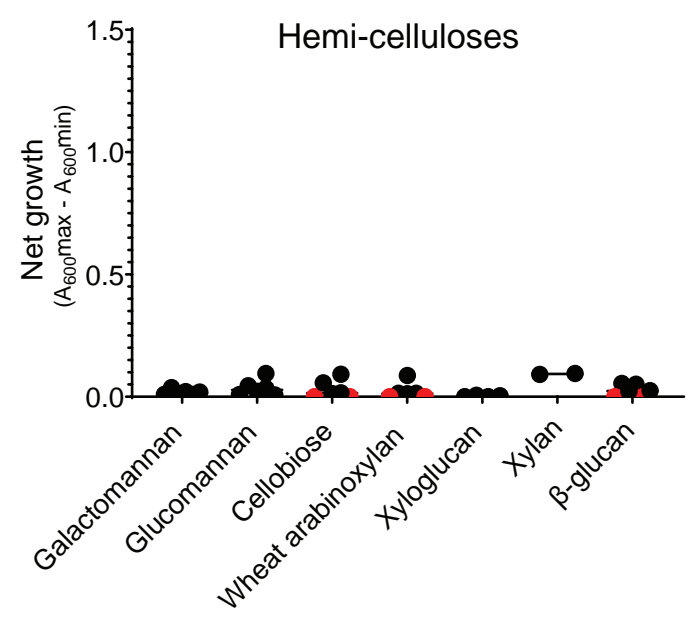
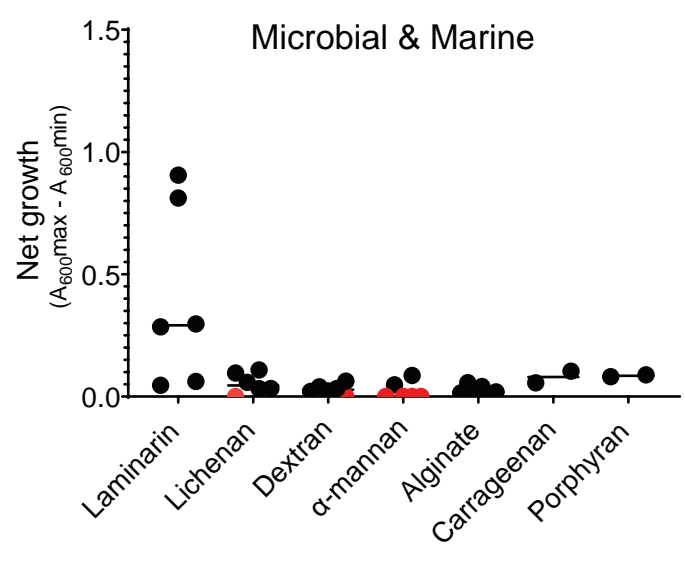
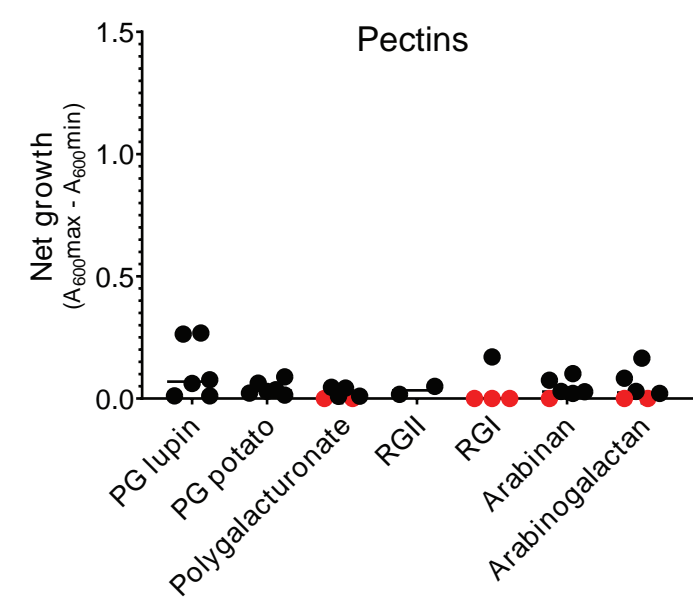
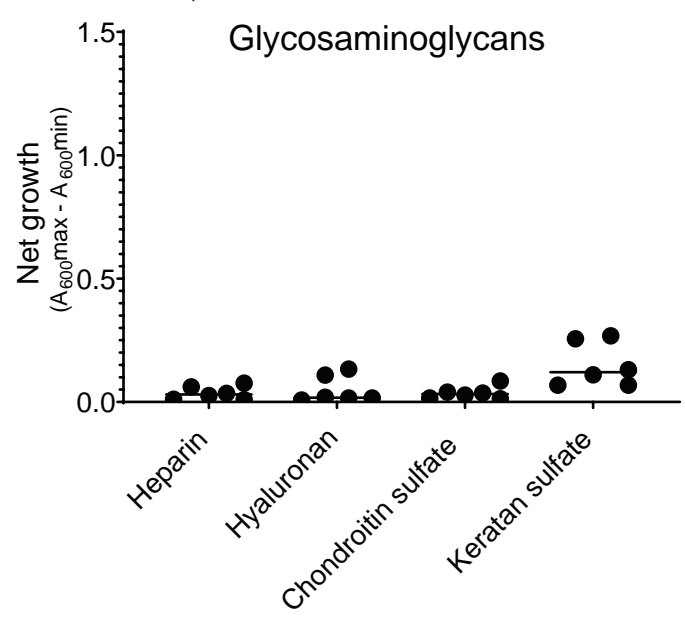
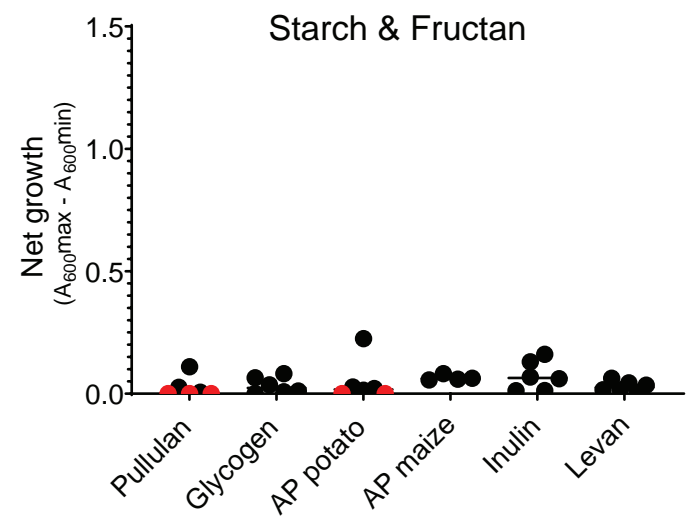
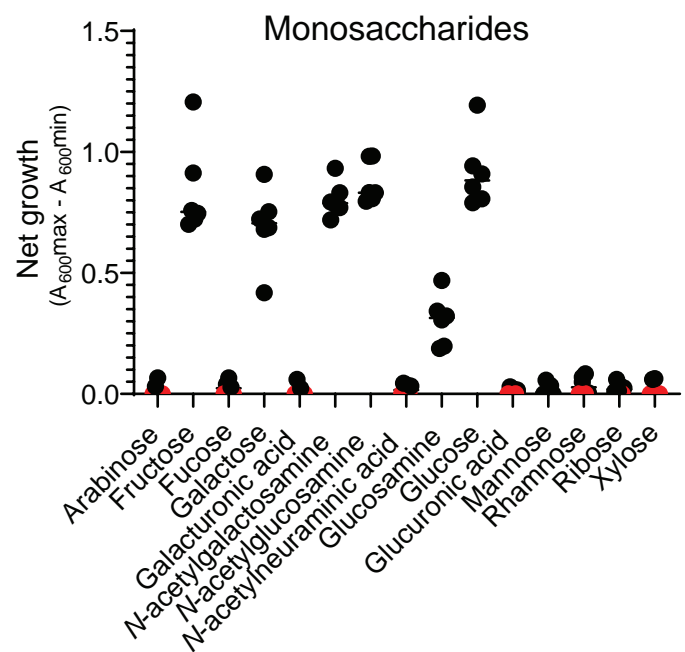
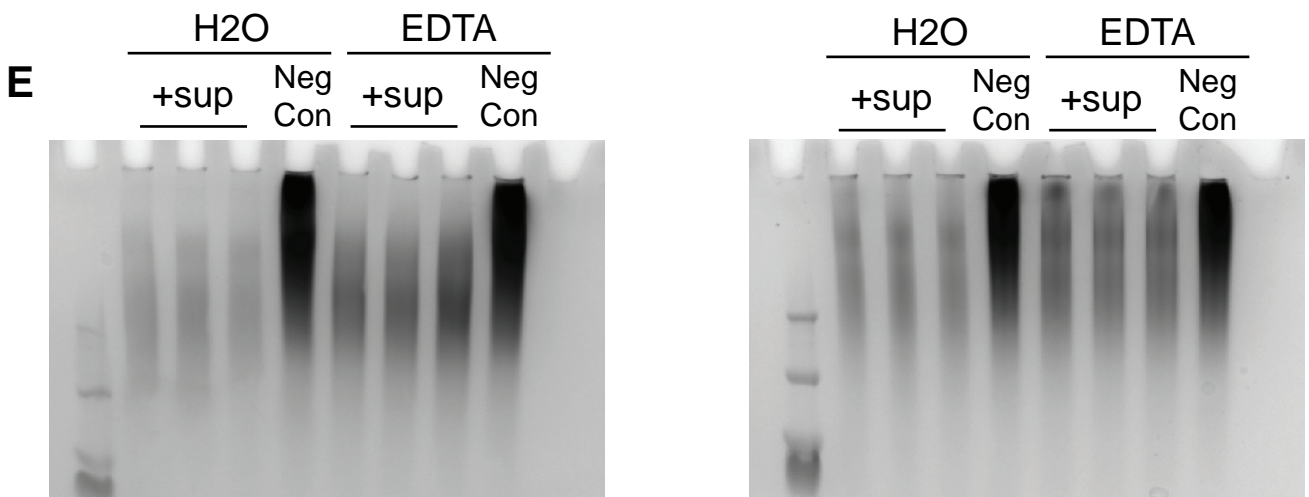
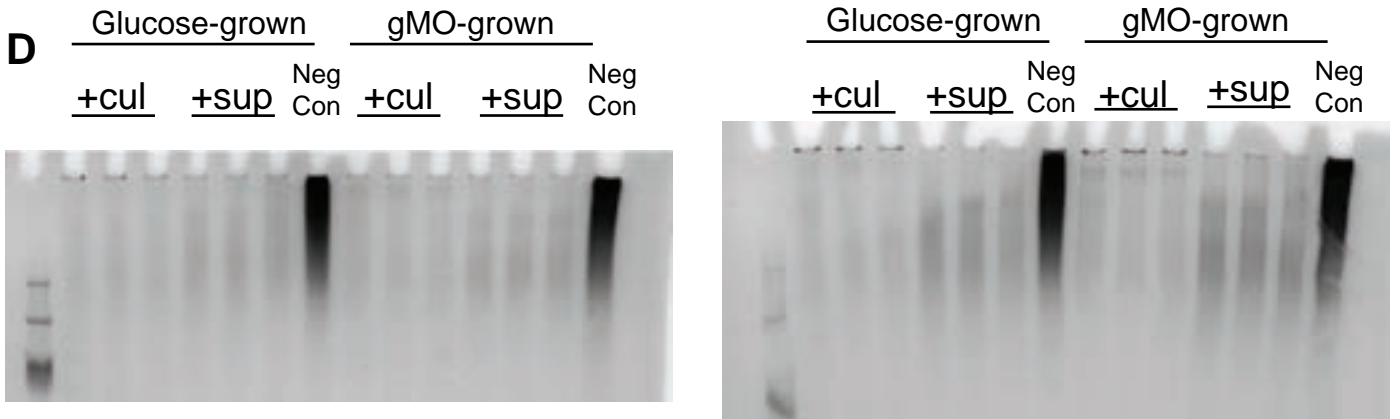
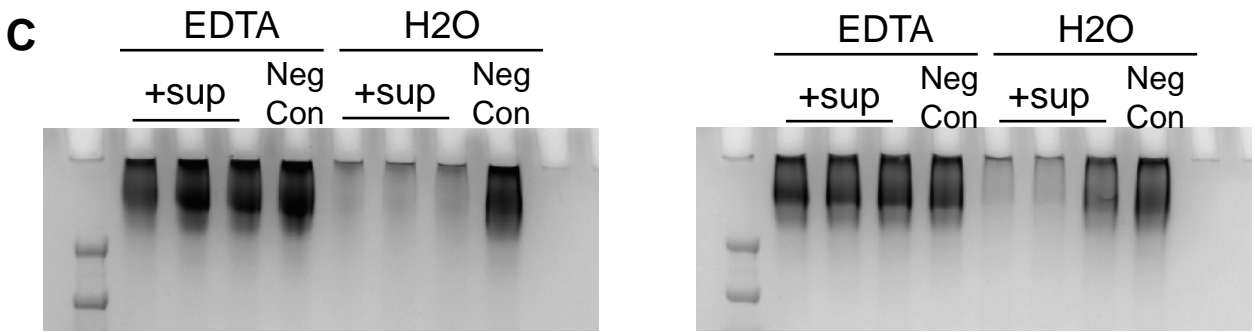
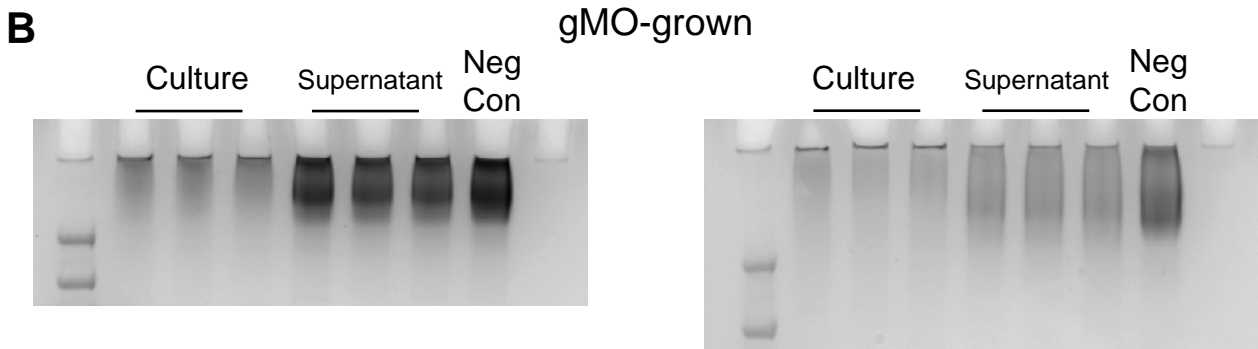
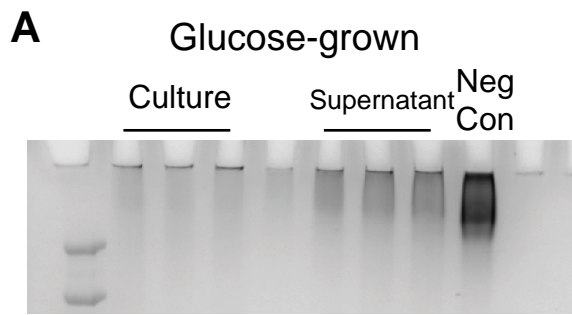


