

Supplementary Materials– Tables 1-7

Table S1. shRNA sequences and CRISPR dCas9 gRNAs

Description	Location (bp upstream of the gene)	Sequence
gRNA1 (617 in mass spec)	<i>Lhb</i> (-3676)-(-3657)	ATCCAGAGCTTCCTGCGGGC
gRNA3 (618 in mass spec)	<i>Lhb</i> (-3800)-(-3781)	CTCCCTAATGCCGGCATTAA
gRNA4 (p594)	<i>Lhb</i> (-3565)-(-3546)	TTAAAACCTGGACCGTTGCTG
shRNA for eRNAs		
#1 (p591)	<i>Lhb</i> (-3754)-(-3736)	AGAAGATAAATCAAGGCG
#2 (p549)	<i>Lhb</i> (-4872)-(-4854)	TTCCCAAGCCTGGACCTCA
#3 (p544)	<i>Lhb</i> (2632)-(-2614)	CCAGGGGCAACGGTCACAA
#4 (p548)	<i>Lhb</i> (2632)-(-2614)	TTGTGACCGTTGCCCTGG
shRNA for Hmgb2	<i>Hmgb2</i> (+817)-(+835) exon2	CCAAGAAATGCTCCGAGAG

Table S2. U1 site mutations

LncRNA site	Sequence	Mutated sequence
End of exon 1	CAG GTGAGA	CAG CCGAGA
End of exon 2	AAG GTAGGG	AAG CCAGGG
End of exon 3	CAG GTAGAG	CAG CCAGAG
End of exon 4	AAG GTGAGC	AAG CCGAGC
Mid intron 1 (-2330)	AAG GTGGAG	AAG CCGGAG
Mid intron 3 (-1473)	CAG GTAGGA	CAG CCAGGA

Table S3. Primers

Primer #	Gene (<i>Lhb</i> unless noted) and position	Sequence (5' to 3')
Primers used for CHIP (many used also for PCR)		
1490	-5228F	GTGAGCTGGCAGTATGCCGA
1491	-5086R	CGTCTCGAAGAAGTACTGAC
596	-5005F	GCTCTCAGAATGCAAGGCTA
614	-4796R	GGTCCTTTCAGCATCCAGCT
595	-4435F	CTCCATCCTGAACATCATAAAC
1518	-4326R	CAGACAAGGCTGGCTTCAAC
594	-4275F	TATGTACTGTAATCCTAATGCTTA
613	-4076R	CAGGTGGACTTGTCTGGTG
1583	-3775F	TACATCACCCTGTGTGGCT
1584	-3705F	TGTATTATGCAGTCAGTGCTG
1535	-3662R	CAGGAAGCTCTGGATTCCAT
1536	-3638R	TTCAGATGAGGGAGCTCCTG
1619	-3610F	CCTTCCAATGCTTTGGGAC
1527	-3567R	CCAGCCTGAGCTTGGTGAG
1620	-3533R	TTAAGGTCATCCTCAGCAAC
546	-3485F	TTGAGTGTTTTGCCTGCATG

1521	-3458F	GGGTCTTCATGCATGGGTGT
612	-3286R	TGATCTACTCTACTCGCACAC
450	-3257F	CCTCCTTGGTGTGGAGAAA
451	-3102R	GAGAGTGGGAGGTGGCTAGA
1384	-2802F	TGTATTCCCTCACCTCACCT
1385	-2633R	TAACAGAACCTGCCACTCT
572	-2432F	TCTGCCTACTCTCCAGGTGA
548	-2255F	CTGATCTATGCGCCTCACA
112	-2196R	TTTCGAGACTGCAGGCAGGG
562	-2096R	TGGGGTCCTTGTGCTCTGGT
1943	-635F	GGCCTACTGCACTGAGAATT
907	-520F	GACACACCCCTTACTTCCAGAG
1944	-474R	GATCAGTAAATTGCGCCTGG
909	-424F	ATCCTGATTAGGGGCTGGGAG
908	-421R	GGATGCCTGGATTGGGTCCAA
910	-325R	GGCACAGCTTCAACCTTAGGTT
913	-235F	CTCCCCTTACCTTGTTTCCCGT
1937	-225F	CCTTGTTTCCCGTGCTTCCA
915	-141F	TCCCTGGCTTCCCTGACCTTG
914	-136R	CAGGGACTAGCTCCAGTGTC
1938	-79R	TGGGTAACCTAGACACTAATC
1935	-68F	CACTACTTAGTGGCCTTGCC
917	-44F	CCACAACCCGCAGGTATAA
916	-42R	TGGGGGTGGCAAGGCCACTA
919	+51F	AATCCCACCAAACCCATAGATG
918	+54R	GATTGGGAAGTACCCTGGG
1936	+61R	TGGTGGGATTGGGAAGTACC
1939	+62F	AACCCATAGATGGACAGCCT
955	+106F	AGAGGAGGGGGTCTCCTA
920	+150R	GTTTATCTCCTGTTTGAAGTCCAC
1940	+250R	TCACTCTACCCCTGACCTG
966	+271R	CCCTGGGACATAGCCAGTG
286	<i>Atoh</i> F	CCCTCACTCAGGTCGCCTG
287	<i>Atoh</i> R	CGTGCGAGGAGCCAATCA
382	mCga -5139R	GCGTTCACAATAACTTGTAACC
440	mCga -5377F	TTGGGAACCTTTGTTTTGACACTTT
3093	G4 positiveF Cga intron1	TGGCAATAGGGGAATTTCCAG
3094	G4 positiveR Cga intron1	TAACCGGATCTGTTTGAATC
3124	G4 negative	CTGAAGTTCATTATGCAATGTTG
3123	G4 negative	CCCCTATATCAGTTGTTATATTC
1959	mZnf2F intron	AGATTGGATGCTCCTGTAGG

