Article

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Detection of transcriptome-wide microRNA-target interactions in single cells with agoTRIBE

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Supplemental Figures



Figure S1. agoTRIBE and Flag/HA-TNRC6B colocalization and lack of detection of endogenous ADAR2 in HEK293T cells. a, Transiently co-expressed agoTRIBE and Flag/HA-TNRC6B are colocalized in the cytoplasm with P-bodies. Anti-ADAR2 detects agoTRIBE (red), the P-bodies are labelled by Flag/HA-TNRC6B (green), a merged image with DAPI. b, Endogenous ADAR2 expression is not detectable in immunolabeled HEK-293T cells. c, Human ADAR2 staining is nuclear (nucleolar) in HEK-293T cells transiently overexpressing human full length ADAR2 WT protein (shown by arrows). The arrowhead shows a neighbouring untransfected HEK-293T cell lacking ADAR2 staining. d, Western blotting analysis of endogenous ADAR2 expression in HEK-293T cells. 40ug of the whole cell extracts of HEK-293T cells transfected with a plasmid expressing hADAR2 WT and un-transfected cells control were separated on SDS-PAGE and simultaneously blotted with polyclonal rabbit α -ADAR2 (GenTex GTX54916) and monoclonal anti- α -Tubulin (Sigma T5168). The antibodies were simultaneously developed with goat anti-rabbit IgG-HRP (Abcam ab6721) and goat antimouse IgG, HRP conjugated (Sigma SAB3701066), and Immobilon Forte Western HRP substrate (Millipore, Merck WBLUF0500). The 81kD and 50kD bands corresponding to hADAR2 and loading control Tubulin are indicated (lane 2 and lanes 2-3, respectively).



Figure S2: miRNA expression change upon agoTRIBE transfection. In total 3 batches of HEK293T cells transfected with agoTRIBE construct and 3 batches of control cells were subjected to small RNA-seq (Methods). DESeq2 identified 13 miRNAs that were differentially expressed in agoTRIBE-transfected cells, using the Benjamini-Hochberg procedure (two-sided). However, these were all lowly abundant, with none of the 13 miRNAs ranking among the top 50 expressed miRNAs in either agoTRIBE or control cells according to the small RNA-seq data.



Figure S3. Positional information in target editing. The observed distance (in nt) between the experimentally reported eCLIP binding site and the editing positions identified by agoTRIBE in the longest mono-exonic 3' UTRs per gene. Expected distance is the result of randomizing editing positions following a uniform distribution across the 3' UTRs and comparing them with eCLIP reported binding sites.



Figure S4. T6B peptide design. a, Schematic representation of TNRC6B Ago-binding domains and T6B, the short TNRC6B-derived protein peptide fused with mCherry. **b**, Transient expression of T6B-mCherry and agoTRIBE in HEK293T cells. In presence of agoTRIBE, T6B-mCherry is localized in cytoplasmic foci similar to P-bodies; highlighted with the arrows. All microscopy experiments were performed in at least two replicates and a representative experiment shown.



Figure S5. De-repression of miRNA targets identified by agoTRIBE and HITS-CLIP upon T6BmCherry transfection. The targets identified by AgoTRIBE and HITS-CLIP are excluded from 'Background'. 'agoTRIBE+seed site' consists of the targets identified by agoTRIBE which are high confidence TargetScan targets of at least one of the 10 most abundant miRNA in HEK-293T cells. Similarly, 'HITS-CLIP+seed site' consists of the targets identified by HITS-CLIP which are high confidence TargetScan targets of at least one of the 10 most abundant miRNA in HEK-293T cells.



Figure S6. Modular agoTRIBE. a, Nanobody fusion allows specific interaction with an EGFP-fused ADAR2DD in the cytosol of live cells. HEK-293T cells transfected with constructs transiently expressing GFP-nanobody fused with Ago2 (GNb_Ago2) and EGFP-tagged ADAR2DD. In presence of GNb_Ago2, EGFP-ADAR2DD segregates into cytoplasmic foci similar to P-bodies, highlighted with an arrow. **b**, Inducible heterodimerization of Ago2 and ADAR2DD by E3 and K3, the synthetic coiled-coil tags. Transient co-expression and localization of E3_Ago2 with K3_ADAR2DD, immunolabeled with anti-ADAR2 (red) and anti-Ago2 (green). **a**-**b** merged images with DAPI. All microscopy experiments were performed in at least two replicates and a representative experiment shown.

a A>G editing levels on agoTRIBE nanobody expression

b A>G editing levels on agoTRIBE heterodimer expression



Figure S7. A>G editing in 3' UTRs for agoTRIBE nanobody and heterodimer designs. a, A>G editing indicative of ADAR2 editing increases in agoTRIBE (eGFP-ADAR2DD and GNb-Ago2) transfected cells compared to control cells (GNb-Ago2) and cells transfected with the ADAR2 editing domain without Argonaute2 (eGPF-ADARDD: 'ADAR-only' controls), however the ADAR-only control in nanobody design gives a relatively higher background. b, A>G editing indicative of ADAR2 editing increases in agoTRIBE (K3-ADAR2DD and E3-Ago2) transfected cells compared to control cells (E3-Ago2) and cells transfected with the ADAR2 editing domain without Argonaute2 (K3-ADARDD: 'ADAR-only' controls).





