

## **Expanded View Figures**

## Figure EV1. Modest differences in talazoparib sensitivity observed in vitro compared to in-vivo settings (Fig. 1) in Brca1-def and Bard1-def breast tumor cells.

(A, B) In vitro cell viability assay comparing the talazoparib sensitivity between talazoparib-sensitive ("Sen") *Brca1*-def and *Bard1*-def breast tumor cells and their talazoparib-resistant ("Res") derivative lines. Cell viability values are normalized to the DMSO-treated control (considered as 100% live cells) and presented as mean values  $\pm$  SEM. *P* values were determined by a two-tailed, unpaired, Welch's test. For the *Brca1*-def model, *n* = 3 for each concentration tested, \*\* at 0.3 nM indicates *P* = 0.0028, \* at 1 nM indicates *P* = 0.0104, \* at 3 nM indicates *P* = 0.0104, and \* at 100 nM indicates *P* = 0.0357. For the *Bard1*-def model, *n* = 3 for each tested concentration, and \* at 3.0 nM indicates *P* = 0.0256. ns; not significant.





(A, B) Immunostained tumor sections from Fig. 1 were quantified using automated QuPath software to identify positively stained cells. Data were presented as mean values  $\pm$  SEM. *P* values were determined by a two-tailed, unpaired, Welch's test. n = 4 mice for talazoparib-sensitive ('Sen'') and -resistant ("Res") groups from both models. *P* values are listed in parentheses following the protein analyzed. For the *Brca1*-def model in (A): CD8 $\alpha$  (0.0375), CD4 (0.0329), CD11C (0.0061), F4/80 (0.0027), and FOXP3 (0.0007). For the *Bard1*-def model in (B), CD8 $\alpha$  (0.0293), CD4 (0.0186), CD11C (0.0048), F4/80 (0.0016), FOXP3 (ns), and S100A9 (0.0182). ns; not significant. (C) Quantitation of immunostained Res tumor sections using antibodies against CD31 (see treatments in Fig. 21-K) using automated QuPath software to identify positively stained cells. n = 5 mice from both groups from both models. Data were presented as mean values  $\pm$  SEM. *P* values were determined by a two-tailed, unpaired, Mann-Whitney test. \*\* indicates P = 0.0079 for the *Brca1*-def model, and \* indicates 0.0159 for the *Bard1*-def model, n = 4 tumors for the lsotype + Veh group, n = 4 tumors for the lsotype + Tal group. n = 6 tumors for Isotype + Tal group, n = 6 tumors for Isotype + Tal group, n = 6 tumors for Isotype + Tal group, n = 6 tumors for Isotype + Tal group. n = 6 tumors for Isotype + Tal group. A n = 5 test. P values were determined by a two-tailed, unpaired, unpaired, Student's t-test, comparing endpoint tumor weights between the Isotype + Tal and anti-VEGFR2 + Tal groups. \* indicates P = 0.0222 for the *Bard1*-def model.



