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Supplemental information

Specialized cells that sense tissue mechanics to regulate *Drosophila* morphogenesis

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SUPPLEMENTAL ITEMS

Supplemental figures

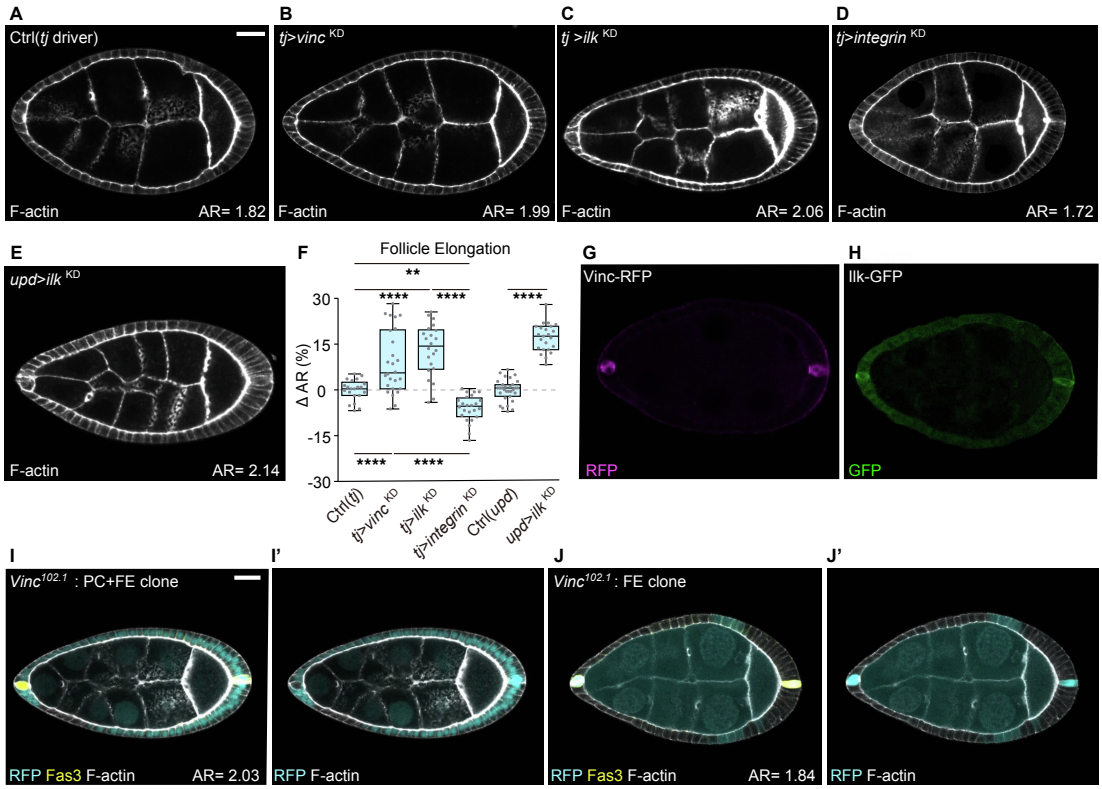

Figure S1. Focal adhesion component expression and function in follicles (Related to Fig. 1).

(**A**-**D**) Follicle-wide (via *tj*-GAL4, control in **A**, n=21) depletion of Vinc (**B**, n=26) or Ilk (**C**, n=22) exhibit opposite effects on tissue elongation compared to Integrin (**D**, n=24).

(E) PC-specific depletion of Ilk also causes hyperelongation (n=23).

(F) Quantitation of aspect ratios in A-E; Statistics used one-way ANOVA.

(**G**, **H**) Endogenous fluorescent tags in Vinc (**G**, n=9) and Ilk (**H**, n=7) reveal expression enriched in PCs.

(**I**, **J**) Follicles containing *vinc*$^{102.1}$ mitotic clones (marked by absence of RFP, cyan) that cover both PCs (labeled by Fas3) and part of follicle epithelium are hyperelongated (**I**, n=9). Follicles containing *vinc*$^{102.1}$ mutant clones limited to the follicle epithelium elongate normally (**J**, n=14).

