane domain. (B) Sequence of the recombinant long secreted form (expected M_r 63 kDa) used for X-ray crystallography; the N-terminal disordered region absent from the recombinant short secreted form (expected M_r 57 kDa) is italicized. (C) Alignment of *T. suis* and *C. elegans* HEX-2 (36% identity over 508 residues).

	41	5,0	60	7.0	8,0	9.0 10.0
TsHEX-2 CeHEX-2 Hs_HexA Hs_HexB Dm_fdl OfHex1	YSQNLKEP QEVKIERP PRFP PRFS PKFR PVYP	PPSNVFIP SRDNEFY. HRGLLLDT HRGILIDT YRGLMLDT YRGILLDT	KRRIVHLDLKGA KNVVIHFDLKGA SRHYLPLSSILD SRHYLPVKIILK SRHFFSVESIKR ARNYYSIESIKR	APKPQHFRAFI PPKVDYFLDLI TLDVMAYNKLI TLDAMAFNKFI TIVGMGLAKMI TIEAMAAVKLI	YEYFV <mark>RI</mark> GAT LRLIAKGGAT VVFHWHLVDD VVLHWHIVDD VLHWHIVDD VTFHW <mark>HITD</mark> S	GILIEWEDMFPY GILLEWEDMFPW PSFPYESFTFPE QSFPYQSITFPE QSFPYISRYYPE QSFPFVTTK <mark>RP</mark> N
TSHEX-2 CeHEX-2 Hs_HexA Hs_HexB Dm_fdl OfHex1	EGRLSDLR TGKLEQFK LMRKGSYN LSNKGSYS LAVHGAYS LYKFGALS	110 N.GDAYSAI N.TDAYSE PVTHIYTA LSHVYTPI E.SETYSE P.QKVYTK	120 DVRMILSTADQ SDVDMILSEATK QDVKEVIEYARL NDVRMVIEYARL QDVREVAEFAKI AAIREVVRFGLE	130 LRLEVIPLVQJ RGIRVLAEFDJ RGIRVLAEFDJ YGVQVIPEIDZ RGVRVLPEFDZ	140 FIGHLEW. FFGHLEW. FPGHT.LSWG FPGHT.LSWG APAHAGNGWD APAH	LLKTH PGLLTP KGQKDLLTP WGPKRGMGELAM DTDLTV
TSHEX-2 CeHEX-2 HS_HexA HS_HexB Dm_fdl OfHex1	150 KFYSFREN EMRKYREN CYSGSEPS CYSRQNKL CINQQPWS CFKAEPWK	160 PRNPQSVC DAYPQVLC GTF DSF FYCGEPPC SYCVEPPC	170 VSNAEAVDLVLH LGNEEGVEFVRE SPVNPSLNNTYE GPINPTLNTTYS GQLNPKNNYTYL GQLNPTKDELYQ	180 LVDQVMAFHKI MIRQVAKKHAH FMSTFFLEVSS FLTTFFKEISH ILQRIYEELLQ YLEDIYSDMAH	190 DYG.QFVH XYGIPFFH SVF.PDFYLH SVF.PDQFIH DHTGPTDFFH SVFDTTDIFH Cata	200 IGADEVYQYG IGADEAFEFG LGGDEVDFTCWK LGGDEVFKCWE LGGDEVNLDCW. MGGDEVSEACWN Ilytic site
TSHEX-2 CeHEX-2 Hs_HexA Hs_HexB Dm_fdl OfHex1	210 ECSRCV VCQESLDW SNPEIQDF SNPKIQDF .AQYFNDT SSDSIQNF	ARMNKENLI IKKNGKNGJ MRKKGFG. MRQKGFG. DLRGLWC. MMQNRWDLI	220 RREDLLLRHI RKQLLALAHL EDFKQLESFY .TDFKKLESFY DFML DKESFLKLWNYF	230 VNVSKHVKTKS KAIAEFAKQQT IQTLLDIVSS IQKVLDIIAT QAMARLKLANN QQKAQDKAYKA	240 (GKNVLM TGDSTHQILA (GKGYVV INKGSIV IGVAPKHVAV AFGKKLPLIL	250 WHDMIANIDASL WHDMLKDFDSRL WQEVFDNK WSSALTNTK WTSTLTNYKHID
TSHEX-2 CeHEX-2 HS_HexA HS_HexB Dm_fdl OfHex1	260 AEKYDLKN IKNLELGQ .VKIQPDT .AKLAPGT .CLPNSQF DYLNKDDY	270 LVEPVLWN IIQPVVWD IQVWRED IVEVWKDS TVQVWGGS IIQVWTTG	280 YAEDLEAFIPMG YSENIIT.LNDY IPVNYMKELE AYPEEIS IWQENYDLLDNG VDPQIKGLLEKG	290 IWETFSAMVPS IFSALAENFPS LVTKAGFRAI .RVTASGFPVJ YNVIFSHVDAV YRLIMSNYDAI	300 YMWGSSAFKG TMWASSAYKG LLSAPW¥LNR LLSAPW¥LDL V¥LDCGFGSW L¥FDCGYGAW	310 ADSPTRYHSNVK ANYPSASTSEVR ISYGPDWKDFYV ISYGQDWRKYYK RATGDAACAPYR VGAGNNWCSPYI
TSHEX-2 CeHEX-2 HS_HexA HS_HexB Dm_fdl OfHex1	320 HYLENHIS HYETNNRN VEPLAFEG VEPLDFGG TWQNVYKH GWOKVYDN	330 WIKQMS' WIRTKQNQ TP TQ RPWERMRL SPAVI	340 FASEKREFRGLI ERKFKNGFQGII EQKALVIGGE KQKQLFIGGE DKKRKKQVLGGE	350 FTGWQRYDHF# VTGWQRYDHL# ACMWGEYVDNT ACLWGEYVDAT VCMWTEQVDEN AALWSEOSDTS	360 AVLCEFL.PI AGLCETL.PI INLVPRLWPR INLTPRLWPR GLDRRLWPR	370 GIPSLTVNMLTI GTASMMLQMQIA AGAVAERLWSNK ASAVGERLWSSK FAALAERLWAEP

