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Supplemental information

Dynamic remodeling of septin structures

fine-tunes myogenic differentiation

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Supplementary Figures

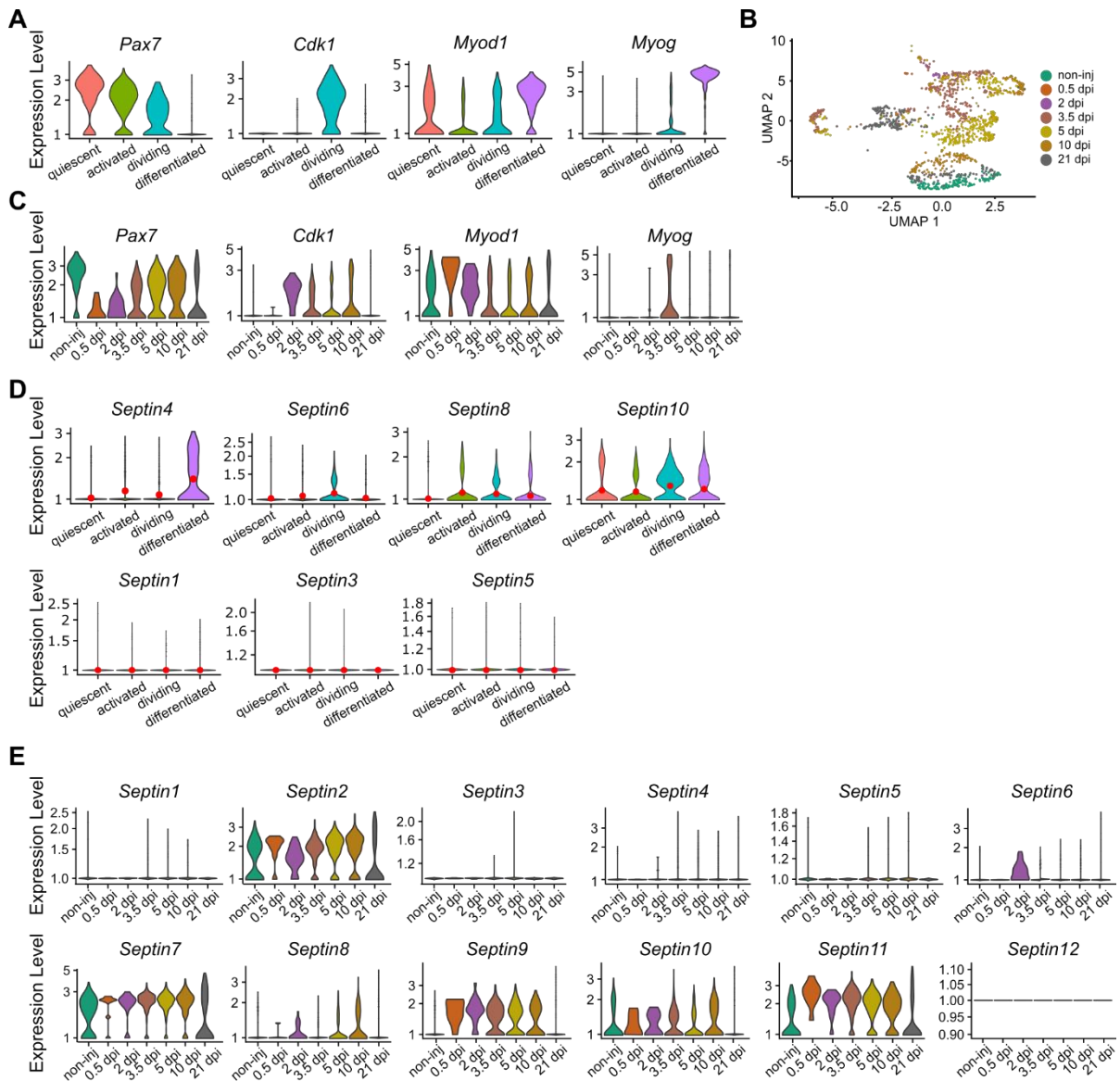


Figure S1 Transcriptional landscape of septins in the population of MuSCs during muscle regeneration, Related to Figure 1.

Single cell RNA sequencing data of non-injured tibialis anterior muscle and additional six time points following cardiotoxin injury (CTX)¹. **A** Violin plots representing subcluster-specific gene expression colored by cluster identity. **B** UMAP visualization of the sub-cluster MuSC in regenerating musculature colored by time points within the sub-cluster. **C** Violin plots representing subcluster-specific gene expression colored by time points. **D** Violin plots showing time point specific expression of septins during muscle regeneration. **D** Violin plots representing gene expression profiles of *Septin1*, *3*, *4*, *5*, *6*, *8* and *10* in MuSC subclusters. **E** Violin plots representing subcluster-specific gene expression of all septins colored by experimental time points.