1960	mZnr2R intron	CCTGAGAACAAGCCACAGC
3C Primers		
598	-6005F	CCAGCAGCCAAGTCAGTCTC
597	-5105F	GTCAGTACTTCTTCGAGACG
596	-5005F	GCTCTCAGAATGCAAGGCTA
595	-4435F	CTCCATCCTGAACATCATAAAC
594	-4275F	TATGTACTGTAATCCTAATGCTTA
546	-3485F	TTGAGTGTTCCTGCATG
111	-2513F	GGTACTTGGAACCTGGCCCA
593	-2375F	ACTTGTGTGCCAGGAGGAAA
59	-1991F	GAATCTTTGGGCTCCCTC
549	-1355F	CCACTAAGTGTTGGATAGG
545	+379F	TCTGCCAGTCTGCATCACC
44	+344F	GCCTGTCAACGCAACTCTGG
UMI-4C primers		
<i>Lhb</i> downstream (nested)		GTCCAAAATAGGTGTGGTTTCAGC
<i>Lhb</i> upstream (outer)		CCCTAATCAGGATGCCTGGA
Additional Primers for eRNA and lncRNA characterization		
1387	-4856R	CCCAAGCCTGGACCTCAG
563	-2825F	AGCCTGTCTGTCTGTGTCT
564	-2606R	CCGGCTTGTGTGACCGTTG
1424	-2655F	TTCAGAGTGGGCAGGTTCTG
2276	-2575F	ATGGCAGGTGCAACCCTAGC
2286	-2152R	GGGGTGTATGTAGACTGTGA
2278	-2232F	TCTATGGTCAAGGCACCCCT
2279	-2061R	CTTTCTGCCAAAGACTTGATG
2280	-1966F	CTAAGAAGACATGAGGGACG
2281	-1803R	CTGAGAAGGTCCCCGCGA
60	-1743R	CAGGCTCCTCCTGCAGTGC
2282	-1235F	GCCCTCATCCCCTGGACGA
2283	-1117R	CTTGTGAAGTCACTTGCAGTA
55	-952F	ATCTGAACTCTGCTGCACTT
2284	-903F	GAACGTGTCTGGATATCCCC
2285	-582R	TCACAGCAGCTCCGCCCTG
367	-394R	GCACCACCGTCTCCAGCCC
181	-151R	GTGTCCCAGTGAATTGGCCTCA
2311	-863R	GCCCATCACCGTTTCTGGT
2312	-2593F	CAGAGAAGGTGAGGACCATG
2313	-2611F	AGCCGTTTCTTGGCTTCC
2314	-1106R	CACAGGCTCACCTTGTGAA
2315	-1088R	TCACCAGCGCTGCTGGG
2316	-880R	GGTGGGGATATCCAGACAC
2324	-814R	CTCGGTAACTTGCCCGATC

2326	-729R	GTGCACCAAGGGTCCGTAG
2321	-643R	ACACTGTAAAGGCCAGCAC
612	-3286R	TGATCTACTCTACTCGCACAC
2300	-3685R	CAGCACTGACTGCATAATACA
2317	+691R	CCCGGTGGGCAGCCAGG
493	+572R	AAGCCCTCTTCTGAGTGTC
Gene expression levels PCR		
46	<i>Lhb</i> (+601)F	TGCCGGCTGCTTTGCCTCCT
45	<i>Lhb</i> (+796)R	CAGGCCATTGGTTGAGTCCT
350	<i>Lhb</i> (2 nd +3 rd) exon (+590)F	CTAGCATGGTCCGAGTACTG
351	<i>Lhb</i> 3 rd exon (+727)R	CTGAGGGCTACAGGAAAGGA
184	<i>Rplp0</i> F	GCGACCTGGAAGTCCAACCTA
185	<i>Rplp0</i> R	ATCTGCTTGGAGCCCACAT
1384	eRNA1 F <i>Lhb</i> (-2802)	TGTATTCCCTCACCTCACCT
1385	eRNA1 R <i>Lhb</i> (-2633)	TAACAGAACCTGCCACTCT
1583	eRNA1 F <i>Lhb</i> (-3775)	TACATCACCCTGTGTGGCT
1534	eRNA1 R <i>Lhb</i> (-3682)	CCCCAGCACTGACTGCATAA
1961	<i>Znrf2</i> F cDNA	ACAGGACTCAGTGCACAGC
1962	<i>Znrf2</i> R cDNA	TGAGCATACCGGGCACTTA
1923	<i>Hmgb2</i> F	CGACTCGTGGTGAACCTTCG
1924	<i>Hmgb2</i> R	CTTCGGAGCATTGGGGTCTT
1672	<i>Greb1</i> 3 rd exon F	GCCGAGCAGACAATGAGGAA
1673	<i>Greb1</i> 4 th exon R	CAGGCTGGGAGACTTAGCAC

