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Supplemental information

De novo missense variants in HDAC3 leading

to epigenetic machinery dysfunction are associated

with a variable neurodevelopmental disorder

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HDAC3 (NM_003883.4): c.328G>A, p.Ala110Thr

Figure S1. Confirmation of *de novo HDAC3* **variants using DNA Sanger sequencing.**

Sanger sequencing was conducted to validate the *de novo* variants using the primers (Table S2) for individuals 1, 2 and their parents.

Figure S2. Characterization of NCoR1 DAD domain-HDAC3 interaction and HDAC3 mutagenesis.

(A) Sequence alignment of NCoR1 and NCoR2 DAD (Deacetylase Activating Domain) domains. This illustrates a 75.8% sequence similarity and a 91.3% similarity at the HDAC3 binding site. Alignment was conducted using the UniProt Align tool. (B) Mutagenesis of HDAC3. Six specific HDAC3 variants were introduced by PCR, utilizing mutagenic primers listed in Table S2. Sanger sequencing confirmed the presence of p.Ala110Thr, p.Gly267Ser, p.Leu266Ser, p.Arg359Cys, p.Asp93Asn, and p.Pro201Ser variants, indicating successful site-directed mutagenesis. (C) Purification of the NCoR1 DAD domain in complex with HDAC3 WT or variant proteins. These proteins, along with the NCoR1 DAD domain, were co-accumulated in Sf9 cells using a baculovirus system. Complexes were purified using Ni-NTA Agarose and Anti-FLAG M2 Affinity Gel 64hrs post-infection, as visualized on the SDS-PAGE gel, indicating no defects in their interaction with the NCoR1 DAD domain.

Figure S3. Histone deacetylase activity of HDAC3-NCoR1 DAD domain complex on acetylated mononucleosomes.

(A) HDAC assay on H3K27ac mononucleosomes. This assay investigated the deacetylase activity of the NCoR1 DAD domain in complex with wild-type (WT) HDAC3 and its variants (p.Ala110Thr, p.Gly267Ser, p.Leu266Ser, and p.Arg359Cys). Mononucleosomes acetylated at lysine 27 on histone H3 (H3K27ac) served as substrates. Immunoblotting with an H3K27ac-specific antibody quantified the deacetylation, revealing decreased activities in the p.Gly267Ser and p.Leu266Ser variants as denoted by the red boxes. (B) HDAC assay on H3ac/H4ac mononucleosomes. This panel assesses the deacetylase activities of the NCoR1 DAD domain-HDAC3 complex, both WT and variant forms, on mononucleosomes with acetylations at various lysine residues on histones H3 (H3K4,9,14,18ac) and H4 (H4K5,8,12,16ac). Antibodies specific to acetylated H3 (H3ac) and H4 (H4ac) detected the HDAC activities. The p.Gly267Ser and p.Leu266Ser variants exhibit notably reduced deacetylase activity, as highlighted in the red boxes across all replicates.

Figure S4. Defective HDAC activity and decreased interactions with NCoR1/2 and KDM1A in *HDAC3* **p.Asp93Asn and p.Pro201Ser variants.**

