

Supplemental information

PI3Kgamma promotes neutrophil extracellular trap formation by noncanonical pyroptosis in abdominal aortic aneurysm

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Figure S1. Construction strategies and identification of PI3Kgamma knockout (*PI3K γ -*) mice.

(A) PI3K γ knockout mice were generated by cutting exon2-exon9 of the *Pik3cg* gene using CRISPR/Cas9 technology.

(B) Genotyping was detected by tail preparation of breeding mice.

(C) Representative western blot images showing PI3K γ protein expression in the aorta at different point time after PPE surgery. n=3.

(D) Blood cell counts of WT and PI3K γ mice were measured. n=6.

(E) Serum inflammatory factors of WT and PI3K γ mice were detected. n=6.

WT, wild type; Heter-, heterozygote; Homo-, homozygote. WBC, white blood cell; Lymph, lymphocytes; Mono, monocytes; Gran, granulosa cells; PLT, platelet; IL1B, interleukin-1 β ; IL6, interleukin-6; TNFA, Tumor Necrosis Factor a; CCL2, C-C motif chemokine ligand 2.

Figure S2. PI3K γ knockout reduces arterial wall damage and inflammation.

(A) Representative immunohistochemistry staining images and quantitative comparison of neutrophil infiltration in WT mouse abdominal aorta after PPE-induced AAA. Neutrophil infiltration was assessed by Ly6G+ neutrophil numbers per high-power field (40X). n=5 in control group and n=6 in AAA group. Scale bars, 50 μ m.

(B) Representative VVG staining images and quantitative comparison of WT or *PI3K γ* ^{-/-} mouse abdominal aorta in PPE-induced AAA. n = 6. Scale bars, 50 μ m.

(C) QPCR analysis of contractile, synthetic, inflammation, and matrix metalloproteinase genes expression normalized to the mean expression of housekeeping gene (*Actb*) in mouse abdominal aorta after PPE-induced AAA. n=6.

(A and B) Two-tailed unpaired student's t test. (C) Two-way ANOVA followed by Bonferroni test.

* $P<0.05$, ** $P<0.01$, and *** $P<0.001$. AAA, abdominal aorta aneurysm; WT, wild type; *PI3K γ* ^{-/-}, phosphoinositide 3-kinase γ knockout.

Figure S3: Purity isolated neutrophils from mice marrow stimulated by different concentration of LPS.

- (A) Representative image of the characteristic morphological appearance of isolated neutrophils stained with Wright-Giemsa stain (40X). Scale bars, 50 μ m.
- (B) Representative image of the characteristic morphological appearance of isolated neutrophils stained with DAPI (40X). Scale bars, 50 μ m.
- (C) Representative scatter plot of isolated neutrophils analyzed by flow cytometry. Neutrophils were identified by double positive staining for CD11b and Ly6G.
- (D) Representative western blot images and quantitative comparison of Cit H3 protein expression in isolated neutrophils stimulated with different concentration of LPS. n=3.

(D) One-way ANOVA followed by Bonferroni test.

* $P<0.05$, ** $P<0.01$, and *** $P<0.001$. Cit H3, citrullinated histone 3; LPS, lipopolysaccharide.

Figure S4: Pharmacological inhibition of PI3K γ reduces NETs formation in neutrophils.

(A and C) Representative immunofluorescence staining images and quantitative comparison of NETs produced by isolated neutrophils from WT mice with different treatments: PBS, LPS (5 μ g/mL), LPS + IPI549 (5 μ M); PBS, TNFa (50ng/mL), TNFa + IPI549 (5 μ M). NETs were detected using immunofluorescent staining of DNA (DAPI, blue), Citrullinated histone 3 (Cit H3, Green), and Myeloperoxidase (MPO, red). NETs expression was calculated by NETs cell numbers/total cell numbers per high-power field (40X). n=6. Scale bars, 50 μ m.

(B and D) Representative western blot images and quantitative comparison of Cit H3 protein expression in each group of neutrophils with different treatments as described

in (A) and (C). n=3.

(A-D) One-way ANOVA followed by Fisher's LSD post hoc test.

* $P<0.05$, ** $P<0.01$, and *** $P<0.001$. PBS, phosphate buffered saline; LPS, Lipopolysaccharide; TNFa, tumor necrosis factor alpha; Cit H3, citrullinated histone 3; NETs, neutrophil extracellular traps.

Figure S5: The inhibitory effect of PI3K γ deficiency on NET formation in neutrophils is independent of ROS signaling.

(A) Representative western blot images and quantitative comparison of Cit H3 and NOX2 protein expression in the neutrophils with different treatments: PBS (WT-derived neutrophils), LPS (5 μ g/mL, WT-derived neutrophils), LPS (PI3K γ -/- derived neutrophil), LPS + DPI (10 μ M, WT-derived neutrophils), LPS + DPI (PI3K γ --derived neutrophils). n=3.