Table S4 Oligonucleotides used for CD

Description	Sequence
<i>Lhb</i> G-quadruplex	GGAAAGGTGGGGGTGGGGGTGGGGTGGGGTGGGG
<i>Lhb</i> G4 kill4	GGAAAGGTGGTGGTGGTGGTGGGGTGGAGTGGTG
<i>Lhb</i> G4 kill5	GGAAAGGTGGTGGTGGTGGTGGTGTGGTGTGGTG
<i>Lhb</i> G4 kill7	GGAAAGGTGGAGGTGGAGGTGGTGTGTGATGAGT
<i>Lhb</i> G-quadruplex mutated	GGAAAGGTGTTTGTGTTTGTGTTGTGTTGTGTTG
<i>c-MYC</i> G-quadruplex (human)	GGGGAGGGTGGGGAGGGTGGGGAAGGTGGGG
<i>Lhb</i> i-motif	CCCCACCCACCCACCCACCCACCCCTTTCC
<i>Lhb</i> i-M kill4	CTCCACACCACCCACCTCCACCTCCACCTTTCC
<i>Lhb</i> i-M kill7	TCACAACCTCACTCCACCACCACCACCTTTCC
<i>Lhb</i> i-motif mutated	CTTCACTTCACTTCACTTTCACCTTTCC
<i>c-MYC</i> i-motif (human)	CCCCACCTTCCCACCTCCCACCTCCCC

Table S5 Constructs for protein pull down

#	Description
507	Control construct, dCas9 without any gRNA
617	dCAS9 with gRNA1 (Table S3) targeting (-3676) -(3658) upstream of <i>Lhb</i>
618	dCAS9 with gRNA3 (Table S3) targeting (-3800) -(3781) upstream of <i>Lhb</i>

Table S6. Sequences upstream of the *Lhb* gene predicted to form G-quadruplex (by both pqsfinder and G4 Hunter). All are shown 5' to 3'.

Description	Strand of G4	Putative G4 forming sequence	Pqsfinder score	G4 Hunter score
<i>Lhb</i> enhancer	(-)	GGGGGTGGGGGTGGGGTGGGGTGGGG	115	3.385
LncRNA promoter	(-)	GAGTGGGAGGTGGCTAGAGGGTGGG	52	1.48
	(-)	AACGGGGGAGGGGAGGGGGTATGGG	94	2.56
	(-)	TGGGTATGGGGGGCTGGGGAGACTG	70	1.96
LncRNA 1 st intron	(+)	GGGTGGAGGGTGAGATAGGAAAGGGGG	57	1.76
	(+)	GGGGTCTGGGAGCCTGGATACCAGGAGTTGGAGGAAG	53	1.4
LncRNA 3 rd Exon	(-)	GGTGGTCAGCGAGCTCTGGGGGCTGGGGCTCTGCAA GAGGGGGGCACTC	81	1.84
<i>Lhb</i> 1 st intron	(-)	ATGGGTTTGGTGGGATTGGGAAGTACCCTGGG	52	1.28
	(+)	GGGGGTTGGAGGAGGGAGAGGGGGG	103	2.16

Table S7. Proteins detected (≥ 3 peptides) following pull-down with both probes (617 and 618) and not with the control (507)

Gene names	Σ Coverage	Σ # Unique Peptides	Σ # Peptides	Σ # PSMs	507	617	618
					A2: Area	B2: Area	C2: Area
Hmgb2	18.1	4	4	9	0.00E+00	2.59E+07	3.36E+06
Afdn	2.88	4	4	5	0.00E+00	4.53E+06	2.12E+06
Gss	6.33	3	3	4	0.00E+00	1.70E+07	2.11E+06
Trove2	5.76	3	3	5	0.00E+00	1.29E+07	2.73E+06
Stoml2	11.05	3	3	6	0.00E+00	3.50E+06	4.18E+06
Tecr	8.53	3	3	4	0.00E+00	3.90E+07	6.76E+06
Camk2d	25	3	3	5	0.00E+00	6.22E+06	1.95E+06

The peptides were filtered with high confidence and passed the 1% FDR threshold. Semi quantitation was performed by calculating the peak area of each peptide: the area is the average of the three most intense peptides from each protein.