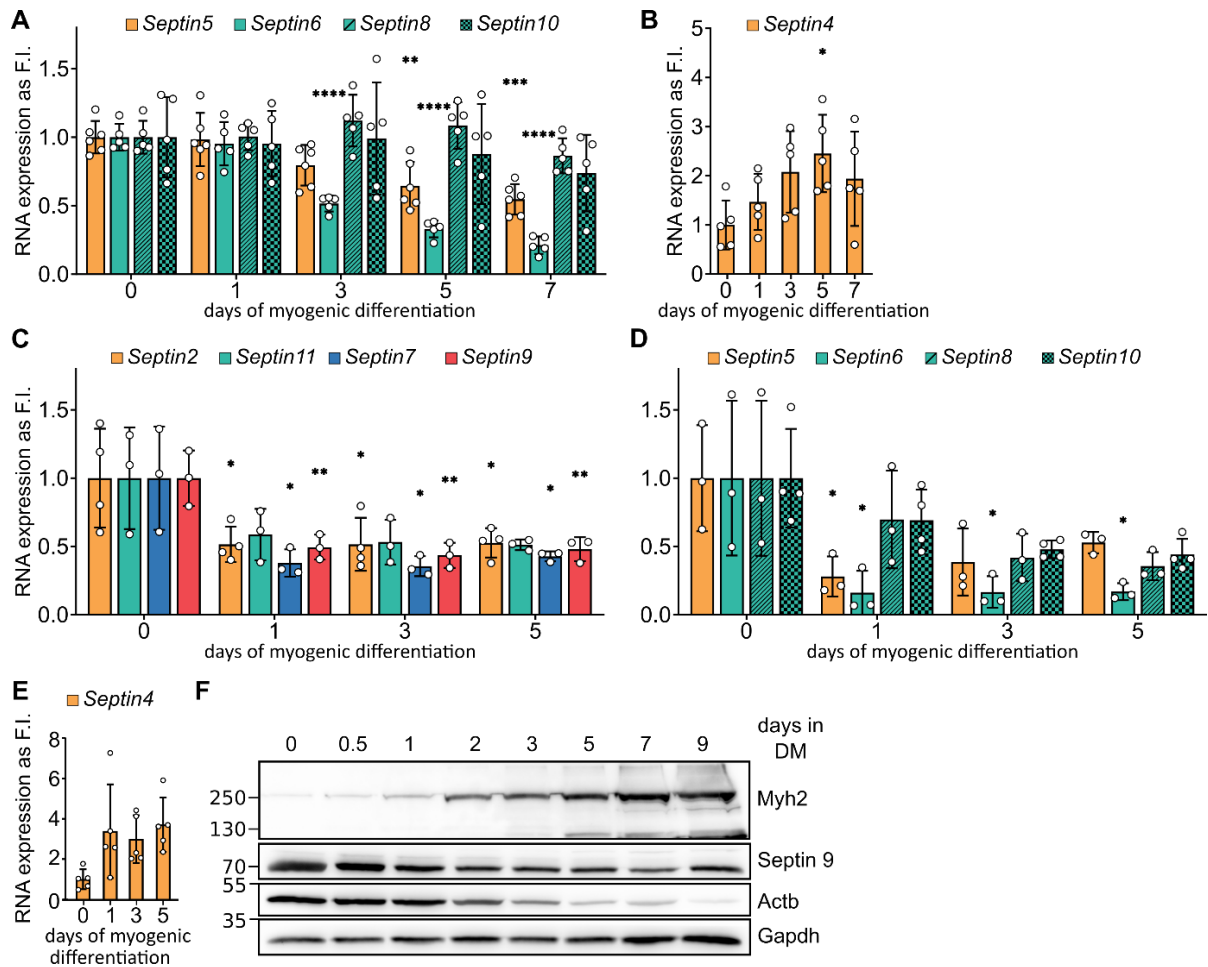


Figure S2 Expression of septins during myogenic differentiation in C2C12 cells and primary myoblasts, Related to Figure 1.

A-B mRNA expression levels of *Septin5*, *6*, *8*, *10* and *4* during C2C12 differentiation. **C-E** mRNA expression levels of core myogenic septins (C) and other expressed septin subunits (D-E) during 5 days of myogenic differentiation in primary myoblasts. **F** Representative western blot of Septin9 during 9 days of myogenic differentiation in primary myoblasts. Data represent mean \pm standard deviation (SD), * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$ from one way ANOVA followed by Dunnett's multiple comparison test.

