Supplementary Information for Multiscale heterogeneity of white matter morphometry in psychiatric disorders

Ashlea Segal^{1,2,3}, Robert E Smith^{4,5}, Sidhant Chopra⁶, Stuart Oldham^{1,2,7}, Linden Parkes⁸, Kevin Aquino⁹, Seyed Mostafa Kia^{10,11,12,13}, Thomas Wolfers^{11,14,15}, Barbara Franke^{16,17}, Martine Hoogman^{17,18}, Christian F. Beckmann^{11,19,20}, Lars T. Westlye^{14,21,22}, Ole A. Andreassen^{21,22}, Andrew Zalesky^{23,24}, Ben J. Harrison²⁵, Christopher G. Davey²⁵, Carles Soriano-Mas^{26,27,28}, Narcís Cardoner^{27,29,30}, Jeggan Tiego^{1,2}, Murat Yücel^{1,31}, Leah Braganza¹, Chao Suo^{1,2,32}, Michael Berk^{33,34,35,36}, Sue Cotton^{34,35}, Mark A. Bellgrove¹, Andre F. Marquand^{10,11,37,38}, Alex Fornito^{1,2}

Correspondence to: Ashlea Segal Wu Tsai Institute, Department of Neuroscience, Yale University, New Haven, United States ashlea.segal@yale.edu

Table of Contents

Supplementary Methods	. 3
Participants	. 3
Anatomical data	. 3
Normative modelling	. 4
Evaluation of model performance	. 5
Characterizing voxel-wise heterogeneity of extreme deviations	. 5
Diffusion-weighted imaging acquisition parameters and processing	. 5
Streamline region assignments	. 6
Identifying streamlines intersecting with WMV deviations	. 7
Supplementary Results	8
Model evaluation	. 8
Analysis of positive deviations	. 8
Supplementary Tables	. 8
Table S2. Balanced classification accuracy scores (%) from the linear support vector machines from the	
held-out controls and cases	. 8
Table S3. Deviation burden at the tract-scale	10
Table S4. Deviation burden at the tract-scale (weight threshold 75%)	10
Table S5. Deviation burden at the region-scale	10
Table S6. Deviation burden at the functional network-scale	11
Table S7. Deviation burden at the functional network-scale (20 network)	11
Supplementary Figures	12
Figure S1. Performance metrics from the normative models	12
Figure S2. Distribution of negative extreme WMV deviation overlap in HC _{test} cohort across scales	13
Figure S3. Connections affected by extreme negative WMV deviations weight threshold of 75%	13
Figure S4. Functional networks affected by extreme negative WMV deviations 20 network parcellation.	14
Figure S5. Distribution of positive extreme WMV deviation overlap in HC _{test} cohort across scales	14
Figure S6. Spatial overlap in extreme positive WMV deviations.	15
Figure S7. Connections affected by extreme positive WMV deviations.	16
Figure S8. Regions attached to connections affected by extreme positive WMV deviations	18
Figure S9. Functional network affected by extreme positive WMV deviations.	18
References	19

Supplementary Methods

Participants

This study included 3746 individuals (1865 healthy controls, HC; 1833 cases across six different diagnostic categories) from 14 separate, independently acquired dataset and 25 scan sites. Full details on study design and clinical characteristics have been described previously for each dataset (see Table 1 for relevant references). Ethnicity was not reported in many of the datasets, and for those datasets that did report ethnicity, most, if not all participants were Caucasian. Each study was approved by the relevant ethics committee and written informed consent was obtained from each participant.

The final sample we examined was taken from a larger pool of individual recruited across the 14 datasets. In addition to the specific quality control procedures used within each study (references in Table 1), we performed a series of additional quality control checks and exclusions for our analysis. Specifically, we excluded participants who:

- Were below 18 years or above 64 years of age (*N*=346)
- Lacked the necessary clinical data (such as a clinical diagnosis or, for healthy controls, the absence of any clinical diagnosis) or demographic information (age and sex, *N*=53)
- T1-weighted structural magnetic resonance imaging (MRI) scans failed the preprocessing pipeline (*N*=63)
- Had T1-weighted MRI scans that did not pass our rigorous manual (*N*=53) and automated quality control procedures (*N*=153), as detailed in Segal et al. (1)
- Were from sites with fewer than 10 individuals in the same group and sex (described in the Normative model section below; *N*=217)

Thus, our final sample for analysis included 2,759 individuals (1,465 controls and 1,294 cases). Table 1 presents demographic details of this cohort.