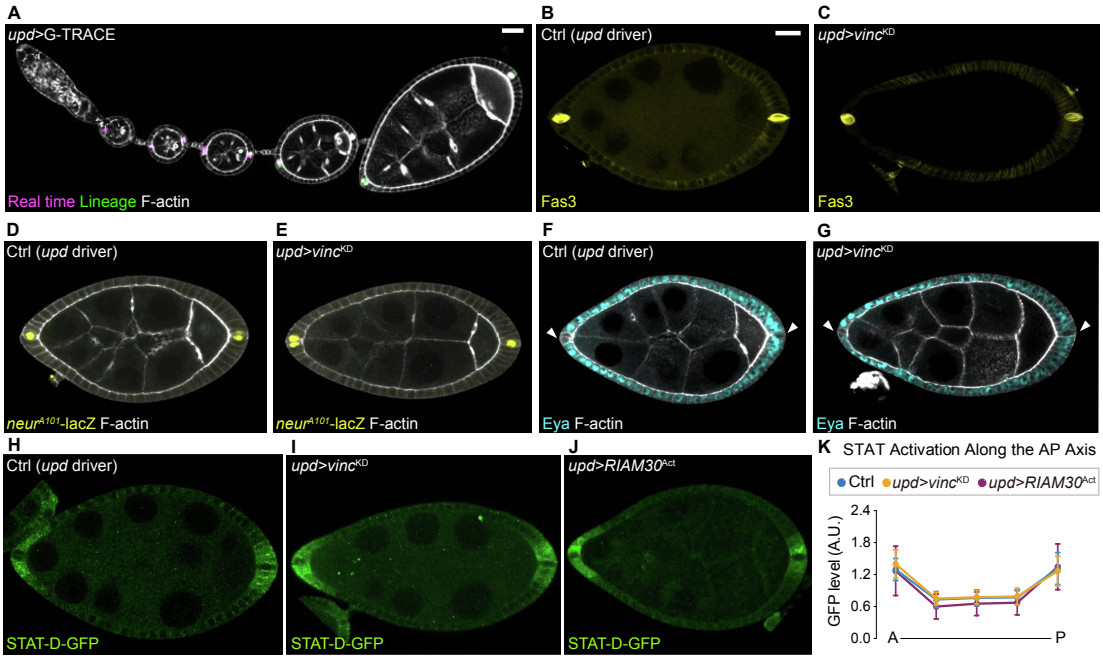

Figure S2. Follicle cell fates are unaltered by PC focal adhesion signaling (Related to Fig. 2).

(**A**) Expression of *upd-GAL4* is limited to PCs throughout follicle development. RFP indicates real-time expression, while GFP indicates past (lineage) expression (n=7).

(**B**-**G**) Markers for the PCs (Fas3 and *neur*^{A101}*-lacZ,* **B-E**) and follicle epithelium (Eya, **F-G**) in control follicles (**B, D, F,** n=11, 14, and 13, respectively) and follicles with PC focal adhesion depletion (**C, E, G** n=15, 10, and 12, respectively).

(**H**-**K**) STAT activation along the A-P axis in control follicles (**H**) is not changed when focal adhesion signaling in PCs drives either hyperelongation (**I**) or hypoelongation (**J**), all sample size =20, quantitation in (**K**). Statistical comparison of mutants to respective control are all non-significant (multiple t test).

