Supplementary Table S1. Cisgene segregation pattern in *HvGS1-1* cisgenic barley lines. A) 12 cisgenic T0 lines were analysed for segregation of the inserted cisgene in the T1 generation using PCR. The resulting ratios were used to estimate the number of cisgene inserts. Cisgene detection was expected to occur at a ratio of 3:1 for a single insert line and 15:1 for a double insert line. B) T1 lines with a single cisgene insert were analysed for cisgene segregation in the T2 generation and the resulting ratios were used to determine whether the lines were homozygous or hemizygous for the inserted cisgene. Cisgene detection was expected to occur at a ratio of 1 for a homozygous line and 3:1 for a hemizygous line.

8 A)

Genetic background T0 line		Observed s	segregation	Expected no. of cisgene neg.				
	No. of T1 samples analysed	Cisgene pos.	Cisgene neg.	Ratio of cisgene pos./neg.	If single insert line	If double insert line	Evaluation	
1	18	10	8	1.25:1	4-5	1-2	Single insert	
2	41	29	12	2.42:1	10-11	2-3	Single insert	
3	40	30	10	3:1	10	2-3	Single insert	
4	41	29	12	2.42:1	10-11	2-3	Single insert	
5	41	40	1	40:1	10-11	2-3	Double/multiple insert	
6	41	33	8	4.13:1	10-11	2-3	Single insert	
7	18	18	0		4-5	1-2	Double/multiple insert	
8	18	18	0		4-5	1-2	Double/multiple insert	
9	41	41	0		10-11	2-3	Double/multiple insert	
10	18	18	0		4-5	1-2	Double/multiple insert	
11	14	11	3	3.67:1	3-4	0-1	Single insert	
12	18	18	0		4-5	1-2	Double/multiple insert	

B)

Genetic background			Observed segregation		Expected no. of cisgene neg.		
T0 line	T1 line	No. of T2 samples analysed	Cisgene pos.	Cisgene neg.	If homo- zygous	If hemi- zygous	Evaluation
2	2.1	14	11	3	0	3-4	Hemizygous
2	2.2	18	18	0	0	4-5	Homozygous
3	3.2	18	18	0	0	4-5	Homozygous
3	3.4	17	13	4	0	4-5	Hemizygous
3	3.10	12	12	0	0	3	Homozygous
4	4.5	19	19	0	0	4-5	Homozygous
4	4.8	19	14	5	0	4-5	Hemizygous
11	11.1	18	18	0	0	4-5	Homozygous
11	11.2	18	15	3	0	4-5	Hemizygous
11	11.3	16	16	0	0	4	Homozygous



19 Supplementary Figure S1



Figure S1 Initial characterization of HvGS1-1 cisgenic T3 lines 2.2, 4.5, 11.1 and 11.3 together with the wild-type. (a) Grain yield, (b) harvest index and (c) nitrogen utilization efficiency (NUE) of plants at maturity growing with medium (0.45 g N L⁻¹ soil) N supply in a greenhouse experiment conducted from December 22nd 2014 to April 10th 2015. Data are presented as means \pm SE (n=20). Different letters indicate significant differences (p<0.05, Fischer LSD) between HvGS1-1 cisgenic lines and wild-type.

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Figure S2 *GS1-1* gene expression and GS1 activity in the two youngest fully developed leaves of 28–day-old plants of the two T3 *HvGS1-1* cisgenic lines 2.2 and 4.5. (a) GS1-1 gene expression with values expressed relative to wild-type plants. (b) GS1 enzyme activity. Data represents mean values \pm SE (n=4). Asterisks (*) indicates significant differences in GS1 activity between the cisgenic lines and the wild-type.



Figure S3 Separate activities of cytosolic GS (GS1) and chloroplastic GS (GS2) in leaves of 28-day-old wild-type plants. GS1 and GS2 were separated on a Mono Q 5/50 GL anion column using Fast protein liquid chromatography (FPLC). Their activities in the different elution fractions were measured by the transferase assay and the product γ -GHA (γ -glutamyl hydroxamate) was quantified spectrophotometrically at 540 nm using synthetic GHA to prepare calibration standards. The first peak corresponds to GS1 and the second peak to GS2, as confirmed by western blotting using an anti-GS antibody [GS1, lower band, 40 kDa; GS2, upper band, 45 kDa].

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105 Supplementary Figure S4



106 Schematic drawing of the structure of the *HvGS1.1* gene used for transformation.