

Supporting Table 1. PVY dependent HR in *N. benthamiana* plants expressing *contig 630*

Virus	<i>A. tumefaciens/c630</i>	<i>A. tumefaciens/ -</i>
O (LW)	++	–
N (Ny)	++	–
N-Wilga	++	–
NTN (Slovenia)	++	–
PVX (O strain)	–	–
TMV (U1)	–	–

The fully developed leaves of *N. benthamiana* plants were inoculated with a suspension of *A. tumefaciens* carrying pICSLUS0003-*c630*, or with *A. tumefaciens* without any plasmid as a control. Two weeks before the *Agrobacterium* infiltration the plants were infected with four different isolates of PVY(O, N, N-Wilga and NTN), PVA or other viruses as a control. Description: -no HR, +/- -weak HR, + HR, ++ strong HR (72 hpi). Experiment was repeated three times.

Supporting Table 2. ELISA titers of PVY in transgenic *35S::Ry_{sto}* *S. tuberosum* cv. Maris Piper plants.

Genotype	A ₄₀₅ values ^a
Maris Piper MOCK	0,035±003
Maris Piper PVY	1,957±0,22
MP_630_A	0,016±0,017
MP_630_B	0,010±0,004
MP_630_C	0,005±0,001
MP_630_D	0,004±0,002
MP_630_E	0,009±0,003
MP_630_F	1,713±0,093
MP_630_G	0,013±0,003
MP_630_H	1,896±0,188
MP_630_I	0,030±0,029
MP_630_J	0,011±0,001
MP_630_K	0,022±0,007
MP_630_L	0,015±0,009

^aMean ELISA absorbance values ± standard errors for three plants each. The plants were tested 3 weeks after inoculation with PVY^{NTN}. ELISA readings were taken after 2 h of incubation with substrate at room temperature as described by Syller (1991).

Supporting Table 3. ELISA titers of PVY in transgenic 35S::Ry_{sto} *S. tuberosum* cv. Russet Burbank plants.

Genotype	A ₄₀₅ values ^a
Russet Burbank MOCK	0,046 ±0,002
Russet Burbank PVY	2,495±0,181
RB_630_A	0,006±0,000
RB_630_B	0,005±0,001
RB_630_C	0,005±0,001
RB_630_D	0,004±0,002

^aMean ELISA absorbance values ± standard errors for three plants each. The plants were tested 3 weeks after inoculation with PVY. ELISA readings were taken after 2 h of incubation with substrate at room temperature as described by Syller (1991).

Supporting Table 4. ELISA titers of PVY in grafted transgenic 35S::Ry_{sto} *S. tuberosum* cv. Maris Piper plants.

Genotype	A ₄₀₅ values ^a
Nicola* MOCK	0,046 ±0,002
Nicola* PVY	3,303±0.278
MP_630_A	0,005±0,001
MP_630_N	0,005±0,0002
MP_630_R	0,005±0,001

* PVY-susceptible potato cultivar

^aMean ELISA absorbance values ± standard errors for three plants each. The plants were tested 3 weeks after inoculation with PVY. ELISA readings were taken after 2 h of incubation with substrate at room temperature as described by Syller (1991)