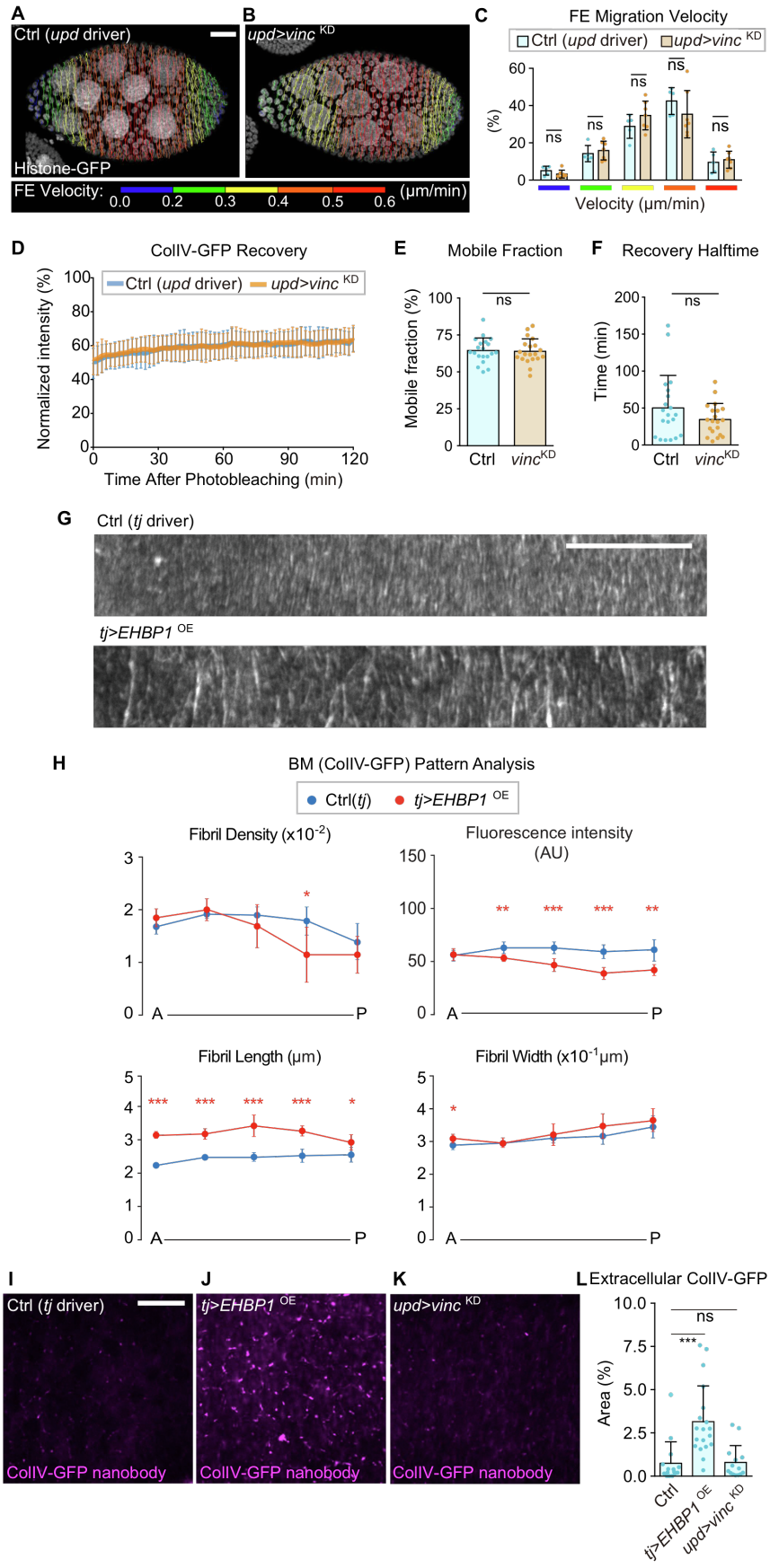

Figure S3. PC focal adhesion signaling does not affect follicle rotation nor BM secretion (Related to Fig. 3).

(**A**-**B**) Tracking of Histone His2Av-GFP in follicle epithelium cells during tissue rotation in control (**A**, n=5, 1983 cells) and PC focal adhesion-depleted follicle (**B**, n=7, 3524 cells) show comparable migration velocities. Each line indicates an individual cell trajectory and is color-coded according to its velocity. See **Video S2**.
(**C**) Quantitation of follicle epithelium migration velocity from (**A-B**). Statistical comparisons (multiple t test) between control and PC focal adhesion-depleted follicles are non-significant.
(**D**-**F**) ColIV deposition in control (n=20) and PC focal adhesion-depleted follicles (n=20) measured using FRAP assay on ColIV-GFP. Mobile fractions (**E**) and recovery halftimes (**F**) showed no significant difference; statistics used Kolmogorov-Smirnov test. Images were acquired every 2 m.
(G) ColIV-GFP patterns from the anterior regions of control (*tj* driver, n=6) and hyperleongated *tj>EHBP1*^{OE} follicles (n=6).
(**H**) Quantitation of follice-wide pattern. Statistics used multiple-t-tests.
(**I**-**L**) Extracellular ColIV-GFP staining in control (**I**, n=15), *tj>EHBP1*^{OE} (**J**, n=19), and *upd* > *vinc*^{KD} (**K**, n=14). Quantitation in (**L**). Statistics used Kolmogorov-Smirnov test.

