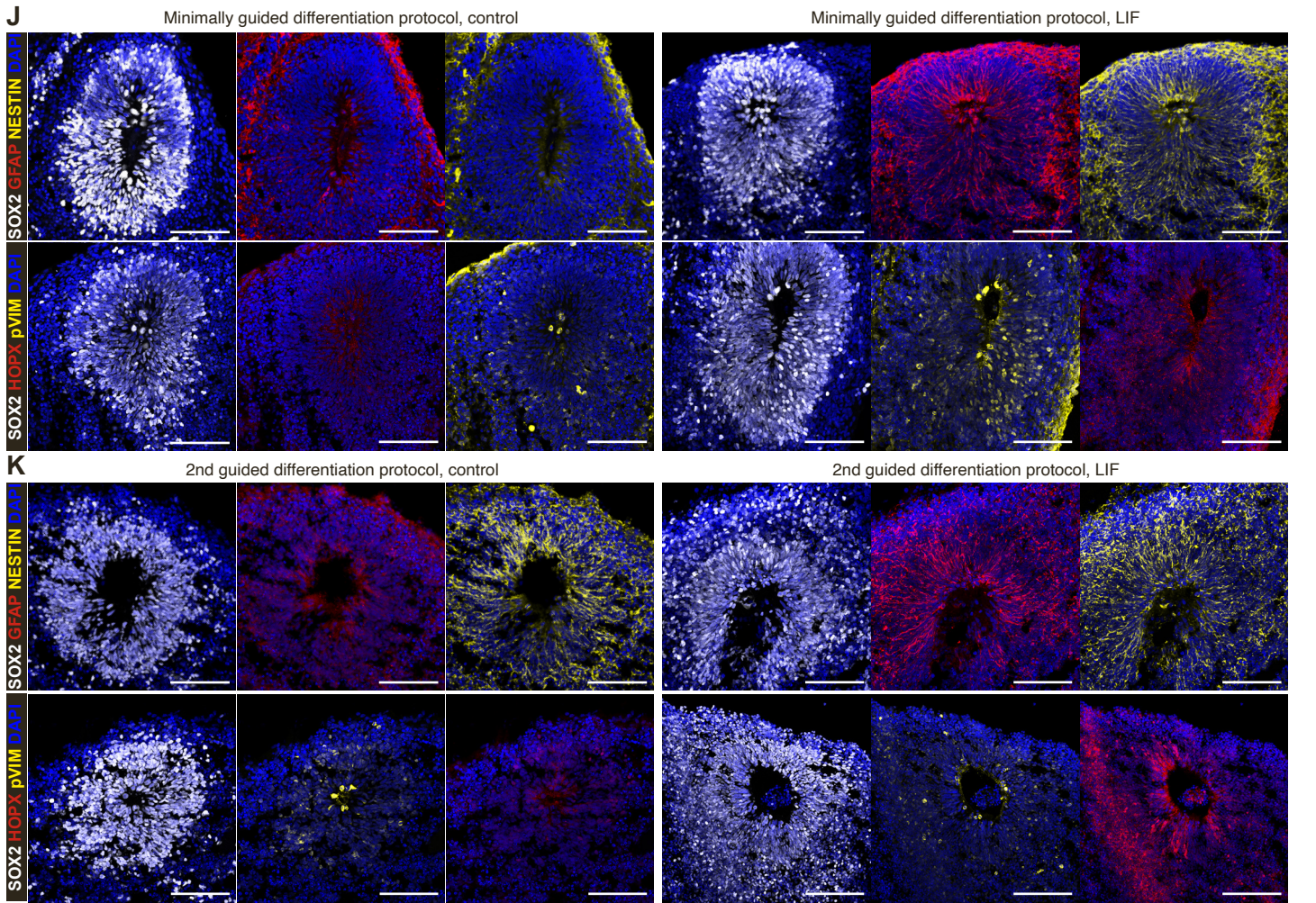