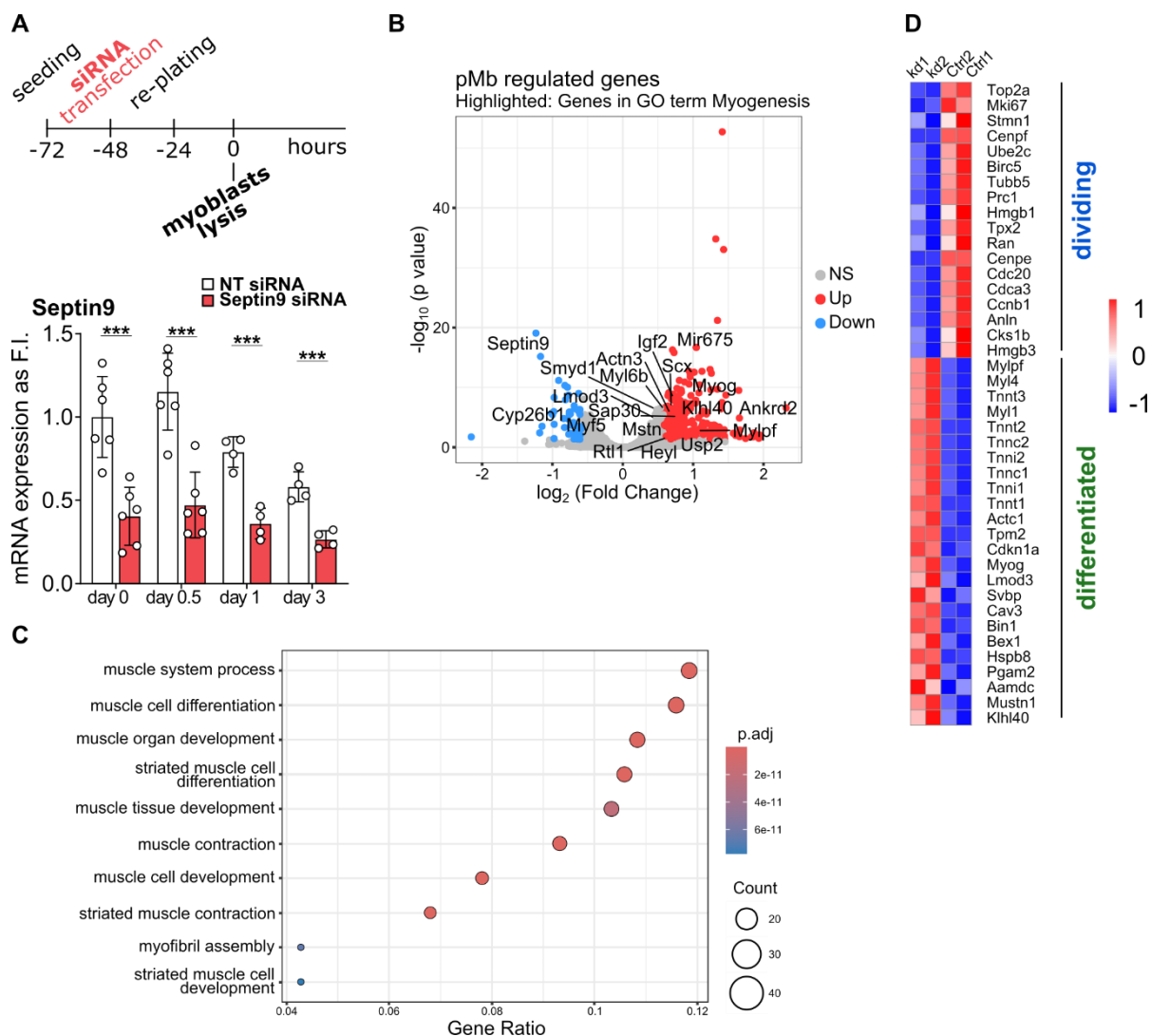


Figure S3 Septin9 depletion induces transition from cycling towards committed cells in primary myoblasts, Related to Figure 2.

A Schematic representation of the experimental setup. Total RNA was isolated from proliferating myoblasts 48 hours after Septin9 knockdown. Septin9 knockdown efficiency validation over 3 days of *in vitro* myogenic differentiation via qRT-PCR in primary myoblasts. **B** Volcano plot of DE genes (adjusted p value < 0.05; $-0.585 \leq \log_2FC \leq 0.585$) of primary myoblasts transfected with either non-targeting siRNA (control) or Septin9 siRNA after in growth medium, genes corresponding to Gene Ontology term “Myogenesis” are highlighted. **C** GO enrichment analysis of genes upregulated in Septin9-deficient primary myoblasts. **D** Heat map depicting marker genes for dividing and differentiated MuSCs defined by scRNAseq data depicted in Figure 1. Genes are filtered for significant regulation in primary myoblasts (adjusted p value < 0.05). Data represent mean \pm standard deviation (SD), *p < 0.05, **p < 0.01, ***p < 0.001, ****p < 0.0001 from two-sided unpaired t test.