(B) Representative western blot images and quantitative comparison of Cit H3 and NOX2 protein expression in isolated neutrophils from WT mice with different treatments: PBS, LPS (5 μ g/mL), LPS + IPI549 (5 μ M), LPS + DPI (10 μ M), LPS + DPI + IPI549. n=3.

(A and B) One-way ANOVA followed by Fisher's LSD post hoc test.

* $P<0.05$, ** $P<0.01$, and *** $P<0.001$. LPS, Lipopolysaccharide; PI3K γ -, phosphoinositide 3-kinase γ knockout; DPI, diphenyleneiodonium chloride; Cit H3, citrullinated histone 3; NOX2, nicotinamide adenine dinucleotide phosphate oxidase 2.

Figure S6: Effects of PI3K γ on the NETs formation of neutrophils via the noncanonical pyroptosis signaling pathways.

(A) Representative western blot image and quantitative analysis of GSDMD protein expression in human AAA tissue and adjacent relatively normal abdominal aortic tissue.

n = 7.

(B) Representative western blot image and quantitative comparison of the expression of IL-1 β and GSDMD in the isolated neutrophils from WT mice with different treatments: PBS, LPS (5 μ g/mL), LPS + IPI549 (5 μ M). n=3.

(C) Representative western blot image and quantitative comparison of the expression of Cit H3, IL-1 β , GSDMD, Caspase11, and Caspase1 in the isolated neutrophils from WT mice with different treatments: PBS, LPS (100ng/mL), LPS + Nigericin (10 μ M), LPS + Nigericin + IPI549 (5 μ M); PBS, Pam3CSK4 (1 μ g/mL), Pam3CSK4 + tLPS (10 μ g/mL, transfer LPS), Pam3CSK4 + tLPS + IPI549 (5 μ M). n=3. Scale bars, 50 μ m.

(A) Two-tailed paired student's t test. (B and C) One-way ANOVA followed by Fisher's LSD post hoc test.

*P<0.05, **P<0.01, and ***P<0.001. Adjacent AA, adjacent abdominal aorta; AAA, abdominal aorta aneurysm; Cit H3, citrullinated histone 3; GSDMD, Gasdermin D; pro-, prosoma; cle-, cleaved; IL-1 β , interleukin 1 beta; Casp1, caspase 1; Casp11, caspase11; PBS, phosphate buffered saline; LPS, Lipopolysaccharide; Nig, nigericin; Pam, Pam3CSK4; tLPS, transfer Lipopolysaccharide.

Figure S7: The inhibitory effect of PI3K γ deficiency on NETs formation in neutrophils

is independent of PI3K/AKT signaling.

(A) Representative western blot images and quantitative comparison of p-AKT, t-AKT and Cit H3 protein expression in the neutrophils with different treatments: PBS (WT-derived neutrophils), LPS (5 μ g/mL, WT-derived neutrophils), LPS (PI3K γ -/- derived neutrophil), LPS + SC79 (4 μ g/mL, PI3K γ -/- derived neutrophils). n=3.

(B) Representative western blot images and quantitative comparison of p-AKT, t-AKT and Cit H3 protein expression in isolated neutrophils from WT mice with different treatments: PBS, LPS (5 μ g/mL), LPS + IPI549 (5 μ M), LPS + IPI549+ SC79 (4 μ g/mL). n=3.

(A and B) One-way ANOVA followed by Fisher's LSD post hoc test.

*P<0.05, **P<0.01, and ***P<0.001. LPS, Lipopolysaccharide; PI3K γ -, phosphoinositide 3-kinase γ knockout; Cit H3, citrullinated histone 3.

Figure S8: PI3K γ regulates neutrophil noncanonical pyroptosis via cAMP/PKA signaling pathways.

(A) Representative western blot image and quantitative comparison of the expression of Cit H3, GSDMD, and Caspase11 in the isolated neutrophils from WT mice with different treatments: PBS, LPS (5 μ g/mL), LPS + IPI549 (5 μ M), LPS + IPI549 + H89 (20 μ M), LPS + IPI549 + MDL12330A (10 μ M). n=3.

(B) Representative immunofluorescence staining images and quantitative comparison of NETs produced by neutrophils with different treatments as described in (A). NETs were detected using immunofluorescent staining of DNA (DAPI, blue), Citrullinated

histone 3 (Cit H3, Green), and Myeloperoxidase (MPO, red). NETs expression was calculated by NETs cell numbers/total cell numbers per high-power field (40X). n=6.

Scale bars, 50 μ m.

(A and B) One-way ANOVA followed by Fisher's LSD post hoc test.