Figure S1



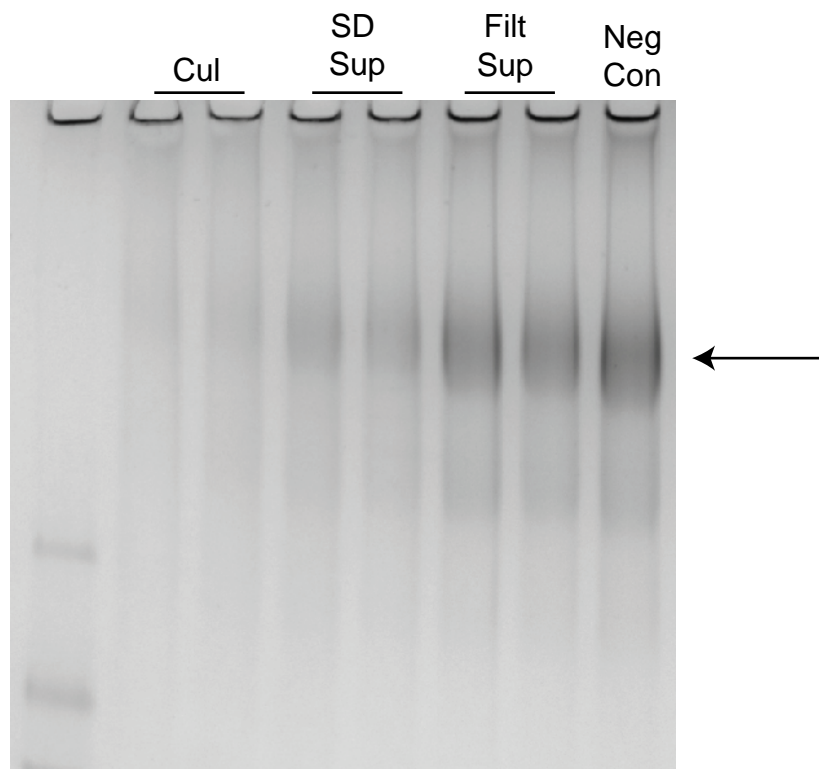
Supplemental Figure S1. *R. torques* growth on various carbohydrates. *R. torques* was grown on 46 substrates, each as the main carbohydrate source in YCFA medium. Growth was measured by absorbance at 600 nm for ~92 hours. Growth curves were analyzed to identify minimum and maximum absorbance values for each curve (excluding values representing artifacts due to brief disturbances of the plates or baseline noise), and the net growth (max absorbance – min absorbance) was calculated. For growths in which the absorbance consistently decreased throughout the growth curve with no apparent increase in absorbance, net growth was reported as zero, and these points are displayed in red. AP = amylopectin, PG = pectic galactan, RG = rhamnogalacturonan (type I or type II specified).

Figure S2



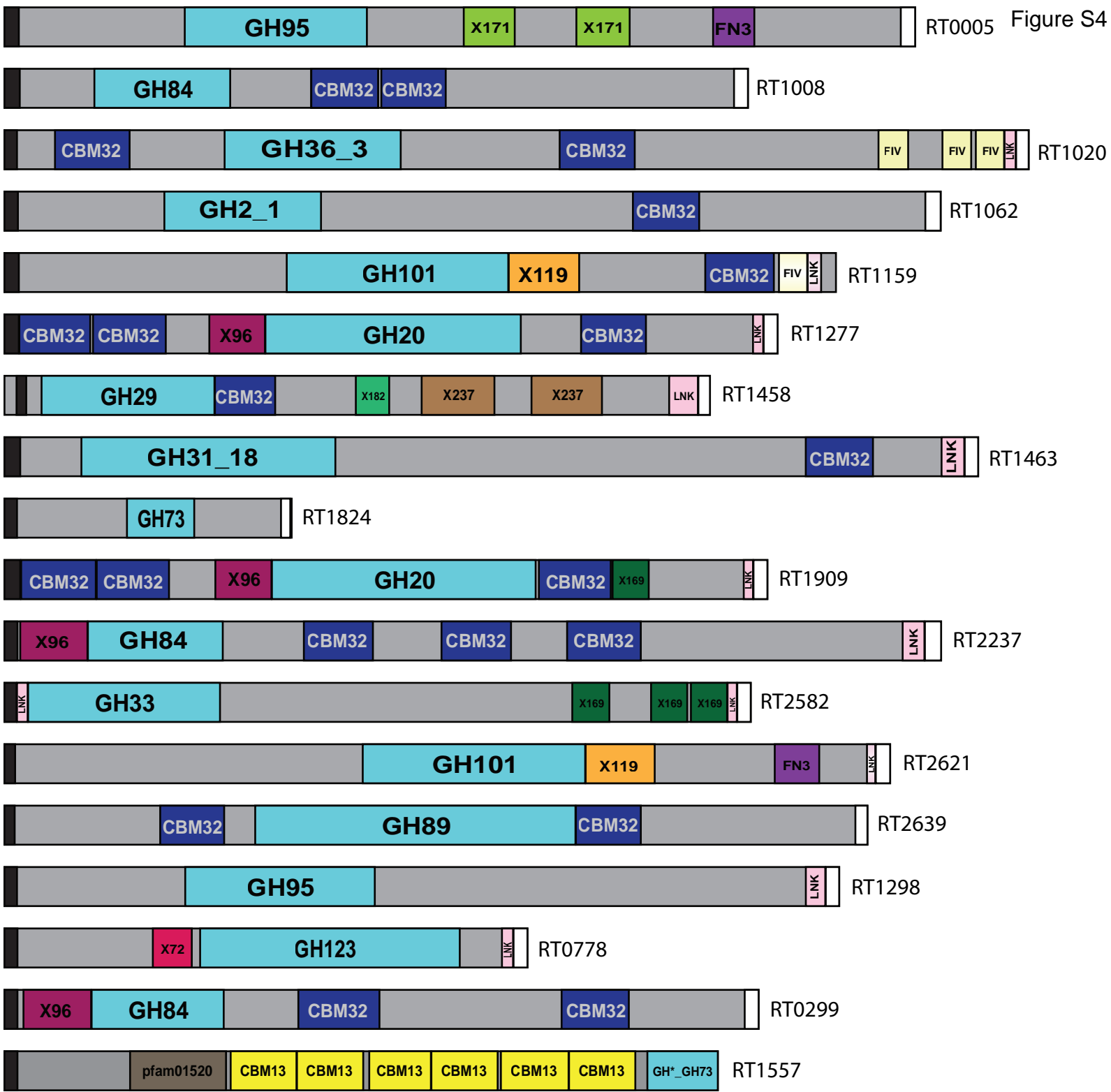
Supplemental Figure S2. Periodic acid-Schiff-stained gels to visualize degradation of mucin glycoproteins by *R. torques*. Culture (*i.e.*, cells and supernatants) or cell-free supernatant samples were collected from *R. torques* after overnight growth on glucose or gMO. Samples were incubated with cMUC2 or PGM anaerobically for 2 days and electrophoresed through 4-12% Bis-tris gels and stained with periodic acid-Schiff stain to visualize mucins. **A-C**, Degradation of cMUC2 (2.5 mg/ml final concentration) by glucose-grown samples (**A**), gMO-grown samples (**B**), or 10mM EDTA treated samples (**C**). **D-E**, degradation of PGM (10 mg/ml final concentration) by glucose or gMO-grown samples (**D**) or 10 mM EDTA treated samples (**E**).

