

Supplementary Information

Inducible auto-phosphorylation regulates a widespread family of nucleotidyltransferase toxins

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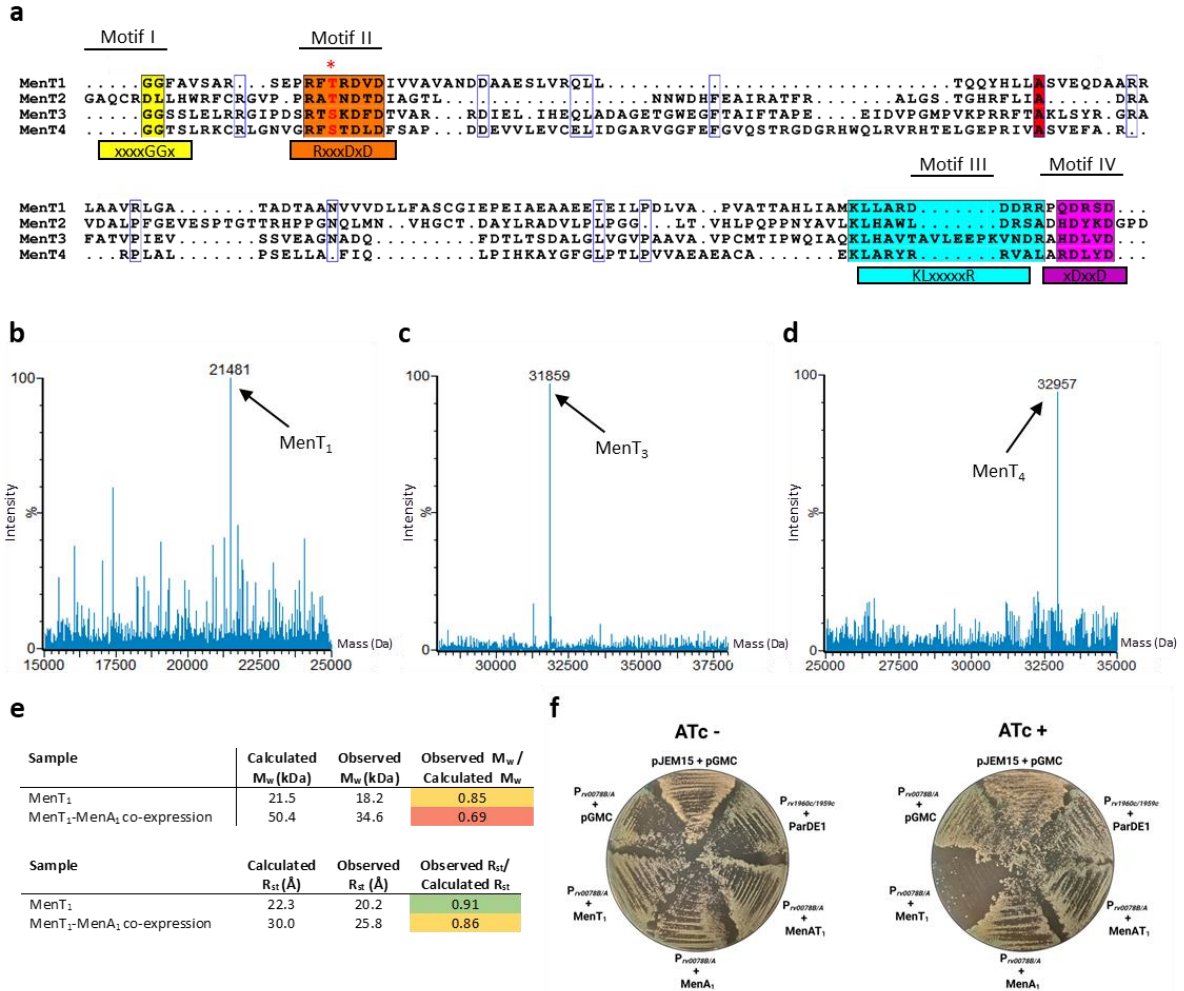
Supplementary Figure 8. Phosphorylation blocks heterotrimeric complex formation.

Supplementary Figure 9. MD simulations support the existence of a MenA₁:MenT₁-p heterodimer in solution.

Supplementary Table 1. Plasmids used in this study.