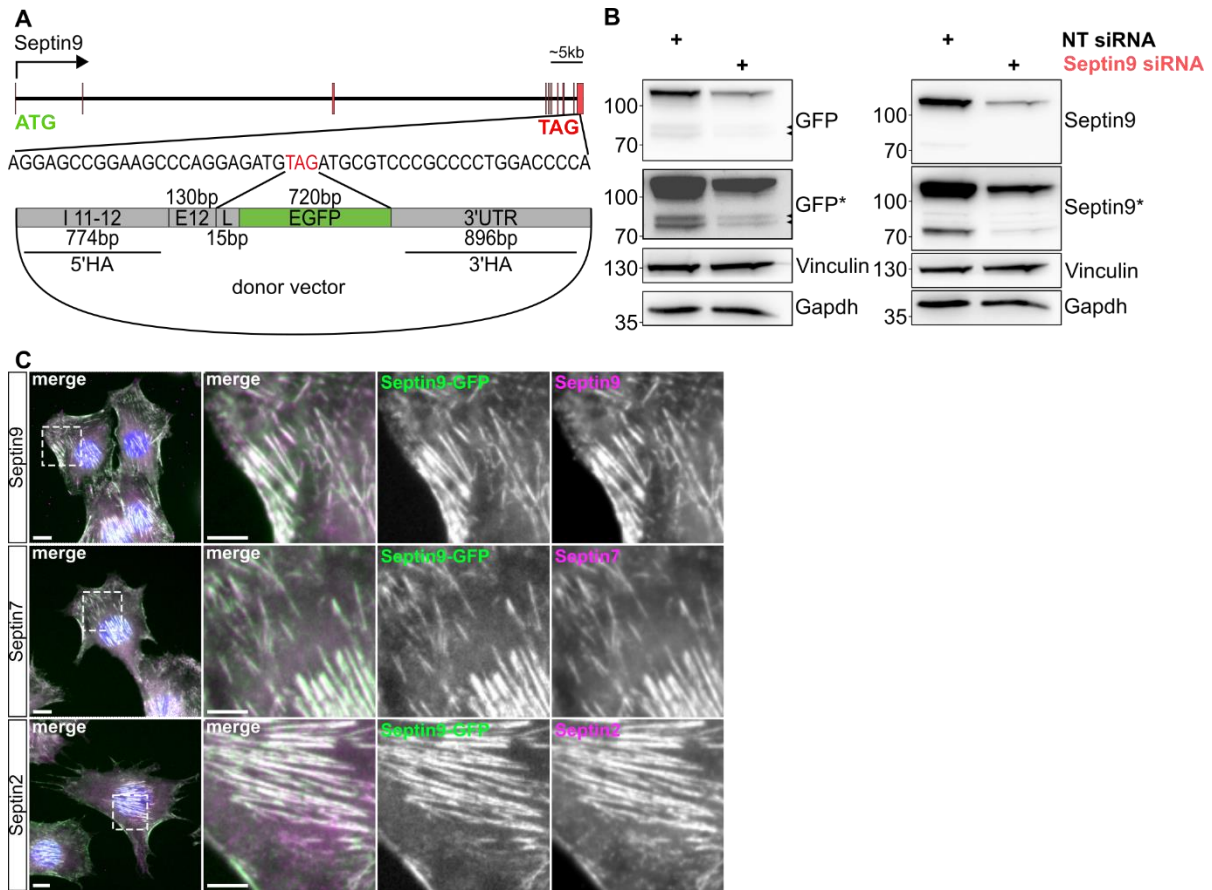


Figure S4 Generation and validation of a Septin9-GFP C2C12 cell line, Related to Figure 4.

A Cloning strategy introducing a 15bp linker (L) and an EGFP sequence instead of the stop codon of Septin9 in C2C12 cells. C-terminal strategy targets all Septin9 isoforms. Several clones were generated and tested. **B** Clone 32 was transfected with either non-targeting siRNA or Septin9 siRNA and subjected to a SDS PAGE analysis followed by a western blot. Antibodies against long Septin9 isoforms (run above 70kDa) and GFP were used to validate the knock-in. The molecular weight shift of about 30kDa (GFP) is visible with both antibodies. Probing the membrane with anti-GFP antibody might reveal the expression of the short isoforms (black arrow heads). Lanes marked with an asterisk (GFP* and Septin9*) represent longer exposure. **C** Representative epifluorescent images of proliferating Septin9-GFP C2C12 cells (clone#32) visualizing co-localization of Septin9-GFP with endogenous septin structures. Septin9-GFP shows knock-in GFP signal, Septin9, Septin7 and Septin2 are stained with antibodies. Scale bar 10 μ m, insets 5 μ m.