Supplementary Figure S3: Alignment of T. suis and C. elegans HEX-2 (both subfamily 1), human HEXA and HEXB, Drosophila melanogaster FDL and Ostrinia furnacalis OfHex1 (subfamily 2). Sequences were shortened to the region of highest homology and aligned using Multalin, http://multalin.toulouse.inra.fr/ - the file was processed on https://espript.ibcp.fr/ESPript/ESPript/Crystal structures exist for human hexosaminidases A and B as well as OfHex1, while HEX-2 and FDL specifically remove the β 1,2GlcNAc from the α 1,3-mannose of N-glycans. Only the catalytic site as well as a region towards the N-terminus is well conserved between the two subfamilies. Highlighted in blue are selected key subfamily-specific residues identified in this and other structural studies (other than those conserved across the families in red).



Supplementary Figure S4: (A) Expression of the recombinant secreted 'long' form of *T. suis* HEX-2 (expected M_r 63 kDa; N-terminal His/FLAG-tag) as detected by anti-FLAG Western blotting of *Pichia* culture supernatants after 24, 48 or 72 hours of induction. (B) Native gel electrophoresis of bovine serum albumin (BSA) and *T. suis* HEX-2 followed by either Coomassie or silver staining suggests that HEX-2 exists as in multimeric forms; BSA has monomeric/dimeric/trimeric molecular masses of 66, 132 and 198 kDa, whereas the dimeric form of HEX-2 would have a mass of 150 kDa as compared to the 75 kDa observed on the reducing/denaturing SDS-PAGE and Western blot. (C) Activity towards pNP-β-GalNAc and pNP-β-GlcNAc of *T. suis* HEX-2 and *C. elegans* HEX-4 expressed in *Pichia pastoris*; the turnover of the aryl GalNAc substrate is some three- or fourfold higher than that of the aryl GlcNAc in keeping with data on other subfamily 1 GH20 hexosaminidases. (D) Activity of *T. suis* HEX-2 at different pH values towards GnGn-PA; the appearance of the GnM-PA product of later RP-HPLC elution time (31.5 mins; see also *Figure 4* of the main text) is most pronounced at pH 6 and 7, in keeping with the optimum found with the aryl GalNAc substrate (see *Figure 3* of the main text). (E and F) Activity towards pNP-β-GalNAc of *T. suis* HEX-2 in the absence or presence of up to 10 mM EDTA or ZnCl₂, indicating minimal effect of divalent cations.



Supplementary Figure S5: (A) Electron density maps in the disordered area of the *T. suis* HEX-2 structure, visualized with Coot (version 0.9.8.3). Difference map Fo-DFc map is represented in green (positive) and red (negative) at 3 sigmas whilst 2Fo-DFc is represented in blue at 1 sigma (0.23 eÅ³). (B) Cartoon and surface representation of crystal structure of T. suis HEX-2 at 2.55 Å. The grey ball represents the Zn²⁺ ion, the binding site pocket is colored in orange with presumed catalytic amino acids displayed in sticks. (C) Comparison of the *T. suis* HEX-2 X-ray structure and the Alpha Fold model, coloured by RMSD (blue specifying the minimum pairwise RMSD and red indicating the maximum, unaligned residues are coloured grey), visualized with pymol (free version 2.5.0, Schrodinger LLC). See also *Figures 6 and 7* of the main text for other visualisations.



Supplementary Figure S6: Superimposition of *T. suis* HEX-2 (8QK1) and *S. pneumoniae* GH20C (5A6J) monomeric crystal structures coloured by RMSD as visualized with pymol (free version 2.5.0, Schrodinger LLC). The distances between aligned C-alpha atom pairs are stored as B-factors of these residues, which are coloured by a spectrum ranging from blue specifying the minimum pairwise RMSD and red indicating the maximum. Unaligned residues are coloured grey. The barrels containing the binding pocket superimpose well, but the N- and C-termini differ (in this view, the N-terminus of HEX-2 is hidden under the barrel). Sequence identity of HEX-2 and GH20C is 29%. See also *Figure 6* of the main text for other visualisations.