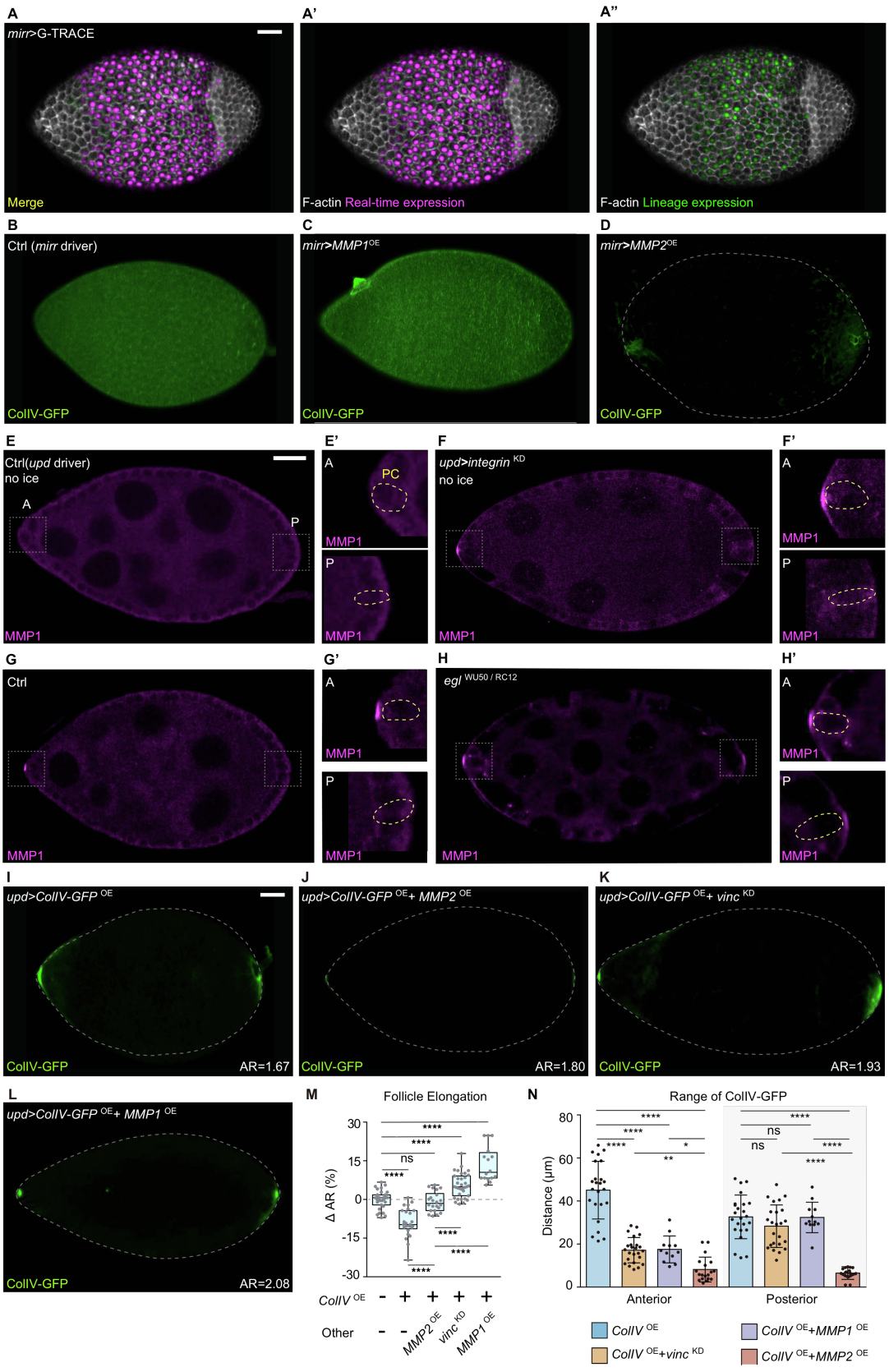

Figure S4. MMP1 does not lead to CollV degradation and is restricted to anterior PCs (Related to Fig 5).

(**A**) Expression of *mirr-GAL4* is limited to the central follicle epithelium throughout follicle development. RFP indicates real-time expression, while GFP indicates past (lineage) expression, n=6.
(**B-D**) ColIV-GFP pattern of control follicles (**B**, n=7) compared to follicles in which MMP1 (**C**, n=5), or MMP2 (**D**, n=4) is overexpressed in the central follicle epithelium.
(**E**-**F**) Compared to control (**E**, n=16), MMP1 is upregulated in anterior PCs (higher magnifications in **E'**, **F'**) when focal adhesions are depleted from these cells (**F**, n=14). Non-ice preps.
(**G**) Ice preps allow MMP1 detection in anterior but not posterior PCs control follicles (n=11).
(**H**) Anterior duplicated follicle (*egl*$^{WU50/RC12}$) shows MMP1 expression at both poles (arrows, n=5). Ice prep.
(**I-L**) ColIV-GFP overexpressed from PCs spreads a few cells from its source (**I**, n=24). MMP2 overexpression eliminates ColIV-GFP signal (**J**, n=19). focal adhesion depletion (**K**, n=24) or MMP1 overexpression (**L**, n=12) significantly limits anterior but not posterior ColIV-GFP spreading.
(**M**) Quantitation of follicle elongation in **I**-**L.** Statistics used one-way ANOVA.
(**N**) Range of ColIV-GFP expressed from anterior or posterior PCs in **I**-**L.** Statistics used one-way ANOVA.