Anatomical data

Quality assurance procedures and the data processing pipeline, which was applied to all raw T1weighted images obtained for each dataset, are provided in detail in Segal et al. (1), with image acquisition parameters for each dataset provided in Table S1 in Segal et al. (1).

Briefly, voxel-wise WMV was estimated using the Computational Anatomy Toolbox(2) (CAT12 r113, <u>http://dbm.neuro.uni-jena.de/cat/</u>), which is included as an extension of Statistical Parametric Mapping software (SPM12, <u>http://www.fil.ion.ucl.ac.uk/spm/software/spm12</u>) in MATLAB v9.8 using the default settings.

First T1-weighted images were corrected for bias-field inhomogeneities, segmented into grey matter, white matter, and cerebrospinal fluid, then spatially normalised to the SPM MNI IXI555 template (https://brain-development.org/ixi-dataset/). The normalised segmented white matter images were modulated and spatially smoothed with an 8mm full width at half maximum (FWHM) Gaussian smoothing kernel. This approach allowed us to assess voxel-wise differences in the absolute amount (volume) of white matter, taking the extent of voxel-wise volumetric adjustments needed to align each participant with the registration template into account. Finally, to exclude non-white matter voxels from our analysis, we generated a mean image from all the normalized unmodulated white matter maps and retained voxels with a tissue probability ≥ 0.2.

Normative modelling

Normative models estimate the centiles of variation around the mean (referred to as the normative range) of a response variable such as WMV from a set of clinically relevant covariates, such as age and sex, across a large control sample (defined here as a group of people without a psychiatric diagnosis), referred to as a *training set*. These estimates are then used to quantify deviations from the normative range of individuals in the *test set*, which typically consists of cases of interest and a held-out set of controls for comparison.(3–5)

As per Segal et al.(1), the training set (HC_{train}, n = 1,465) for the normative model was created by randomly selecting from each scanner site either 90% of controls or all controls if the sample size for that site was less than 30. The test set consisted of all clinical data (n = 1,294), as well as the remaining controls from each scan site (HC_{test}, n = 269), which offered a normative benchmark for assessing case-specific model deviations.

B-spline basis expansion over age was used to model non-linear effects, and likelihood warping was used to model non-Gaussian and heteroscedastic imaging phenotypes. To handle the site effect, we included scan site as a covariate (6), as in (7).

To identify positive and negative extreme deviations in white matter voxel (WMV) estimates from the normative model, we thresholded the deviation maps at |z| > 2.6 (p < .005) and applied a 10-voxel extent threshold (Figure 1d). This method was used for each individual in both clinical and control (HC_{test}) groups. We chose this approach for consistency with previous work (1,8) and because it provides a uniform definition of extreme deviations, unlike adaptive thresholding methods like the false discovery rate (FDR) (9), which vary per individual.

4

Evaluation of model performance

We assessed model fit for each brain region by evaluating five performance metrics: (i) explained variance (EV); (ii) the standardized mean-squared error (SMSE); (iii) the mean standardized log-loss (MSLL); (iv) skew; and (v) kurtosis. We removed voxels with poor performance (SMSE > 1.5, MSLL > 0.5, or skew > 1.5), which amounted to 1.7% voxels in total (Figure S1). We also evaluated the model's efficacy in partitioning site-related variance in the data using linear support vector machines. Specifically, we used a series of one-versus-all linear support vector machines (with default slack parameter = 1) trained on the deviation z-maps from the HC_{test} subset to classify scan sites. For each site, we ran a 2-fold SVM classifier to obtain the mean balanced accuracy score for the given site. In this analysis, a balanced accuracy close to chance-level (50%) indicates that the resulting deviations were not contaminated by residual site effects (Table S2)