## Figure EV3. FLT1 promotes PARPi-resistance in the Brca1-def and Bard1-def breast cancer models.

(A) Representative images of IHC for total FLT1 expression in tumor sections from the mice described in Fig. 1. Scale bars, 20 µm. (B) Immunostained sections from (A) were quantified using automated QuPath software to identify positively stained cells. n = 5 talazoparib-sensitive ("Sen") tumors, and n = 4 talazoparib-resistant ("Res") tumors from both Brca1-def and Bard1-def models. Data were presented as mean values ± SEM. P values were determined by a two-tailed, unpaired, Mann-Whitney test. \* indicates P = 0.0159 for both models. (C), Representative images of IHC for FLT4 in tumor sections from Fig. 1. Scale bars, 20 µm. (D) Immunostained sections from (C) were quantified using automated QuPath software to identify positively stained cells. n = 4 Sen and Res tumors for both Brca1-def and Bard1-def models. Data were presented as mean values ± SEM. P values were determined by a two-tailed, unpaired, Welch's test. ns; not significant. (E) qRT-PCR results of Flt1 repression of the indicated groups in the Brca1-def model for both gRNAs. n = 6 (consisting of two independent experiments for each triplicate testing) for both Lenti-Con and Flt1i for gRNA1 and n = 3 (one triplicate testing) for both Lenti-Con and Flti for gRNA2. Data were presented as mean values ± SEM. P values were determined by a two-tailed, unpaired, Welch's test: \*\*\*\* indicates P < 0.0001 for gRNA1 and \* indicates 0.0285 for gRNA2. (F) qRT-PCR results of Flt1 repression of the indicated groups in the Bard1-def model. n = 6 (two independent triplicate testing) for both Lenti-Con and *Flt1* if or gRNA1 and n = 3 (one triplicate testing) for both Lenti-Con and *Flt1* if or gRNA2. Data were presented as mean values ± SEM. P values were determined by a two-tailed, unpaired, Welch's test: \*\*\*\* indicates P < 0.0001 for gRNA1 and \* indicates P = 0.0273 for gRNA2. (G) gRT-PCR results of Flt1 expression of the indicated groups in the Brca1-def model. n = 3 (one triplicate testing) for Flt1i and Flt1i + Flt1 overexpression ("o/e") groups. Data were presented as mean values ± SEM. P values were determined by a two-tailed, unpaired, Welch's test: \*\* indicates P = 0.0016. (H) qRT-PCR results of Flt1 expression of the indicated groups in the Bard1-def model. n = 3 (one triplicate testing) for Flt1i and Flt1i + Flt1 o/e groups. Data were presented as mean values ± SEM. P values were determined by a two-tailed, unpaired, Welch's test: \*\* indicates P = 0.0067. (I) Tumor weights from Fig. 3D were plotted at endpoint. For the Brca1-def model, n = 6 tumors for Lenti-Con + Veh, n = 8 tumors for Lenti-Con + Tal, n = 5 tumors for Flti (gRNA1) + Veh or Tal and Flti (gRNA2) + Veh, and n = 7 tumors for Flti (gRNA2) + Tal treatment groups. For the Bard1-def model, n = 5 tumors for Lenti-Con + Veh or Tal, Flt1i (gRNA1) + Veh or Tal, and Flt1i (gRNA2) + Tal, and n = 3 tumors for Flt7i (gRNA2) + Veh. Data were presented as mean values ± SEM. P values were determined by a two-tailed, unpaired, Mann-Whitney test, comparing endpoint tumor weights between Lenti-Con + Tal and Flt1i (gRNA1 or gRNA2) + Tal groups. For the Brca1-def model, \*\* indicates P = 0.0016 between Lenti-Con + Tal and Flt1i (gRNA1) + Tal and \*\*\* indicates P = 0.0003 between Lenti-Con + Tal and Flt1i (gRNA2) + Tal. For the Bard1-def model, \*\* indicates P = 0.0079 between Lenti-Con + Tal and Flt1i (gRNA1 or gRNA2) + Tal. (J) Schematic representation of the experiment designed to test whether Flt1 re-expression rescues talazoparib-resistance in Brca1-def and Bard1-def mammary tumors with Flt1 repression. The generation of Brca1- and Bard1-def cancer cells were described in Fig. 3. To stably re-express Flt1, we transduced Flt1-repressed cells with lentiviral particles carrying Flt1 cDNA. For the Brca1-def model, randomized mice received either vehicle ("Veh") or talazoparib ("Tal") treatment starting at 2 weeks after tumor-cell injection and were euthanized at 4 weeks following injection. For the Bard1-def model, randomized mice received treatment at 1 week after tumor-cell injection and were euthanized at 3 weeks following injection. (K) Tumor growth curves comparing Flt1i + Tal and Flt1i-Flt1 o/e + Veh or Tal. For the Brca1def model, n = 7 mice for *Flt1i* + Tal, n = 4 mice for *Flt1i* - *Flt1i* + 0/e + Veh, and n = 3 mice for *Flt1i* - *Flt1i* + 0/e + Tal. For the *Bard1*-def model, n = 5 mice for *Flt1i* + Tal, n = 4mice for Flt1i-Flt1 + o/e + Veh, and n = 6 mice for Flt1i-Flt1 + o/e + Tal. Data were presented as mean values ± SEM. P values were determined with a one-way ANOVA test, comparing endpoint tumor volumes between the *Flt1i* + Tal and *Flt1i*-*Flt1* + o/e + Tal groups. For the *Brca1*-def model, \*\*\* at 4 weeks indicates *P* = 0.0001 and for the Bard1-def model, \*\* at 3 weeks indicates P = 0.0074. (L) Tumor weights from K were plotted at endpoint. For the Brca1-def model, n = 7 tumors for Flt1i + Tal, n = 4 tumors for Flt1i-Flt1 + o/e + Veh, and n = 3 tumors for Flt1i-Flt1 + o/e + Tal. For the Bard1-def model, n = 5 tumors for Flt1i + Tal, n = 4 tumors for Flt1i-Flt1 + o/e + Veh, and n = 6tumors for Flt1i-Flt1 + o/e + Tal. Data were presented as mean values ± SEM. P values were determined by a two-tailed, unpaired, Mann-Whitney test, comparing endpoint tumor weights between the Flt1i + Tal and Flt1i-Flt1 + o/e + Tal groups. For the Brca1-def model, \* indicates P = 0.0167 and for the Bard1-def model, \*\* indicates P = 0.0043.