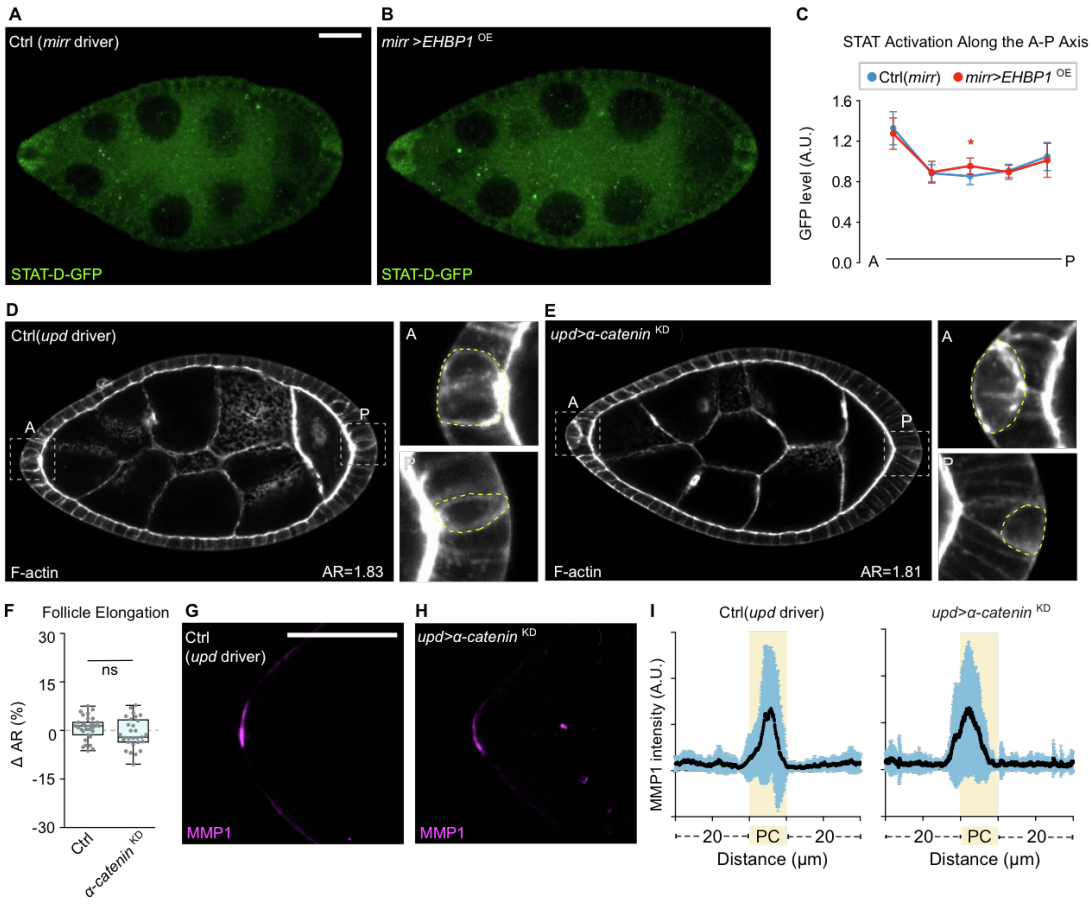

Figure S5. The PC response to distant BM stiffening is not through Upd signaling nor cell-cell junctions (Related to Fig. 6).

(**A-B**) Control (**A**, n=17) and follicles with central BM stiffening (**B**, n=20) show comparable STAT signaling activity, quantitated in (**C**). Statistics used multiple t tests.

(**D-F**) Compared to control (**D**, n=31), depleting α-catenin from PCs (**E**, n=31, apical retraction marked by yellow dashed lines) does not alter follicle elongation, quantitated in (**F**). Statistics used unpaired Mann-Whitney test.

(**G-H**) MMP1 levels are comparable in control and *α-catenin* mutant (n=16 and 17, respectively). (**I**) Quantitation of MMP1 signal along the anterior PCs (highlighted in yellow) for 50 μm.

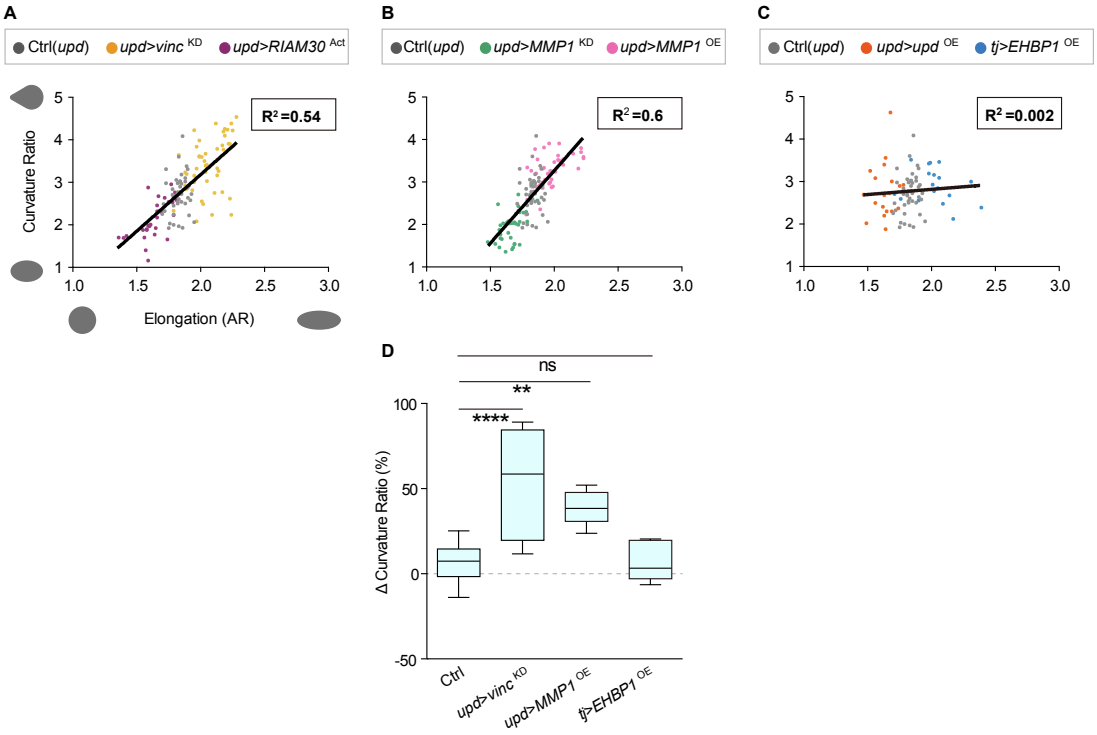

Figure S6: MMP1 regulates anterior follicle shape (Related to Fig. 3).
(A-C) Quantitation of 'pointiness' in fixed follicles, measured by ratio of curvature of anterior to posterior pole (y axis) versus degree of tissue elongation (x axis). Pointiness correlates highly with elongation in hyperelongated or hypoelongated follicles induced by changing FA signaling **(A)** or MMP1 manipulation in PCs **(B)**, but not in hyperelongated or hypoelongated follicles where STAT signaling or ColIV fibril secretion is manipulated **(C)**. The same dataset of control follicles is shown for each graph.
(D) Change in pointiness of follicles in bursting assay. Hyperelongated follicles induced by FA depletion or MMP1 overexpression show strong increases compared to control, while hyperelongated follicles induced by increased ColIV fibril secretion do not. Statistics used one-way ANOVA.