Characterizing voxel-wise heterogeneity of extreme deviations

We characterized the heterogeneity of thresholded person-specific extreme deviation maps for each disorder using a non-parametric approach, following Segal et al. (1). This procedure involved subtracting the HC_{test} overlap map from each disorder's overlap map, resulting in an overlap difference map for each disorder, separately for positive and negative extreme deviations. For each disorder, we then permuted group labels (i.e., HC, case) and repeated the procedure 10,000 times to derive an empirical distribution of overlap difference map under the null hypothesis of random group assignment. For each white matter voxel, we obtained *p*-values as the proportion of null values that exceeded the observed difference. The tails of the null distribution (i.e., values associated with p<0.10) were approximated using a generalized Pareto distribution (10), as implemented in the Permutation Analysis of Linear Models software package (PALM alpha116) (11), to allow inference at arbitrarily high levels of precision. Statistically significant effects were identified using an FDR-corrected (9) threshold of p_{FDR} <.05, two-tailed.

Diffusion-weighted imaging acquisition parameters and processing

We used diffusion tractography to identify the axonal pathways that intersected each WMV deviation (Figure 1f-h). We then used this information to build matrices encoding the specific tracts affected by WMV deviations (Figure 1i-k). The deviation-related structural circuitry was mapped in an independent cohort as high-quality DWI data is essential for accurate estimate of

tractograms and such data was not available for all clinical samples. This logic precisely aligns with the logic of standard lesion network mapping studies of neurological patients (12–16). We focused on the same sub-sample of 150 people in the HCP dataset as Segal et al (1) to minimise the computational burden.

Data were acquired on a customized Siemens 3 T Connectome Skyra scanner at Washington University in St. Louis, MO, USA, using a multishell protocol for the diffusion weighted imaging with the following parameters: 1.25-mm^3 voxel size; repetition time (TR) = 5520 ms; echo time (TE) = 89.5 ms; field of view (FOV) of 210 mm by 180 mm; 270 directions with *b* = 1000, 2000, 3000 s/mm² (90 per *b* value); and 18 *b* = 03 volumes. Structural T1-weighted data were acquired with 0.7-mm voxels, TR = 2400 ms, TE = 2.14 ms, and an FOV of 224 mm by 224 mm.

The diffusion data used underwent the HCP minimal pre-processing pipeline,(17) which included normalization of mean b = 0 images across diffusion acquisitions and correction for echo-planar imaging susceptibility and signal outliers, eddy current-induced distortions, slice dropouts, gradient nonlinearities, and participant motion. T1-weighted data were corrected for gradient and readout distortions before being processed with FreeSurfer.(18) The details of this pipeline are provided in more detail elsewhere.(19) Using the corrected diffusion data, we estimated fibre orientation distributions (FODs) using multishell Constrained Spherical Deconvolution,(20) which formed the basis for probabilistic tractography with the Fibre Orientation Distributions (iFOD2) algorithm, as implemented in MRtrix3.(21–24) We applied Anatomically Constrained Tractography (ACT) to improve the biological accuracy of the structural networks.(25) ACT uses multi-tissue segmentation to ensure streamlines are beginning, traversing, and terminating in anatomically plausible locations. A total of 10 million streamlines were generated using dynamic seeding along with default MRtrix3 parameters.(21–24,26)

Streamline region assignments

We examined streamline estimates of connectivity between 132 regions, defined by combining well-validated parcellations of the cortex (Schaefer 100 region 7 network parcellation)(27) and subcortex (Tian Subcortical Atlas Scale II, 32 region parcellation).(28) The atlas was then transformed from the template surface to the surface of each individual in the HCP_{150} sample based on the spherical registration procedure implemented in FreeSurfer.(18) Next, the parcellation was projected to a volumetric image and resampled to the same resolution as the HCP_{150} diffusion data using *FSL's applywarp* with nearest-neighbour interpolation.(29) The

parcellation was subsequently combined with each person's tractogram and streamlines were assigned to the nearest brain region within 5 mm of the streamline endpoints using *MRtrix3* (v 3.0) *tck2connectome* function.(24,30)

The tractograms were transformed to MNI IXI555 space to facilitate interpretation with respect to the individual-specific WMV deviation maps. To this end, we used *FSL(v* 6.0.6) *FLIRT* and *FNIRT*(29,31) to generate and compose transforms for co-registering each HCP₁₅₀ participant's diffusion data to their T1-weighted volume, and their T1-weighted volume to the SPM MNI IXI555 template. Finally, we used *MRtrix's tcktransform* to warp the tractogram to MNI IXI555 space using the composed warp.