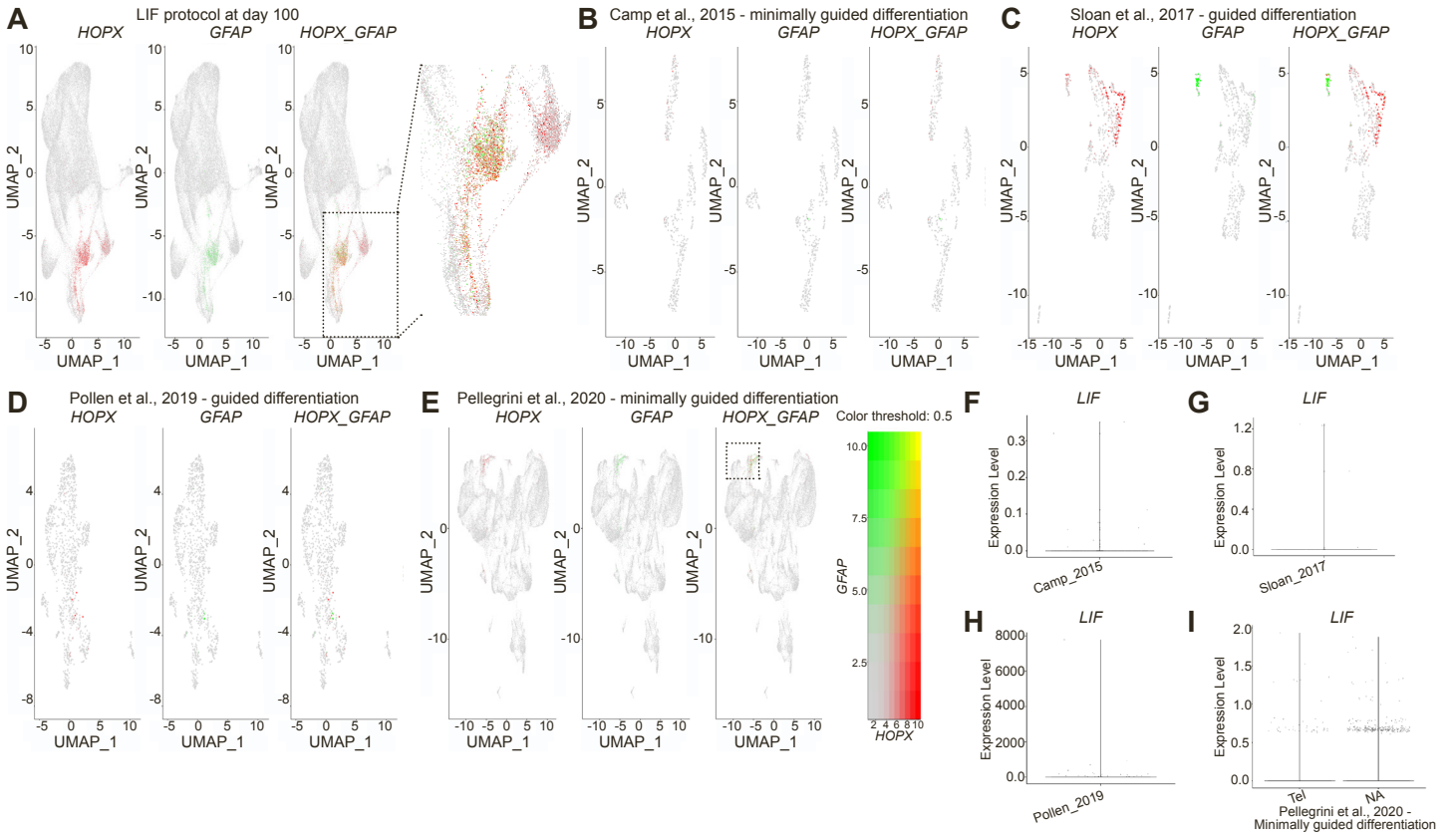