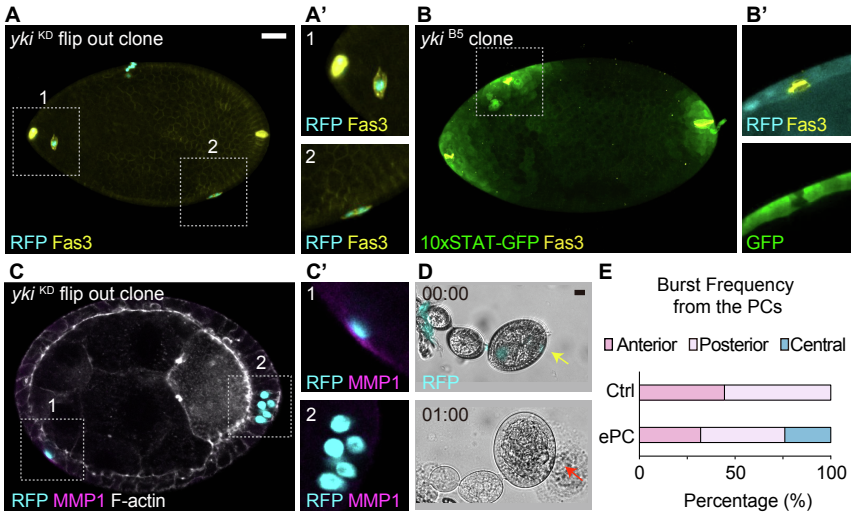

Figure S7. Ectopic PCs are sufficient to alter tissue mechanics (Related to Fig. 7).

(**A**) Follicle epithelium to PC fate transformation in *yki*-depleted clones (marked by RFP, cyan; Fas3: PC marker, yellow, n=23).

(**B**) Ectopic PCs (marked by Fas3, yellow, without RFP) activate STAT signaling (green) in surrounding follicle epithelium cells, n=23.

(**C**) MMP1 upregulation in ectopic anterior but not posterior PCs (marked by RFP, cyan), compare box 1 (anterior) and 2 (posterior), n=44.

(**D**) Ectopic PCs (yellow arrow) in the central follicle induce local mechanical failure (burst position, red arrow), as revealed by central as well as polar bursting under hypotonic stress. **Video S7**.

(**E**) Quantitation of bursting position in follicles with central PCs as compared to control, n= 27 and 25 respectively.

Supplementary Table 1. Detailed Genotypes and Experimental Conditions. Related to all main and supplemental figures.

