Expanded View Figures



Figure EV1. Mis18 α and Mis18 β contain two domains capable of oligomerising.

(A, B) Domain architecture and amino acid conservation of (A) Mis18 α and (B) Mis18 β . Alignments include *Homo sapiens* (*hs*), *Bos taurus* (*bt*), *Mus musculus* (*mm*) and *Gallus gallus* (*gg*). The conservation score is mapped from red to cyan, where red corresponds to highly conserved and cyan to poorly conserved. Secondary structures as annotated/predicted by Conserved Domain Database [CDD] and PsiPred, http://bioinf.cs.ucl.ac.uk/psipred. Multiple sequence alignments were performed with MUSCLE (Madeira et al, 2019) and edited with Aline (Bond and Schüttelkopf, 2009). Dashed boxes highlight Yippee domains whilst solid boxes highlight C-terminus α -helices. (C) Superposition of Mis18 β_{Yippee} structures predicted by AlphaFold (light pink) and RaptorX (green). RaptorX generated five models and the model with the lowest estimated error (1.9 Å) is shown here. The AlphaFold and RaptorX models superpose well with an RMSD of 0.95 Å. (D) The PAE plot corresponding to the Mis18 $\alpha/\beta_{\text{Yippee}}$ AlphaFold model shown in Fig. 1D.



Figure EV2. SAXS analysis of Mis18 $\alpha/\beta \Delta N$, Mis18 α/β and Mis18_{core}.

(A) SAXS scattering curves of Mis18 $\alpha/\beta \Delta N$, Mis18 α/β and Mis18_{core}. (B) Guinier Plot showing *Rg* of 53 Å, 60 Å, and 63 Å for Mis18 $\alpha/\beta \Delta N$, Mis18 α/β and Mis18_{core}, respectively. (C) Modified Guinier Plot showing *Rc* of 26 Å, 30 Å, and 31 Å for Mis18 $\alpha/\beta \Delta N$, Mis18 α/β and Mis18_{core}, respectively. (D) SAXS *P(r)* distributions showing maximum dimensions of 190 Å, 215 Å, and 230 Å for Mis18 $\alpha/\beta \Delta N$, Mis18 α/β and Mis18_{core}, respectively.



Figure EV3. Structural characterisation of the Mis18_{core} complex.

(A) Representative micrograph of negative staining EM of the Mis18a/Mis18B/Mis18BP1₂₀₋₁₃₀ (Mis18_{core}) complex cross-linked using GraFix (Kastner et al, 2008; Stark, 2010). Beneath is the corresponding SDS-PAGE analysis of fractions from GraFix, fractions 8 and 9 were used to make grids. (B) Two models (Class II-III) generated for Mis18_{core} from negative staining EM analysis. All show that the overall shapes of the Mis18_{core} cresemble a telephone handset with 'ear' and 'mouth' pieces assuming different relative orientations. (C) Cartoon representation of the model of Mis18_{core} complex generated in Fig. 2B. Zoomed in panel shows interaction between Mis18α and Mis18β Yippee domains using the second interface. Important residues for this interaction highlighted in pink and purple. (D) SEC profile of Mis18a_{WT}/Mis18β_{WT} (red) and Mis18α_{C154R/D160R}/Mis18β_{WT} (black) and corresponding SDS-PAGE analysis of the fractions. Samples were analysed using Superdex 200 increase 10/300 in 20 mM Tris-HCl pH 8.0, 250 mM NaCl and 2 mM DTT.



Figure EV4. Structural and biochemical characterisation of Mis18α C-terminal helix.

(A) Cartoon representation of the crystal structure of Mis18 α_{C-term} /Mis18 β_{C-term} (PDB ID: 75FY). Mis18 α is shown in purple and Mis18 β in light pink. Potential residues involved in the interaction are highlighted. Mis18 α (purple) and Mis18 β (light pink). Right panel shows SDS-PAGE analysis of cobalt and amylose pull-down of His-MBP-Mis18 $\beta_{188-229}$ wT with His-SUMO-Mis18 $\alpha_{191-233}$ mutants. SDS-PAGE shows protein bound to nickel resin as input (I) and protein-bound to amylose resin to assess interaction (P). Control with WT proteins shown in Fig. 4A. (B) Western blot analysis of co-immunoprecipitation (Co-IP) experiments using Mis18 α antibody to test interaction of mCherry as a control, Mis18 α_{-m} Cherry with and without mutations in the C-terminal α -helices and Mis18 β_{-GFP} . Top panel shows blot against mCherry, middle panel shows blot against GFP, and bottom panel shows blot against tubulin as loading control. (C) SEC-MALS of His-SUMO-Mis18 $\alpha_{188-233}$ L121 $\alpha_{1212A/L215A}$ Normalised absorption at 280 nm (mAU, left y-axis) and molecular mass (kDa, right y-axis) are plotted against elution volume (ml, x-axis). Measured molecular weight (MW) and the calculated subunit stoichiometry based on the predicted MW. Samples were analysed using a Superdex 75 increase in 50 mM HEPES pH 8.0, 150 mM NaCl and 1 mM TCEP. (D) Representative immunoblots showing expression levels of transiently expressed tagged proteins after transfection.