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Supplementary Fig. 1 | Characterization and structural analysis of EfpA. a, Phylogenetic tree of EfpA
 homologues. *M. tuberculosis* and other pathogenic species of mycobacterium are highlighted with black
 star and empty stars, respectively. b, Size-exclusion chromatography profile of *Mt*EfpA with C-terminal

5 GFP tag after solubilized by LMNG-CHS. Source data are provided as a Source Data file. c, SDS-PAGE with 6 GFP fluorescence scan (left) and Coomassie stain (right). Source data are provided as a Source Data file. 7 d, Flow cytometry assay of *Mt*EfpA topology on HEK plasma membrane. HEK_{emp}, HEK293 cells without MtEfpA-GFP expressed. HEK_{MtE}, HEK293 cells with MtEfpA-GFP overexpressed. S-HEK_{emp}/S-HEK_{MtE}, cells 8 9 incubated with APC anti-GFP antibody without permeabilization. Intra-HEK_{emp}/Intra-HEK_{MtE}, cells 10 incubated with APC anti-GFP antibody after permeabilization. The oval on the right is represent the 11 plasma membrane of cells. Green represents GFP signal, and red represents APC signal. Source data are 12 provided as a Source Data file. e, Lipidomics and small-molecule analysis of dimeric MtEfpA-LMNGCHS 13 and monomeric *Mt*EfpA-DDMCHS using negative mode (top) and positive mode (bottom). One biological 14 repeat was performed for this experiment. Source data are provided as a Source Data file. f, Electrostatic 15 surface and hydrophobic surface of *Mt*EfpA. **g**, Dimeric interface of *Mt*EfpA dimer.

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Supplementary Fig. 2 | Cryo-EM reconstruction of dimeric MtEfpA. a, Representative cryo-EM micrograph of MtEfpA. b, 2D class averages of MtEfpA with a box size of 266 Å. c, Flowchart for processing cryo-EM data of MtEfpA. d, Particle angle distribution for the final map of dimeric MtEfpA. e, Local resolution of the final cryo-EM map. f, Gold standard Fourier shell correlation (FSC) curve for the final reconstruction. Half map #1 vs. half map #2 of MtEfpA with masked shown in black. Model vs. refined map with masked shown in purple. g, Close-up view of all structure elements fitted in densities with contour level at 5σ (blue).

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Supplementary Fig. 3 | Negative stain analysis of *MtEfpA* and mutants. a, Representative micrographs of monomeric *MtEfpA* purified with DDMCHS (upper) and dimeric *MtEfpA* purified with LMNGCHS

- 31 (bottom). **b**, Size exclusion chromatography (SEC) profile of *Mt*EfpA wide type and mutants purified with
- 32 LMNGCHS. The mutants forming monomers were marked with stars. Source data are provided as a Source
- 33 Data file. **c**, Negative stain 2D averages of *Mt*EfpA wild type (WT) and mutants purified with LMNGCHS.

Supplementary Fig. 4 | Cryo-EM reconstruction of monomeric MsEfpA. a, Representative cryo-EM micrograph of MsEfpA. b, 2D class averages of MsEfpA with a box size of 180.4 Å. c, Flowchart for processing cryo-EM data of MsEpfA. d, Particle angle distribution for the final map of monomeric MsEfpA.
e, Gold standard Fourier shell correlation (FSC) curve for the final reconstruction. f, Close-up view of all helixes (olive) fitted in densities with contour level at 5σ (blue) and a molecular of PE (brown) fitted into the density at lipid-binding site B. g, Superimposing the model of MsEfpA (olive) to MtEfpA (purple).

Supplementary Fig. 5 | Lipids binding channel in MtEfpA. a, The cartoon model of MtEfpA (purple and 46 47 orange) binding with cardiolipin (green) and two PE (wheat and yellow) from side view (left) and top view (right). **b**, Superimposition of phosphatidylethanolamine (PE_B) and the density at site B at a contour level 48 of 6 σ , presented in top view (left upper), side view (left bottom), and the *Mt*EfpA residues interacting with 49 50 PE_B within 4 Å (right). c, The density at site C (6 σ) fitted with the model of phosphatidylethanolamine (PE_c) showed in top view (left upper), side view (left bottom), and the residues of MtEfpA interacting with PE_c 51 52 within 4 Å (right). d, Cross-sectional views of lipids binding cavities in MtEfpA. Left, the cavities of 53 cardiolipin (CDL, green) and phosphatidylethanolamine (PE_B, wheat). Right, the cavities of 54 phosphatidylethanolamines (PE_B and PE_C).

