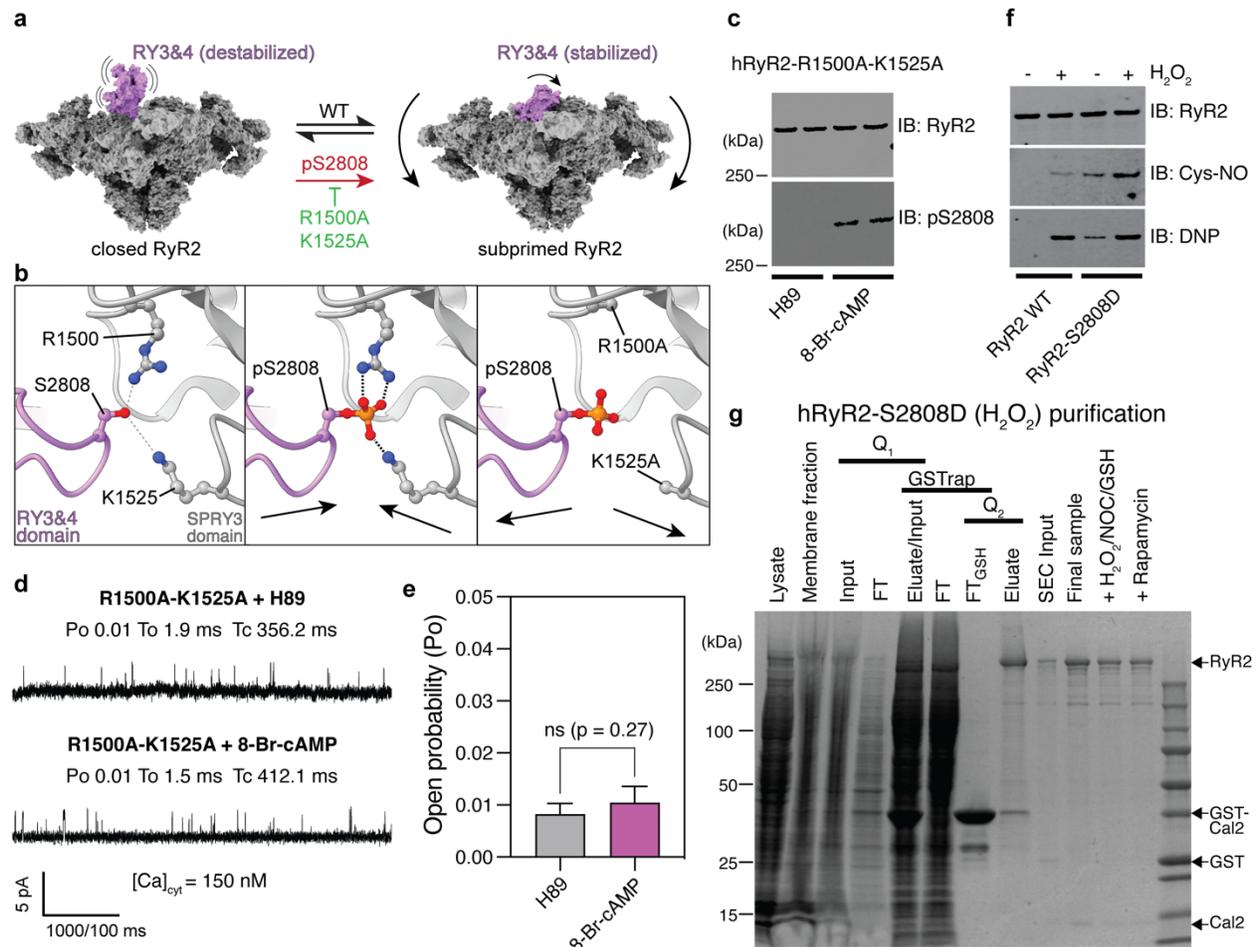


Supplementary information



Supplementary Fig. 1. Phosphorylation of residue RyR2-S2808 induces the subprimed state and post-translational modifications.

a Schematic equilibrium of the different conformations of RyR2. Panel adapted from Marco C. Miotto et al., Structural analyses of human ryanodine receptor type 2 channels reveal the mechanisms for sudden cardiac death and treatment. *Sci. Adv.* 8, eabo1272(2022). DOI:10.1126/sciadv.abo1272 under a CC BY license:

<https://creativecommons.org/licenses/by/4.0/>.¹⁰ The conformation of the destabilized RY3&4 domain (phosphorylation domain) was modelled with Coot to illustrate our hypothesis.

b Close-up view of the phosphorylation loop (residues 2803-2819 modelled with Coot) of the WT RyR2 (left), PKA-phosphorylated RyR2 (middle), and RyR2-R1500A-K1525A (right).