Identifying streamlines intersecting with WMV deviations

To construct the dysconnection overlap matrices, we used the extreme WMV deviation cluster map (Figure 2a) of each patient/control to filter each HCP₁₅₀ individual's tractogram (Figure 2b-c) to retain only streamlines intersecting with at least one WMV cluster from the deviation maps. This generated a filtered streamline count matrix (C), indicating pairwise connections affected by deviations. We binarized C', setting C'_{ij} to 1 if the filtered edge's streamline count exceeded 10, and 0 otherwise (Figure 2d).

The *C*' matrices were aggregated across HCP₁₅₀ individuals to create a consensus matrix (*C*^{*G*}), which encoded the proportion of participants for whom each edge was present in *C*'. This matrix estimates the probability that a given connection is associated with observed WMV deviations in that participant in the clinical group (or HC_{test} group), given individual variability in normative connectome architecture. A separate *C*^{*G*} matrix was obtained for each individual in the clinical and HC_{test} groups. We binarized *C*^{*G*}, setting edges with weights less than 50% to zero, to encode connections affected by a WMV extreme deviation (termed extreme edge deviation matrix; Figure 2e). We repeated the analysis with a 75% threshold to validate our findings (see Figure S3).

Finally, we aggregated these binary *C^G* matrices for each clinical and HC_{test} group, separately for positive and negative deviations, creating group-specific overlap matrices. Each element in these matrices shows the proportion of individuals in each group affected by an extreme deviation in that pairwise connection, referred to as the dysconnection overlap matrix (Figure 2f).

To evaluate group differences using permutation-based inference, similar to the voxelscale deviation analysis, we subtracted the HC_{test} group's dysconnection overlap matrix from each clinical group's matrix and performed mass univariate testing across connectome edges.

7

Significant differences in dysconnection overlap were identified using an empirical null distribution from 10,000 permutations and a generalized Pareto tail approximation (10), with a threshold of pFDR < .05, two-tailed. This was done separately for positive and negative extreme deviations.

Supplementary Results

Model evaluation

The results from the linear support vector machine indicated that the balanced accuracy scores ranged between 49.71 and 58.53% across all sites, suggesting that resulting deviations were not contaminated by residual site effects (Table S2).

Rank correlations indicated that scan quality (as measured by CAT12's IQR) was weakly associated with extreme deviation burden for the cases (*rho* = 0.07, *p*=.02), but not for HC_{test} (*rho* = -0.04, *p*=.55), however this difference was not statistically significant (Fisher's z-transformation, *z* = 1.64, *p* = 0.10).

Analysis of positive deviations

Despite both cases and controls generally showing more extreme positive than negative deviations (Figure S6), there were few differences in the extent of overlap, with <1% of edges show significantly greater overlap in ASD and SCZ (Figure S7) compared to controls, and 76.52% of regions and all networks showing significantly greater overlap in ASD, relative to controls (Figure S8 and Figure 9 respectively). No other disorders at any of the spatial scales showed significantly greater overlap compared to controls after correcting for multiple comparisons.

Supplementary Tables

Table S2. Balanced classification accuracy scores (%) from the linear support vector machinesfrom the held-out controls and cases

Dataset	Site	Held-out controls (HC _{test})
ABIDE I	CALTECH	N/A
	CMU	N/A
	LEUVEN_1	N/A
	MAX-MUN	50.00
	NYU	50.00
	PITT	N/A
	SBL	N/A
	USM	50.00
ABIDE II	BNI	50.00

	IU	N/A
ASRB	BRIS	50.00
	MELB	54.81
	PERT	N/A
	SYDN	50.00
FEMS		50.00
MON		56.42
IMPACT		58.53
KANMDD		N/A
MITASD		50.00
OCDPG		50.00
RUSMDD		50.00
SPAINOCD		49.71
TOP15		51.44
WASHASD		50.00
YoDA		56.25

*N/A = collection sites where data for HCs was < 30, therefore all HC data was included in training set.