* $P<0.05$, ** $P<0.01$, and *** $P<0.001$. Cit H3, citrullinated histone 3; GSDMD, Gasdermin D; pro-, prosoma; cle-, cleaved; Casp11, caspase11; PBS, phosphate buffered saline; LPS, Lipopolysaccharide; MDL, MDL12330A; NETs, neutrophil extracellular traps.

Table S1: the key reagent and resource used in the present study.

Reagent or Resource	Source	Identifier
Antibodies		
Rabbit anti-Histone H3 (citrulline R2 + R8 + R17)	Abcam	Cat# ab5103
Mouse anti- PI3-kinase p110 gamma	Santa Cruz Biotechnology	Cat# sc-166365
Rabbit anti-GSDMD	Abcam	Cat# ab210070
Rabbit anti-GSDMD	Abcam	Cat# ab219800
Rabbit anti-AKT	Abcam	Cat# ab179463
Rabbit anti-p-AKT	Abcam	Cat# ab192623
Rabbit anti-NOX2	Abclonal	Cat# A12430
Rabbit anti-Ly6G	Abclonal	Cat# A22270
Rabbit anti-IL-1 β	Abcam	Cat# ab254360
Rabbit anti-Caspase11	Abcam	Cat# ab180673
Rabbit anti-Caspase1	Abclonal	Cat# A0964
Goat anti-MPO	R&D	Cat# AF3667-SP
Zombie AquaTM dye	Biolegend	Cat# 423101
TruStain FcX™ PLUS (anti-mouse CD16/32)	Biolegend	Cat# 156603
Antibody		
APC anti-mouse Ly-6G	Biolegend	Cat# 127613
FITC anti-mouse/human CD11b	Biolegend	Cat# 101205
Alexa Fluor 594-conjugated donkey anti-goat IgG	Invitrogen	Cat# A32758
Alexa Fluor 488-conjugated donkey anti-rabbit	Invitrogen	Cat# A32790
IgG		
Rabbit anti- β -Actin	Abclonal	Cat# AC038
Mouse anti-GAPDH	Abcam	Cat# ab181602
Chemicals		
Precision Plus Protein Dual Color Standards	Bio-Rad	Cat# 1610374
Prestained Protein Marker VII	Servicebio	Cat# G2087
100 bp DNA Ladder	Beijing Tsingke Biotech Co.,	Cat# TSJ100-100

	Ltd.	
Sodium citrate-EDTA antigen repair buffer	Beyotime	Cat# P0086
Mouse/Rabbit Enhanced Polymer test system	ZSGB-BIO	Cat# PV9000
DAB Substrate Kit	ZSGB-BIO	Cat# ZLI-9018
LPS-O111:B4	Sigma-Aldrich	Cat# L3024
Porcine pancreatic elastase	Sigma-Aldrich	Cat# E1250
Opti-MEM	Gibco	Cat# 31985070
TNF-alpha	MedChemExpress	Cat# HY-P7058
IPI549	MedChemExpress	Cat# HY-100716
Cl-amidine hydrochloride	MedChemExpress	Cat# HY-100574A
Disulfiram	MedChemExpress	Cat# HY-B0240
Diphenyleneiodonium chloride	MedChemExpress	Cat# HY-100965
Pam3CSK4	MedChemExpress	Cat# HY-1
Nigericin	MedChemExpress	Cat# HY-1
SC79	MedChemExpress	Cat# HY-18749
H89	MedChemExpress	Cat# HY-15979
MDL12330A	MedChemExpress	Cat# HY-1
DOTAP Liposomal Transfection Reagent	Roche	Cat# 11202375001
DAPI Fluoromount-G	Southernbiotech	Cat# 0100-20
10x Poly-L-lysine Solution	Solarbio	Cat# P2100
RNAiso Plus	Takara	Cat# 9109
RIPA Lysate	Beyotime	Cat# P0013B
Protease and phosphatase inhibitor	Beyotime	Cat# P1045
SDS-PAGE protein loading buffer	Beyotime	Cat# P0015L
Immobilon Western HRP substrate Luminol Millipore		Cat# WBKLS0500
Reagent		
Wright-Giemsa Stain Solution	Solarbio	Cat# P1020

Critical commercial assay		
TransDirect® Mouse Genotyping Kit	TransGen Biotech Co., Ltd.	Cat# AD501-01

Verheff's Elastic Stain Kit	Abiowell	Cat# AWI0267b
Hifair® AdvanceFast One-step RT-gDNA Yeasen		Cat# 11151ES60
Digestion SuperMix		
Veterinary fully automatic hematology analyzer	Mindray	Cat# BC2800vet
Mouse IL1B ELISA Kit	Renjie Biotech	Cat# RJ16944
Mouse TNFA ELISA Kit	Renjie Biotech	Cat# RJ17929
Mouse IL6 ELISA Kit	Abclonal	Cat# RK00008
Mouse CCL2 ELISA Kit	Abclonal	Cat# RK00381
Mouse cAMP ELISA Kit	Jianglai biotech	Cat# JL13362
PKA Kinase Activity Assay Kit	Abcam	Cat# ab139435
2X Universal SYBR Green Fast qPCR Mix	Abclonal	Cat# RK21203
Mouse marrow neutrophil isolation kit	Solarbio	Cat# P8550