## Supplemental information

### Generation of human cerebral organoids with a structured outer subventricular zone

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**Supplemental Figure 1 related to Figure 1 and Figure 2**



**Figure S1. LIF in minimally guided and guided cortical differentiation protocols**

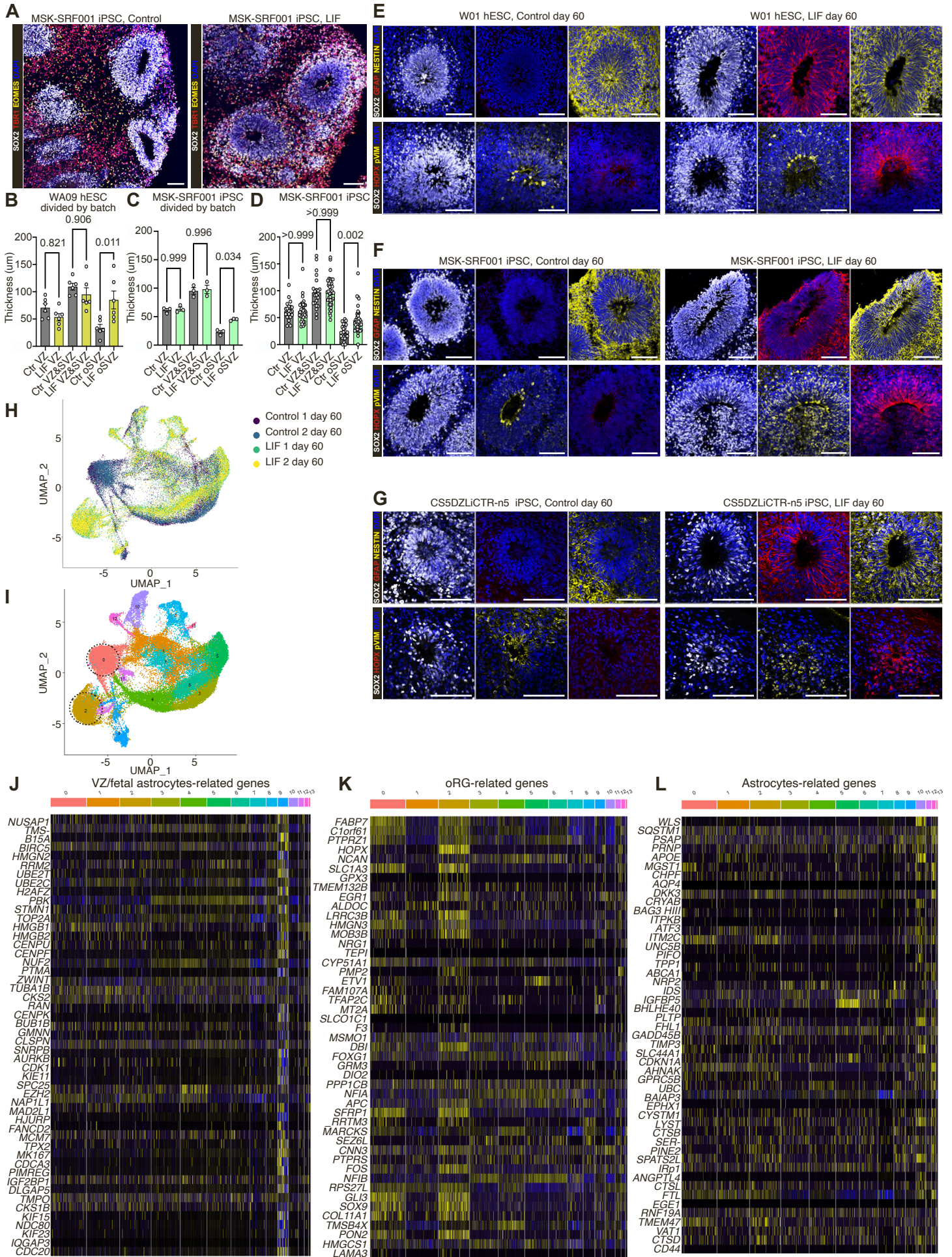
**(A-E)** Feature plots from the scRNA-seq datasets depicting *HOXP*, *GFAP*, and co-expression of *HOXP* and *GFAP* in our study at day 100 of differentiation (**A**); their expression in other two guided differentiation protocols (**C** and **D**) and two minimally guided differentiation protocols (**B** and **E**).

**(F-I)** Expression levels of *LIF* in various cerebral organoid models. In a minimally guided differentiation protocol, telencephalon (Tel) cells do not express *LIF*, while not-assigned (NA) cell types do (**I**).

**(J)** SOX2 (gray), GFAP (red), NESTIN (yellow), phospho-VIMENTIN (pVIM) (yellow) and HOPX (red) staining in day 60 brain organoids derived from a minimally guided differentiation protocol +/- LIF. Cell nuclei stained with DAPI. Scale bars 100  $\mu$ m.