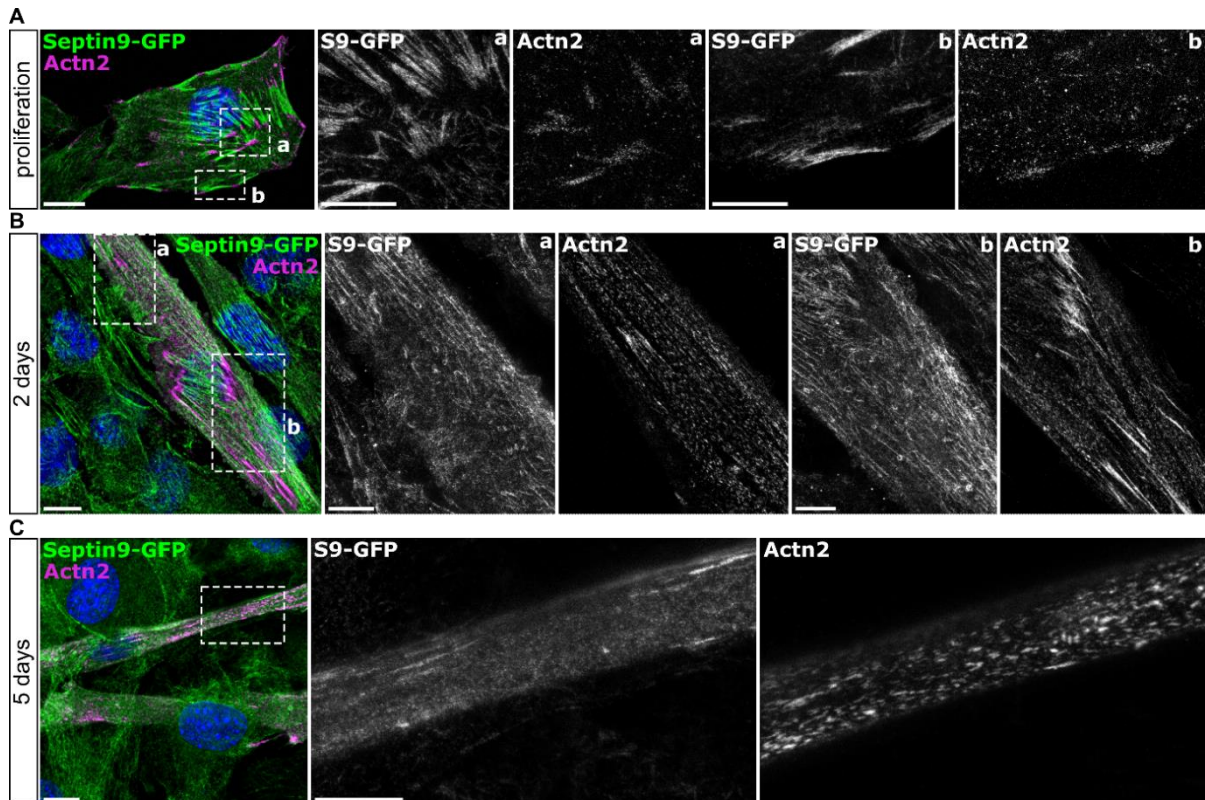


Figure S5 Septin9-GFP does not colocalize with α -actinin-2 during *in vitro* myogenesis. Related to Figure 4.

A Confocal (merge) and STED (insets) fluorescent micrographs of proliferating Septin9-GFP knock-in myoblasts. α -actinin-2 localizes to the focal adhesions and Septin9-GFP is excluded from these structures. **B** Representative images capture cytoskeletal reorganization in the nascent myotube after 2 days of differentiation. Septin9-GFP is no longer organized in long, straight filaments, but in short rods and rings, showing no colocalization with α -actinin-2. **C** The mature myotube shows no colocalization between residual Septin9-GFP and α -actinin-2, that resembles primitive sarcomeres. Scale bars 10 μ m, insets in cells 5 μ m.