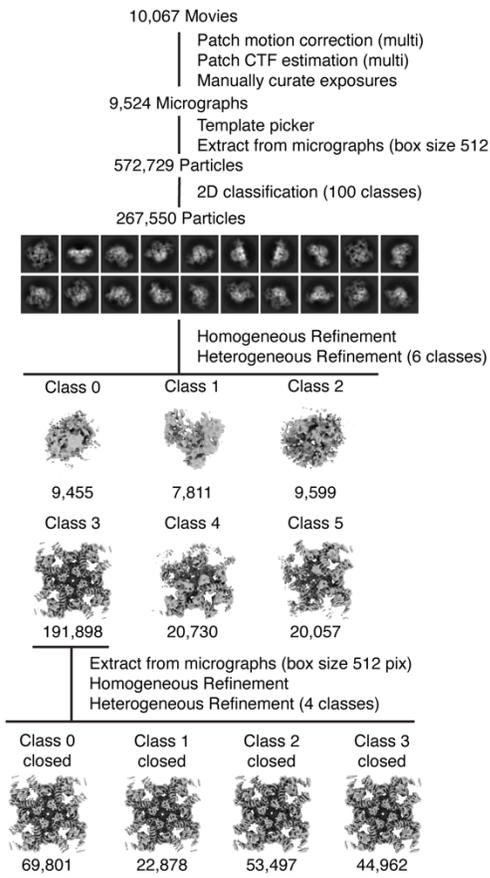
The dashed lines represent the hydrogen and ionic bonds, and the arrows represent the expected domain movements.

c Immunoblots of microsomes prepared from HEK293 expressing RyR2-R1500A-K1525A treated with either 30 μ M H89 overnight or 100 μ M 8-Br-cAMP for 30 min, showing phosphorylation of residue S2808. **d,e** PKA phosphorylation fails to increase the open probability (P_o) of single RyR2-R1500A-K1525A channels. Single-channel current recordings traces from recombinant RyR2-R1500A-K1525A at Ca^{2+} 150 nM, with H89 or 8-Br-cAMP treatment.

Opening events are shown as upward deflections. P_o : opening probability, T_o : meantime open, T_c : meantime closed. **f** Immunoblots of microsomes prepared from HEK293 expressing WT RyR2 or RyR2-S2808D treated with 1 mM H_2O_2 for 30 min, showing oxidation and nitrosylation of RyR2. **g** SDS-PAGE of human recombinant RyR2-S2808D purified from HEK293 cells treated with 1 mM H_2O_2 for 30 min.

a

RyR2-S2808D (H₂O₂)



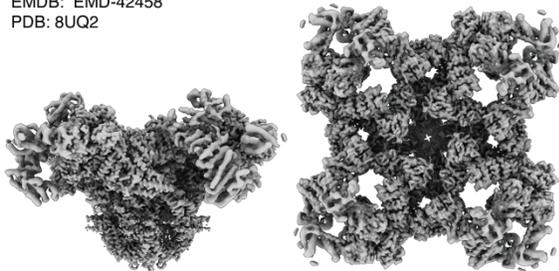
191,898 Particles

Non-uniform Refinement (2.98 Å)
Symmetry expansion (C4)

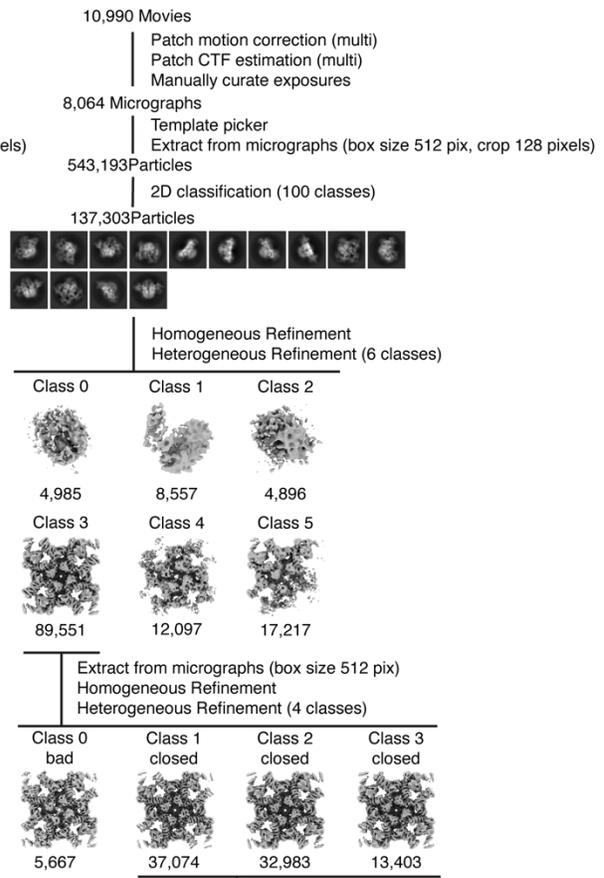
Local Refinement (big masks)
- TaF+TM+CTD (2.81 Å)
- JSol+CSol (2.79 Å)
- calstabin-2 + NTD+SPRY (2.79 Å)
- BSol (3.21 Å)