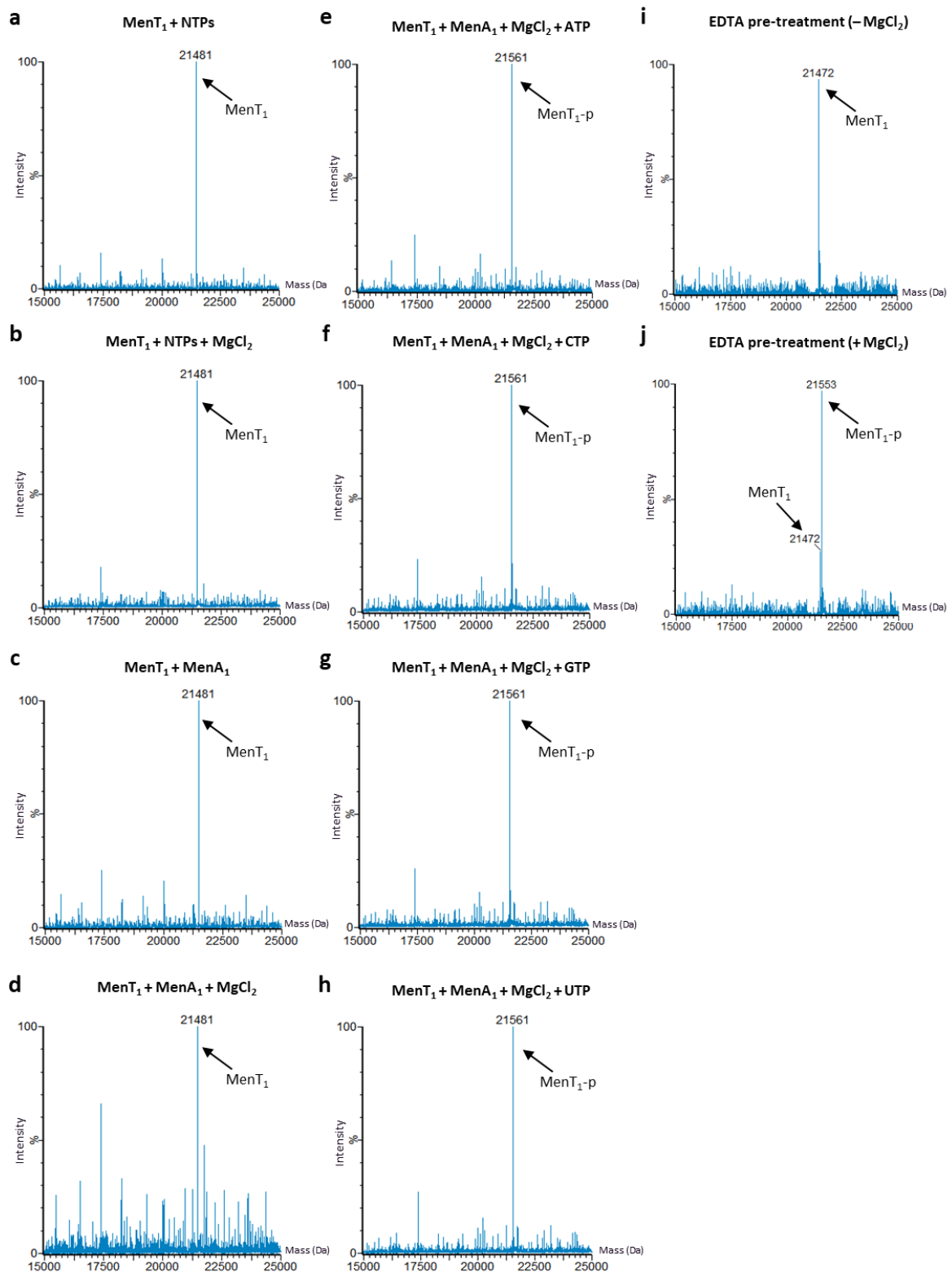
Supplementary Figures

Supplementary Figure 1



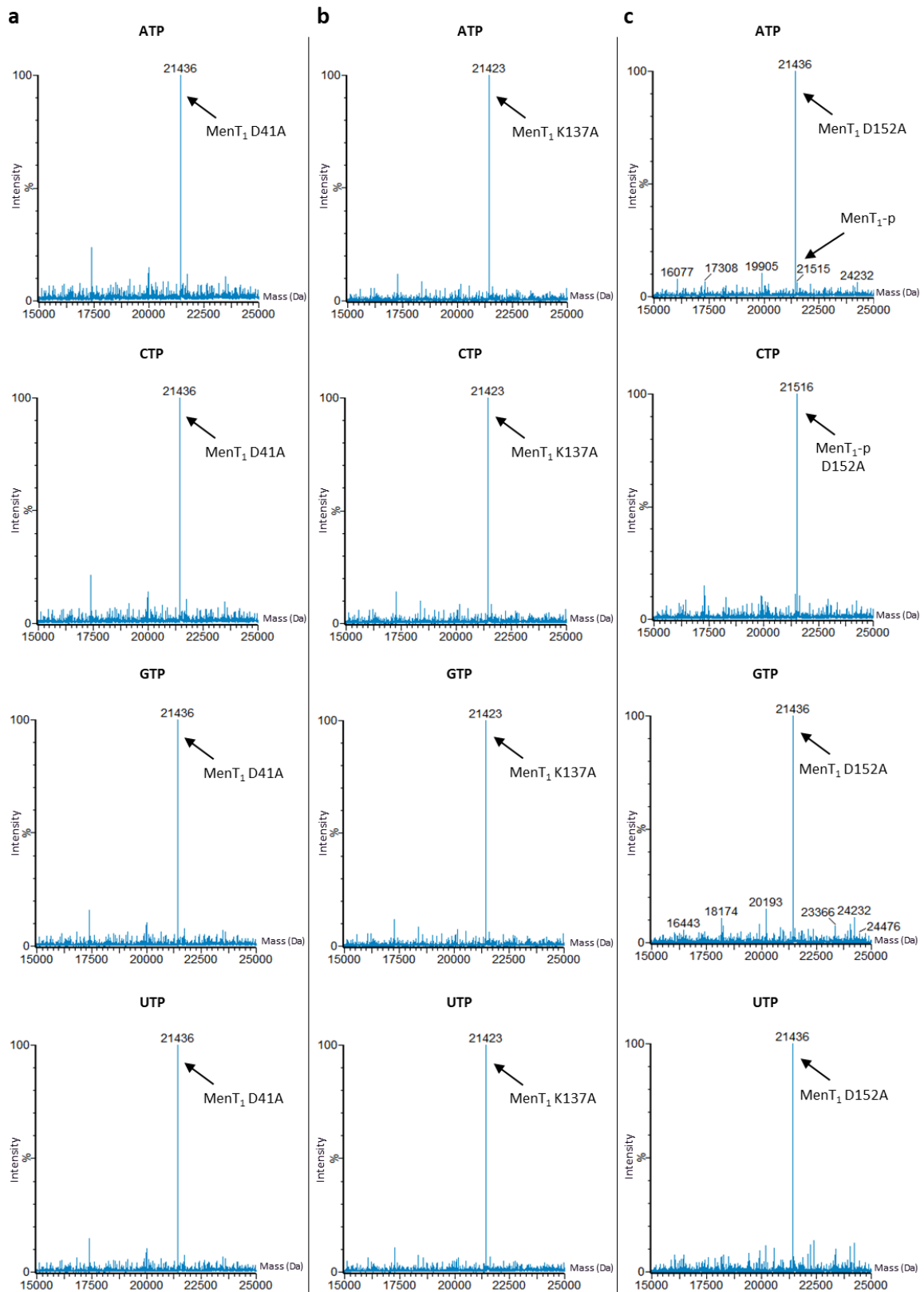
Supplementary Figure 1. MenT₁ is phosphorylated when co-expressed with MenA₁ *in vivo*. **a** Sequence alignment of the four MenT toxins from *M. tuberculosis* with conserved DUF1814 motifs coloured yellow (motif I), orange (motif II), cyan (motif III), and purple (motif IV). The site of MenT₁ and MenT₃ phosphorylation is denoted by a red asterisk. **b-d** ES²-ToF MS of purified MenT₁ (**b**), MenT₃ (**c**) and MenT₄ (**d**) expressed in the absence of cognate MenA antitoxins. **e** Observed/calculated Molecular Weight (M_w) and Stokes radii (R_{st}) corresponding to MenT₁ expressed in the absence and presence of MenA₁. Observed/calculated ratios are coloured green if $\leq 10\%$, yellow if > 10 and $\leq 20\%$, and red if $> 20\%$ deviation from expected values. Samples were analysed using a HiPrep™ 16/60 Sephacryl® S-200 HR column. **f** Blue/white colony screening of *M. smegmatis* mc²-155 co-transformed with pJEM15 vector-only, pJEM15-*rv0078B/A* containing a 1000 bp promoter insert (P_{rv0078B/A}), or pJEM15-*rv1960c/1959c* containing a 1000 bp promoter insert (P_{rv1960c/1959c}), and either pGMC -vector, -MenT₁, -MenA₁, -MenAT₁, or -ParDE1. Screening was performed by plating strains onto LB agar plates supplemented with Km (50 $\mu\text{g}\cdot\text{ml}^{-1}$), Sp (100 $\mu\text{g}\cdot\text{ml}^{-1}$), Tween-80 (0.05% v/v), IPTG (1 mM), and 5-bromo-4-chloro-3-indolyl β -D-galactopyranoside (X-Gal; 40 $\mu\text{g}\cdot\text{ml}^{-1}$), in the presence or absence of the pGMC inducer anhydrotetracycline (ATc; 100 $\text{ng}\cdot\text{ml}^{-1}$). Pictures were taken after 4 days incubation at 37 °C. Data are representative of three independent biological replicates.