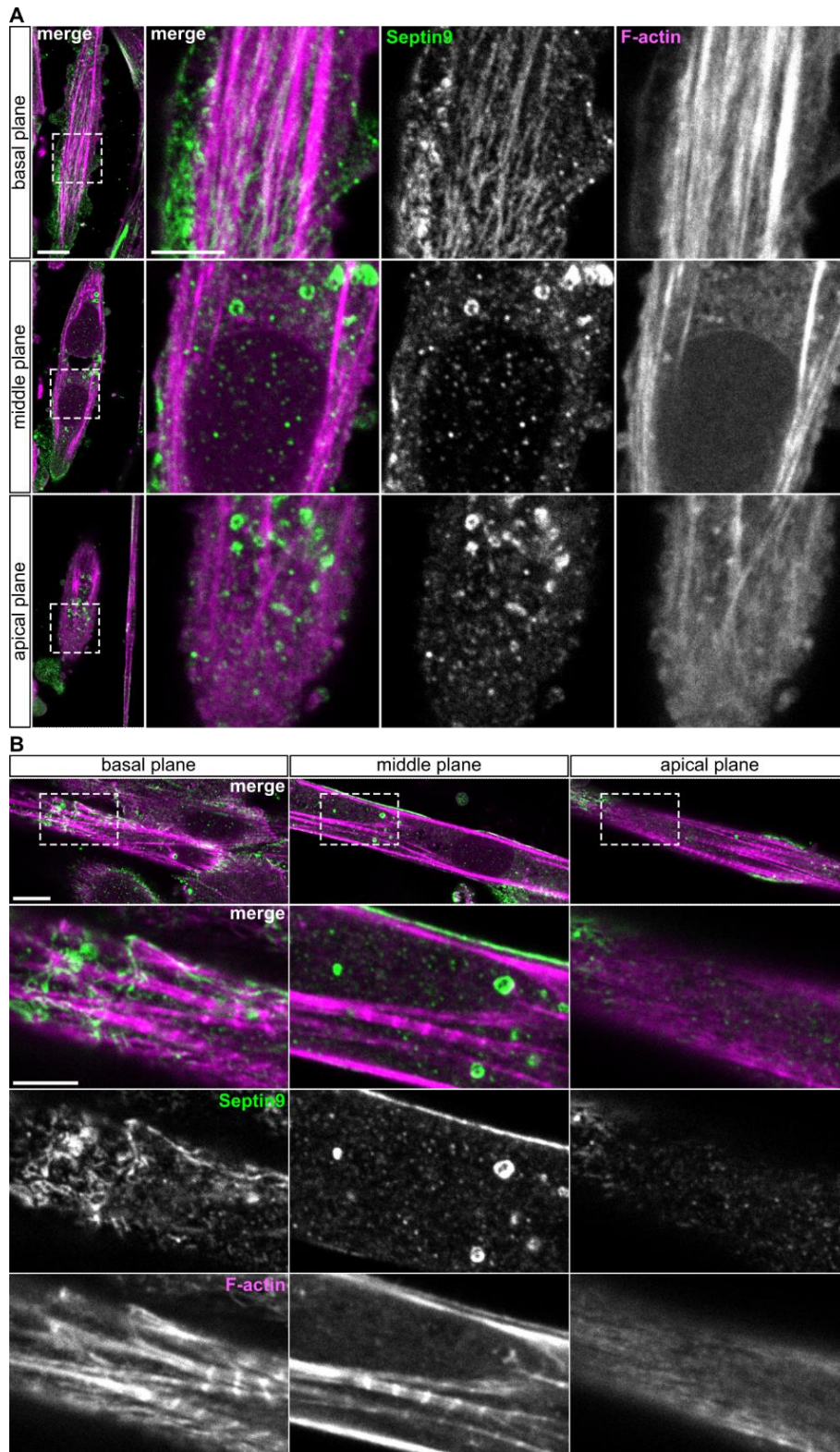


Figure S6 Sub-cellular distribution of native Septin9 in a myotube, Related to Figure 4.

Confocal images showing sub-cellular distribution of endogenous Septin9 after prolonged differentiation for 5 days. **A** Representative confocal images of a nascent myotube without apparent sarcomeric structures visualized with phalloidin. Increments in $6\mu\text{m}$ steps at the bottom, middle ($6\mu\text{m}$) and upper ($12\mu\text{m}$) plane of the wild type C2C12 cells. **B** Representative confocal images of a mature myotube with sarcomeres visualized with phalloidin. The bottom, middle ($5\mu\text{m}$) and upper ($11\mu\text{m}$) plane in of the wild type C2C12 cells. Magnified insets show Septin9 and Actin (phalloidin) organization in the boxed area of the myotube. Scale bar $10\mu\text{m}$, inset $5\mu\text{m}$.

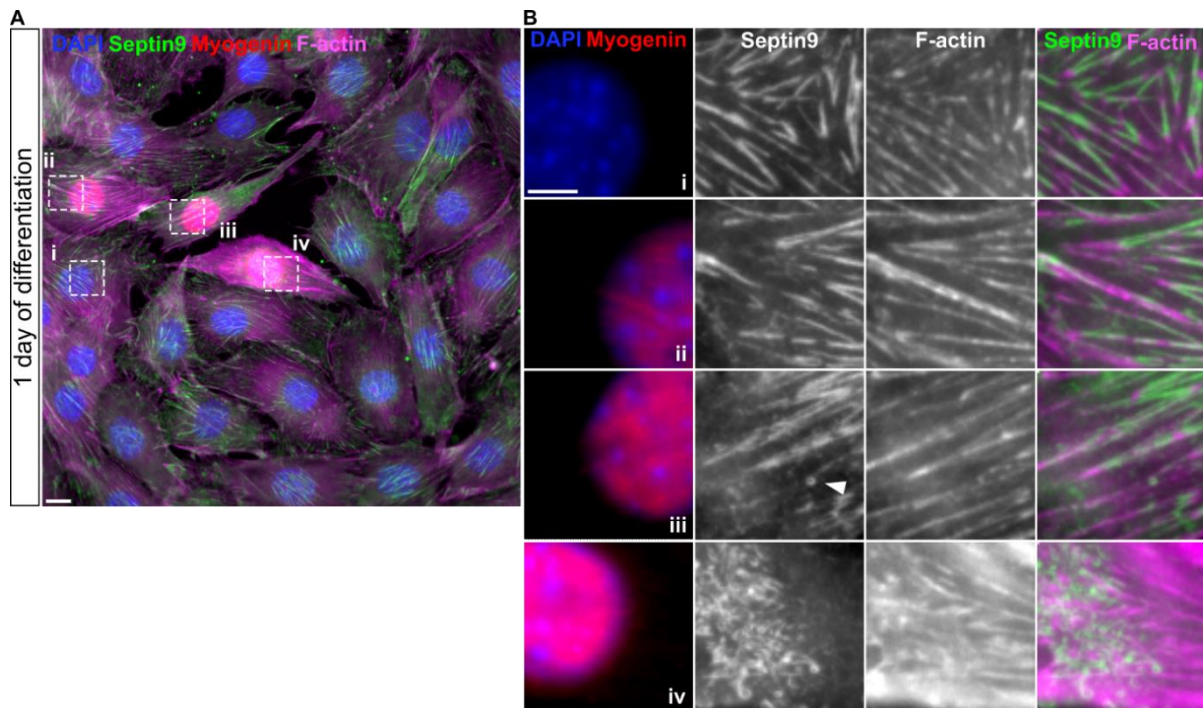


Figure S7 Septin9 reorganization is subsequent to Myogenin expression, Related to Figure 4.

A Representative epifluorescence image of wild type C2C12 cells differentiated for 1 day, visualizing Septin9, Myogenin and F-Actin via phalloidin. **B** Magnified insets from A show Septin9 organization in Myogenin-negative (inset i) and Myogenin positive cells (insets ii-iv). Myogenin-positive cells that display increased actin levels (inset iv) show the strongest Septin9 reorganization. White arrowhead shows a septin ring. Scale bar 10 μm , inset 5 μm .