Local Refinement (small masks)
- RY1&2 (3.36 Å)
- RY3&4 (3.37 Å)
- BSol2 (3.79 Å)

Composite map: human RyR2-S2808D (H₂O₂) in the subprimed state
EMDB: EMD-42458
PDB: 8UQ2



RyR2-S2808D (H₂O₂) + ARM210



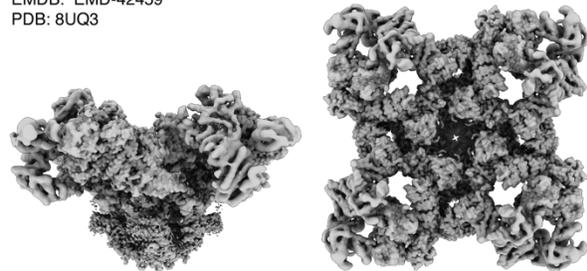
83,460 Particles

Non-uniform Refinement (3.18 Å)
Symmetry expansion (C4)

Local Refinement (big masks)
- TaF+TM+CTD (2.94 Å)
- JSol+CSol (2.94 Å)
- calstabin-2 + NTD+SPRY (2.92 Å)
- BSol (3.55 Å)

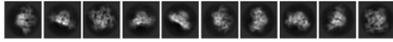
Local Refinement (small masks)
- RY1&2 (3.76 Å)
- RY3&4 (3.75 Å)
- BSol2 (4.16 Å)

Composite map: human RyR2-S2808D (H₂O₂) + ARM210 in the closed state
EMDB: EMD-42459
PDB: 8UQ3



b RyR2-S2808D (H₂O₂) + NOC-12 + GSH

13,908 Movies
 Patch motion correction (multi)
 Patch CTF estimation (multi)
 Manually curate exposures
 13,908 Micrographs
 Template picker
 Extract from micrographs (box size 512 pix, crop 128 pixels)
 889,547 Particles
 2D classification (150 classes)
 83,074 Particles



Homogeneous Refinement
 Heterogeneous Refinement (6 classes)

Class 0	Class 1	Class 2
4,540	7,566	4,257
Class 3	Class 4	Class 5
37,559	12,477	16,675

Extract from micrographs (box size 512 pix)
 Homogeneous Refinement
 Heterogeneous Refinement (4 classes)

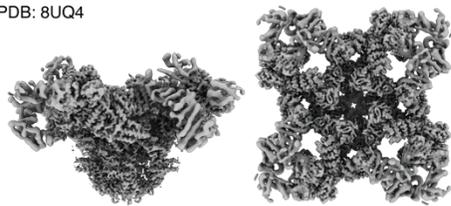
Class 0 closed	Class 1 bad	Class 2 bad	Class 3 bad
19,753	7,174	4,356	6,092

19,753 Particles
 Non-uniform Refinement (3.64 Å)
 Symmetry expansion (C4)

Local Refinement (big masks)
 - TaF+TM+CTD (3.34 Å)
 - JSol+CSol (3.28 Å)
 - calstabin-2 + NTD+SPRY (3.22 Å)
 - BSol (4.26 Å)

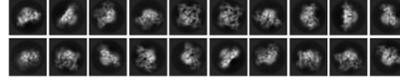
Local Refinement (small masks)
 - RY1&2 (4.39 Å)
 - RY3&4 (4.72 Å)
 - BSol2 (5.63 Å)

Composite map: human RyR2-S2808D (H₂O₂)
 + H₂O₂ + NOC-12 + GSH in the subprimed state
 EMDB: EMD-42460
 PDB: 8UQ4



RyR2-S2808D (H₂O₂) + Rapamycin

18,491 Movies
 Patch motion correction (multi)
 Patch CTF estimation (multi)
 Manually curate exposures
 16,985 Micrographs
 Template picker
 Extract from micrographs (box size 512 pix, crop 128 pixels)
 783,191 Particles
 2D classification (100 classes)
 320,951 Particles



Homogeneous Refinement
 Heterogeneous Refinement (6 classes)

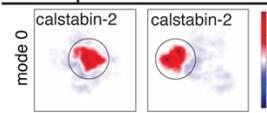
Class 0	Class 1	Class 2	Class 3	Class 4	Class 5
15,685	14,212	14,859	172,878	28,047	75,270