Supplementary Figure 2



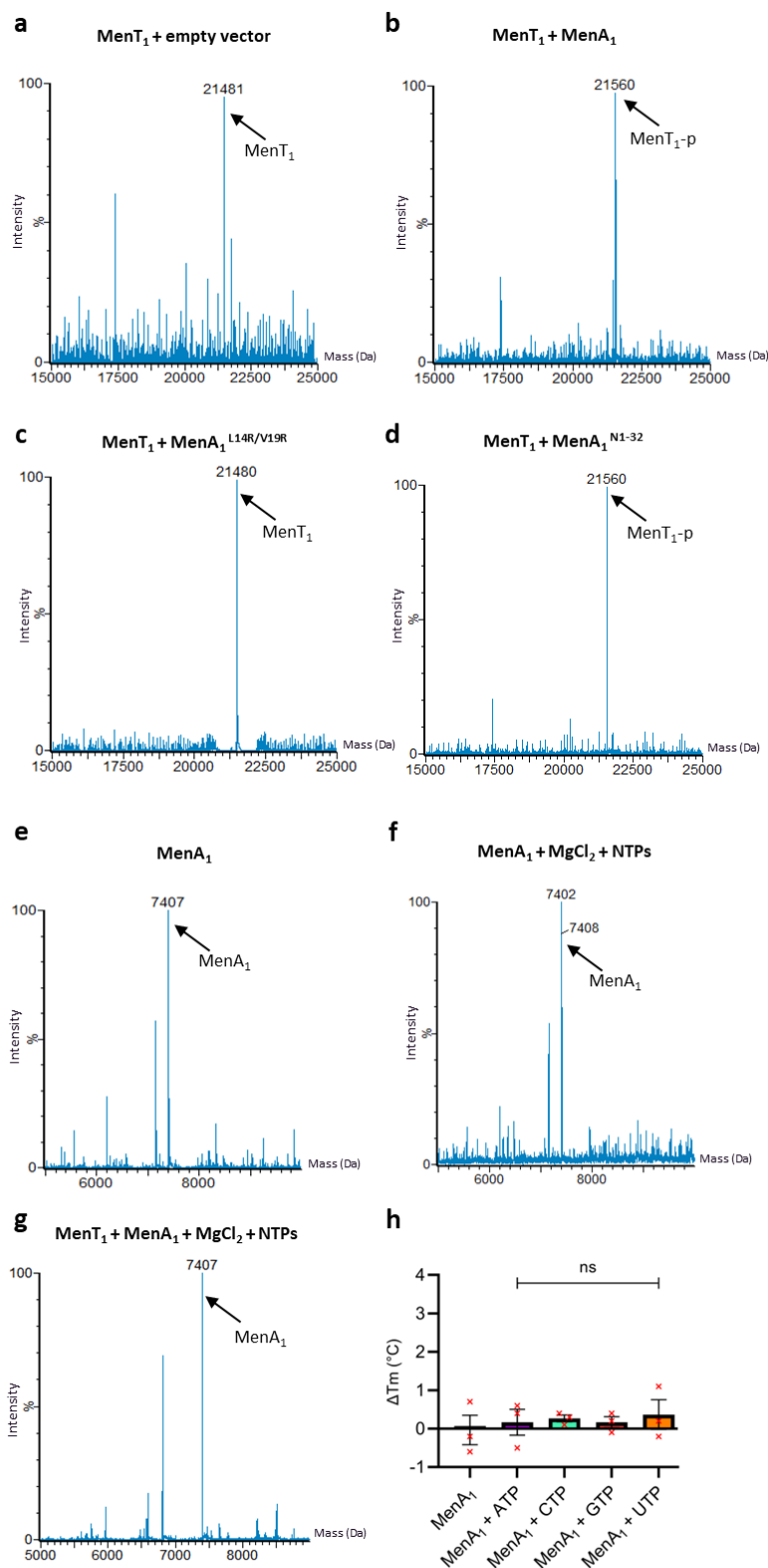
Supplementary Figure 2. MenT₁ is phosphorylated when incubated with each nucleotide in the presence of MenA₁ and MgCl₂. a-h ES⁺-ToF MS of MenT₁ co-incubated with NTPs in the absence (a) or presence (b) of MgCl₂, MenT₁ co-incubated with MenA₁ in the absence (c) or presence (d) of MgCl₂, and MenT₁ co-incubated with MenA₁, MgCl₂, and either ATP (e), CTP (f), GTP (g), or UTP (h). i-j ES⁺-ToF MS of MenT₁ co-incubated with MenA₁ and EDTA for 1 h, prior to the addition of CTP in the absence (i) or presence (j) of MgCl₂. Data are representative of three independent biological replicates.