(A) HDAC assays were conducted using H3K27ac-mononucleosomes or H3/4Kac-mononucleosomes as substrates at various concentrations (0, 30, 60, 120 nM), in conjunction with 100 nM of acetylated mononucleosomes. The deacetylation activities of complexes comprising either the NCoR1 DAD domain-HDAC3 WT or variant forms were measured for histone H3K27ac. H3ac, and H4ac in triplicate ($n=3$ /data point). The results are plotted as mean \pm SD. The p.Asp93Asn and p.Pro201Ser variants do not result in histone acetylation levels comparable to the WT, indicating defective HDAC activity. (B) Western blot images for HDAC activity assessment using the HDAC3-NCoR1 DAD domain complex are provided. The analysis was performed in triplicate, with both short and long exposures displayed. (C) Silver-stained SDS-PAGE analysis demonstrating the protein complexes co-immunoprecipitated (IP) with FLAG-tagged HDAC3 from HEK293T cells. Tested conditions include an empty vector (EV), wild-type (WT), and HDAC3 p.Asp93Asn and p.Pro201Ser variants. The band intensities corresponding to the NCoR complex (red arrow) and KDM1A (blue arrow) are reduced in the variant forms. Specifically, the p.Pro201Ser variant shows a remarkable decrease in the NCoR complex band intensity (red rectangle), and all variants exhibit diminished KDM1A bands (second band in blue rectangle). (D) Western blot analyses confirm the differential co-immunoprecipitation of NCoR1, NCoR2, and KDM1A with HDAC3 variants, using an anti-FLAG antibody for immunoprecipitation. FLAG-tagged HDAC3 and GAPDH serve as a reference for protein expression and loading control, respectively. (E) Quantification of coimmunoprecipitated NCoR1, NCoR2, and KDM1A, normalized to WT HDAC3 levels, based on the Western blot data in Panel B. **P* < 0.05, ****P* < 0.001, *****P* < 0.0001. Data are plotted as mean \pm SD ($n=3$ /data point).

Figure S5. Differential protein interaction profiles of HDAC3 variants in HEK293T cells.

(A) Venn diagram analysis. This panel presents a Venn diagram that quantifies the number of proteins exhibiting significantly altered interactions (*P* < 0.05) with each of four *HDAC3* variants (p.Ala110Thr, p.Leu266Ser, p.Gly267Ser, and p.Arg359Cys) in comparison to the wild-type (WT) HDAC3 in HEK293T cells. (B) Consistent interaction changes across the *HDAC3* variants. This table enumerates specific proteins that display consistent changes in interaction levels across all *HDAC3* variants tested. The proteins KDM1A, GANAB, AMER1, AKAP8L, and EIF3B show uniformly reduced interactions across the variants, indicating a possible common pathway or functional disruption. In contrast, HNRNPR, which is associated with proteasome degradation pathways, exhibits an increased interaction. (C). Pathway analysis for down-regulated GO terms. Bar chart of Gene Ontology (GO) terms enriched among proteins with decreased interaction with at least two variants, indicating decreased histone deacetylation processes (GO:0016575). (D) Network of downregulated GO terms. A network diagram of the down-regulated GO terms connected with biological processes, revealing potential pathways impacted by the altered protein interactions. (E) Pathway analysis for up-regulated GO terms among proteins with increased interaction with at least two variants. The increased interaction with ATP-dependent protein folding chaperone (GO:0016575) molecules indicated increased protein degradation. (F) Network of up-regulated GO terms. A network diagram displaying the interconnected GO terms and molecular functions that are up-regulated, providing insight into the potential compensatory mechanisms or effects of the variant interactions.

 \mathbf{A}

B

C

Phenotypes present in multiple matching patients

Figure S6. Copy-number variations (CNVs) involving *HDAC3* **from the DECIPHER database.**

(A) CNV deletions (loss) and duplications (gain) encompassing *HDAC3*. Intellectual disability and seizure were the most prevalent phenotypes reported among the eleven patients (see Table S6). (B) Pie chart depicting the inheritance patterns of *HDAC3* CNVs, predominantly occurring *de novo*. (C) The size distribution of the CNVs. (D) The overlapping regions (chr5:141518803-142023518, 504Kb) encompassing *HDAC3* across 11 patients, as cataloged in the DECIPHER database. This region included ten genes, with *DIAPH1* and *PCDH12* identified as morbid OMIM genes (indicated in dark green). *DIAPH1* is associated with 'Deafness, autosomal dominant 1, with or without thrombocytopenia, autosomal dominant (MIM: 124900)', and 'Seizures, cortical blindness, microcephaly syndrome, autosomal recessive (MIM: 616632)', while *PCDH12* is linked to 'Diencephalic-mesencephalic junction dysplasia syndrome 1, autosomal recessive (MIM: 251280)'. The probability of haploinsufficiency (pHaplo) and triplosensitivity (pTriplo) scores for *HDAC3* are 0.62 and 1.00, respectively, ranking as the third and the highest among the genes within this region. This finding highlights *HDAC3*'s significant role in these CNVs, while also acknowledging the potential impact of gene dosage for the other genes present.