Experimental model

C57BL/6J mice	SPF Hunan SJA Laboratory	N/A
Animal Co., Ltd.		
PI3K γ -KO/- mice	GemPharmatech Co. Ltd	Cat# T009406

Table S2: the patient clinical information in the present study. (n=7)

Patient	Gender	Age	Smoking	Diameter(mm)	HL	HTN	CAD
ID1	Male	72	no	43	yes	yes	no
ID2	Female	63	no	42	no	yes	no
ID3	Male	65	yes	89	yes	no	no
ID4	Male	64	yes	43	no	no	no
ID5	Male	77	yes	71	yes	yes	no
ID6	Male	58	yes	40	no	yes	yes
ID7	Male	22	no	68	no	yes	no

HL: Hyperlipidemia; HTN: Hypertension; CAD: Coronary artery disease.

Table S3: The primer sequences used in the present study.

Gene name	Primer Sequence (5'-3')
Pik3cg-WT	Forward: ACGCTGAGCCTCTCTCCATC Reverse: CCATAACCCTGGAAACAATGGC
Pik3cg-KO	Forward: GCTGCGCTGCCTTATTAACTTT Reverse: CTGCGAGAGTTAGGGACTCAGGTGT
Tagln	Forward: CCAACAAGGGTCCATCCTACG Reverse: ATCTGGGCGGCCTACATCA
Cnn1	Forward: GACGGGATCATTCATTGCGAA Reverse: CCCCATACTGGTAATGGCTTG
Acta2	Forward: GTCCCAGACATCAGGGAGTAA Reverse: TCGGATACTTCAGCGTCAGGA
Myh11	Forward: ATGAGGGTGGTCGTGGAGTTG Reverse: GCCTGAGAAGTATCGCTCCC
Klf4	Forward: GTGCCCGACTAACCGTTG Reverse: GTCGTTGAACCTCCTCGGTCT
Spp1	Forward: AGCAAGAAACTCTCCAAGCAA Reverse: GTGAGATTCGTCAGATTCATCCG
IL6	Forward: TACCACTTCACAAGTCGGAGGC Reverse: CTGCAAGTGCATCATCGTTGTTTC
Ccl2	Forward: GCTACAAGAGGATCACCAGCAG Reverse: GTCTGGACCCATTCTTCTTGG
TNF alpha	Forward: CAGGCGGTGCCTATGTCTC Reverse: CGATCACCCCGAAGTTTAGTAG
IL-1β	Forward: GAAATGCCACCTTTGACAGTG Reverse: TGGATGCTCTCATCAGGACAG
Mmp2	Forward: ACCTGAACACTTCTATGGCTG Reverse: CTTCCGCATGGTCTCGATG
Mmp9	Forward: GCAGAGGCATACTGTACCG Reverse: TGATGTTATGATGGTCCCACTTG