Figure	Panel	Genotype	Condition
Figure 1	B	*w^{1118}*	25 °C; 3 D
	C	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-nls-GFP*	30 °C; 3-4 D
	D	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}*	30 °C; 3-4 D
	E	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-β integrin^{KD}*	30 °C; 3-4 D
	F	*upd-GAL4/+; UAS-cg25c-GFP/+; UAS-vkg-GFP/ tub-GAL80^{TS},UAS-Dicer2*	30 °C; 3-4 D
	G	*upd-GAL4/+; UAS-mCherry-RIAM30-RAP1^{CAAX}/+ ; tub-GAL80TS,UAS-Dicer2/+*	30 °C; 6 D
	H	*upd-GAL4/+; UAS-cg25c-GFP/ tub-GAL80^{TS} ; UAS-vkg-GFP/UAS-vinc^{KD}*	30 °C; 3-4D
	I	*upd-GAL4/+; UAS-mCherry-RIAM30-RAP1^{CAAX}/+ ; tub-GAL80^{TS}/UAS-β integrin^{KD}*	30 °C; 6 D
	J	Ctrl:*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		Others: from D-I	
	K	Ctrl: *upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-GFP^{KD}*	30 °C; 3-4 D
		Hyperelongated: *upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}*	30 °C; 3 D
		Round: *upd-GAL4/+; UAS-cg25c-GFP/+; UAS-vkg-GFP/ tub-GAL80^{TS},UAS-Dicer2*	30 °C; 3 D
Figure 2	A	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-Upd^{KD}*	30 °C; 6 D
	B	*upd-GAL4/+; UAS-Upd ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 4 D
	C	*upd-GAL4/+; UAS-Upd / tub-GAL80^{TS} ; UAS-vinc^{KD}/+*	30 °C; 4 D
	D	Ctrl:*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		Others: from A-C	
	E	*upd-GAL4/+; 10xSTAT92E-dGFP/ + ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		upd-GAL4/+; 10xSTAT92E-dGFP/ + ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}	30 °C; 3-4 D
		upd-GAL4/+; 10xSTAT92E-dGFP/UAS-mCherry-RIAM30-RAP1^{CAAX} ; tub-GAL80^{TS},UAS-Dicer2/+	30 °C; 6 D
Figure 3	B	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
	C	*tj-GAL4,vkg-GFP/UAS-vkg^{KD}; tub-GAL80^{TS}/UAS-cg25c^{KD}*	30 °C; 3 D
	D	*tj-GAL4,vkg-GFP/+; tub-GAL80^{TS}/UAS-FLAG-EHBP1*	25 °C; 3-4 D
	E	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}*	30 °C; 3-4 D
	F	*upd-GAL4/+; vkg-GFP/UAS-mCherry-RIAM30-RAP1^{CAAX} ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 6 D
	G	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{KD}*	30 °C; 3-4 D
	H	integrin^{KD}: *upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-β integrin^{KD}*	30 °C; 3-4 D
		MMP1^{OE}: *upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{OE}*	30 °C; 1-2 D
		Others as indicated in B-G	
	I, J, L	Ctrl: *upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		tj-GAL4,vkg-GFP/+; tub-GAL80^{TS}/UAS-FLAG-EHBP1	25 °C; 3-4 D
		upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}	30 °C; 3-4 D
		upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{OE}	30 °C; 1-2 D
Figure 4	A	Ctrl: *upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}	30 °C; 3-4 D
	B, F	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
	C, F	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}*	30 °C; 3-4 D
	D, F	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{OE}*	30 °C; 1-2 D
	E, F	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{KD}*	30 °C; 3-4 D
Figure 5	A-C	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
	D	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{KD}*	30 °C; 3-4 D
	E	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}*	30 °C; 3-4 D
	F	*upd-GAL4/+;UAS-mCherry-RIAM30-RAP1^{CAAX} /+ ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 6 D
	G	*upd-GAL4/+; UAS-cg25c-GFP/+; UAS-vkg-GFP/ tub-GAL80^{TS},UAS-Dicer2*	30 °C; 3 D
	I	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{OE}*	30 °C; 1-2 D
	J	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{OE}*	30 °C; 1-2 D
	K	*upd-GAL4/+; tub-GAL80^{TS}/ +; UAS-integrin^{KD}/UAS-MMP1^{KD}*	30 °C; 3-4 D
	L	Ctrl: *upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		others: as indicatedin D,I, and J	
Figure 6	A	*tub-GAL80^{TS}/vkg-GFP; mirr-GAL4/UAS-FLAG-EHBP1*	25 °C; 3-4 D
	B	*tub-GAL80^{TS}/UAS-cg25c-GFP; mirr-GAL4/UAS-vkg-GFP*	30 °C; 2-3 D
	C	*tub-GAL80^{TS}/+; mirr-GAL4/UAS-FLAG-EHBP1*	25 °C; 3-4 D
	D-F	*Vinc$^{102.1}$, FRT19A / hs-FLP^{122}, ubi-mRFP-nls, FRT19A; tub-GAL80^{TS}/UAS-vkg^{KD}; mirr-GAL4/UAS-cg25c^{KD}*	see Methods