Table S3. Deviation burden at the tract-scale

	Negative extreme deviations				Positive extreme deviations			
	Participants with deviation, %	Max overlap, %	Significant loci at p<.05 (p _{FDR} <.05), %		Participants with deviation, %	Max overlap, %	Significant (p _{FDR} <.05),	t loci at <i>p<.05</i> ,%
			PAT	HC	, / ·		PAT	HC
HC _{test}	19.7	8.18			58.36	30.86		
ADHD	16.34	6.54	0	0.31 (0)	67.97	35.95	0.32 (0)	0.16 (0)
ASD	31.68	11.88	37.89 (6.08)	0	70.79	40.1	17.26 (0.02)	0
BP	19.74	6.14	0.60 (0)	0.01 (0)	58.77	32.02	0.18 (0)	1.83 (0)
MDD	19.25	6.21	0.21 (0)	0	50.93	22.36	0	7.72 (0.01)
OCD	10.18	4.19	0	1.86 (0)	62.87	29.94	0	4.18 (0)
SCZ	35.77	15.4	19.99 (3.33)	0	63.19	39.16	2.59 (0.10)	0

Table S4. Deviation burden at the tract-scale (weight threshold 75%)

	Negative extreme deviations				Positive extreme deviations					
	Participants with deviation, %	Max overlap, %	Significant connections at <i>p<.05</i> (p _{FDR} <.05), %		Significant connections at p<.05 (p _{FDR} <.05), %		Participants with deviation. %	Max overlap, %	Significant c at p<.05 (p _{FL}	connections _{DR} <.05), %
			PAT	HC			PAT	HC		
HC _{test}	19.7	6.69			58.36	28.62				
ADHD	16.34	6.54	0.01	0.14 (0)	67.97	33.99	0.17 (0)	0.21 (0)		
ASD	31.68	10.40	27.68 (1.21)	0	70.3	37.13	11.41(0)	0		
BP	19.74	4.82	0.71 (0)	0	58.77	29.39	0.13 (0)	1.62 (0)		
MDD	19.25	5.59	0.26 (0)	0	50.93	19.88	0	5.60 (0.03)		
OCD	10.18	3.59	0	0.69 (0)	62.87	25.15	0	3.83 (0)		
SCZ	35.77	13.84	12.53 (0)	0	63.19	36.55	1.65 (0.06)	0.05 (0)		

Table S5. Deviation burden at the region-scale

	Negative extreme deviations				Positive extreme deviations			
	Participants with deviation, %	Max overlap, %	Significant reg p<.05 (p _{FDR} <.03	Significant regions at F p<.05 (p _{FDR} <.05), % v d		Max overlap, %	Significant p<.05 (p _{FDR} •	regions at <.05), %
			PAT	HC			PAT	HC
HC _{test}	19.7	14.87			58.36	53.53		
ADHD	16.34	15.03	0	1.51 (0)	67.97	60.13	10.61 (0	0
ASD	31.68	27.23	86.36 (84.85)	0	70.79	64.85	83.33 (75)	0
BP	19.74	17.11	0	0	58.77	56.58	0	0
MDD	19.25	13.04	0	0	50.93	46.58	0	30.30 (0)

OCD	10.18	8.98	0	31.82	62.87	58.08	0	0
				1.51				
SCZ	35.77	33.42	92.42(90.91)	0	63.19	59.79	8.33 (0)	0

Table S6. Deviation burden at the functional network-scale

	Negative extreme deviations				Positive extreme deviations			
	Participants with deviation, %	Max overlap, %	Significant networks at p<.05 (p _{FDR} <.05), %		Participants with deviation, %	Max overlap, %	Significan at p<.05 (µ	t networks o _{FDR} <.05), %
	·		PAT	HC			PAT	HC
HC _{test}	19.7	18.86			58.36	57.25		
ADHD	16.34	16.34	0	0	67.97	66.67	0	0
ASD	31.68	31.68	100 (100)	0	70.79	69.31	100 (100)	0
BP	19.74	19.74	0	0	58.77	58.77	0	0
MDD	19.25	18.01	0	0	50.93	50.93	0	0
OCD	10.18	10.18	0	90 (90)	62.87	61.68	0	0
SCZ	35.77	35.25	100 (100)	0	63.19	63.19	0	0