Supplementary Figure 3



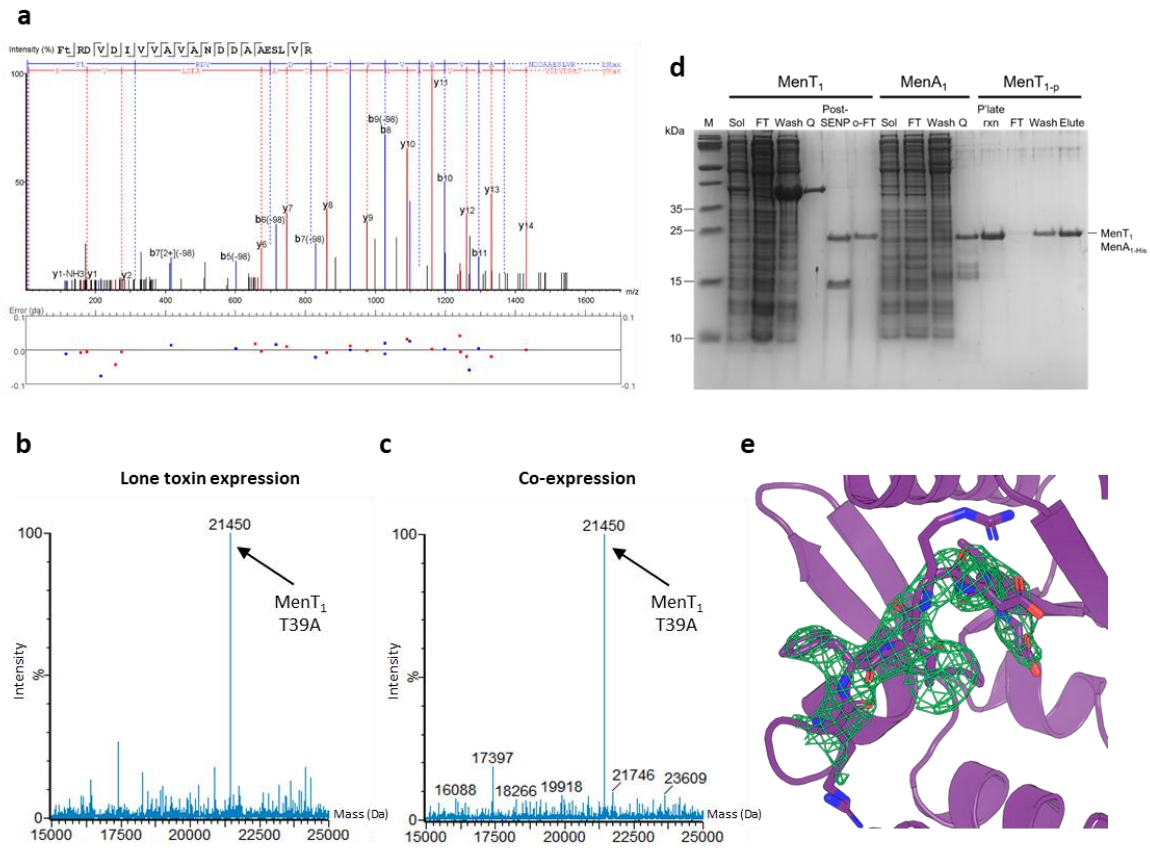
Supplementary Figure 3. D41 and K137 are essential for phosphorylation. a-c ES⁺-ToF MS of MenT₁ mutants D41A (a), K137A (b), and D152A (c) co-incubated with MgCl₂, MenA₁, and either ATP, CTP, GTP, or UTP. Data are representative of three independent biological replicates.

Supplementary Figure 4



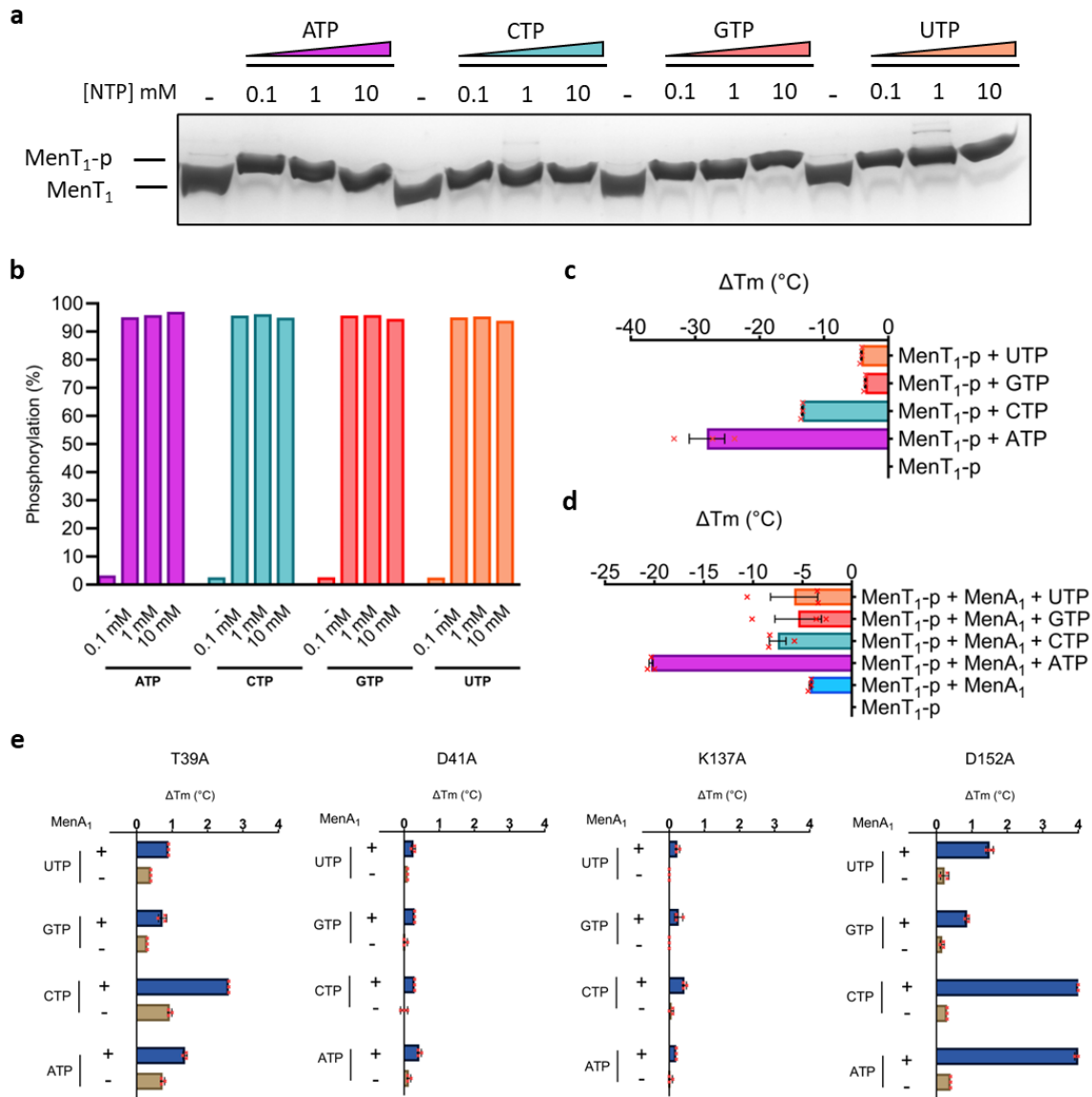
Supplementary Figure 4. Phosphorylation activity is localised to the MenA₁ N-terminus. **a-d** ES⁺-ToF MS of purified MenT₁ co-expressed with either empty vector (**a**), wild-type MenA₁ (**b**), or L14R/V19R (**c**) and N1-32 (**d**) mutants. **e-g** ES⁺ ToF MS of purified MenA₁ alone (**e**), or MenA₁ co-incubated with NTPs and MgCl₂, either in the absence (**f**) or presence (**g**) of MenT₁. **h** Mean changes in melting temperature following overnight incubation of MenA₁ with MgCl₂ and either ATP, CTP, GTP, or UTP (one-way ANOVA, $p = 0.903$). Data are representative of three independent biological replicates and bars display mean values \pm SEM.