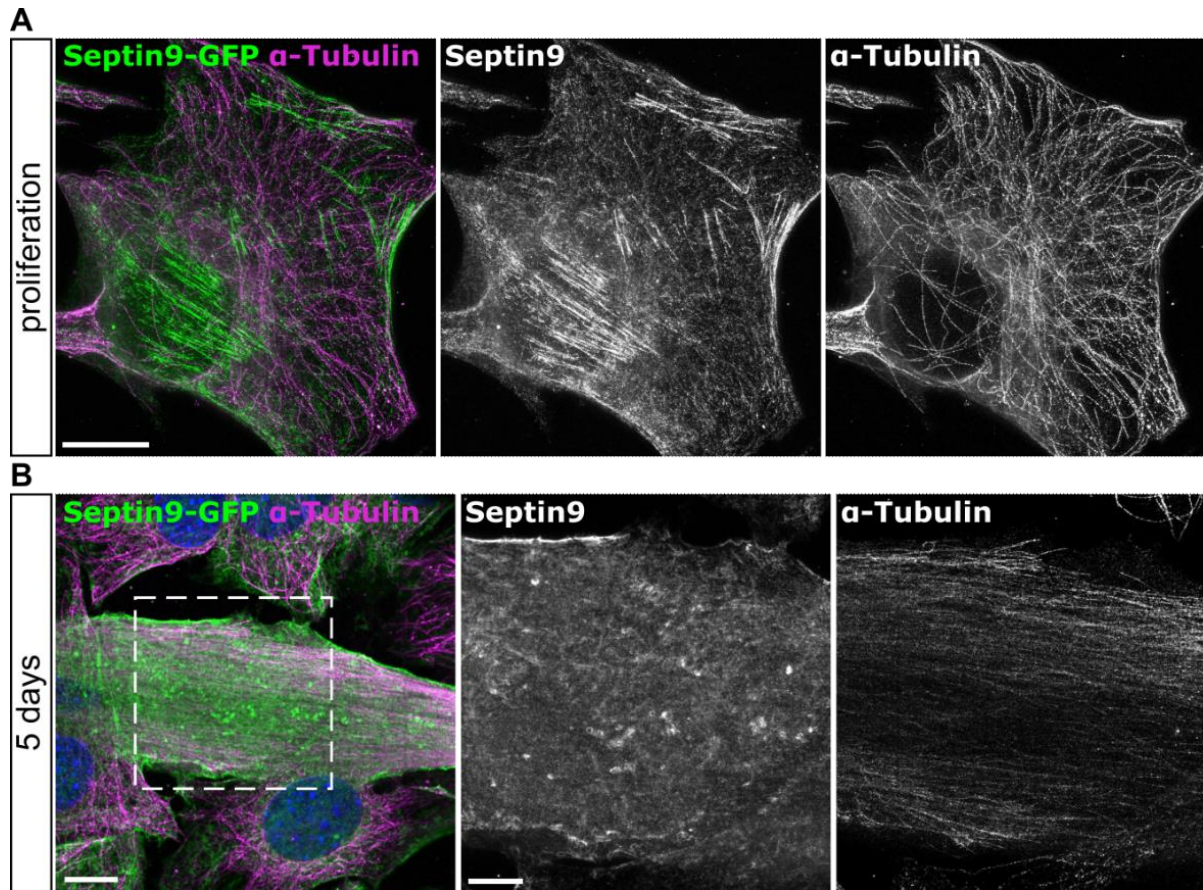


Figure S8 Microtubules in C2C12 cells are mostly devoid of Septin9, Related to Figure 4.

A Confocal (merge) and STED (insets) fluorescent micrographs of proliferating and differentiating for 5 days **B** Septin9-GFP knock-in myoblasts. Microtubules are visualized using α -Tubulin antibody. Septin9 appears to colocalize only occasionally at the cell periphery. Scale bars 10 μ m, insets in cells 5 μ m.