Table S7. Deviation burden at the functional network-scale (20 network)

	Negative extreme deviations				Ро	sitive extrem	ne deviatior	IS
	Participants with deviation, %	Max overlap, %	Significant networks at <i>p</i> <.05 (<i>p</i> _{FDR} <.05), %		Participants with deviation, %	Max overlap, %	Significan at p<.05 (p	t networks 0 _{FDR} <.05), %
			PAT	HC			PAT	HC
HC _{test}	19.7	18.96			58.36	57.25		
ADHD	16.34	16.34	0	0	67.97	66.67	0	0
ASD	31.68	31.68	100 (100)	0	70.79	69.31	95 (95)	0
BP	19.74	19.74	0	0	58.77	58.33	0	0
MDD	19.25	16.15	0	0	50.93	50.93	0	10 (0)
OCD	10.18	10.18	0	80 (80)	62.87	61.68	0	0
SCZ	35.77	35.25	100 (100)	0	63.19	63.19	0	0



Supplementary Figures

Figure S1. Performance metrics from the normative models. These metrics measure the accuracy with which our normative model estimated the relationship between GMV and age, sex, and site. The distributions of i) explained variance (EV; higher is better), ii) Standardized mean squared error (SMSE; lower is better), iii) Mean standardized log-loss (MSLL; lower is better), iv) Skew (close to zero is better), v) Kurtosis (close to zero is better) across WMV voxels. The left panel presents data for the HC_{test} cohort, the right panel presents data for the cases.

a Overlap maps



Figure S2. Distribution of negative extreme WMV deviation overlap in HC_{test} cohort across scales.



Figure S3. Connections affected by extreme negative WMV deviations, using a weight threshold of 75%. (a) Matrices showing the tract-scale dysconnection for each clinical group and the held-out control group (HC_{test}). (b) Histograms showing the distribution of overlap percentages observed across all interregional connections. (c) Matrices showing edges structurally connected to extreme negative WMV deviations (Z < - 2.6, cluster threshold=10) with significantly greater overlap in cases, compared to



controls (top), and significantly greater overlap in controls, compared to cases (bottom, p < .05, two-tailed).

Figure S4. Functional networks affected by extreme negative WMV deviations using 20 network parcellation. (a) Network maps quantifying the proportion of individuals showing a significant deviation in each network, for each diagnostic group. (b) The network-scale difference in percent overlap for extreme negative WMV deviations (z < - 2.6, cluster threshold=10) between each clinical group and the control cohort. ** corresponds to p_{FDR} < .05, two-tailed, * corresponds to p < .05, two-tailed. The solid black line indicates $-\log_{10} p = 1.6$ (p=.05, two-tailed, uncorrected).



Figure S5. Distribution of positive extreme WMV deviation overlap in HC_{test} cohort across scales



Figure S6. Spatial overlap in extreme positive WMV deviations. (a) Spatial map which quantifies the proportion of individuals showing significant an extreme deviation in a given voxel, yielding an extreme deviation overlap map. (b) Statistical plots showing voxels with significantly greater overlap in cases, compared to controls (left) and significantly greater overlap in controls, compared to cases in extreme positive deviations (top), and greater overlap in controls compared to cases (bottom, p < .05, two-tailed).



Figure S7. Connections affected by extreme positive WMV deviations. (a) Matrices showing the tractscale dysconnection for each clinical group. (b) Histograms showing the distribution of overlap percentages observed across all inter-regional connections. Note that the y-axis is broken so can see the distribution of non-zero values. (c) Matrices showing edges structurally connected to extreme positive WMV deviations (Z < 2.6, cluster threshold=10) with significantly greater overlap in cases, compared to controls (top), and significantly greater overlap in controls, compared to cases (bottom, p < .05, twotailed).