Supplementary Figure 5



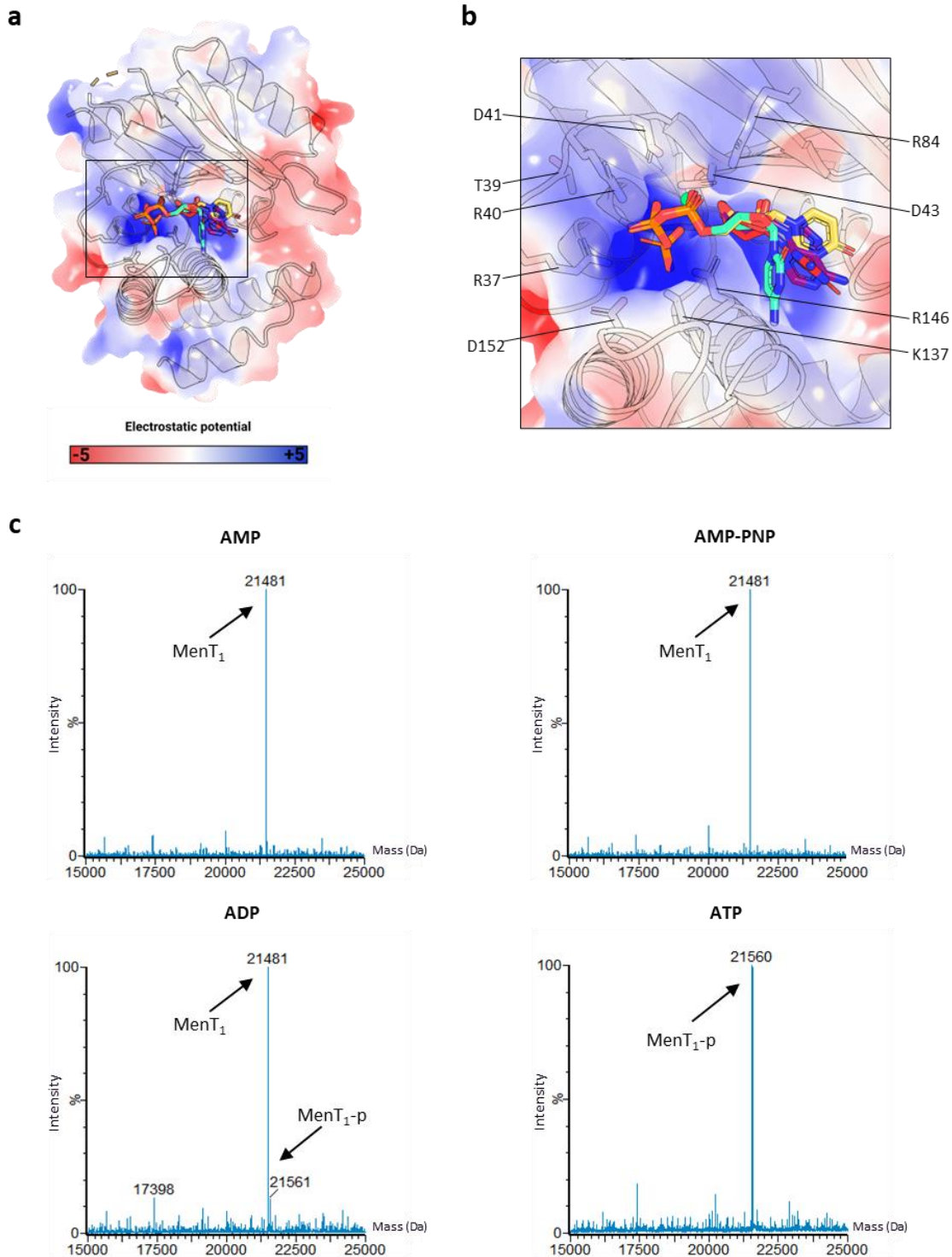
Supplementary Figure 5. MenT₁ is phosphorylated at T39. **a** LC-MS/MS analysis following ProAlanase peptide digests of MenT₁ expressed in the presence of MenA₁ identified T39 as the site of phosphorylation. Comparison of the masses of two identical fragments (₃₇RFIRDVIVAVANDDAESLVR₆₀) from MenT₁ expressed with and without MenA₁ revealed the co-expression sample to be 79.966 Da higher than toxin expressed alone, matching the expected mass of a single phosphate. **b-c** ES²-ToF MS of MenT₁ T39A expressed and purified in the absence (**b**) and presence (**c**) of MenA₁ confirms T39 is the sole site of phosphorylation. Data are representative of three independent biological replicates. **d** SDS-PAGE analysis of MenT_{1-p} fractions during purification. **e** Unbiased F₀-F_c electron density map depicting clear density prior to modelling in of the phosphothreonine.