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Supplementary Fig. 6 | Molecular dynamics simulation analysis of ligands in MtEfpA and Docking of 56 57 BRD-9327. a, Molecular dynamics simulation analysis for MtEfpA in monomer or dimer and RMSD of three 58 lipids. The bottom images present the interaction between MtEfpA and cardiolipin from the final frame 59 of four trajectories. b, Molecular dynamics simulation analysis for MtEfpA-BRD8000.3 in monomer or 60 dimer and RMSD of BRD-8000.3 and two lipids. The bottom images depict the interaction between MtEfpA 61 and BRD8000.3 from the final frame of four trajectories. The hydrogen bond is represented by a blue 62 dashed line. c, The configuration of BRD-8000.3 in one of the trajectories extracted from every 50 ns of 63 simulation (gray) superimposed with the solved structure of MtEfpA-BRD and the location of resistant 64 mutations to BRD-8000.3 (green). d, The location of resistant mutations (orange) to BRD-9327 (light blue) on *Mt*EfpA and the top 5 docking results of BRD-9327 at lipid-binding site B. 65

Supplementary Fig. 7 | Conservation analysis of EfpA homologous in *Mycobacterium* at the hydrophilic
 central pocket and three lipid-binding sites. a, Overall structure of *Mt*EfpA-lipids colored by conservation

69 of 61 EfpA homologues from different species of Mycobacterium. b-e, Zoomed-in view of residue

70 conservation at the hydrophilic central pocket (**b**) and three lipid-binding sites (**c**-**e**).

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- Supplementary Fig. 8 | Cryo-EM reconstruction of dimeric *Mt*EfpA-BRD8000.3. a, Representative cryo EM micrograph of *Mt*EpfA complexed with BRD-8000.3. b, 2D class averages of *Mt*EfpA with a box size of
 252 Å. c, Flowchart for processing cryo-EM data of *Mt*EpfA-BRD8000.3. d, Particle angle distribution for
 the final map of dimeric *Mt*EfpA-BRD8000.3. e, Gold standard Fourier shell correlation (FSC) curve for the
 final reconstruction of *Mt*EfpA-BRD8000.3. f, Close-up view of all structural elements of *Mt*EfpA BRD8000.3 fitted in densities with contour level at 5σ (blue).

82 Supplementary Fig. 9 | Structural comparison of *Mt*EfpA binding with cardiolipin and BRD-8000.3. a,

Superimposing structure of *Mt*EfpA (purple) and *Mt*EfpA-BRD (orange) at binding site A. b, c, Density of
 cardiolipin (dark green) in *Mt*EfpA (b) and BRD-8000.3 (light green) in *Mt*EfpA-BRD (c) with a contour level

85 at 6σ.

Supplementary Fig. 10 | Comparison of the structures of MtEfpA to the structures of MFSD2A. a, 88 89 Superimposing the outward-open structure of MtEfpA (purple) with the outward-open structure of lysophospholipid transporter MFSD2A from Mus musculus (MmusMFSD2A, light orange. PDB ID: 7N98). 90 91 **b**, Superimposing the inward structure of *Mt*EfpA-AF (gray) with the inward structure of MFSD2A from 92 Gallus gallus (GqMFSD2A, blue-green. PDB ID: 7MJS). c, Zoomed-in view of lipid binding site C shows the 93 lipids density in MtEfpA (yellow) and MmusMFSD2A (mesh, EMDB-24252). d, Zoomed-in view of lipid 94 binding site B shows the lipids density in MtEfpA (wheat) and the density of lysophosphatidylcholine (LPC, 95 orange, EMDB-23883) in GgMFSD2A. 96

Supplementary Fig. 11 | Inward and outward state of *Mt*EfpA. a, Superimposing AlphaFold-predicted
 inward-state of *Mt*EfpA (*Mt*EfpA-AF, lightgray-gray-darkgray) with the outward state of *Mt*EfpA (blue lightblue-lightpink). b, The lipid binding site A/B (left) and site C (right) after superimposing. c, Alignment
 of NTD (left), HD (mid) and CTD (right) from *Mt*EfpA and *Mt*EfpA-AF separately.

104 Supplementary Table 1 | Data collection and refinement statistics

	<i>Mt</i> EfpA (EMD-37641) (PDB 8WM5)	<i>Mt</i> EfpA-BRD (EMD-42204) (PDB 8UFD)	<i>Ms</i> EfpA (EMD-42205) (PDB 8UFE)	
Data collection and processing				
Magnification	45,000	29,000	36,000	130,000
Voltage (kV)	200	300	200	300
Electron exposure (e-/ Ų)	54	49	44	70
Defocus range (µm)	1.2-2.5	1.2-2.5	1.2-2.5	1.2-2.2
Pixel size (Å)	0.87	0.788	1.1	0.6632
Final Micrographs processed (no.)	3,031	6,550	3,280	3,554
Initial particle images (no.)	2,424,800	2,014,853	6,546,828	3,142,841
Final particle images (no.)	49,783	52,099	52,150	
Symmetry imposed	C2	C2	C1	
Map resolution (Å)	3.12	3.26	3.68	
FSC threshold	0.143	0.143	0.143	
Refinement				
Model resolution (Å)	3.2	3.5	4.1	
FSC threshold	0.5	0.5	0.5	
Model composition				
Non-hydrogen atoms	7179	7164	3449	
Protein residues	950	950	470	
Ligands	5	6	1	
<i>B</i> factors (Å ²)				
Protein	55.14	101.10	114.57	
Ligand	55.79	102.03	105.33	
R.m.s. deviations				
Bond lengths (Å)	0.009	0.030	0.003	
Bond angles (°)	0.924	1.462	0.640	
Validation				
MolProbity score	1.88	1.90	2.19	
Clashscore	12.86	18.45	12.83	
Poor rotamers (%)	0.00			
Ramachandran plot				
Favored (%)	96.19	97.25	94.44	
Allowed (%)	3.81	2.75	5.34	
Disallowed (%)	0.00	0.00	0.21	