Figure S3



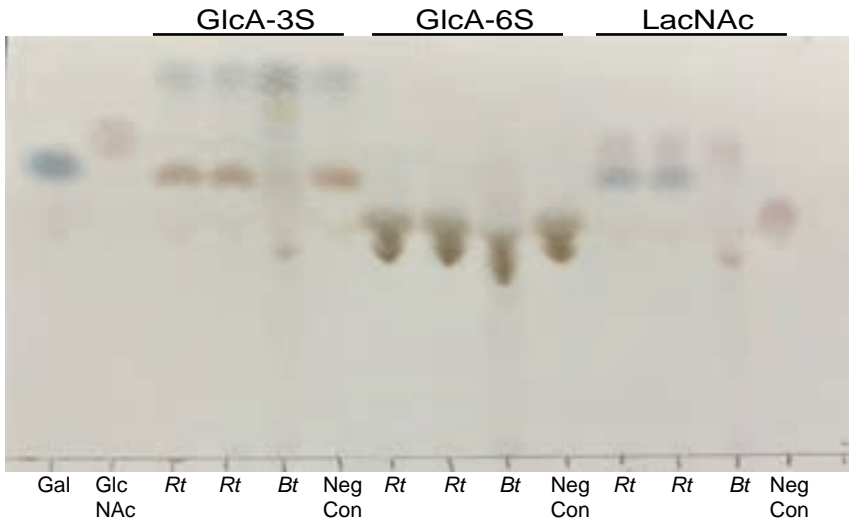
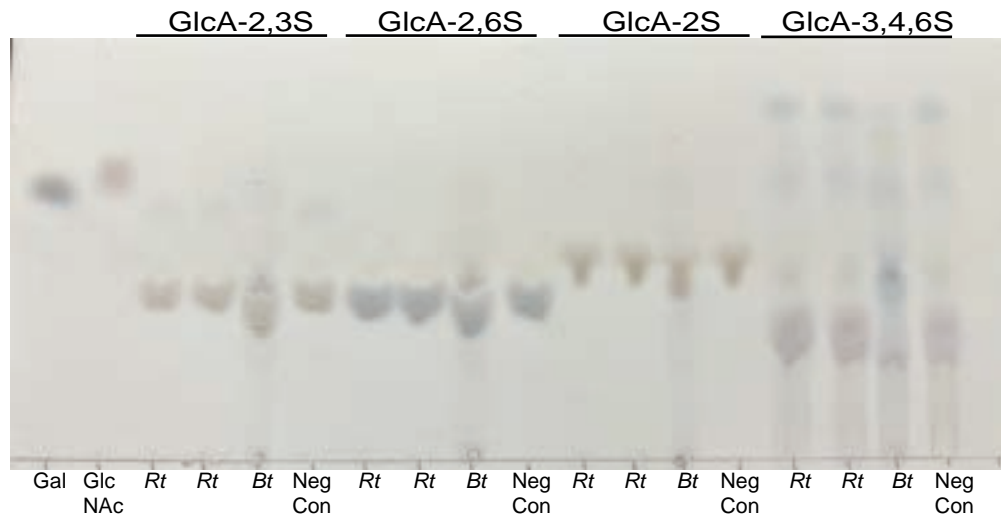
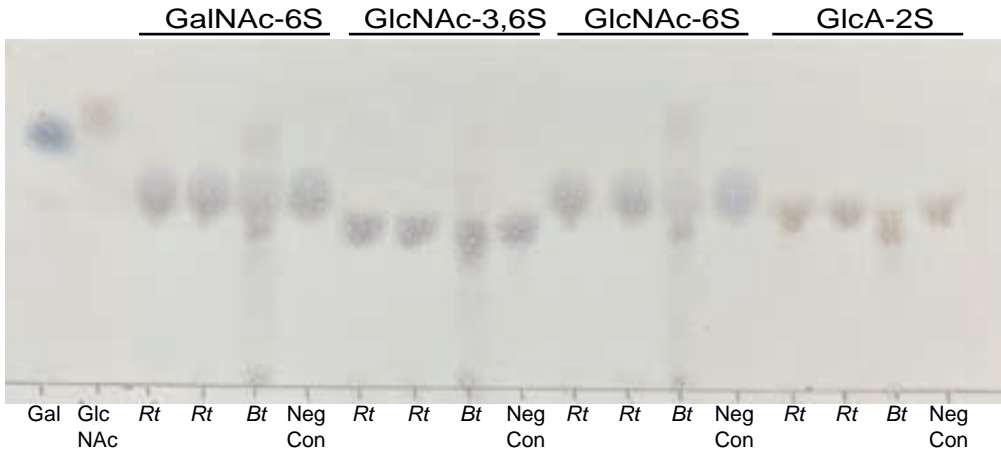
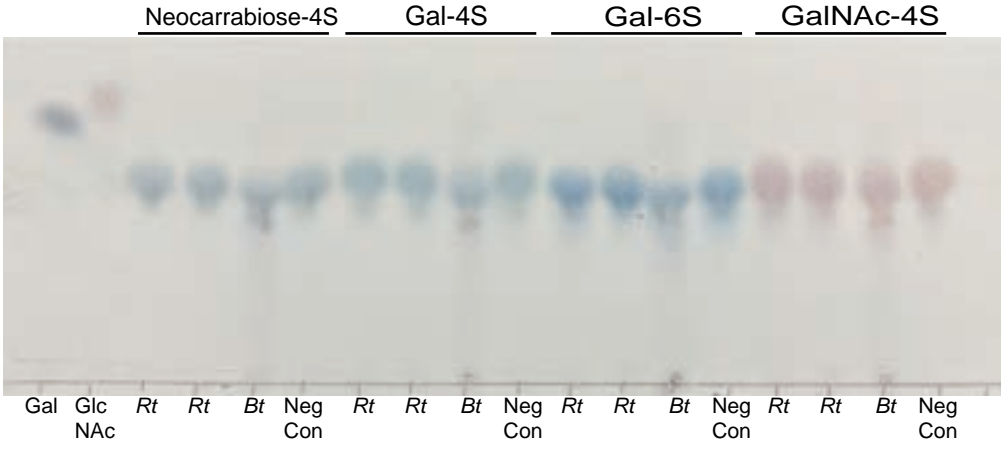
Supplemental Figure S3. Degradation of cMUC2 by *R. torques* supernatants after

centrifugation or filtering. *R. torques* was grown overnight in YCFA with glucose and samples were either centrifuged or filtered (0.22 um PVDF filters) to collect supernatants. Supernatants were then incubated with cMUC2 anaerobically for 2 days. Samples were analyzed by electrophoresis through a 4-12% Tris-Glycine gel and stained with periodic acid-Schiff stain to visualize mucin (major band indicated by arrow). Cul = whole culture (cells + supernatant), SD sup = spun-down (centrifuged) supernatant, Filt sup = filtered supernatant, Neg con = negative control (YCFA with glucose added in lieu of *R. torques* cultures or supernatant).