Extract from micrographs (box size 512 pix)
 Homogeneous Refinement
 Heterogeneous Refinement (4 classes)

Class 0 closed	Class 1 closed	Class 2 closed	Class 3 closed
38,840	24,470	73,393	35,373

172,076 Particles
 Non-uniform Refinement (3.29 Å)

Symmetry expansion (C4)
 688,304 Expanded Particles
 Local Refinement (SPRYs + NSol + calstabin-2 mask)
 3D Variability (3 modes, same mask)



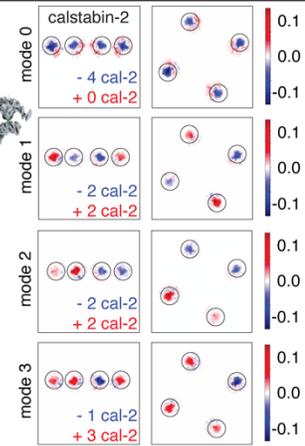
Cluster 1 & 4 (conservatively chosen)
 234,334 Expanded Particles
 (at least 34% of expanded particles contain no calstabin-2)

Local Refinement (big masks)
 - TaF+TM+CTD (3.32 Å)
 - JSol+CSol (3.11 Å)
 - NTD+SPRY (3.12 Å)
 - BSol (3.81 Å)

Local Refinement (small masks)
 - RY1&2 (4.02 Å)
 - RY3&4 (4.20 Å)
 - BSol2 (5.62 Å)

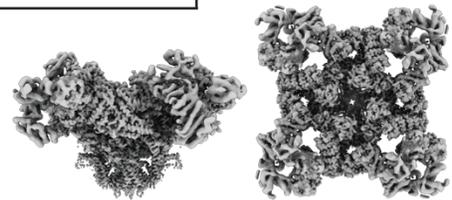
Composite map: human RyR2-S2808D (H₂O₂)
 + Rapamycin in the primed state
 EMDB: EMD-42461
 PDB: 8UQ5

Local Refinement (four calstabin-2 mask)
 3D Variability (4 modes, same mask)



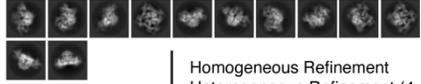
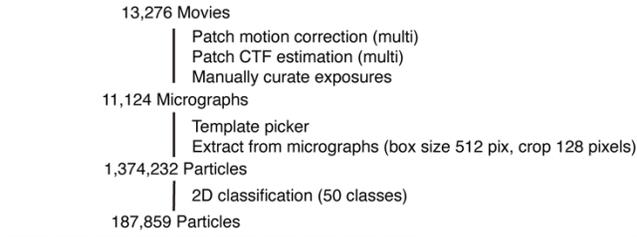
Cluster 3 (conservatively chosen)
18,232 Particles
 (at least 10% of initial particles contain no calstabin-2)

Non-uniform Refinement (3.96 Å)

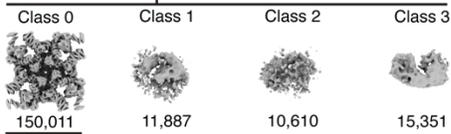


c

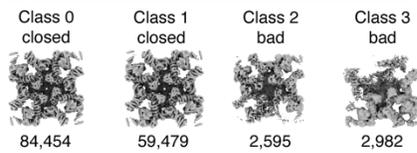
RyR2-R420Q



Homogeneous Refinement
 Heterogeneous Refinement (4 classes)



Extract from micrographs (box size 512 pix)
 Homogeneous Refinement
 Heterogeneous Refinement (4 classes)

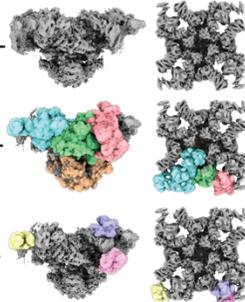


143,933 Particles

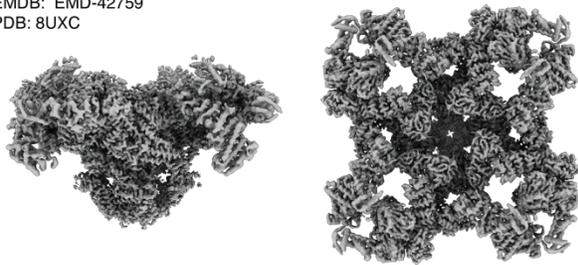
Non-uniform Refinement (2.86 Å)
 Symmetry expansion (C4)

Local Refinement (big masks)
 - TaF+TM+CTD (2.65 Å)
 - JSol+CSol (2.64 Å)
 - calstabin-2 + NTD+SPRY (2.64 Å)
 - BSol (2.91 Å)