Actb

Forward: GTGCTATGTTGCTCTAGACTTCG

Reverse: ATGCCACAGGATTCCATACC

Figure S1

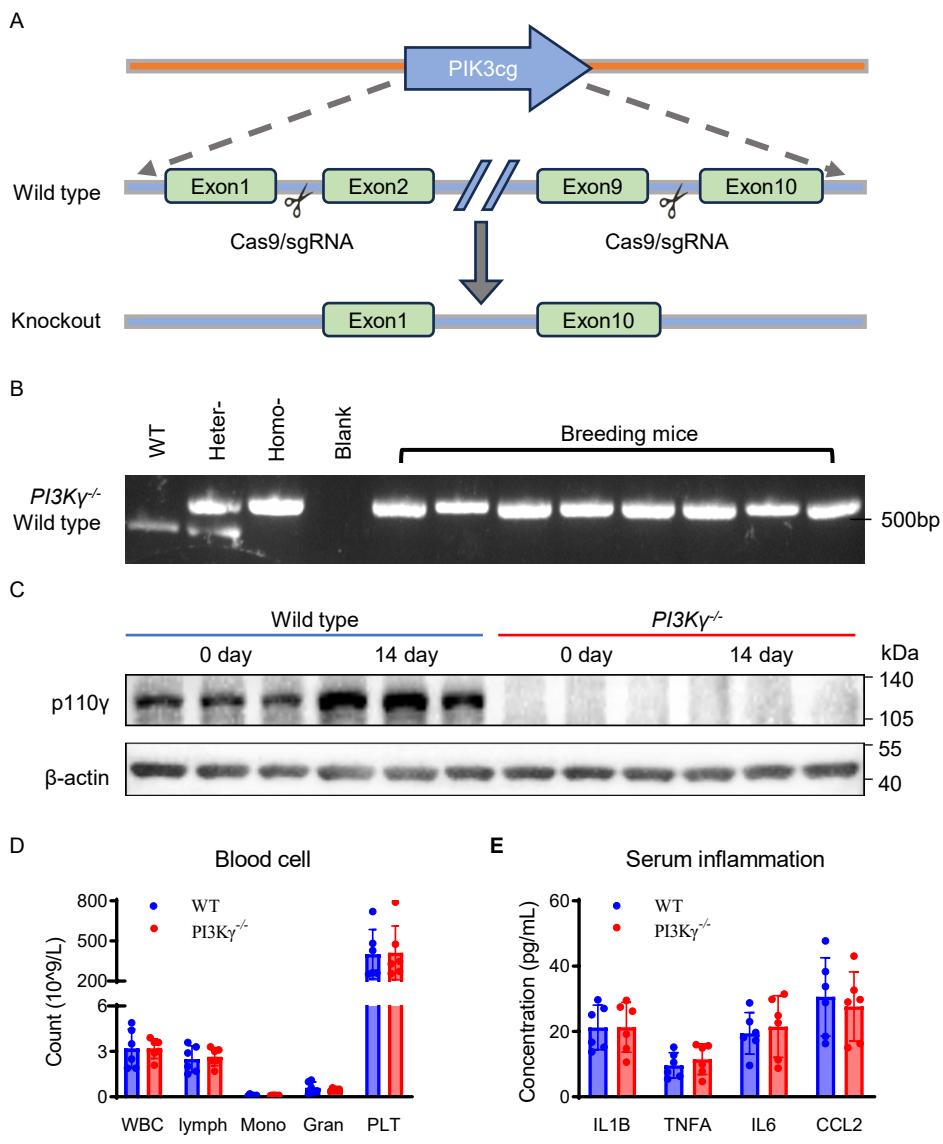


Figure S2

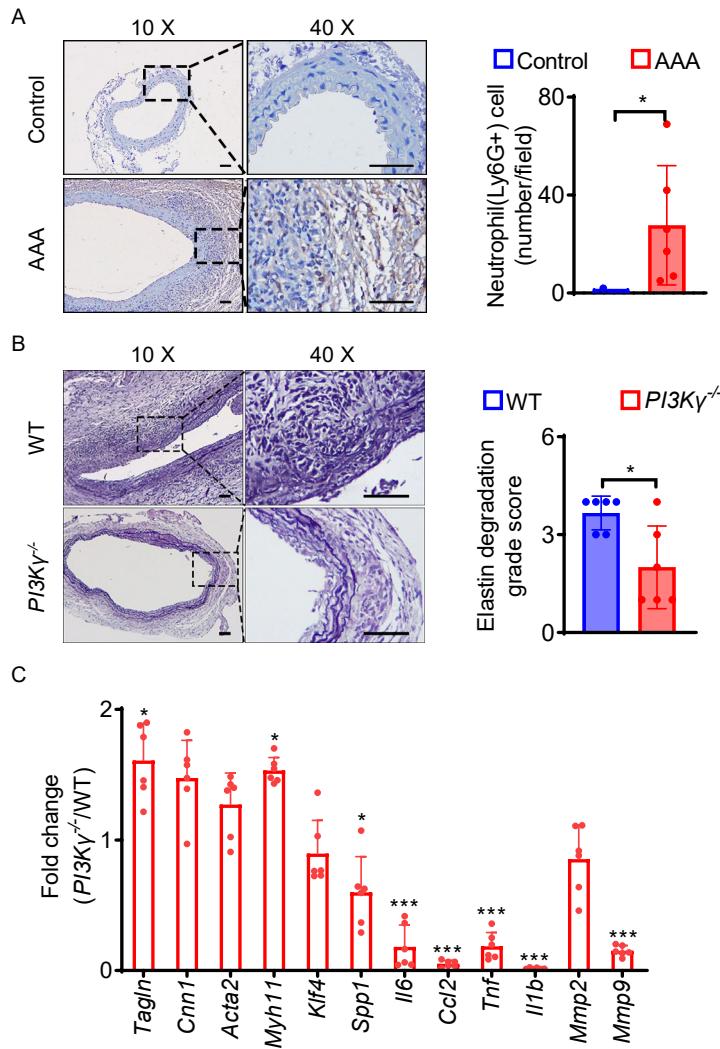
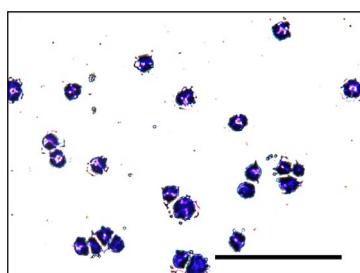
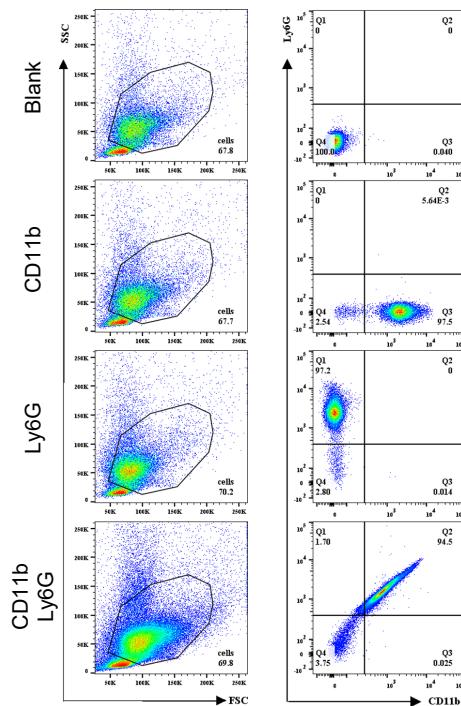