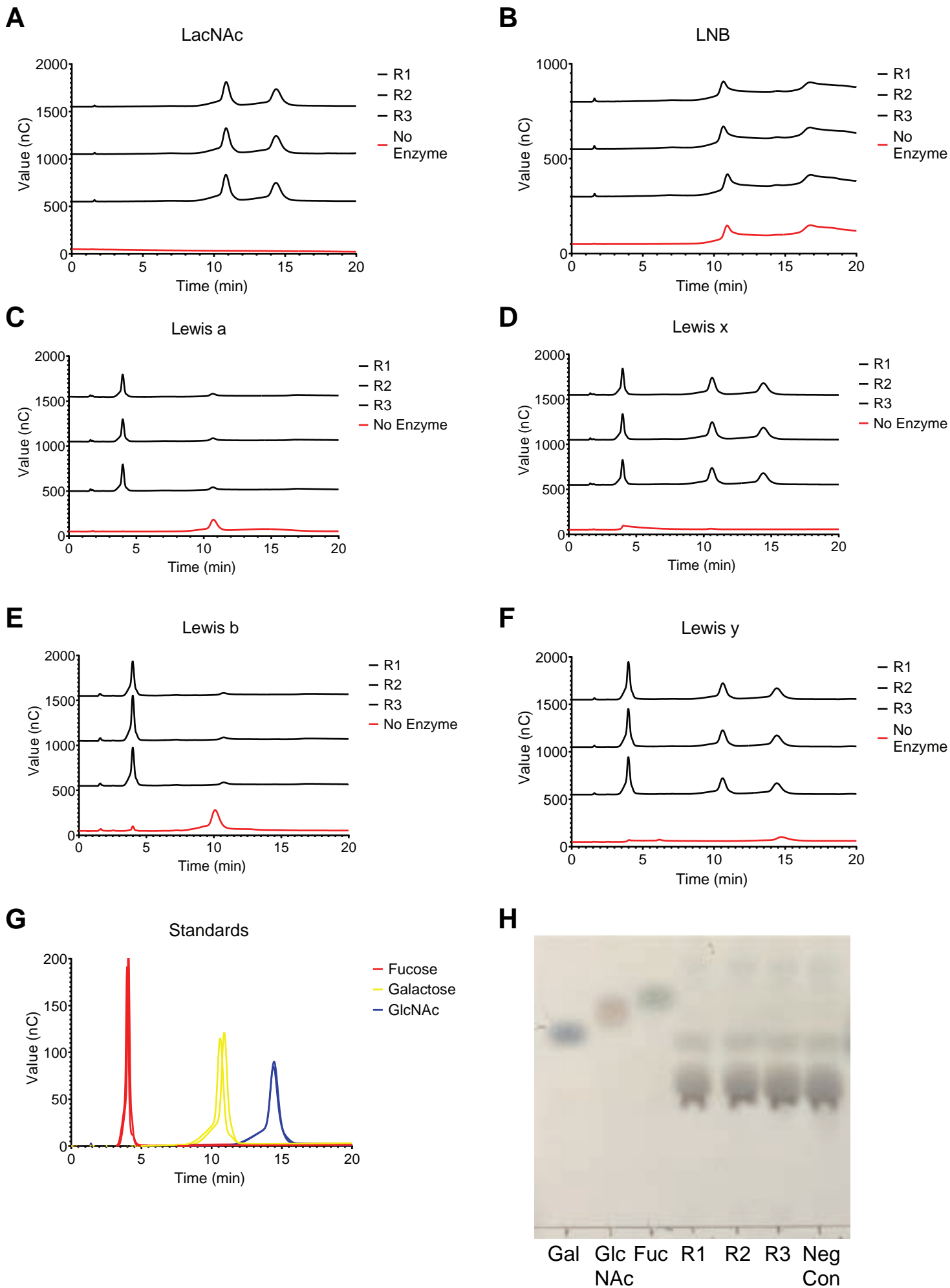
Supplemental Figure S4. Predicted domain architecture of highly expressed and transcribed CAZymes. Domain architecture predictions were generated using CAZy. Figure made with Illustrator for Biological Sequences⁷⁰. Black = signal peptide; white = transmembrane domain; teal = CAZyme domains; blue = CBM domains. GH*_GH73 denotes a distant relationship of that domain to the GH73 family.

Figure S5



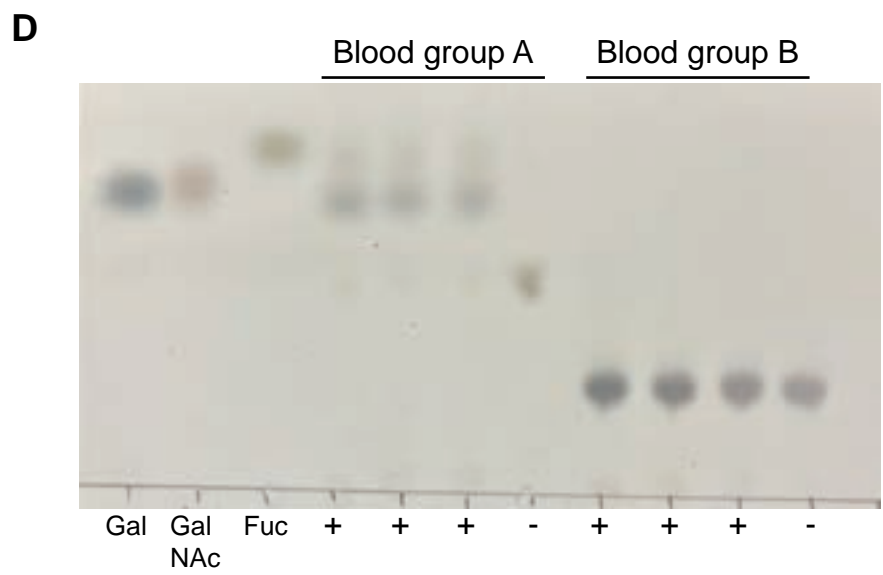
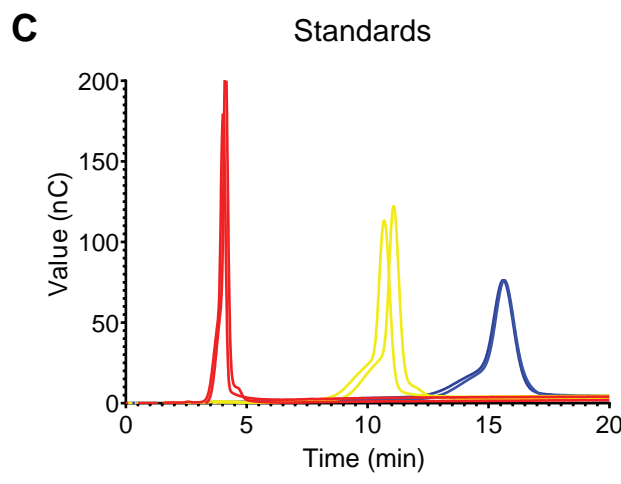
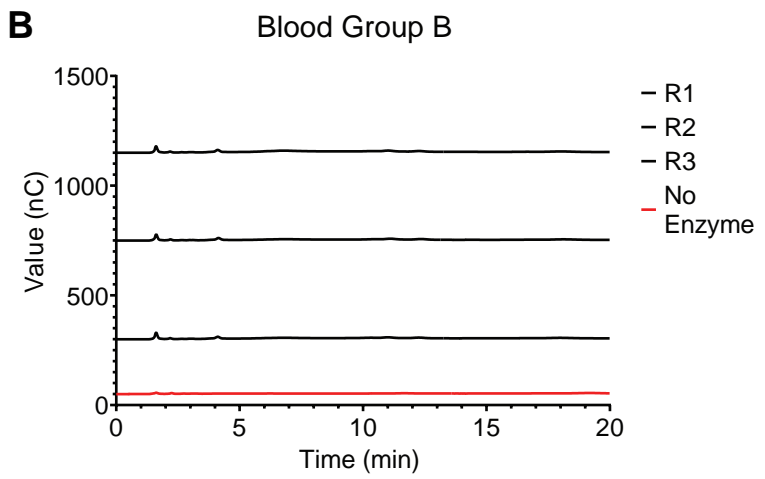
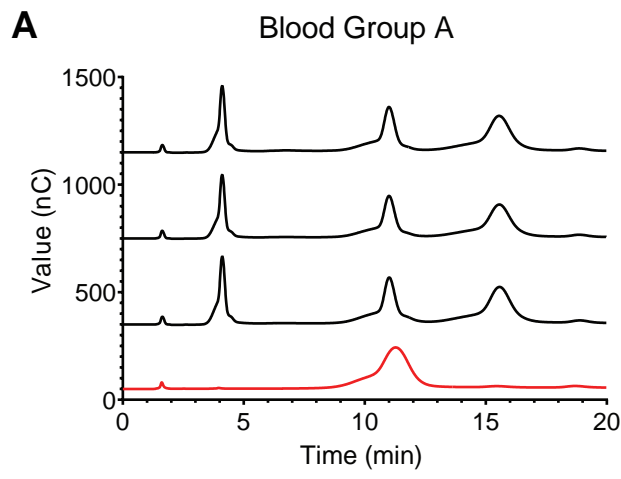
Supplemental Figure S5. *R. torques* fails to degrade sulfated sugars. Proteins were ammonium sulfate precipitated from the supernatant of *R. torques* overnight cultures after growth in YCFA with glucose and incubated with respective sugars (5mM final concentration, except for GlcA-6-O-sulfate which was 20 mM final concentration). Reactions were analyzed by thin layer chromatography using diphenylamine-aniline-phosphoric acid developer. *Rt* indicates samples where *R. torques* precipitated proteins were added, *Bt* indicates samples where sonicated *B. thetaiotaomicron* cultures exposed to keratan sulfate was added, and Neg Con indicates negative control samples that contained buffer (10 mM MES and 5 mM CaCl₂). Gal = galactose; GalNAc = *N*-acetylgalactosamine; GlcNAc = *N*-acetylglucosamine; GalA = galactosamine; GlcA = glucosamine.