Supplementary Figure 6



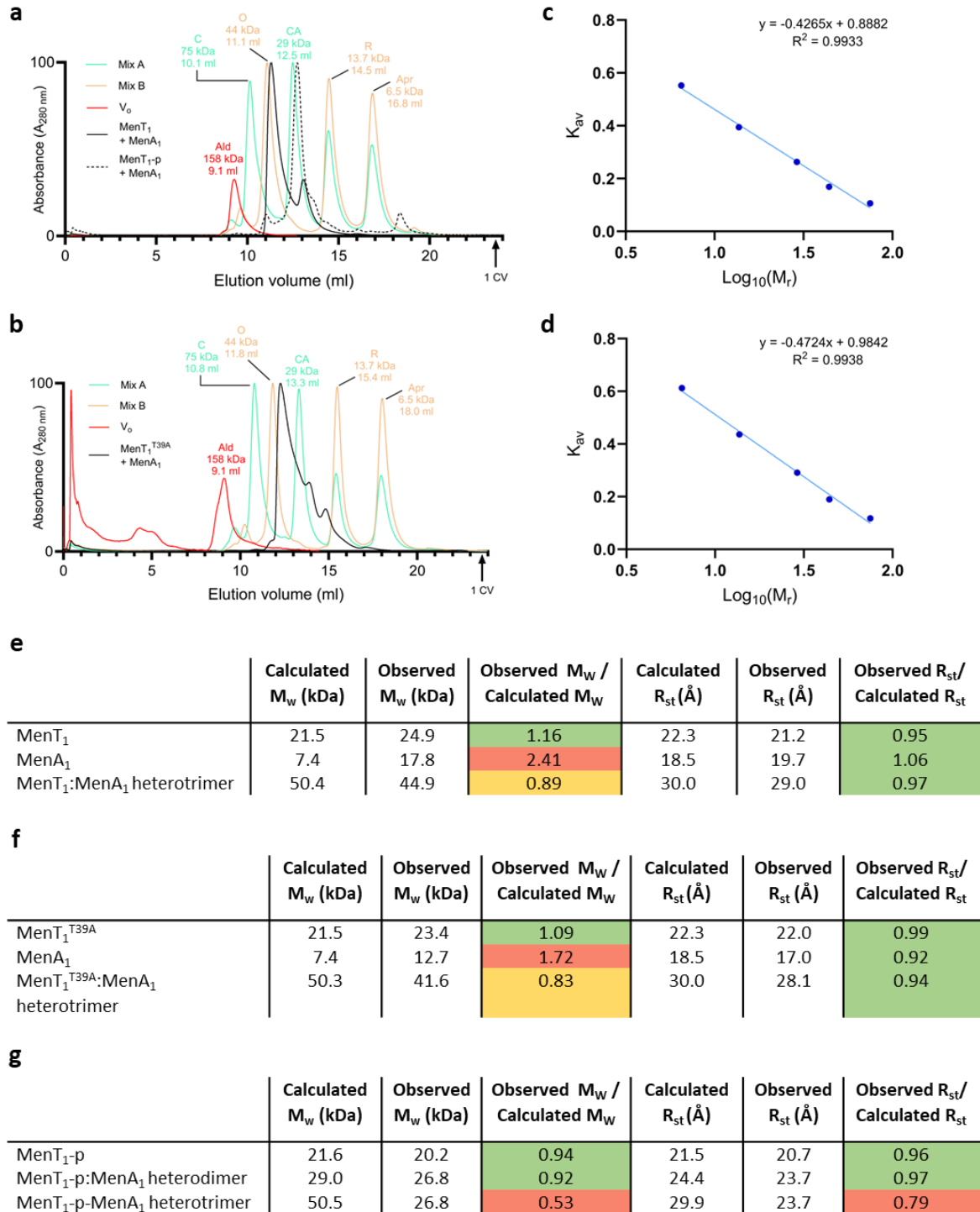
Supplementary Figure 6. All four NTPs are viable substrates for phosphorylation activity. **a** Phos-Tag SDS-PAGE showing levels of MenT₁-p following incubation with MenA₁ and MgCl₂ in the absence and presence of 0.1 mM, 1 mM, and 10 mM of each NTP. **b** Densitometric analysis of **(a)** reveals a lack of phosphorylation substrate specificity. Data are representative of two independent biological replicates. **c-d** Mean changes in melting temperature following overnight incubation of MenT₁-p with MgCl₂ and either ATP, CTP, GTP, or UTP, in the absence **(c)** or presence **(d)** of MenA₁. **e** Mean changes in melting temperature during thermal shift assays following incubation of MenT₁ catalytic mutants T39A, D41A, K137A, or D152A with MgCl₂ and ATP, CTP, GTP, or UTP, in the absence or presence of MenA₁. Data for panels **c-e** are representative of three independent biological replicates and bars display mean values +/- SEM.

Supplementary Figure 7



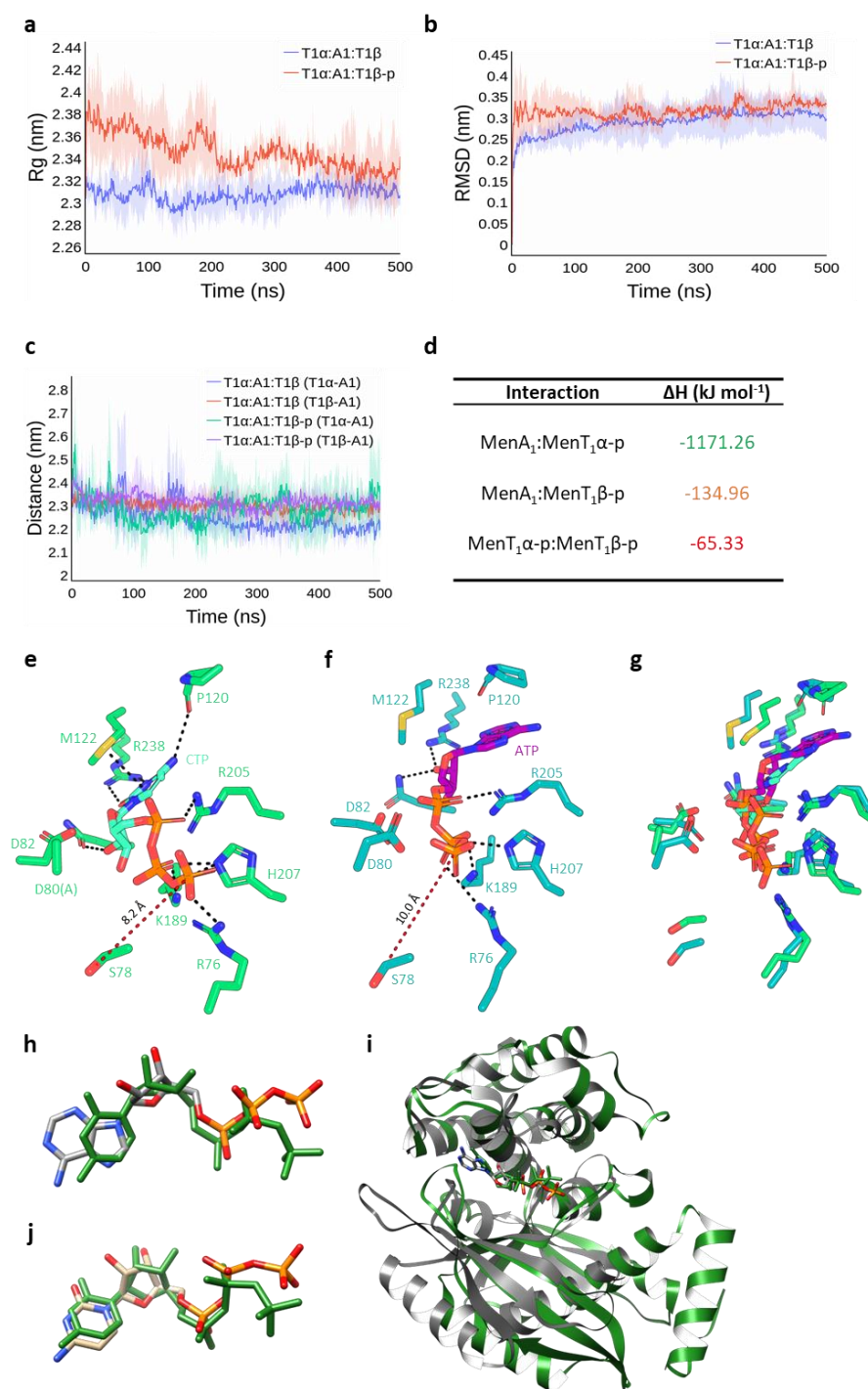
Supplementary Figure 7. Molecular docking predicts a major role of the phosphate tail in nucleotide binding. **a** Electrostatic surface colouring of MenT₁ and structural overlay of best-scored poses for ATP (purple), CTP (teal), GTP (red), and UTP (yellow) following molecular docking of nucleotides and Mg²⁺ to the MenT₁ active site. **b** Close-up view as in **(a)**. Residues of interest are shown as sticks, with surface colouring depicting electrostatic potential from $-5 \text{ kBT}e^{-1}$ (red) to $+5 \text{ kBT}e^{-1}$ (blue), where e is the electron, T is temperature and kB is the Boltzmann constant. Electrostatics were generated using default settings for the APBS plugin (PyMol). **c** ES⁺-ToF MS of MenT₁ co-incubated with MgCl₂, MenA₁, and either AMP, ADP, or AMP-PNP. Data are representative of three independent biological replicates.