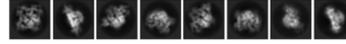
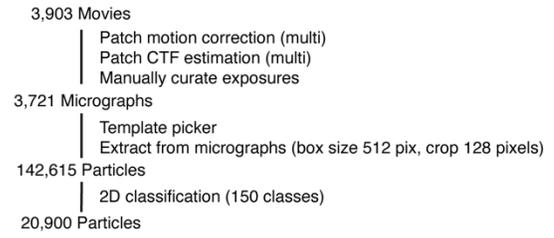
Local Refinement (small masks)
 - RY1&2 (3.11 Å)
 - RY3&4 (3.22 Å)
 - BSol2 (3.35 Å)



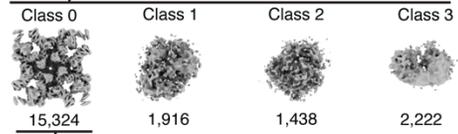
Composite map: PKA-phosphorylated human RyR2-R420Q in the primed state
 EMD: EMD-42759
 PDB: 8UXC



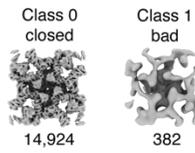
RyR2-R420Q + ARM210



Homogeneous Refinement
 Heterogeneous Refinement (4 classes)



Extract from micrographs (box size 512 pix)
 Homogeneous Refinement
 Heterogeneous Refinement (2 classes)



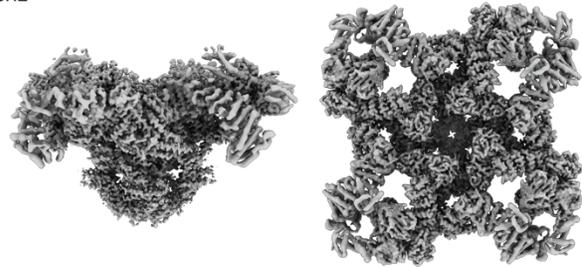
14,924 Particles

Non-uniform Refinement (3.53 Å)
 Symmetry expansion (C4)

Local Refinement (big masks)
 - TaF+TM+CTD (3.18 Å)
 - JSol+CSol (3.10 Å)
 - calstabin-2 + NTD+SPRY (3.15 Å)
 - BSol (3.76 Å)

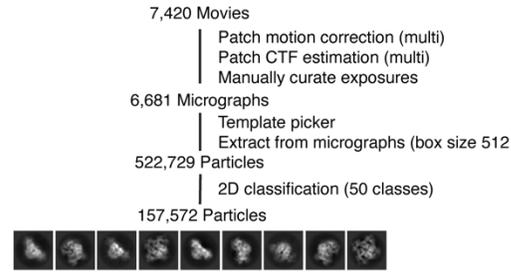
Local Refinement (small masks)
 - RY1&2 (3.95 Å)
 - RY3&4 (4.39 Å)
 - BSol2 (5.02 Å)

Composite map: PKA-phosphorylated human RyR2-R420Q+ ARM210 in the closed state
 EMD: EMD-42761
 PDB: 8UXE



d

RyR2-R420W



Homogeneous Refinement
 Heterogeneous Refinement (4 classes)

Class 0	Class 1	Class 2	Class 3
115,505	13,183	10,228	18,656

Extract from micrographs (box size 512 pix)
 Homogeneous Refinement
 Heterogeneous Refinement (4 classes)

Class 0 closed	Class 1 closed	Class 2 bad	Class 3 bad
32,235	70,243	3,877	8,625

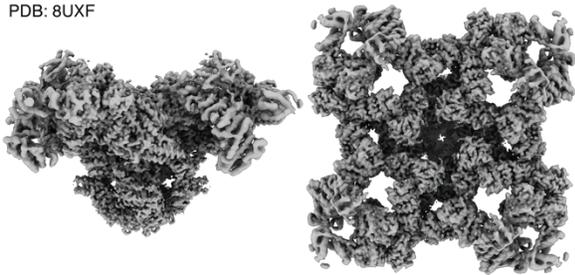
102,478 Particles

Non-uniform Refinement (3.13 Å)
 Symmetry expansion (C4)

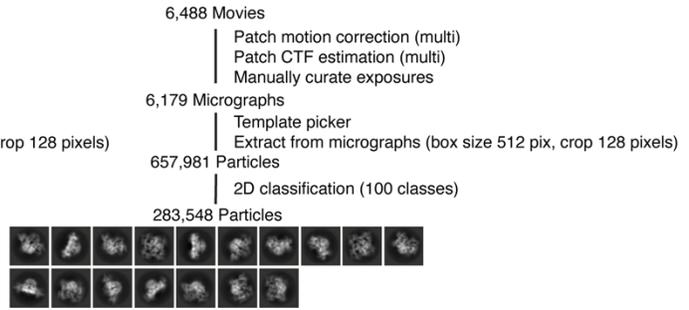
Local Refinement (big masks)
 - TaF+TM+CTD (2.91 Å)
 - JSol+CSol (2.91 Å)
 - calstabin-2 + NTD+SPRY (2.90 Å)
 - BSol (3.13 Å)