Figure EV4. Tumor and body weight analysis of mice treated with vehicle ("Veh"), talazoparib ("Tal"), and/or axitinib ("Axi").

(A) Tumor weights from Fig. 4B were plotted at endpoint. n = 6 Veh-treated tumors, n = 7 Tal-treated tumors, n = 5 Axi-treated tumors, and n = 5 tumors treated with Tal + Axi. Data were presented as mean values ± SEM. *P* values were determined by a two-tailed, unpaired, Mann-Whitney test, comparing endpoint tumor weights between the Tal and Tal + Axi groups. \*\* indicates P = 0.0051. (B) Tumor weights from Fig. 4E were plotted after collection at endpoint. n = 5 Veh-treated tumors, n = 6 Tal-treated tumors, n = 5 Axi-treated tumors, n = 7 tumors treated with Tal + Axi. Data were presented as mean values ± SEM. *P* values were determined by a two-tailed, unpaired Mann-Whitney test comparing endpoint tumor weights between the Tal and Tal + Axi groups. \*\*\* indicates P = 0.0006. (C, D) Body weight from treatment initiation until endpoint from Fig. 4A, D for each experiment. For the *Brca1*-def model, n = 6 Veh-treated tumors, n = 5 Axi-treated tumors, n = 5 Axi-treated tumors, n = 5 Axi-treated tumors, n = 5 tumors treated with Tal + Axi. For the *Bard1*-def model, n = 5 Veh-treated tumors, n = 5 Axi-treated tumors, and n = 7 tumors treated with Tal + Axi.



## Figure EV5. Quantitation of phosphorylated STAT3 (pSTAT3) levels in Brca1-def and Bard1-def breast tumor cells and tumor tissue sections.

(A, B) Immunoblot analysis was performed using antibodies against pSTAT3, STAT3 and  $\beta$ -actin (loading control) using lysates from *Flt1*-expressing (Con) and -deficient (*Flt1*i), talazoparib-resistant ("Res") *Brca1* and *Bard1*-def tumor cells, treated with 50 ng/mL of mouse PGF protein that were used in Fig. 5A, B. (C) Representative images of IHC for pSTAT3 staining in tumor sections from the mice described in Fig. 1 comparing talazoparib-sensitive ("Sen") tumors to talazoparib-resistant ("Res") tumors. Scale bars, 20 µm. (D) Immunostained sections from (C) were quantified using automated QuPath software to identify positively stained cells. For the *Barc1*-def model, n = 4 tumors for both Sen and Res. For the *Bard1*-def model, n = 5 Sen tumors, and n = 4 Res tumors. Data were presented as mean values ± SEM. *P* values were determined by a two-tailed, unpaired, Mann-Whitney test. ns; not significant.