Figure S8. Regions attached to connections affected by extreme positive WMV deviations. (a) Spatial maps quantifying the proportion of individuals showing significant structural connectivity in each region, yielding a region-scale dysconnection map for each diagnostic group. (b) Histograms showing the distribution of overlap percentages observed across all regions. (c) Statistical maps showing regions structurally connected to extreme positive WMV deviations (z < -2.6, cluster threshold=10) with significantly greater overlap in cases, compared to controls (top), and significantly greater overlap in controls compared to cases (bottom, p < .05, two-tailed).



Figure S9. Functional network affected by extreme positive WMV deviations. (a) Network maps which quantify the proportion of individuals showing a significant deviation in each network, yielding a network-scale dysconnection map for each diagnostic group. (b) The network-scale difference in percent overlap for extreme positive WMV deviations (Z > 2.6, cluster threshold=10) between each clinical group and the control cohort. ** corresponds to $p_{FDR} < .05$, two-tailed, * corresponds to p < .05, two-tailed

References

- Segal A, Parkes L, Aquino K, Kia SM, Wolfers T, Franke B, et al. (2023): Regional, circuit and network heterogeneity of brain abnormalities in psychiatric disorders. Nat Neurosci 1– 17.
- 2. Gaser C, Dahnke R, Thompson PM, Kurth F, Luders E, Initiative ADN (2022, June 13): CAT A Computational Anatomy Toolbox for the Analysis of Structural MRI Data. bioRxiv, p 2022.06.11.495736.
- 3. Marquand AF, Rezek I, Buitelaar J, Beckmann CF (2016): Understanding Heterogeneity in Clinical Cohorts Using Normative Models: Beyond Case-Control Studies. *Biological Psychiatry* 80: 552–561.
- 4. Marquand AF, Kia SM, Zabihi M, Wolfers T, Buitelaar JK, Beckmann CF (2019): Conceptualizing mental disorders as deviations from normative functioning. *Mol Psychiatry* 24: 1415–1424.
- 5. Rutherford S, Kia SM, Wolfers T, Fraza C, Zabihi M, Dinga R, *et al.* (2022): The normative modeling framework for computational psychiatry [no. 7]. *Nat Protoc* 17: 1711–1734.
- 6. Fraza CJ, Dinga R, Beckmann CF, Marquand AF (2021): Warped Bayesian linear regression for normative modelling of big data. *NeuroImage* 245: 118715.
- 7. Rutherford S, Fraza C, Dinga R, Kia SM, Wolfers T, Zabihi M, *et al.* (2022): Charting brain growth and aging at high spatial precision ((C. I. Baker, B. Taschler, O. Esteban, & T. Constable, editors)). *eLife* 11: e72904.
- 8. Wolfers T, Doan NT, Kaufmann T, Alnæs D, Moberget T, Agartz I, *et al.* (2018): Mapping the Heterogeneous Phenotype of Schizophrenia and Bipolar Disorder Using Normative Models. *JAMA Psychiatry* 75: 1146–1155.
- Benjamini Y, Hochberg Y (1995): Controlling the False Discovery Rate : A Practical and Powerful Approach to Multiple Testing Yoav Benjamini ; Yosef Hochberg Journal of the Royal Statistical Society . Series B (Methodological), Vol. 57, No. 1. (1995), pp. Royal Statistical Society 57: 289–300.
- 10. Winkler AM, Ridgway GR, Douaud G, Nichols TE, Smith SM (2016): Faster permutation inference in brain imaging. *NeuroImage* 141: 502–516.
- 11. Winkler AM, Ridgway GR, Webster MA, Smith SM, Nichols TE (2014): Permutation inference for the general linear model. *NeuroImage* 92: 381–397.
- 12. Boes AD, Prasad S, Liu H, Liu Q, Pascual-Leone A, Caviness VS, Fox MD (2015): Network localization of neurological symptoms from focal brain lesions. *Brain* 138: 3061–3075.
- 13. Fox MD (2018): Mapping Symptoms to Brain Networks with the Human Connectome. *New England Journal of Medicine* 379: 2237–2245.
- 14. Kuceyeski A, Maruta J, Relkin N, Raj A (2013): The Network Modification (NeMo) Tool: Elucidating the Effect of White Matter Integrity Changes on Cortical and Subcortical Structural Connectivity. *Brain Connectivity* 3: 451–463.
- 15. Kuceyeski A, Boes A (2022): Lesion-Network Mapping: From a Topologic to Hodologic Approach. In: Pustina D, Mirman D, editors. *Lesion-to-Symptom Mapping: Principles and Tools*. New York, NY: Springer US, pp 149–166.
- 16. Sperber C, Griffis J, Kasties V (2022): Indirect structural disconnection-symptom mapping. *Brain Struct Funct* 227: 3129–3144.
- 17. Glasser MF, Sotiropoulos SN, Wilson JA, Coalson TS, Fischl B, Andersson JL, *et al.* (2013): The minimal preprocessing pipelines for the Human Connectome Project. *NeuroImage* 80: 105–124.
- 18. Fischl B (2012): FreeSurfer. NeuroImage 62: 774–781.
- 19. Arnatkeviciute A, Fulcher BD, Oldham S, Tiego J, Paquola C, Gerring Z, *et al.* (2021): Genetic influences on hub connectivity of the human connectome. *Nature Communications* 12. https://doi.org/10.1038/s41467-021-24306-2