Figure 7	**A**	*upd-GAL4/+; Fas3-GFP/+; tub-GAL80^{TS},UAS-Dicer2/+*	25 °C; 3 D
	B-C	*w^{1118}*	25 °C; 3 D
	E	FE: *Indy-GFP* and PC: *Fas3-GFP*	25 °C; 3 D
	F	st6/st8 anterior and posterior: *upd-GAL4/+; Fas3-GFP/+; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		tub-GAL80^{TS}/Fas3-GFP; mirr-GAL4/UAS-FLAG-EHBP1	25 °C; 3 D
		upd-GAL4/+; Fas3-GFP/+; tub-GAL80TS,UAS-Dicer2/UAS-vinc^{KD}	30 °C; 3-4 D
		upd-GAL4/+; Fas3-GFP/UAS-mCherry-RIAM30-RAP1CAAX ; tub-GAL80^{TS}/+	30 °C; 6 D
		upd-GAL4/+; Fas3-GFP/UAS-mbs^{KD}; tub-GAL80TS,UAS-Dicer2/+	30 °C; 3-4 D
		upd-GAL4/+; Fas3-GFP/UAS-zip^{KD}; tub-GAL80TS,UAS-Dicer2/+	30 °C; 3-4 D
	H, J	*upd-GAL4/+; UAS-mbs^{KD}/+; tub-GAL80TS,UAS-Dicer2/+*	30 °C; 3-4 D
	I, J	*upd-GAL4/+; UAS-zip^{KD}/+; tub-GAL80TS,UAS-Dicer2/+*	30 °C; 3-4 D
Figure S1	**A**	*tj-GAL4, tub-GAL80^{TS}/+ (II)*	30 °C; 3-4 D
	B	*tj-GAL4, tub-GAL80^{TS}/+ ; UAS-vinc^{KD}/+*	30 °C; 3-4 D
	C	*tj-GAL4, tub-GAL80^{TS}/+ ; UAS-ilk^{KD}/+*	30 °C; 3-4 D
	D	*tj-GAL4, tub-GAL80^{TS}/+ ; UAS-β integrin^{KD}/+*	30 °C; 3-4 D
	E	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/ UAS-ilk^{KD}*	30 °C; 3-4 D
	G	*Vinc-RFP(II)*	25 °C; 3 D
	H	*Ilk-GFP(III)*	25 °C; 3 D
	I-J	*Vinc$^{102.1}$, FRT19A / hs-FLP^{122}, ubi-mRFP-nls, FRT19A*	see Methods
Figure S2	**A**	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-G-TRACE*	30 °C; 3-4 D
	B, F	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
	C, G	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}*	30 °C; 3-4 D
	D	*upd-GAL4/+; tub-GAL80^{TS}/+ ; neur^{A101}-lacZ/ +*	30 °C; 3-4 D
	E	*upd-GAL4/+; tub-GAL80^{TS}/+ ; neur^{A101}-lacZ/ +/UAS-vinc^{KD}*	30 °C; 3-4 D
	H, K	*upd-GAL4/+; 10xSTAT92E-dGFP/ + ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
	I, K	*upd-GAL4/+; 10xSTAT92E-dGFP/ + ; tub-GAL80^{TS},UAS-Dicer2/UAS-vincKD*	30 °C; 3-4 D
	J, K	*upd-GAL4/+; 10xSTAT92E-dGFP/UAS-mCherry-RIAM30-RAP1^{CAAX} ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 6 D
Figure S3	**A**	*upd-GAL4/+; tub-GAL80^{TS}/+;His2Av-GFP/+*	30 °C; 3-4 D
	B	*upd-GAL4/+; tub-GAL80^{TS} ;His2Av-GFP/UAS-vinc^{KD}*	30 °C; 3-4 D
	D-F	*Ctrl: upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}	30 °C; 3-4 D
	G-H	*tj-GAL4, vkg-GFP/+; tub-GAL80^{TS}*	25 °C; 3-4 D
		tj-GAL4, vkg-GFP/+; tub-GAL80^{TS} / UAS-FLAG-EHBP1	25 °C; 3-4 D
	I	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
	J	*tj-GAL4,vkg-GFP/+; tub-GAL80^{TS}/UAS-FLAG-EHBP1*	25 °C; 3-4 D
	K	*upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}*	30 °C; 3-4 D
Figure S4	**A**	*tub-GAL80^{TS}/+; mirr-GAL4/UAS-G-TRACE*	30 °C; 3-4 D
	B	*tub-GAL80^{TS}/vkg-GFP; mirr-GAL4/+*	30 °C; 1 D
	C	*tub-GAL80^{TS}/vkg-GFP; mirr-GAL4/UAS-MMP1*	30 °C; 1 D
	D	*tub-GAL80^{TS}/vkg-GFP; mirr-GAL4/UAS-MMP2*	30 °C; 8 h
	E, G	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
	F	*upd-GAL4/+; + ; tub-GAL80^{TS},UAS-Dicer2/UAS-β integrin^{KD}*	30 °C; 3-4 D
	H	*egl^{IWU50}/egl^{IRC12}*	25 °C; 2 D
	I	*upd-GAL4/+; UAS-cg25c-GFP/+; UAS-vkg-GFP/ tub-GAL80^{TS},UAS-Dicer2*	30 °C; 3 D
	J	*upd-GAL4/+; UAS-cg25c-GFP/tub-GAL80^{TS}; UAS-vkg-GFP/ UAS-MMP2^{OE}*	30 °C; 2 D
	K	*upd-GAL4/+; UAS-cg25c-GFP/tub-GAL80^{TS}; UAS-vkg-GFP/ UAS-vinc^{KD}*	30 °C; 3 D
	L	*upd-GAL4/+; UAS-cg25c-GFP/tub-GAL80^{TS}; UAS-vkg-GFP/ UAS-MMP1^{OE}*	30 °C; 2 D
	M-N	as indicated in I-L	
Figure S5	**A**	*tub-GAL80^{TS}/10xSTAT92E-dGFP; mirr-GAL4/+*	25 °C; 3-4 D
	B	*tub-GAL80^{TS}/10xSTAT92E-dGFP; mirr-GAL4/UAS-FLAG-EHBP1*	25 °C; 3-4 D
	D, G, I	*upd-GAL4/+; +; tub-GAL80^{TS},UAS-Dicer2/ +*	30 °C; 3-4 D
	E, H, I	*upd-GAL4/+; +; tub-GAL80^{TS},UAS-Dicer2/ UAS-α-catenin^{KD}*	30 °C; 3-4 D
Figure S6	**D**	Ctrl: *upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/+*	30 °C; 3-4 D
		upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-vinc^{KD}	30 °C; 3-4 D
		upd-GAL4/+; vkg-GFP/+ ; tub-GAL80^{TS},UAS-Dicer2/UAS-MMP1^{OE}	30 °C; 1-2 D
		tj-GAL4,vkg-GFP/+; tub-GAL80^{TS}/UAS-FLAG-EHBP1	25 °C; 3-4 D
Figure S7	**A, C**	*hs-FLP^{122}; + ; Act>stop>GAL4,UAS-His-RFP/ UAS-yki ^{KD}*	see Methods
	B	*hs-FLP^{122}; FRT42D, yki^{B5} / FRT42D,ubi-RFP ; 10XStat92E-GFP/+*	
	D, E	*hs-FLP^{122}; vkg-GFP/+ ; Act>STOP>GAL4,UAS-His-RFP/ UAS-yki ^{KD}*	
Video S6		*hs-FLP^{122}; Fas3-GFP/+ ; Act>STOP>GAL4,UAS-His-RFP/ UAS-yki ^{KD}*	See Methods