Local Refinement (small masks)
 - RY1&2 (3.26 Å)
 - RY3&4 (3.42 Å)
 - BSol2 (3.74 Å)

Composite map: PKA-phosphorlated human RyR2-R420W in the primed state
 EMDB: EMD-42762
 PDB: 8UXF



RyR2-R420W + ARM210



Homogeneous Refinement
 Heterogeneous Refinement (7 classes)

Class 0	Class 1	Class 2	Class 3
9,094	8,690	10,502	26,915
Class 4	Class 5	Class 6	
20,828	110,191	97,328	

Extract from micrographs (box size 512 pix)
 Homogeneous Refinement
 Heterogeneous Refinement (4 classes)

Class 0 closed	Class 1 closed	Class 2 closed	Class 3 bad
79,706	37,000	65,018	24,939

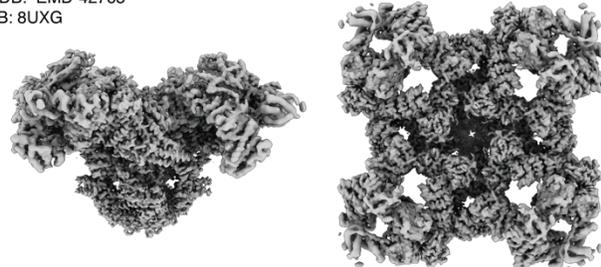
181,724 Particles

Non-uniform Refinement (3.08 Å)
 Symmetry expansion (C4)

Local Refinement (big masks)
 - TaF+TM+CTD (2.93 Å)
 - JSol+CSol (2.97 Å)
 - calstabin-2 + NTD+SPRY (2.91 Å)
 - BSol (3.24 Å)

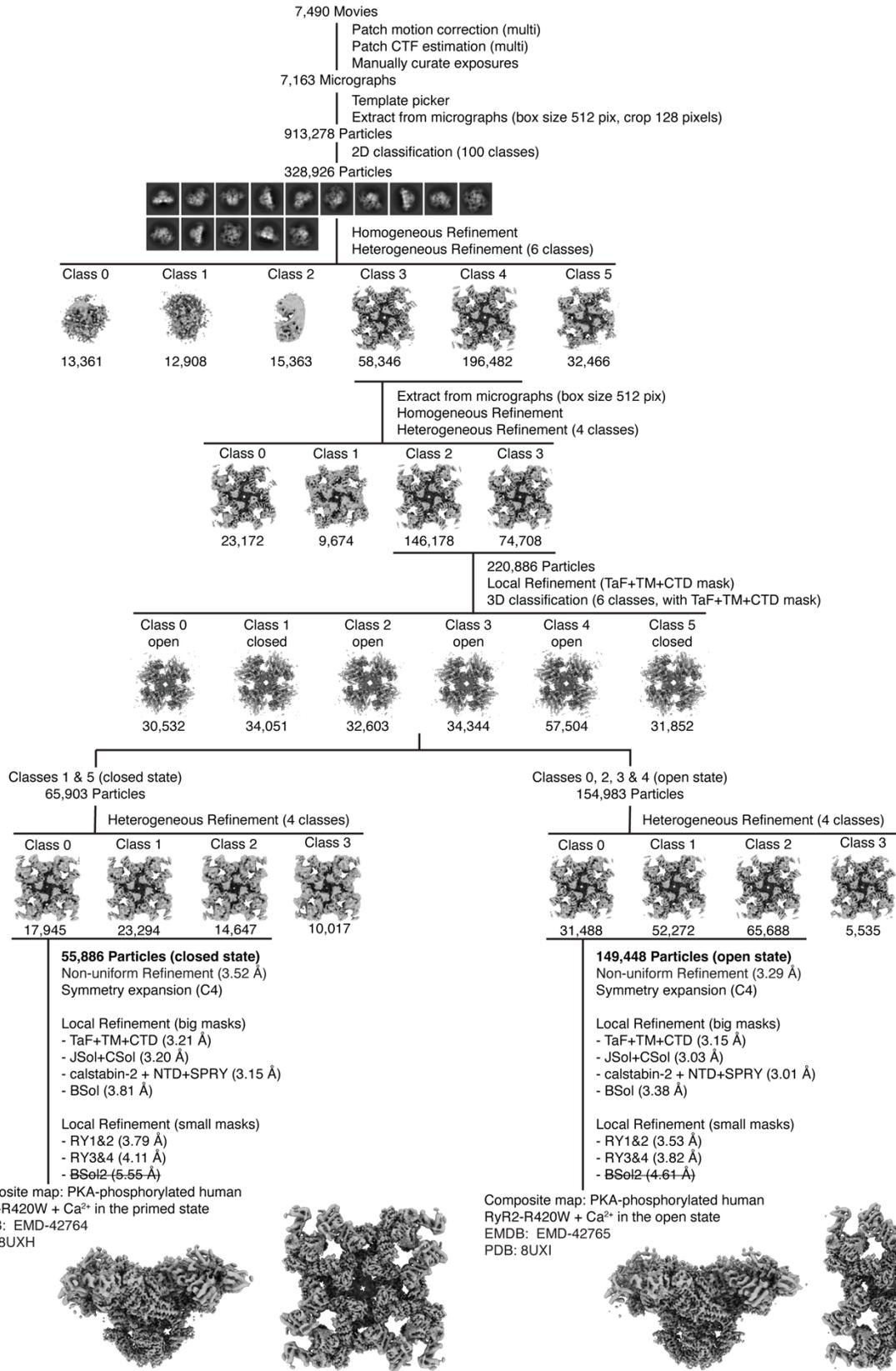
Local Refinement (small masks)
 - RY1&2 (3.38 Å)
 - RY3&4 (3.19 Å)
 - BSol2 (3.29 Å)