**(K)** SOX2 (gray), GFAP (red), NESTIN (yellow), phospho-VIMENTIN (pVIM) (yellow) and HOPX (red) staining in day 60 cortical organoids derived from a second guided differentiation protocol +/- LIF. Cell nuclei stained with DAPI. Scale bars 100  $\mu$ m.

Supplemental Figure 2 related to Figure 1 and Figure 2



**Figure S2. LIF treatment in various hPSC lines and transcriptional profiling**

**(A)** SOX2 (gray), TBR1 (red) and EOMES/TBR2 (yellow) staining in day 60 MSK-SRF001 iPSC control and LIF-treated cortical organoids. Cell nuclei stained with DAPI. Scale bars 100  $\mu$ m.

**(B)** WA09 hESC rosette area quantifications from Figure 1C, showing the inter-batch variability. One-way ANOVA with Tukey's test.

**(C)** MSK-SRF001 iPSC rosette area quantifications. One-way ANOVA with Tukey's test.

**(D)** MSK-SRF001 iPSC rosette area quantifications from **(C)** showing the inter-batch variability. One-way ANOVA with Tukey's test.

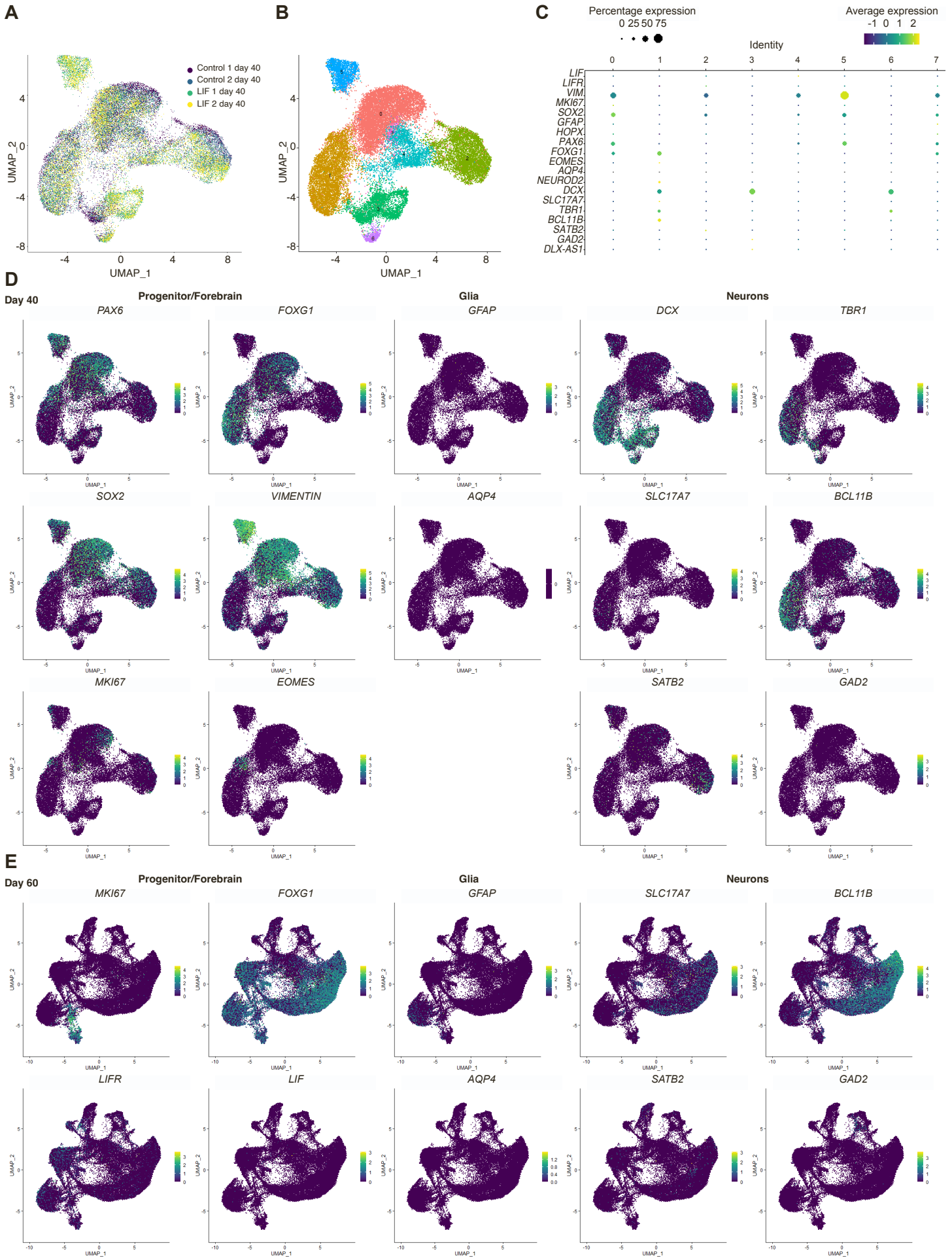
**(E-G)** SOX2 (gray), GFAP (red), NESTIN (yellow), HOPX (red), phospho-VIMENTIN (pVIM) (yellow) stainings in day 60 control and LIF-treated organoids from the WA01 hESC line **(D)**, the MSK-SRF001 iPSC line **(E)** and the CS5DZLiCTR iPSC line **(F)**. Cell nuclei stained with Dapi. Scale bars 100  $\mu$ m.