Supplementary Figure 8



Supplementary Figure 8. Phosphorylation blocks heterotrimeric complex formation. **a-b** Overlaid SEC elution profiles of known molecular weight calibrants against MenT₁ + MenA₁ and MenT₁-p + MenA₁ (**a**), or MenT₁ T39A + MenA₁ (**b**). Samples were analysed using a Superdex™ 75 increase 10/300 GL SEC column. Where significant time had elapsed between experiments, the column was re-calibrated. Calibrants; Ald: Aldolase; C: Conalbumin; O: Ovalbumin; CA: Carbonic Anhydrase; R: Ribonuclease A; Apr: Aprotinin. **c-d** Linear plots of K_{av} against $\text{Log}(M_w)$ presented with equation of line and R^2 value following column calibration using known molecular weight calibrants; (**c**) corresponds to experiment (**a**); (**d**) corresponds to experiment (**b**). **e-g** Observed/calculated Molecular Weight (M_w) and Stokes radii (R_{st}) corresponding to MenA₁, and either MenT₁ (**e**), MenT₁ T39A (**f**), or MenT₁-p (**g**) incubated in the absence and presence of MenA₁. Observed/calculated ratios are coloured green if $\leq 10\%$, yellow if > 10 and $\leq 20\%$, and red if $> 20\%$ deviation from expected values; (**e**) and (**f**) correspond to experiments (**a**) and (**c**); (**g**) corresponds to experiments (**b**) and (**d**).

Supplementary Figure 9



Supplementary Figure 9. MD simulations support the existence of a MenA₂:MenT₁-p heterodimer in solution. **a** Radius of gyration plots for MenT₁ α :MenA₂:MenT₁ β and MenT₁ α :MenA₂:MenT₁ β -p during molecular dynamics simulations depicting overall conformational rigidity of non-phosphorylated and mono-phosphorylated trimers. **b** RMSD plots illustrating overall stability of MenT₁ α :MenA₂:MenT₁ β and MenT₁ α :MenA₂:MenT₁ β -p trimers during MD simulations. **c** Average distances between respective protomer chains in MenT₁ α :MenA₂:MenT₁ β and MenT₁ α :MenA₂:MenT₁ β -p heterotrimer simulations. **d** Protein-protein interaction enthalpies (kJ/mol) calculated for phosphorylated MenT₁:MenA₂:MenT₁ ternary complexes, illustrating stability of the MenA₂:MenT₁ α -p binary complex. **e-f** Close-up binding-site views of the MenT₃-CTP crystal structure (**e**; PDB 8XHR) and MenT₃ bound to ATP following MD simulations (**f**). Residues involved in nucleotide binding are shown as sticks red for oxygen, blue for nitrogen, and orange for phosphorus, with dashed lines shown to depict protein-ligand interactions. **g** Overlay of poses as in **e-f** depicting high overall similarity between known and predicted binding modes for CTP and ATP respectively. **h** Comparison of the best-scoring pose calculated for ATP bound to MenT₁ (colored by element) with the experimental structure of CTP bound to MenT₃ (dark green) after superimposing the protein backbones. **i** Overall binding mode of the calculated MenT₁-ATP complex (dim gray, ATP colored by element) overlaid onto the MenT₃-CTP complex (experimental structure, dark green). **j** The best-scoring pose calculated for CTP binding to MenT₃ (CTP colored by element) superimposed onto the experimental structure of CTP binding to MenT₃ (dark green). The calculation was performed to validate the molecular docking protocol. Plotted data for **a-c** represent three independent replicates.