Composite map: PKA-phosphorlated human RyR2-R420W + ARM210 in the closed state
 EMDB: EMD-42763
 PDB: 8UXG

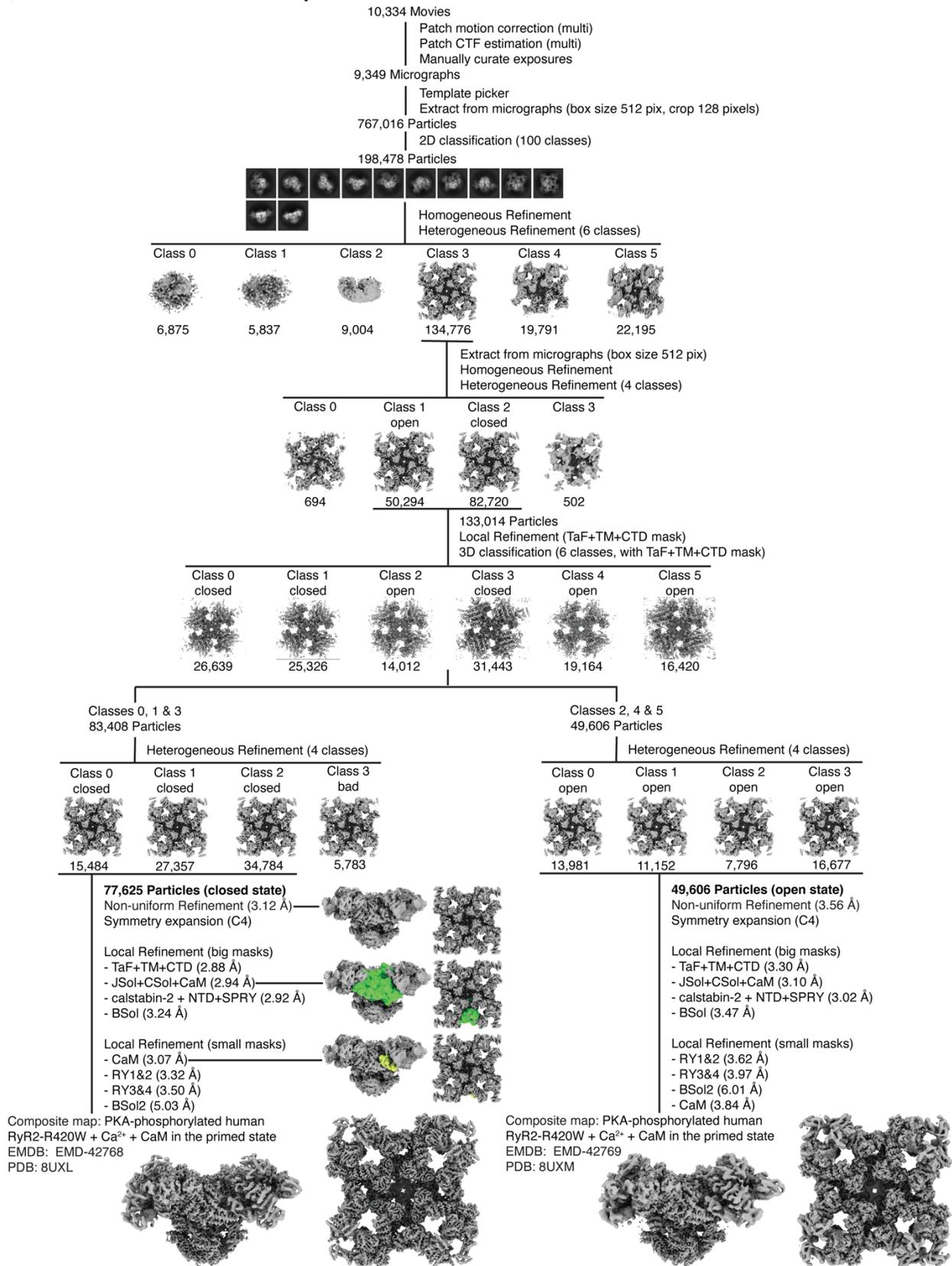


e

RyR2-R420W + Ca²⁺



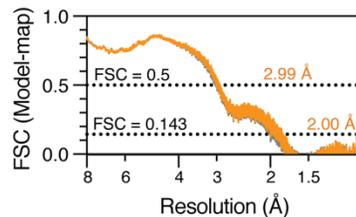
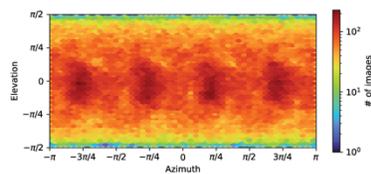
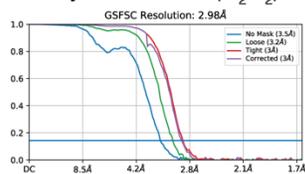
f

RyR2-R420W + Ca²⁺ + CaM

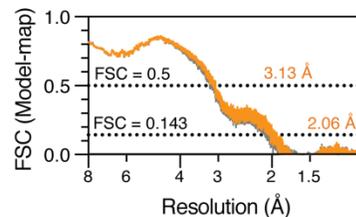
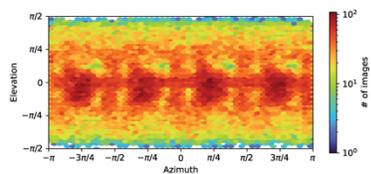
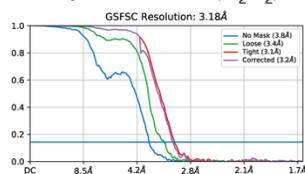
Supplementary Fig. 2. Cryo-EM data processing. a-f Entire cryoSPARC processing of the cryo-EM datasets to obtain final composite maps of RyR2-S2808D, RyR2-R420Q, and RyR2-R420W variants in the different conditions and states. The number of particles