**(H)** Single-cell RNA-seq (scRNA-seq) experiments at day 60 showing plots for original identity.

**(I)** Seurat clusters in control and LIF-treated organoids at day 60. The progenitor cluster (0) from control cortical organoids and the oRG cluster 2 from LIF-treated cortical organoids are highlighted.

**(J-L)** Heatmaps showing the transcriptional signatures of VZ/fetal astrocytes **(J)**, oRG **(K)** and astrocytes **(L)** derived from Sloan et al.<sup>10</sup> and mapping of all the various cellular identities present at day 60 in control and LIF-treated organoids.

Figure S3 related to Figure 1, 2 and 3



**Figure S3. Day 40 and Day 60 scRNA-seq of control and LIF-treated cortical organoids**

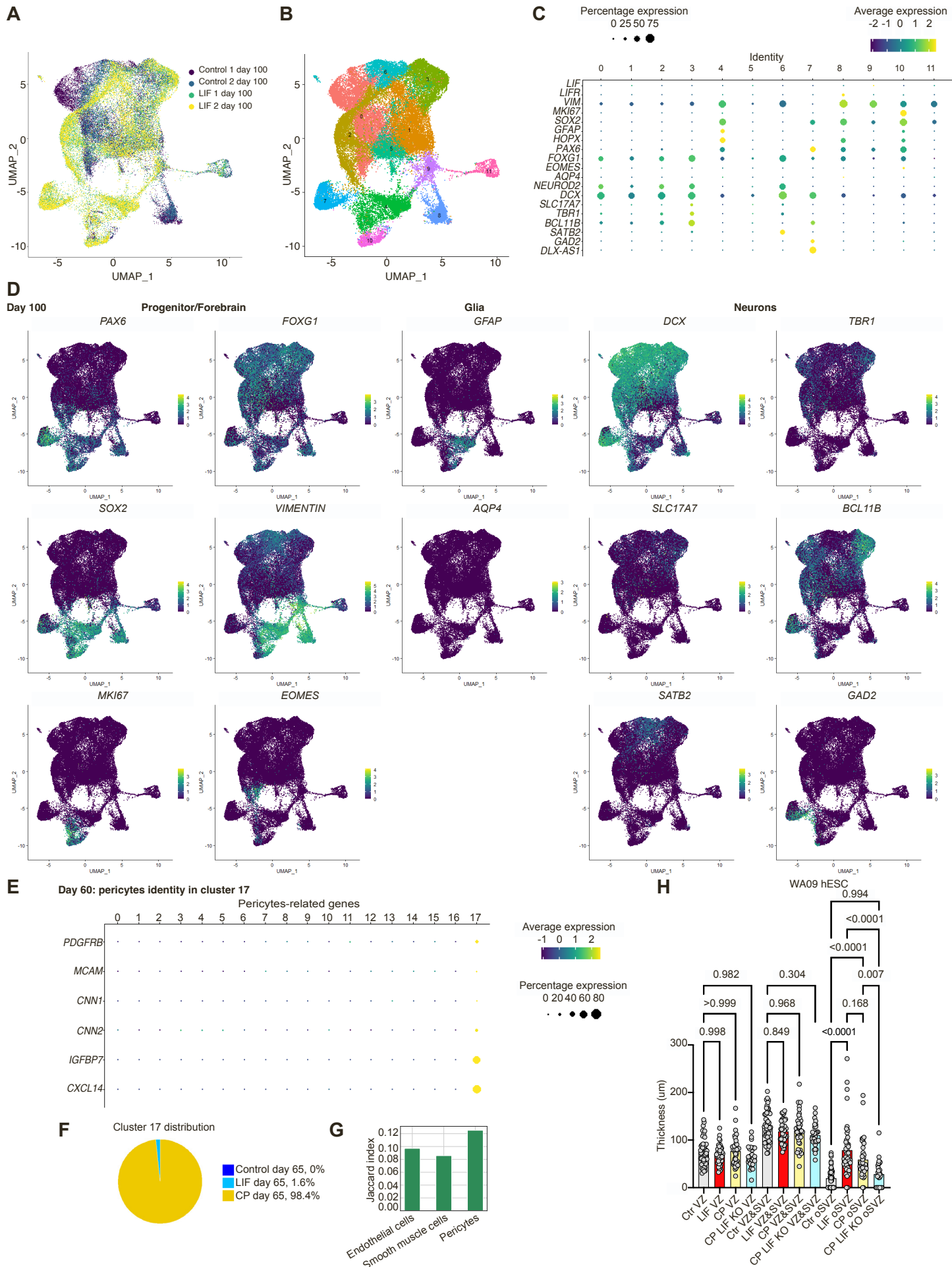
**(A and B)** scRNA-seq experiments showing plots for original identity **(A)** and Seurat clusters **(B)** in control and LIF-treated cortical organoids at day 40. Two independent batches of 10 organoids each were processed and analyzed by scRNA-seq for both control and LIF condition.