Table S1. Overview of *HDAC3* **variants and** *in silico* **predictions.**

Abbreviations: SNUH, Seoul National University Hospital; DDD, Deciphering Developmental Delay; BCH, Boston Children's Hospital; CADD, Combined Annotation Dependent Depletion¹; REVEL, Rare Exome Variant Ensemble Learner²; DANN, Deleterious Annotation of genetic variants using Neural Networks³; GERP, Genomic Evolutionary Rate Profiling⁴; PhyloP, phylogenetic p-values⁵; ACMG/AMP, the American College of Medical Genetics and Genomics and the Association for Molecular Pathology; NLS, nuclear localization signal; IP4, inositol phosphate 4.

Table S2. Primer information.

aMutated nucleotide sites are highlighted in red for mutagenic primers.

Table S3. List of antibodies used in this study.

Table S4. Quantified protein levels measured as iBAQ intensities.

Excel spreadsheets are provided for this table.

Table S5. Functional assessment results of *HDAC3* **variants.**

aPredicted impairment based on structural proximity to critical domains, and previous functional experiments.^{6,7} bsee Figures 2, S4A for the relevant findings. ^csee Figures 3, 4, S4B, S4C, S4D for the relevant findings. dsee Figure 5 for the relevant findings.

Abbreviations: HDAC, histone deacetylase; NCoR, nuclear receptor co-repressor; IP4, inositol phosphate 4; CoREST, co-repressor of repressor element 1 silencing transcription factor; NLS, nuclear localization signal;

Table S6. The complete list of patients involving *HDAC3* **in the DECIPHER database.**

Supplemental References

- 1. Rentzsch P, Witten D, Cooper GM, Shendure J, Kircher M. CADD: predicting the deleteriousness of variants throughout the human genome. *Nucleic Acids Res*. 2019;47(D1):D886-D894. doi:10.1093/nar/gky1016
- 2. Ioannidis NM, Rothstein JH, Pejaver V, et al. REVEL: An Ensemble Method for Predicting the Pathogenicity of Rare Missense Variants. *Am J Hum Genet*. 2016;99(4):877-885. doi:10.1016/j.ajhg.2016.08.016
- 3. Quang D, Chen Y, Xie X. DANN: a deep learning approach for annotating the pathogenicity of genetic variants. *Bioinformatics*. 2015;31(5):761-763. doi:10.1093/bioinformatics/btu703
- 4. Cooper GM, Stone EA, Asimenos G, Green ED, Batzoglou S, Sidow A. Distribution and intensity of constraint in mammalian genomic sequence. *Genome Res*. 2005;15(7):901-913. doi:10.1101/gr.3577405
- 5. Pollard KS, Hubisz MJ, Rosenbloom KR, Siepel A. Detection of nonneutral substitution rates on mammalian phylogenies. *Genome Res*. 2010;20(1):110-121. doi:10.1101/gr.097857.109
- 6. Sun Z, Feng D, Fang B, et al. Deacetylase-Independent Function of HDAC3 in Transcription and Metabolism Requires Nuclear Receptor Corepressor. *Mol Cell*. 2013;52(6):769-782. doi:10.1016/j.molcel.2013.10.022
- 7. Lahm A, Paolini C, Pallaoro M, et al. Unraveling the hidden catalytic activity of vertebrate class IIa histone deacetylases. *Proc Natl Acad Sci USA*. 2007;104(44):17335-17340. doi:10.1073/pnas.0706487104