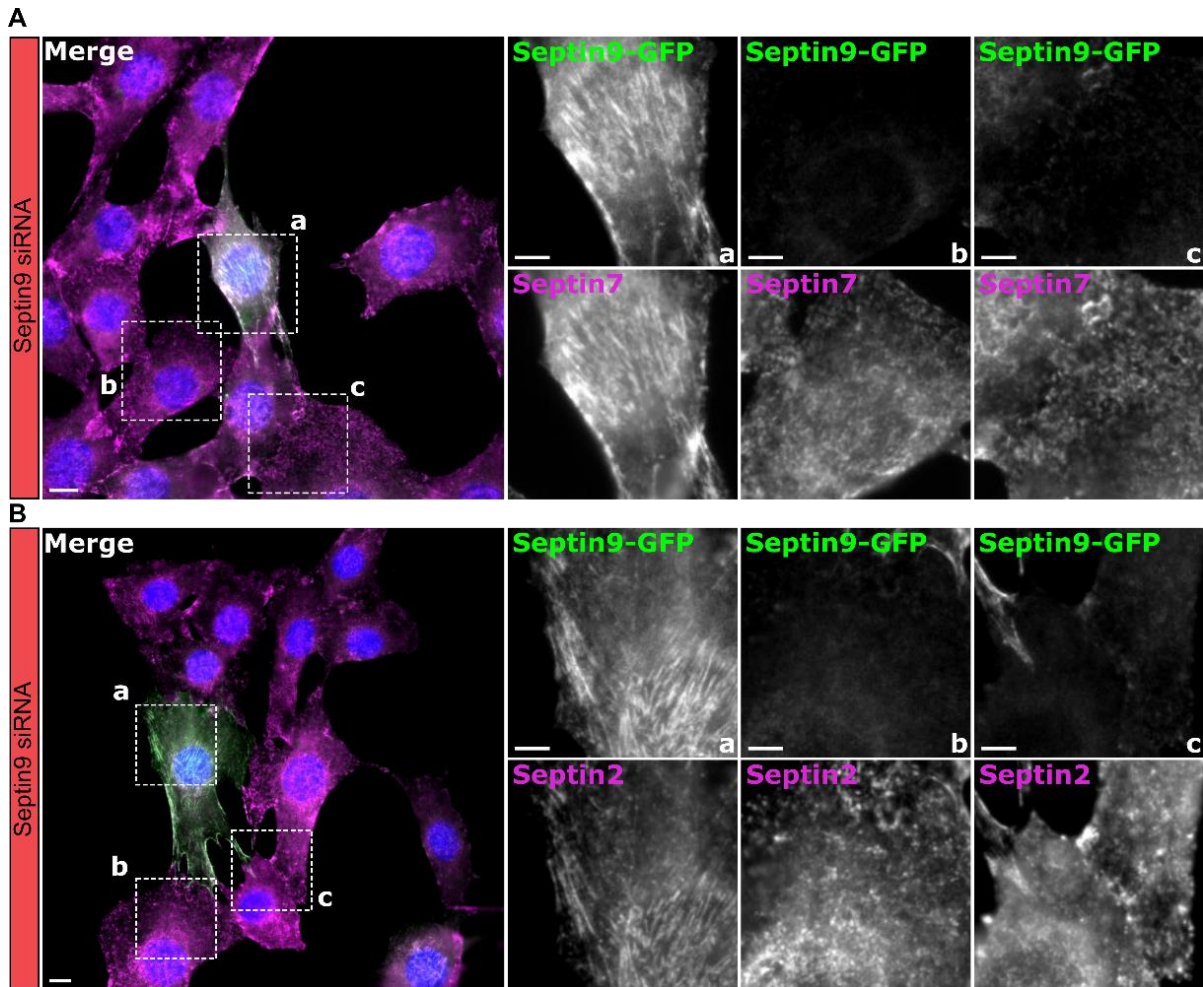


Figure S9 Integrity of septin filaments in absence of Septin9 in Septin9-GFP C2C12 cell line, Related to Figure 3.

A-B Representative epifluorescent images of proliferating Septin9-GFP clone#32 48 hours after siSeptin9 treatment visualizing Septin7 (**A**) and Septin2 (**B**). Septin9-GFP shows knock-in GFP signal, Septin7 and Septin2 are stained with antibodies. Scale bar 10 μm , insets 5 μm .

Supplementary Tables

Table S1 Primers for RT-qPCR, related to STAR Methods

	Forward	Reverse
<i>Myog</i>	CCAAGGTCTCCTGTGCTGATG	TTGGCAAAACCACACAATGC
<i>Myh2</i>	GGAGGCTGAGGAACAATCCA	GTCATTCCACAGCATCGGGA
<i>Myh8</i>	AATGATGTTTCACAGCTGCAGAG	CCATCATGGCGGCATCAGTA
<i>Rna18s1</i>	CGGCTACCACATCCAAGGAA	GCTGGAATTACCGCGGCT
<i>Septin2</i>	GCCTGTCATTGCGAAAGCTG	GGGATACTGGCCTTGAGGAG
<i>Septin4</i>	CATCGTGGAAGTGGAAAACCC	GACAGCAGGGATAGGGAAGTC
<i>Septin5</i>	CAAGCAGTACGTTGGCTTCG	GTTGATGCGTTCCTCAGCAC
<i>Septin6</i>	CCCACAGGACATTGCTTA	AGGTGGGCATTCATGGTTCC
<i>Septin7</i>	GCCAACCTCCCAAATCAAGTG	ACTTTGGATTGCTCCACCTGT
<i>Septin8</i>	AGAGCGAGCTCCACAAGTTC	CAAAGGCAGGTGTGCGTTC
<i>Septin9</i>	GGCTATGTGGGGATCGACTC	CGGCTGATTTTGGACTTGAA
<i>Septin10</i>	GAGATAAAAGAGCATCCCCGC	AATCCAGTCTCCCCACAC
<i>Septin11</i>	AGCCAGTGTGGTAAAGAGC	GTTTCGCAGCTTTCATTAC
<i>Wwtr1</i>	TCCCCACAACCTCCAGAAGAC	CAAAGTCCCGAGGTCAACAT
<i>Fst</i>	CTCTTCAAGTGGATGATTTTC	ACAGTAGGCATTATTGGTCTG

Supplemental References

- [1] Oprescu, S.N., Yue, F., Qiu, J., Brito, L.F., and Kuang, S. (2020). Temporal Dynamics and Heterogeneity of Cell Populations during Skeletal Muscle Regeneration. *iScience* 23. <https://doi.org/10.1016/j.isci.2020.100993>.