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Human (hg38)	Chr7 Chr7:26,211	,443-26,214,823 Go 🕋 4 🕨 🛷 🗖 🗙 🤛		
ENSG00000122565 CBX3 miR-378 target	p222 p213 p211 p152 p143 p141 p123 p121 p111 q1121 q1123 q2111 q2113 q213 q221 q223 q311 q312 q3132 q321 q33 q34 q35 q361			
	4 26 212 000 hp	3 381 bp	26 214 000 bp	
control_TGTAAGTCAC+GAGGAC CGT_dedupped.bam Coverage	P-2E			
control_TGTAAGTCAC+GAGGAC CGT_dedupped.bam		and the second		
trbe_TTGGAGCAAT+GAATTCGC A_dedupped.bam Coverage				
tribe_TTGGAGCAAT+GAATTCGC A_dedupped.bam		And		
	miR-378			
Gene				
Human (hg38)	Chr2 Chr2:65,268	.075-65,271,566 Go 🖀 4 🕨 🖗 🗖 🗶 📁		
ENSG00000138071 ACTR2 miR-16 target	p252 p243 p233 p223 p21 p162 p15	p133 p12 p112 q11.1 q12.1 q13 q142 q21.1 q22.1 q23.1 q24.1	q24.3 q31.2 q32.2 q33.1 q34 q35 q362 q37.1	
	65 269 000 1 1	ър 65 270 000 Бр 1 1	65 271 900 hp. 1 1	
control_TGTAAGTCAC+CCTCGT; AGT_dedupped.bam Coverage	P-21 UNKNOWN			
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nbe_TGTGGTGCTA+TTCTTCGG G_dedupped.bam	miR-16		Muton.	
		11-10-1-		
Gene Human (bg38)	G chr1 G chr1:93.13	6.629-93 139 160 Co 🚔 🖌 🔊 🗖 🗑 🗖 🚽		
	p36.31 p36.13 p35.3 p34.3 p34.1 p32.3 p31.3	p31.1 p22.3 p21.3 p12 q12 q21.1 q22 q21.3	q24.3 q25.3 q31.2 q32.1 q32.3 q42.11 q42.2 q43 q44	
MTF2 miR-10 target		2 533 bp	83 139 600 kp	
Sequence 🔿				
control_TGTAAGTCAC+CCTCGT. AGT_dedupped.bam Coverage		· · · · · · · · · · · · · · · · · · ·		
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A_decupped bare		miR-10		
Gene				



Figure S8. Editing patterns of the miRNA target transcripts. Each screenshot shows the editing pattern in a control cell vs. an agoTRIBE transfected cell for one miRNA targets (similar to Figure 3e). The miRNA binding site for most expressed miRNA is indicated with arrow and name of the miRNA.



Figure S9: editing levels of stable and unstable transcripts. The 20% transcripts with the longest half-lives according to published data (Schueler, Munschauer et al., Genome Biology, 2014) were considered 'stable transcripts' and the 20% with the shortest half-lives were considered 'unstable transcripts'. Boxes indicate median and 25th and 75th quantiles, and the whiskers indicate smallest and largest values. Overall, the stable transcripts had more editing events summed over the 540 HEK293T single cells (599 editing events) than did the unstable transcripts (455 editing events). When normalizing the number of editing events to expression levels (by dividing the number of editing events with the sum of expression read counts over the 540 HEK293T cells) and considering only transcripts that were high-confidence TargetScan targets of the top 10 most abundant miRNAs in this cell line, the stable transcripts (0.11 edits/read counts).



Figure S10: Expression and targeting of stable and unstable transcripts over the cell cycle. The estimated overall miRNA targeting (in yellow) during cell cycle for 20% transcripts with the longest half-lives ('stable transcript') and 20% transcript with the shortest half-lives ('unstable transcripts'). Expression values and editing events were normalized to spike-ins and to the number of cells in each cell cycle stage, similar to Figure 3i (Methods).



Figure S11: Expression of agoTRIBE transcript over the cell cycle. The expression is estimated by the linker reads that are normalized to mRNA read counts, similar to Figure 3c (Methods).



The cell cycle AGOTRIBE protein abundance in HEK293T single cells

cell_phase

Figure S12: Expression of the AGOTRIBE protein over the cell cycle. Protein expression was profiled by fluorescently labeled anti-ADAR2 antibody (Methods).



Figure S13: miRNA expression in agoTRIBE transfected cells across the cell cycle. 10 most highly expressed miRNAs in each cell cycle stages are shown. Expression is normalized to RPM (reads per million values, Methods).

Supplemental Tables

Supplementary Tables are available at https://github.com/vaishnoviS/agoTRIBE/tree/main/SupplementaryTables Table1: Overview of all the bulk datasets Table2: Expression value for all genes for 6 samples in main figure 1 Table 3: Editing positions in 3' UTR for 6 samples in main figure 1 Table 4: Editing events in 3' UTR for 6 samples in main figure 1 Table 5: Overview of the HEK293T single cells - Cell ID, agoTRIBE or control. Seurat inferred cell state, reads mapping to Linker sequence Table6: Expression value for all genes for quality control processed HEK293T single cells Table7: Editing events in 3' UTR for quality control processed HEK293T single cells Table8: Differentially edited genes between cell cycle stages in HEK293T cells Table9: Overview of the K562 single cells – Cell ID, differentiated or undifferentiated, reads mapping to Linker sequence Table10: Expression value for all genes for K562 single cells Table11: Editing events in 3' UTR for K562 single cells Table12: Overview of all the bulk small RNA-seg datasets

Table13: Expression value for all miRNA for all bulk small RNA-seq datasets