**(C)** Genes selected from B to mark selected populations of interest (dividing cells, neuronal cell types, neuronal progenitors, astrocytes, postmitotic neurons and radial glial cells) in control and LIF-treated organoid clusters at day 40.

**(D)** Feature plots depicting the distribution of the expression of key selected progenitor/forebrain, glia, and neuronal genes in control and LIF-treated organoids at day 40.

**(E)** Feature plots depicting the distribution of the expression of key selected progenitor/forebrain, glia, and neuronal genes in control and LIF-treated organoids at day 60.

**Figure S4 related to Figure 3 and 4**





**Figure S4. Day 100 scRNA-seq of control and LIF-treated cortical organoids, and identification and impact of LIF-secreting pericytes in CP assembloids**

**(A and B)** scRNA-seq experiments showing plots for original identity **(A)** and Seurat clusters **(B)** in control and LIF-treated cortical organoids at day 100. Two independent batches of 10 organoids each were processed and analyzed by scRNA-seq for both control and LIF conditions.

**(C)** Genes selected from **(B)** to mark selected populations of interest (dividing cells, neuronal cell types, neuronal progenitors, astrocytes, postmitotic neurons, and radial glial cells) in control and LIF-treated cortical organoid identities at day 100.

**(D)** Feature plots depicting the distribution of the expression of key selected progenitor/forebrain, glia, and cortical neuron genes in control and LIF-treated cortical organoids at day 100.

**(E)** Genes selected from Figure 4L to mark selected key pericyte markers in cluster 17 in CP assembloids at day 65.

**(F)** Pie chart showing cluster 17 distribution in control organoids, LIF-treated organoids and CP assembloids at day 65. Identity 17 is almost exclusively enriched in CP assembloids and represents the pericytes population.

**(G)** Jaccard index for identity 17, specific for the CP assembloids, shows a stronger overlap with fetal brain pericytes compared to endothelial cells or smooth muscle cells.

**(H)** Rosette quantifications of WA09 control and LIF-treated cortical organoids, CP assembloids and CP LIF KO assembloids at day 60, based on the separation of the regions as shown in Figure 1B. One-way ANOVA with Tukey's test. Adjusted p values are shown on the graph.