Figure S6



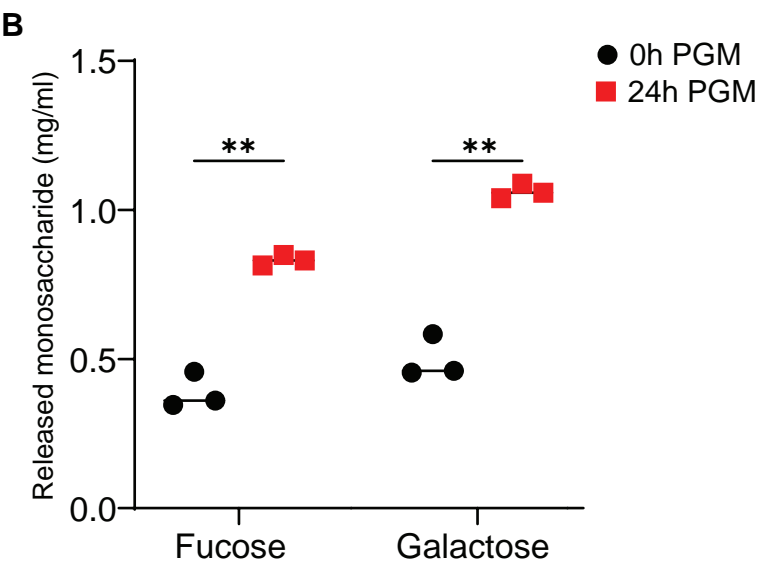
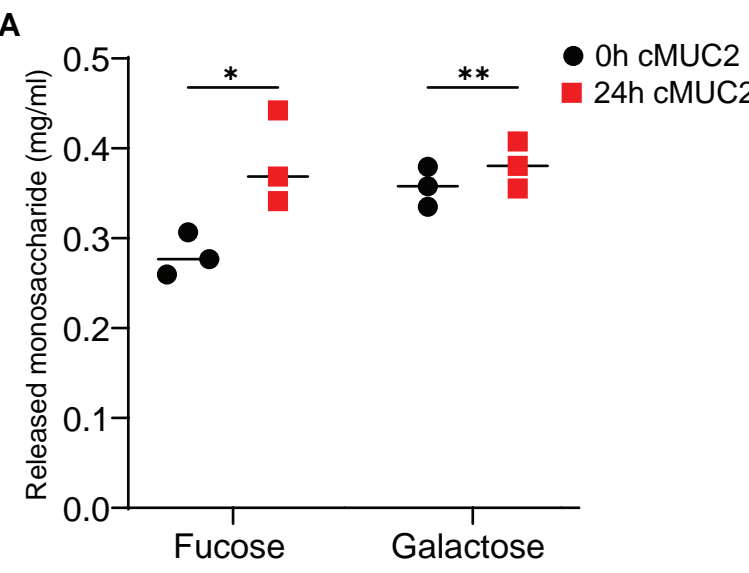
Supplemental Figure S6. HPAEC-PAD analysis of *R. torques* ammonium sulfate precipitated protein activity on model mucin sugars. A-G, Chromatograms collected by HPAEC-PAD analysis of *R. torques* supernatant precipitated protein reactions with model glycans *N*-acetyllactosamine (LacNAc, **A**), and lacto-*N*-biose (LNB, **B**), Lewis a (**C**), x (**D**), b (**E**), y (**F**) compared to single monosaccharide standards (**G**). R1-R3 = replicates reactions with supernatant precipitates from 3 *R. torques* cultures. No enzyme control is shown in red, where PBS was added instead of *R. torques* precipitates. **H**, Thin layer chromatogram of reaction products of activity assay on lacto-*N*-biose (LNB) from A. R1-R3=replicate reactions with *R. torques* supernatant precipitates; Neg Con=no enzyme control reaction with PBS.

Figure S7



Supplemental Figure S7. HPAEC-PAD analysis of *R. torques* ammonium sulfate precipitated protein activity on blood groups A and B. **A-C**, Chromatograms collected by HPAEC-PAD analysis of *R. torques* supernatant precipitated protein reactions with model glycans blood groups A (**A**) and B (**B**) compared to single monosaccharide standards (**C**). R1-R3 = replicates reactions with supernatant precipitates from 3 *R. torques* cultures. No enzyme control is shown in red, where PBS was added instead of *R. torques* precipitates. **D**, Thin layer chromatogram of blood group A and blood group B reactions from A. Gal=galactose, GlcNAc=*N*-acetylglucosamine, Fuc=fucose, R1-R3=replicate reactions with *R. torques* supernatant precipitates, Neg Con=no enzyme control reaction with PBS.

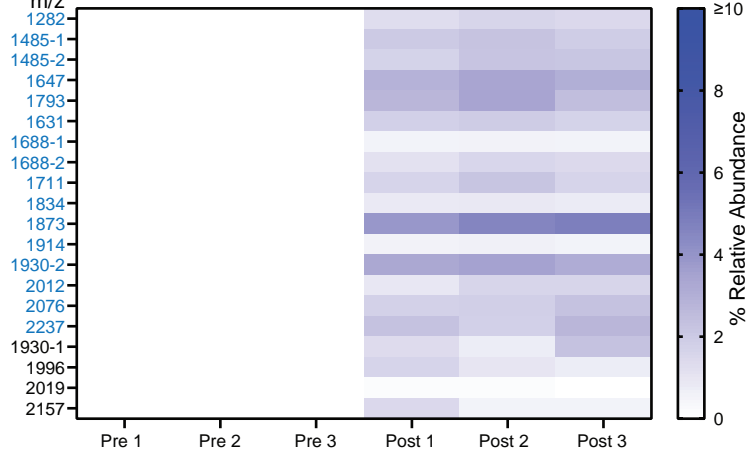
Figure S8



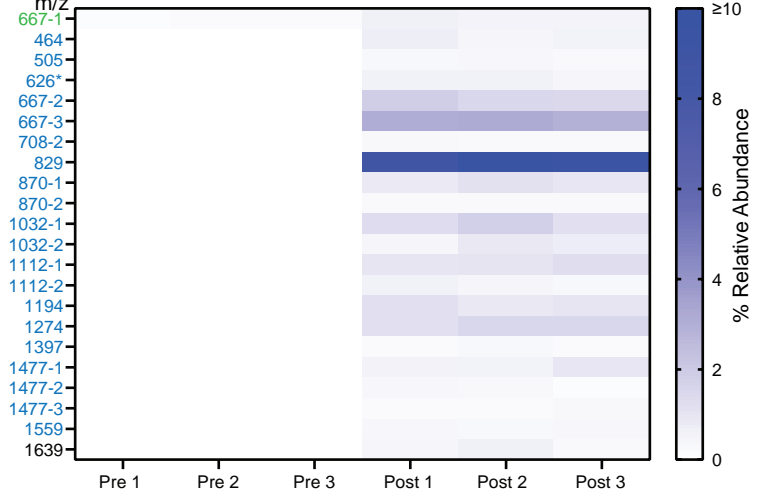
Supplemental Figure S8. Fucose and galactose released from cMUC2 and PGM after digestion by supernatants from *R. torques* grown on gMO. A-B, Concentration of fucose or galactose released after 24-hour incubation of *R. torques* supernatants from cultures grown on gMO with cMUC2 (2.5 mg/ml final; **A**) or PGM (10 mg/ml final; **B**). Statistics were analyzed with paired, two-tailed t tests comparing release of each monosaccharide between substrates in each panel. * $p < 0.05$, ** $p < 0.01$.

Figure S9

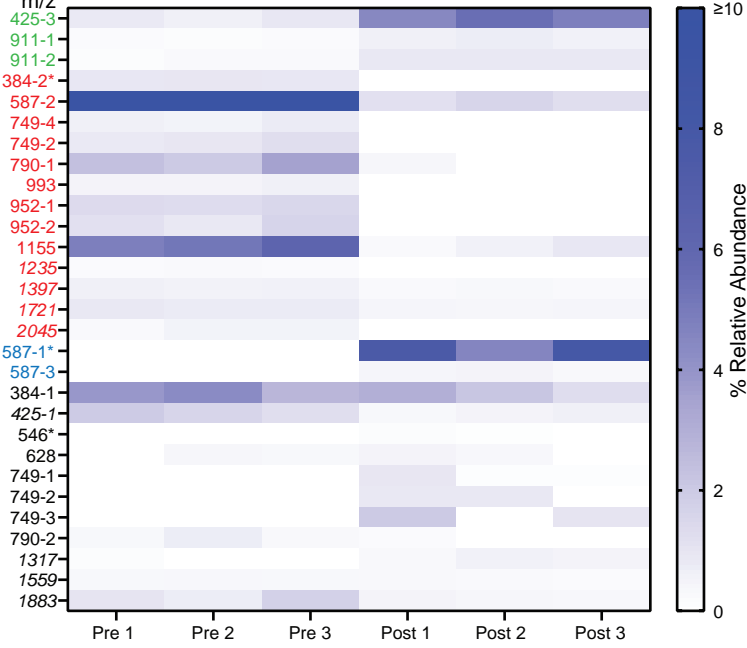
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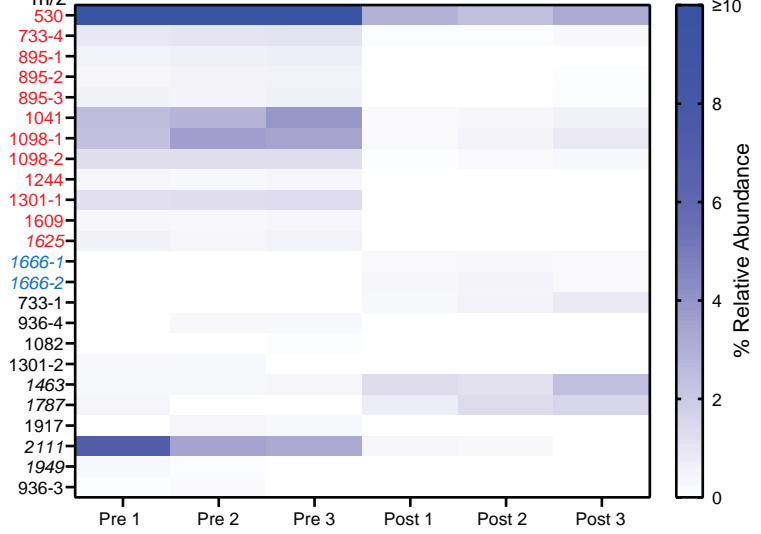
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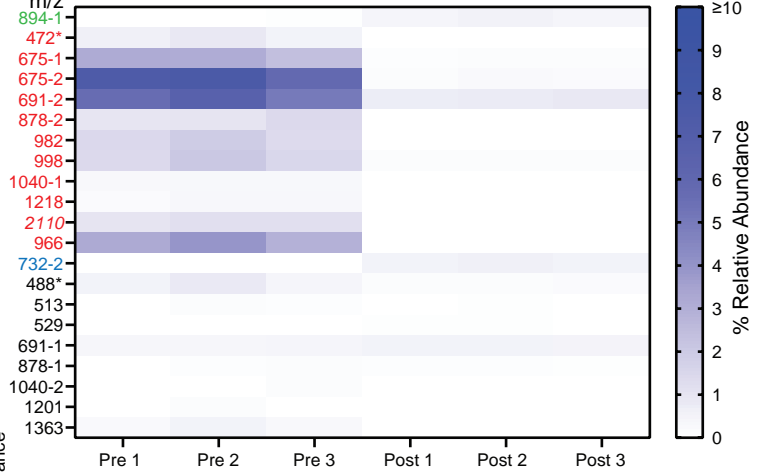
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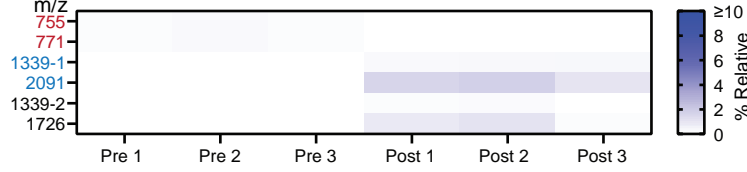
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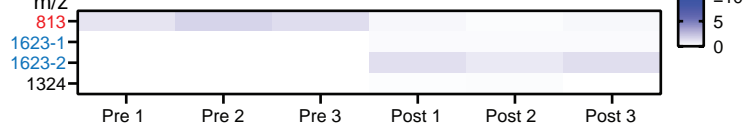
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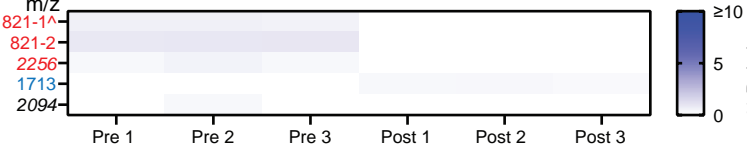
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G Sulfated & Fucosylated