Figure S3

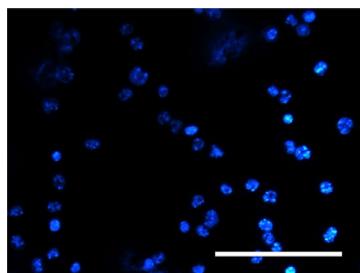
A



B



C



D

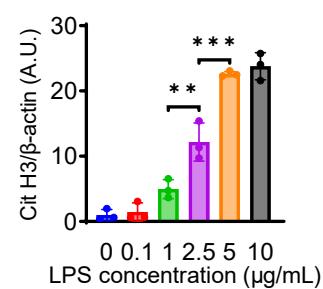
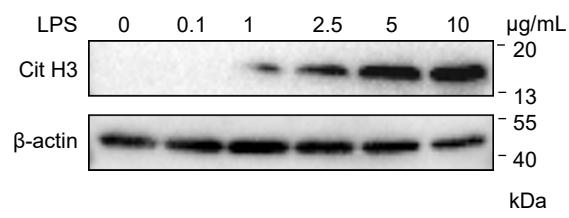


Figure S4

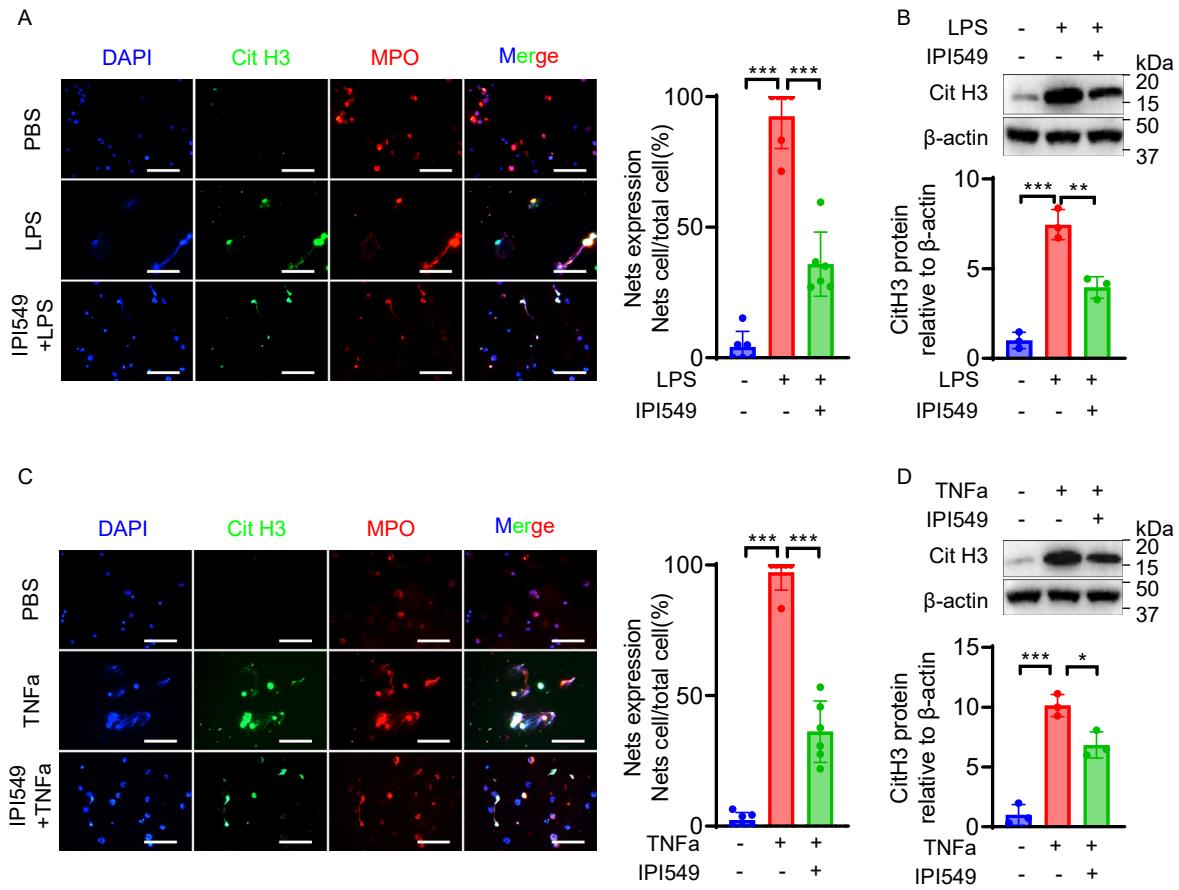


Figure S5

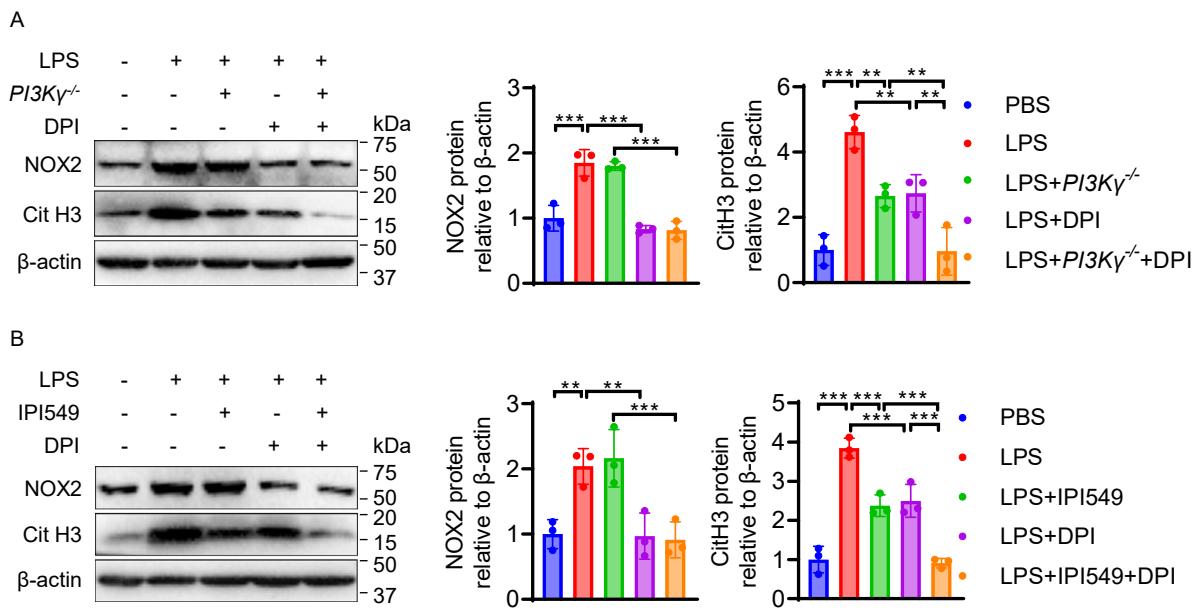


Figure S6