Supplementary Table

Supplementary Table 1 - Plasmids used in this study

Gene name	Primers	Primer sequence (5' to 3')
pGMC-MenA ₁ T ₁	Rv0078B In-Fusion For	GAAGACAGGCTGCCCATGGCAGTTTCCGTCGCTGC
	Rv0078A In-Fusion Rev	TGTATAATAAAGTTGTTACCACTTGGCGGCGAGGC
pGMC-MenT ₁	Rv0078A In-Fusion For	GAAGACAGGCTGCCCATGAACGCTGTGGAGTCGAC
	Rv0078A In-Fusion Rev	TGTATAATAAAGTTGTTACCACTTGGCGGCGAGGC
pGMC-MenA ₁	Rv0078B In-Fusion For	GAAGACAGGCTGCCCATGGCAGTTTCCGTCGCTGC
	Rv0078B In-Fusion Rev	TGTATAATAAAGTTGTTATGTGAACCGTGTGGACG
pGMC-MenT ₁ T39A	Rv0078A T39A For	TCCGAACCACGTTTCGCCCGTACGTGGACATT
	Rv0078A T39C Rev	AATGTCCACGTCACGGGCGAAACGTGGTTCGGA
pGMC-MenT ₁ T39C	Synthesized and cloned by Genscript	
pGMC-ParDE1	Synthesized and cloned by Genscript	
pJEM15-P _{r_v0078B/A}	Synthesized and cloned by Genscript	
pJEM15-P _{r_v1960c/1959c}	Synthesized and cloned by Genscript	
pLAM-MenA ₁	Rv0078B NdeI For	TTGAATTCATATGGCAGTTTCCGTCGCTGCGCAG
	Rv0078B EcoRI Rev	TTGAATCTTATGTGAACCGTGTGGACG
pRARE	Novagen	
pPF656 MenA ₃	pPF1330 Rv1044 For	TTTCATATGCAATTGAGGAGGACAGGGATGTGTGCAAACCGTATCTAA
	pPF1331 Rv1044 Rev	TTTACTAGTCCCGGGCTTGGTCACGCCGATG
pPF657 MenT ₃	pPF1332 Rv1045 For	TTTCATATGCAATTGAGGAGGACAGGGATGACCAAGCCCTATTCGTC
	pPF1333 Rv1045 Rev	TTTACTAGTCCCGGGTTCATCTTTTCGTCGCC
pPF658 MenA ₄	pPF1334 Rv2827c For	TTTCATATGCAATTGAGGAGGACAGGGATGGT GAGCCCAGCCG
	pPF1335 Rv2827c Rev	TTTACTAGTCCCGGGTCCACGCCTTCCGATC
pTRB517 His ₆ -SUMO-MenT ₃	TRB1120	TTTGGTACCAAGAAGGAGATATATCCATGAGTGGC
	TRB1121	GCCTCCCGTCTGCTGTTGAA
	TRB1122	TTCAACAGCAGACGGGAGGCCACCAAGCCCTATTCGTCGCC
	TRB1124	TTTAAGCTTTTATTATCTTTTCGTCGCCGATCAA
pTRB544 His ₆ -SUMO-MenT ₄	TRB1175	TTTCCCGGAAGAAGGAGATATATCCATGAGTGGC
	TRB1176	TTCAACAGCAGACGGGAGGCCCGGTCTGACCCGTGCG
	TRB1177	TTTAAGCTTTTATTAGGACCGCAGCACCCGCCAG
	TRB1121	TTCAACAGCAGACGGGAGGCAACAAAGCTAAATTAACAGC
pTRB597 MenA ₁	TRB1681 Rv0078B For	CAACAGCAGACGGGAGGTAGGAGGACAGGGATGGCAGTTTCCGTCGCTG
	TRB 1682 Rv0078B Rev	GCGAGAACCAAGGAAAGTTTATTATCATGTGAACCGTGTGGACG
pTRB617 His ₆ -SUMO-MenA ₁	TRB1699 Rv0078B For	CAACAGCAGACGGGAGGTGAGTTTCCGTCGCTGCG
	TRB1700 Rv0078B Rev	GCGAGAACCAAGGAAAGTTTATTATGTGAACCGTGTGG
pTRB629 His ₆ -SUMO-MenT ₁	TRB1701 Rv0078A For	CAACAGCAGACGGGAGGTAACGCTGTGGAGTCGACAC
	TRB1702 Rv0078A Rev	GCGAGAACCAAGGAAAGTTTATTACCACTTGGCGGCGAGGCC
pTRB655 His ₆ -SUMO-MenA ₁ N1-32	TRB1883 For	TAATAACCTTCTGTTCTCGCATTC
	TRB1884 Rev	CCGTTACGACCCAGCCT
pTRB669 MenA ₃ D74A	Synthesized and cloned by Genscript	
pTRB670 MenA ₃ S93A	Synthesized and cloned by Genscript	
pTRB671 MenA ₃ H98A	Synthesized and cloned by Genscript	
pTRB672 MenA ₃ D102A	Synthesized and cloned by Genscript	
pTRB673	Synthesized and cloned by Genscript	

MenA ₃ N104A		
pTRB674 MenA ₃ D155A	Synthesized and cloned by Genscript	
pTRB717 MenA ₃ V103A	Synthesized and cloned by Genscript	
pTRB718 MenA ₃ P105A	Synthesized and cloned by Genscript	
pTRB719 MenA ₃ D102A/N104A	Synthesized and cloned by Genscript	
pTRB720 MenA ₃ D102A/V103A/N104A	Synthesized and cloned by Genscript	
pTRB698 His ₆ -SUMO-MenT ₁ T39A	Synthesized and cloned by Genscript	
pTRB699 His ₆ -SUMO-MenT ₁ D41A	Synthesized and cloned by Genscript	
pTRB700 His ₆ -SUMO-MenT ₁ K137A	Synthesized and cloned by Genscript	
pTRB701 His ₆ -SUMO-MenT ₁ D152A	Synthesized and cloned by Genscript	
pTRB704 His ₆ -SUMO-MenA ₁ L14R/V19R	Synthesized and cloned by Genscript	
tRNA primers		
<i>M. tuberculosis</i> tRNA Gly-3	Mtb tRNA Gly-3 For	ATTAATACGACTCACTATAGGCGGGCGTAGCTCAATGGT
	Mtb tRNA Gly-3 Rev	TGGAGCGGGCGACGGGAATC
<i>M. tuberculosis</i> tRNA Met-2	Mtb tRNA Met-2 For	ATTAATACGACTCACTATAGGGCGATGTAGCTCAGTCGGTTAGAGCGA
	Mtb tRNA Met-2 Rev	TGGTAGCGATGGCCGGACTCGAA
T7-tRNA	T7-For	ATTAATACGACTCACTAT
	HDV-Rev	AAACGACGGCCAGTGCCAAG
	HDV short-Rev	CTTCTCCCTTAGCCTACCG
<i>M. tuberculosis</i> tRNA T7-Gly3-HDV	Gly3-HDV For	CGATTCCCCTCGCCCGCTCCAGGGTCGGCATGGCATCTC
	Gly3-HDV Rev	GAGATGCCATGCCGACCCCTGGAGCGGGCGACGGGAATCG
	T7-Gly3 For	ATTAATACGACTCACTATAGCGGGCGTAGCTCAATGGTA
	HDV-Rev	AAACGACGGCCAGTGCCAAG
	Gly3 Mtb For	GCGGGCGTAGCTCAATGGTAGAGCCCTAGTCTTCCAACTAGCGACGCGG GTTTCGATTCCCGTCGCCCCTCCA
	HDVFor	GGGTCGGCATGGCATCTCCACCTCTCGCGGTCCGACCTGGGCTACTTCGG TAGGCTAAGGGAGAAGCTTGGCACTGGCCGTCGTTT
Sequences of T7-tRNA-HDV constructs		
T7-Met-2-HDV attaatacactcactatagcgatgtagctcagtcggttagagcgaacgactcataatcgtaggtcgcgggttcgagtcggccatcgctaccagggtcggcatggcatctccacctctcgcg gtccgacctgggctacttcggttaggtaaggagaagcttggcactggccgtcgttt		