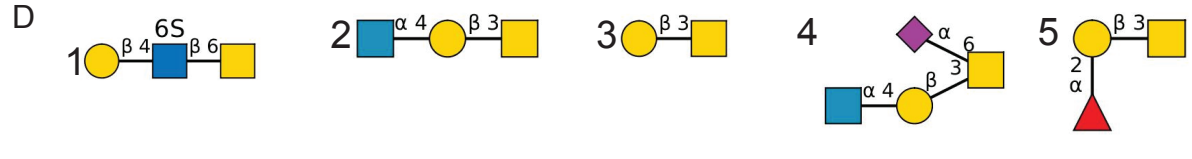
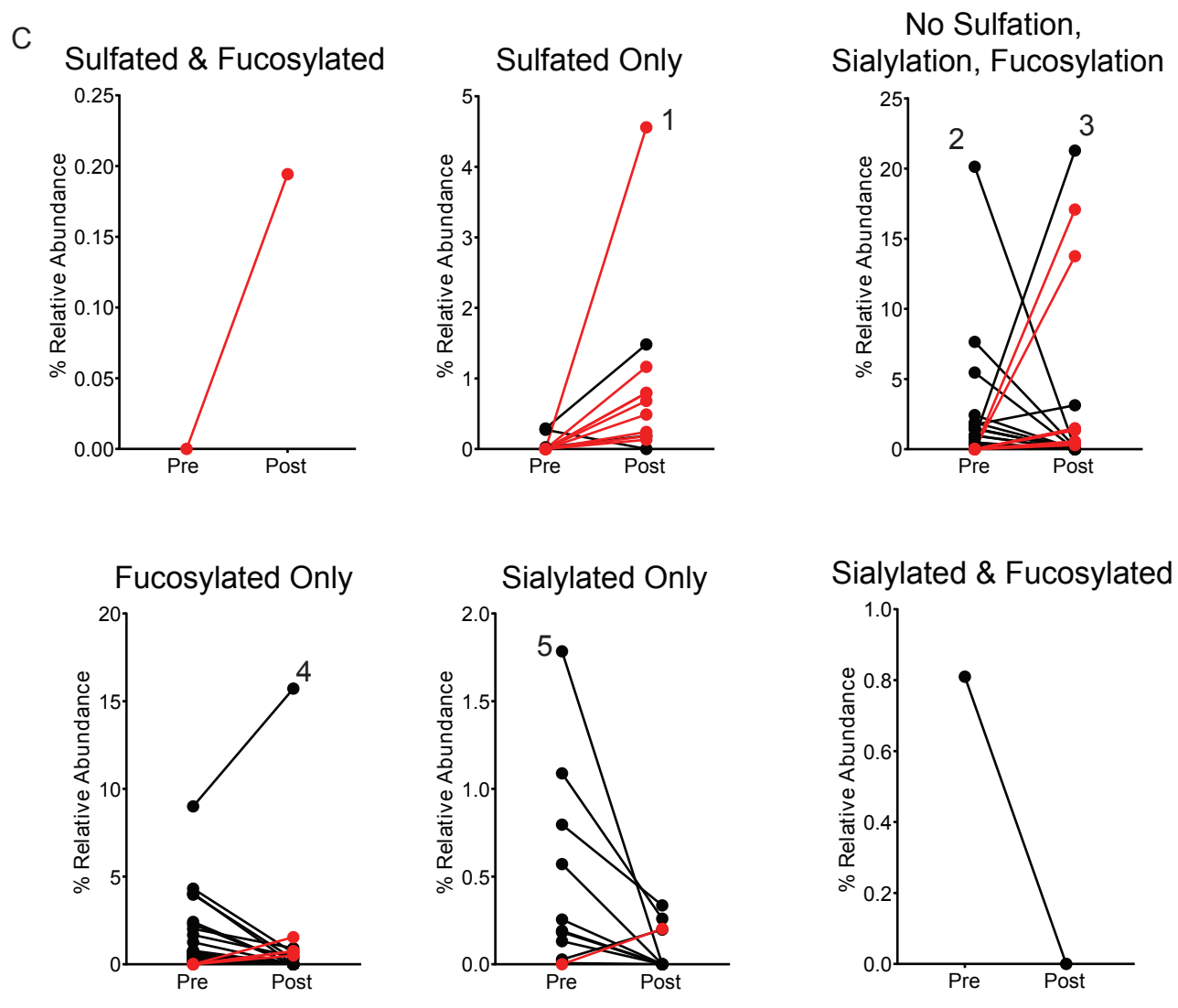
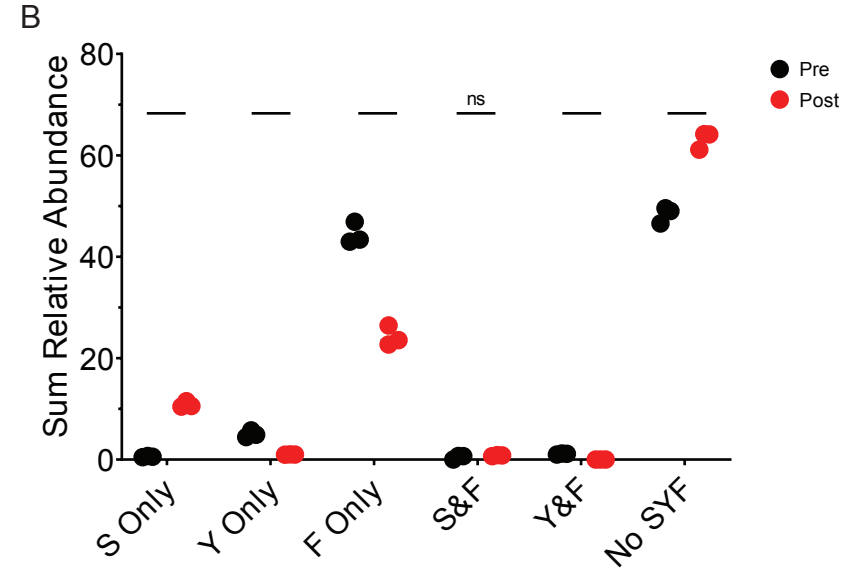
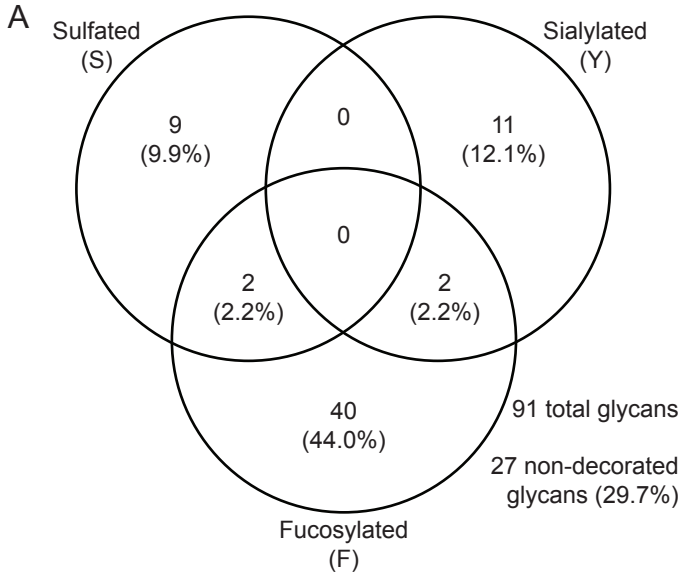