- 20. Jeurissen B, Tournier J-D, Dhollander T, Connelly A, Sijbers J (2014): Multi-tissue constrained spherical deconvolution for improved analysis of multi-shell diffusion MRI data. *NeuroImage* 103: 411–426.
- 21. Tournier J-D, Calamante F, Connelly A (2007): Robust determination of the fibre orientation distribution in diffusion MRI: Non-negativity constrained super-resolved spherical deconvolution. *NeuroImage* 35: 1459–1472.
- 22. Tournier J-D, Calamante F, Connelly A (2010): Improved probabilistic streamlines tractography by 2nd order integration over fibre orientation distributions, vol. 1670. 1670.
- 23. Tournier J-D, Calamante F, Connelly A (2012): MRtrix: Diffusion tractography in crossing fiber regions. *Int J Imaging Syst Technol* 22: 53–66.
- 24. Tournier J-D, Smith R, Raffelt D, Tabbara R, Dhollander T, Pietsch M, *et al.* (2019): MRtrix3: A fast, flexible and open software framework for medical image processing and visualisation. *NeuroImage* 202: 116137.
- 25. Smith RE, Tournier J-D, Calamante F, Connelly A (2012): Anatomically-constrained tractography: Improved diffusion MRI streamlines tractography through effective use of anatomical information. *NeuroImage* 62: 1924–1938.
- 26. Smith RE, Tournier J-D, Calamante F, Connelly A (2015): SIFT2: Enabling dense quantitative assessment of brain white matter connectivity using streamlines tractography. *NeuroImage* 119: 338–351.
- 27. Schaefer A, Kong R, Gordon EM, Laumann TO, Zuo X-N, Holmes AJ, *et al.* (2018): Local-Global Parcellation of the Human Cerebral Cortex from Intrinsic Functional Connectivity MRI. *Cerebral Cortex* 28: 3095–3114.
- 28. Tian Y, Margulies DS, Breakspear M, Zalesky A (2020): Topographic organization of the human subcortex unveiled with functional connectivity gradients. *Nature Neuroscience* 23: 1421–1432.
- 29. Jenkinson M, Beckmann CF, Behrens TEJ, Woolrich MW, Smith SM (2012): FSL. *NeuroImage* 62: 782–790.
- 30. Smith RE, Tournier J-D, Calamante F, Connelly A (2015): The effects of SIFT on the reproducibility and biological accuracy of the structural connectome. *NeuroImage* 104: 253–265.
- 31. Jenkinson M, Bannister P, Brady M, Smith S (2002): Improved Optimization for the Robust and Accurate Linear Registration and Motion Correction of Brain Images. *NeuroImage* 17: 825–841.