Supporting Table 5. Primers used to clone candidate *Ry_{sto}* genes

Gene	Primer	Sequence
c1241	124_1_35S_F	GGCTTAAU CTTTAGCAATATTCTTCAACAATG
	124_1_35S_R	GGTTTAAU TGACTGACATATGGTTCGTATATATC
c124_2	122_1_35S_F	GGCTTAAU GAGAAGTTACCTGTTAGTCTTTAG
	124_2_35S_R	GGTTTAAU AACATATTCTTTCCATACCATTTTATGATAC
c359	359_35S_F	GGCTTAAU CTGTTAGTCTTTAGCAATATTCTTCAACAATGG
	359_35S_R	GGTTTAAU CATGTTTGGTTGAATACAAAATTTAAGAAAGAC
c516	516_35S_F	GGCTTAAU CCTCTTGCAGAAGCATTATAGATTCT
	516_35S_R	GGTTTAAU TGTCACGCTAAGGAGATTAATTTCTC
c535	535_35S_F	GGCTTAAU GGGTCTTCATTGCTGATAAAAATTCTG
	535_35S_R	GGTTTAAU GTGACTTAACACTTAATTTGAGTGTAAATATTTTC
c630	630_35S_F	GGCTTAAU GGTGCTAAGAAGACTTCATATCAG
	630_35S_R	GGTTTAAU GGATACTCAACGTATCTACCTTATTATAA
c660	660_35S_F	GGCTTAAU GTATTGACTTGTCACTTGTTAATTC
	660_35S_R	GGTTTAAU GGGATGAGGCTGAGTGCCTCAAG
c692	692_35S_F	GGCTTAAU CTGTTAGTCTTTAGCAATATTCTTCAACAATGG
	692_35S_R	GGTTTAAU CATGTTTGGTTGAATACAAAATTTAAGAAAGAA
c908	908_35S_F	GGCTTAAU CAAGGACTTTATGCATTGAACTTCATTCA
	908_35S_R	GGTTTAAU GTTCAACATGGATTCAATGTTACTCAATAC
c999	999_35S_F	GGCTTAAU ACGCCTAAATCTCCCTGAATAGTCTATT
	999_35S_R	GGTTTAAU GCAGATGAAGTAACTAATGACAATTATAGTAT
c1459	1459_35S_F	GGCTTAAU GCAGAAGCATCACAGATTCTTTTAGTC
	1459_35S_R	GGTTTAAU GACAACAACCTTTACCGTTACGTCAAAG
c630N	630_genomic_F1	GGCTTAAU CCAGGAAAACCTCGTCCATCAA
	630_genomic_R1	AGCGTACTCUT CCCCGTATGAC ATT TTC TCC
	630_genomic_F2	AGAGTACGCU AACAGCAGAATATGGCTTCC
	630_genomic_R2	GGTTTAAU GGGCCAGAGAAAAAGTTGAGCAATCATC

In bold - extension for USER cloning

Normal font - gene specific sequence

35S - primers to clone gene into expression vector under 35S promoter and OCS terminator

genomic - primers to clone gene into expression vector under the control of native regulatory elements.

Supporting Table 6. Primer sequences for quantitative RT and qRT-PCR

Gene	Primer	Sequence
124_1	qPCR_124_1_35S_F	GCGAGCGATACTGCAATCTC
	qPCR_124_1_35S_R	ATCCTCAGGAAGTCCACTCAATATG
c124_2	qPCR_124_1_35S_F	GCGAGCAAAACTGCAATATACCA
	qPCR_124_2_35S_R	GATGCAAGACAAAACCTTGAGGAATGT
c359	qPCR_359_35S_F	CGCACTTTTTAAAGATGATGAGCG
	qPCR_359_35S_R	GGTGTTTAGAGGAGCTTGCACACATC
c516	qPCR_516_35S_F	GCTCACTTTGAAGAATGGTCTG
	qPCR_516_35S_R	TCTCCTTAAGAAATTTGCAATCCC
c535	qPCR_535_35S_F	TGTCTGCGTGGAATCTCTTG
	qPCR_535_35S_R	CTACATCGAGCGTCGATAAGC
c630	qPCR_630_35S_F	ATGCTTTCAAAGACGATGAGCGGT
	qPCR_630_35S_R	ATCTTTGCATTTTTCTCATGTTGGG
c660	qPCR_660_35S_F	CTTGGCAAACGTGGATGTTAAG
	qPCR_660_35S_R	CTCGTAAGGAGGATAAACCTGATATT
c692	qPCR_692_35S_F	GCAAGTCTGACCCTGAAAGA
	qPCR_692_35S_R	GGCACAACAAGTGAAAGGATAC
c908	qPCR_908_35S_F	CTGATATCTCTTCTCTGAAGTCCA
	qPCR_908_35S_R	GTTATTCTCTCTGGTGATAAAGACCT
c1459	qPCR_1459_35S_F	GAAGAGGAGTTCTCATGGTAAG
	qPCR_1459_35S_R	GAAGACGGATTGGCATTGATCC
EF1	qpcr_EF1_F	GACAAGCGTGTTATTGAGAGG
	qPCR_EF1_R	CACAGTGCAGTAGTACTTAGTG
L23	qPCR_L23_F	AAGGATGCCGTGAAGATGT
	qPCR_L23_R	GCATCGTAGTGGAGTCAAC
Sec3	qPCR_Sec3_F	GCTTGACACGCCATATCAAT
	qPCR_Sec3_R	TGGATTTTACCACCTTCCGCA
PVY_uni	qPCR_PVY_F	CATAGGAGAACCTGAGATGCCAACT
	qPCR_PVY_R	TGGCGAGGTTCCATTTTCA
EDS1	RT_PCR_EDS1_F	CGATGAATTTAGCCCTGAATGACTTAGG
	RT_PCR_EDS1_R	TCACACTTGGTCTGGTACTCCTGTA
NRG1	RT_PCR_NRG1_F	GTTCTCTGCTCAATCTTAAGAGAATCAG
	RT_PCR_NRG1_R	GATCACATGTTTCAGCTGGTACTTCC