H Sialylated & Fucosylated



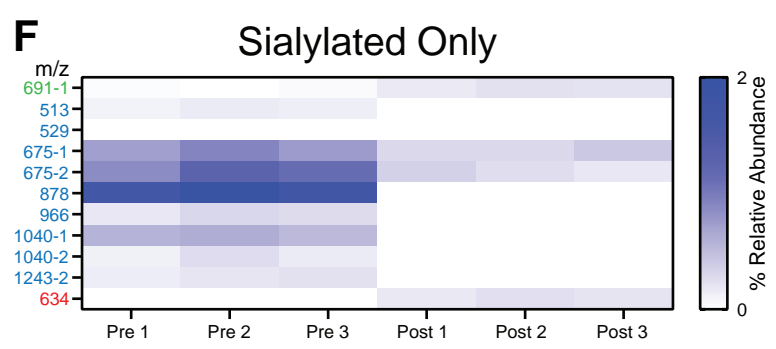
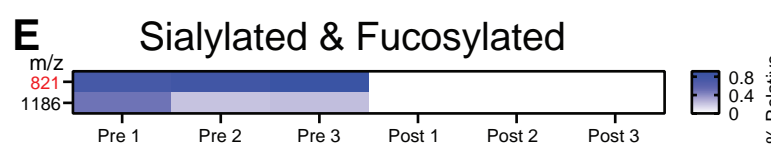
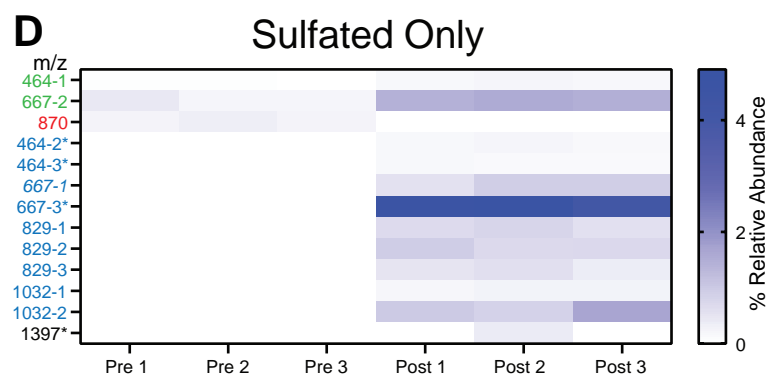
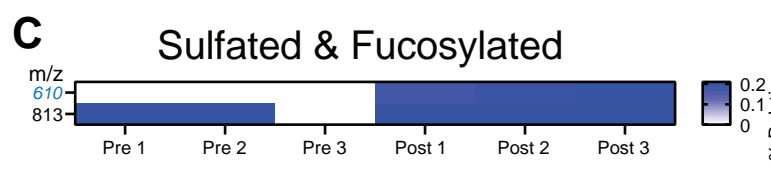
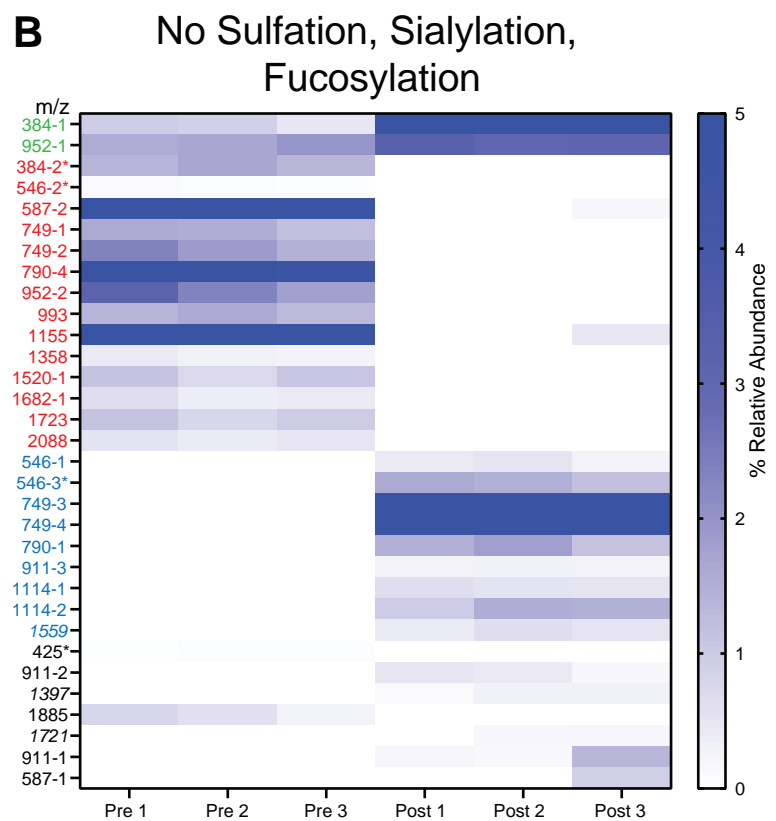
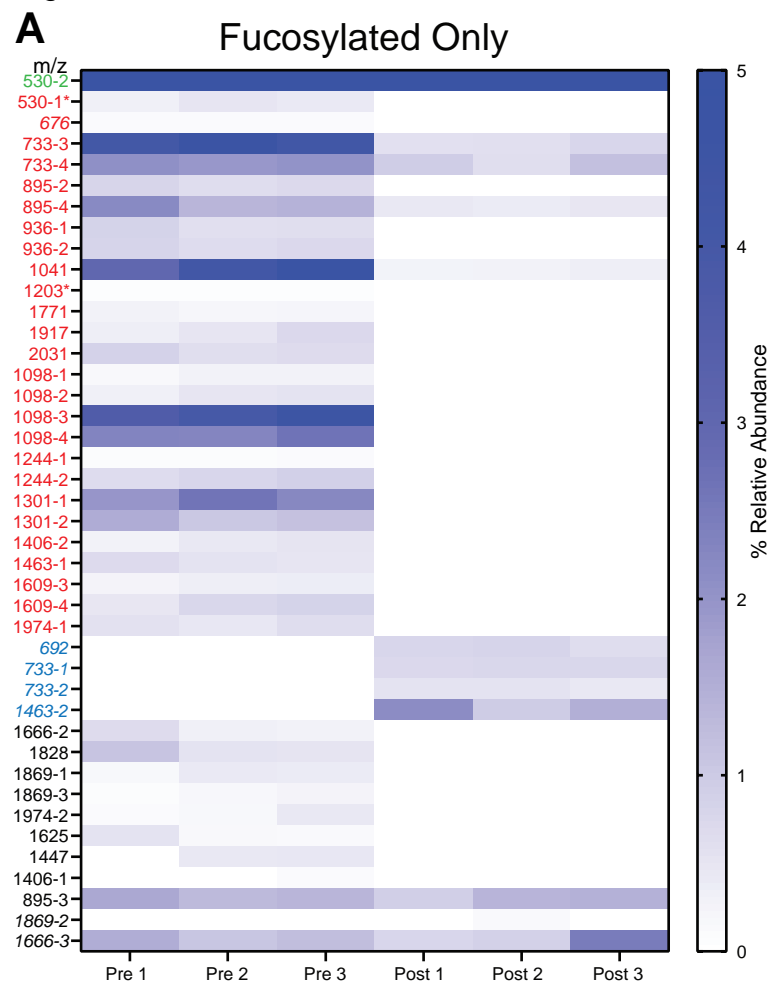
Supplemental Figure S9. LC-MS/MS glycans identified before and after growth of *R.*

***torques* on cMUC2. A-H**, Glycans detected were grouped based on structural features: sulfated, sialylated and fucosylated (**A**), sulfated only (**B**), no sulfation, sialylation, fucosylation (**C**), fucosylated only (**D**), sulfated and sialylated (**E**), sialylated only (**F**), sulfated and fucosylated (**G**), and sialylated and fucosylated (**H**). Heat maps indicate percent relative abundance of each glycan pre- or post-growth of *R. torques*. *M/z* values in green represent glycans detected in at least one pre-growth sample that significantly increase in abundance post-growth; glycans in red decreased in abundance post-growth compared to pre-growth, glycans in blue were undetected pre-growth but were detected post-growth; and glycans in black were not significantly different in abundance pre- or post-growth. Statistics were analyzed using paired, two-tailed t-tests between the three pre- and three post-growth samples for each glycan. * = peeling reaction product, ^ = fuc-ol terminating glycan, italicized *m/z* values = *N*-glycan.



Supplemental Figure S10. Fucosylated and sialylated glycans are degraded after growth of *R. torques* on PGM, but sulfated glycans accumulate. **A**, Number and percentage of total glycans detected in PGM pre- and/or post-growth of *R. torques* displaying indicated structural features by LC-MS/MS. **B**, Sum relative abundance of glycans and peeling reaction products pre- or post-*R. torques* growth on PGM with respective structural features. Statistical analysis was performed with paired, two-tailed t tests between pre- and post-growth samples for each structural feature category. S=sulfated, Y=sialylated, F=fucosylated. **p<0.01, ***p<0.001. **C**, relative abundance of glycans detected that were retained on the mucin polypeptide backbone and were significantly different in pre- or post-growth samples of *R. torques* on PGM. Red lines indicate glycans that were not detected in any of the pre-growth samples but were detected in the post-growth samples. Each point represents the average relative abundance of three samples. **D**, Putative structures of select detected glycans. Numbers refer to corresponding plot in C. Structures generated with DrawGlycan⁶⁹.

Figure S11

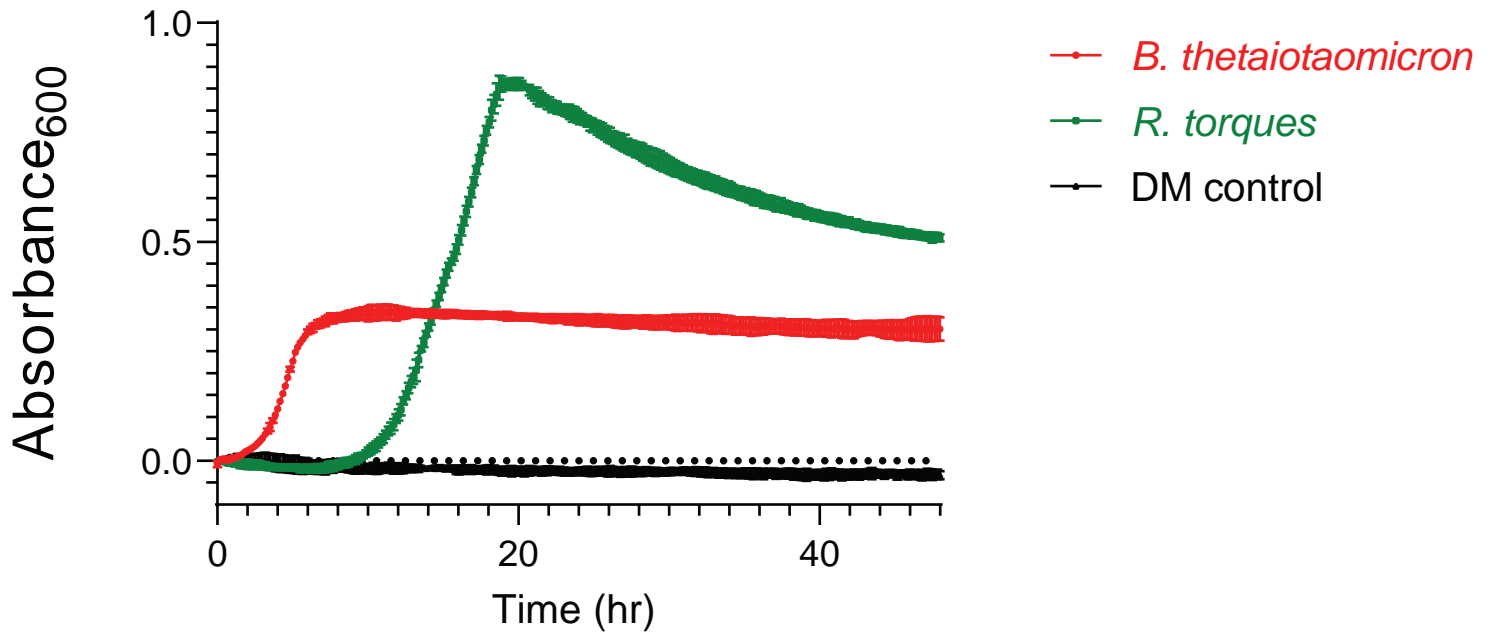


Supplemental Figure S11. LC-MS/MS glycans identified before and after growth of *R.*

***torques* on PGM. A-F**, Glycans detected were grouped based on structural features: fucosylated only (**A**), no sulfation, sialylation, or fucosylation (**B**), sulfated and fucosylated (**C**), sulfated only (**D**), sialylated and fucosylated (**E**), and sialylated only (**F**). Heat maps indicate percent relative abundance of each glycan pre- or post-growth of *R. torques*. Molecular weights in green represent glycans detected in at least one pre-growth sample that significantly increase in abundance post-growth; glycans in red decreased in abundance post-growth compared to pre-growth, glycans in blue were undetected pre-growth but were detected post-growth; and glycans in black were not significantly different in abundance pre- or post-growth. Statistics were analyzed using paired, two-tailed t-tests between the three pre- and three post-growth samples for each glycan. MW = molecular weight. * = peeling reaction product, italicized m/z values = *N*-glycan.