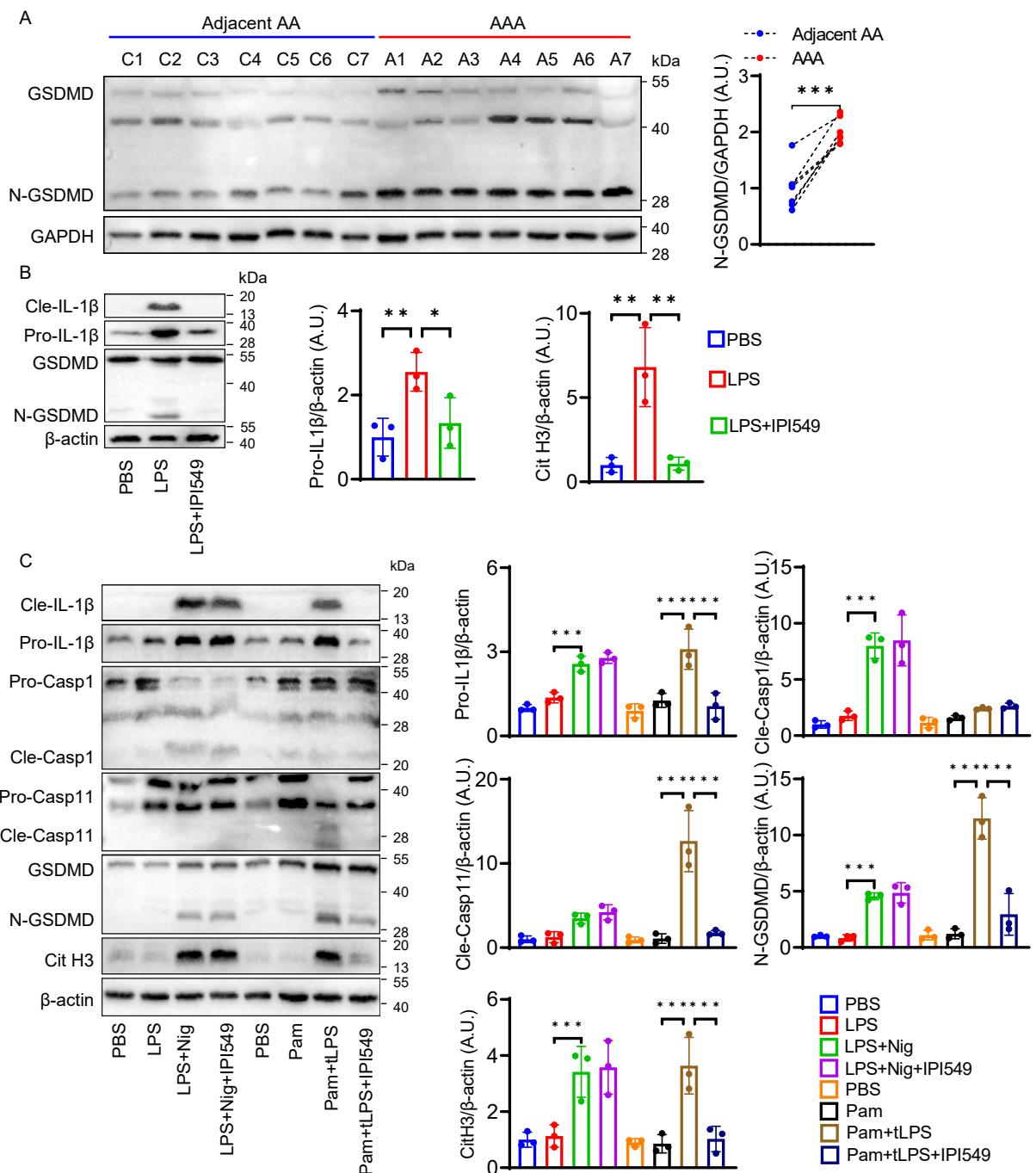
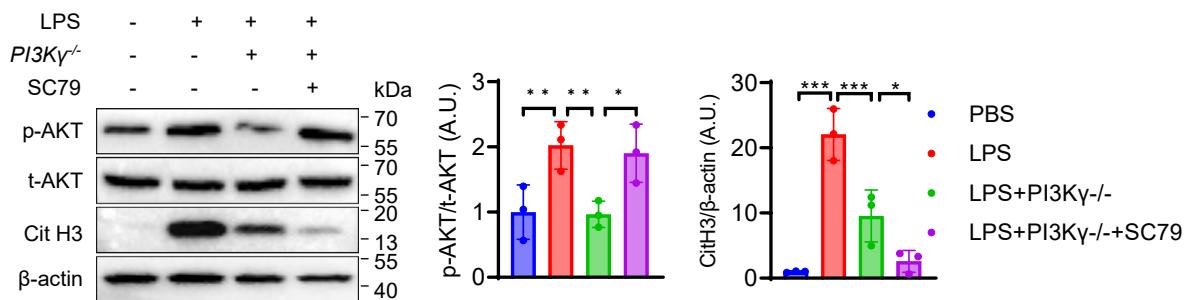


Figure S7

A



B

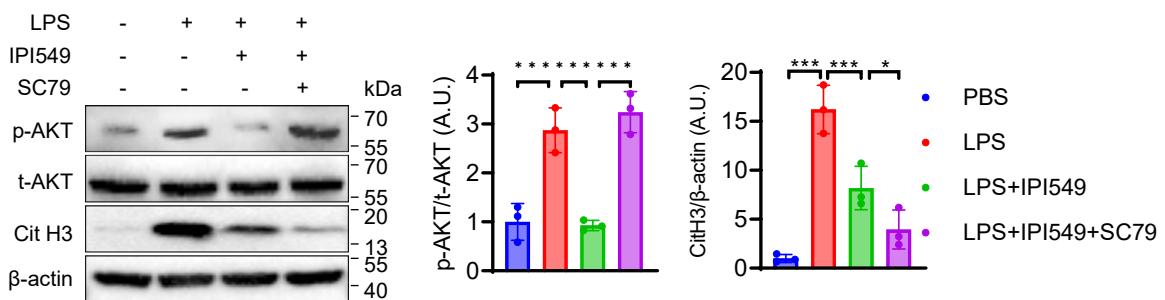


Figure S8