Figure S12

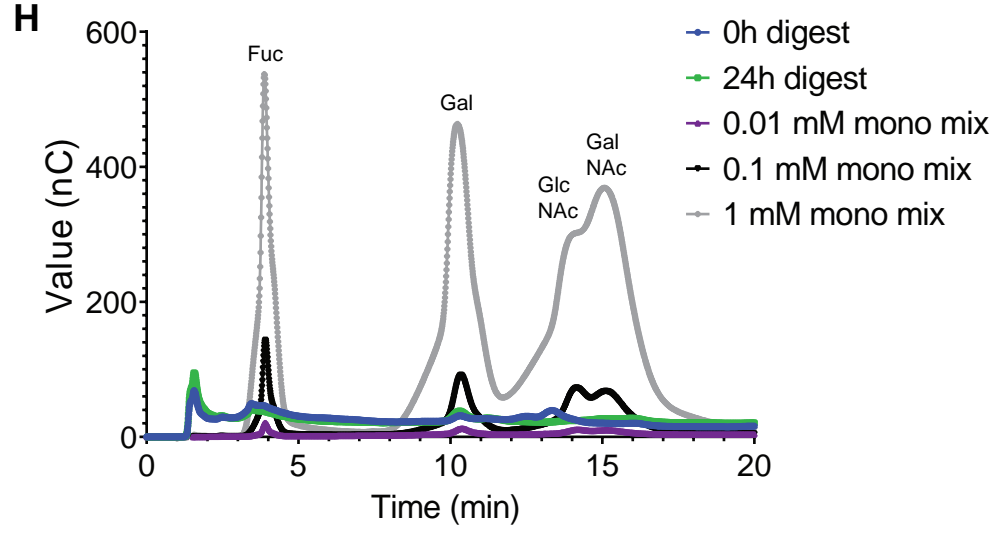
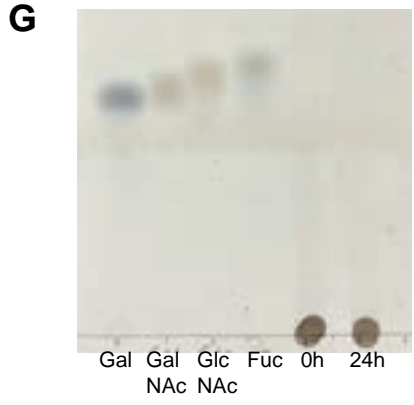
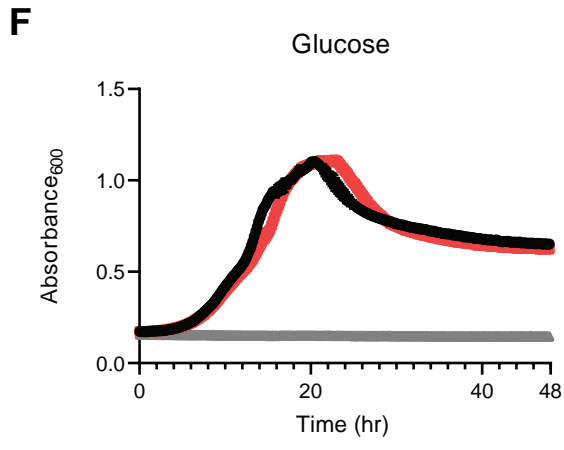
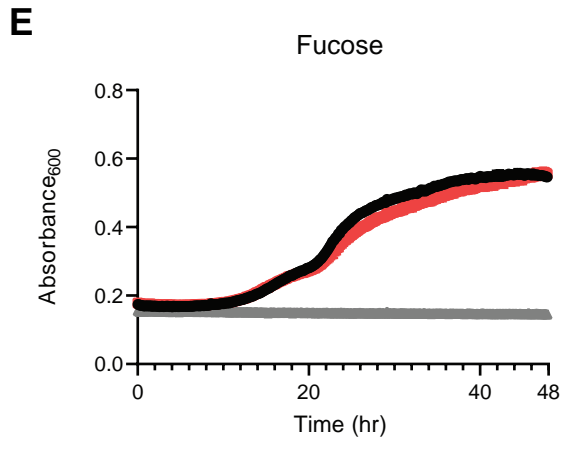
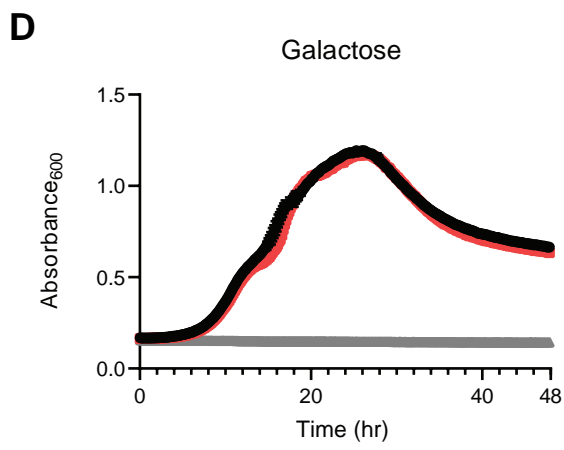
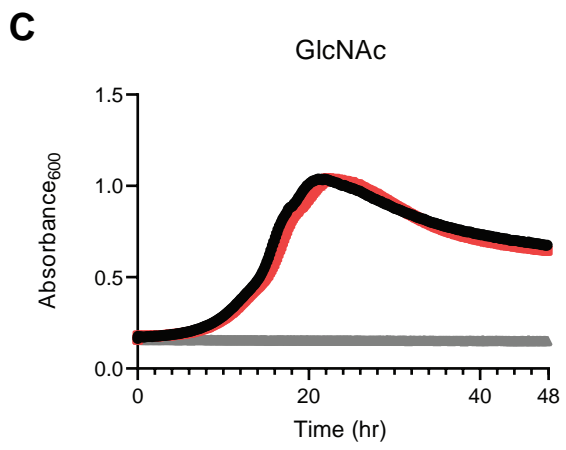
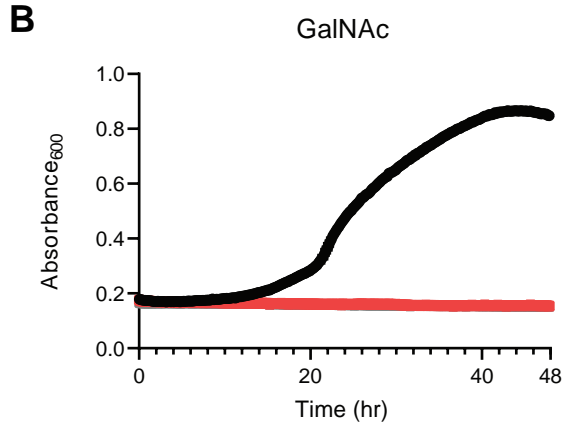
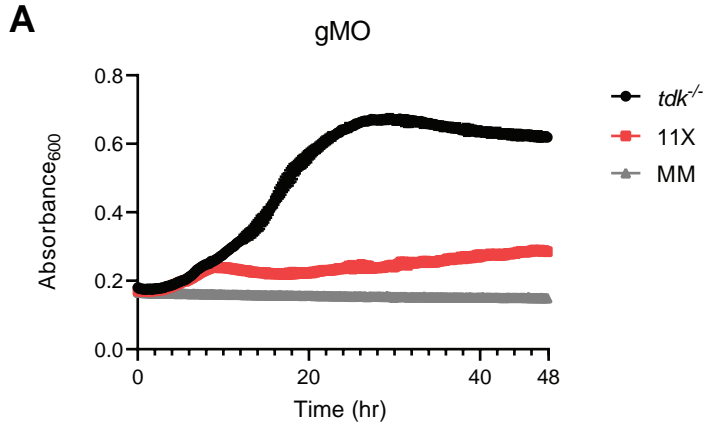
DM+PGM



Supplemental Figure S12. Growth of *B. thetaiotaomicron* and *R. torques* on PGM in partially defined medium (DM). *B. thetaiotaomicron* *tdk*^{-/-} and *R. torques* were grown anaerobically at 37°C in DM with PGM (10 mg/ml final concentration) as the major carbohydrate source and growth was monitored by Absorbance₆₀₀. DM control contained DM+PGM only. Each point represents n=3 and error bars represent SD.

Figure S13

Figure F



Supplemental Figure S13. Growth of *B. thetaiotaomicron* tdk^{-/-} and *B. thetaiotaomicron* 11X mutant on glucose, gMO, and mucin monosaccharides and dialysis validation of *R. torques* PGM digest. **A-F**, Both strains of *B. thetaiotaomicron* were grown in Bacteroides minimal medium with gMO (**A**), *N*-acetylgalactosamine (**B**, GalNAc), *N*-acetylglucosamine (**C**, GlcNAc), galactose (**D**), fucose (**E**), or glucose (**F**). MM = Bacteroides minimal medium negative control. gMOs were at a final concentration of 10 mg/ml, and all others were 5 mg/ml. The first collected A₆₀₀ value (0 h) was omitted from all conditions due to high baseline noise. Each point represents n=3 and error bars represent SD. **G**, thin layer chromatogram of *R. torques*-digested PGM substrates (0h control and 24h digest) to assess efficiency of dialysis in removing monosaccharides from these substrates. Standards at 10 mM were spotted once (3 μl), PGM substrates at 20 mg/ml were spotted twice (3 μl each). Gal=galactose, GalNAc=*N*-acetylgalactosamine, GlcNAc=*N*-acetylglucosamine, Fuc=fucose. **H**, HPAEC-PAD analysis of 0h and 24h PGM digests for presence of monosaccharides after dialysis. Mono mix=fucose (fuc), galactose (gal), *N*-acetylgalactosamine (GalNAc), and *N*-acetylglucosamine (GlcNAc). Concentrations for mono mixes reflect concentration of each monosaccharide in the mix.