and the resolution achieved for each refinement are shown for each step. The masks used are shown only in **a** but are identical in all processing procedures, except those including calstabin-2 (**b**) and CaM (**f**).

a

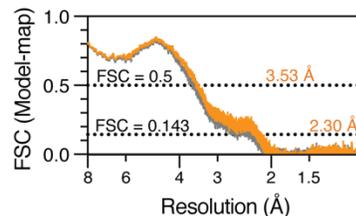
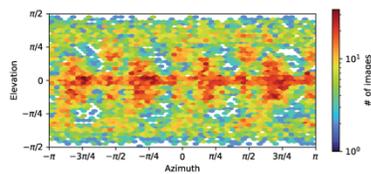
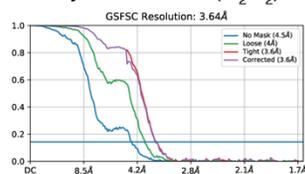
human RyR2-S2808D (H_2O_2) in the subprimed state (PDB: 8UQ2)



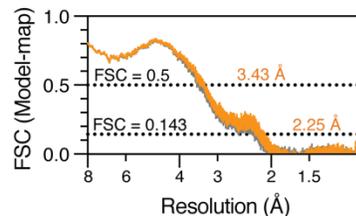
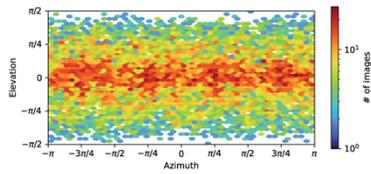
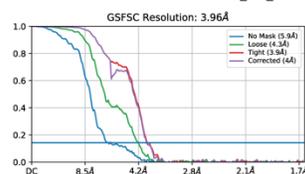
human RyR2-S2808D (H_2O_2) + ARM210 in the closed state (PDB: 8UQ3)



human RyR2-S2808D (H_2O_2) + H_2O_2 /NOC-12/GSH in the subprimed state (PDB: 8UQ4)

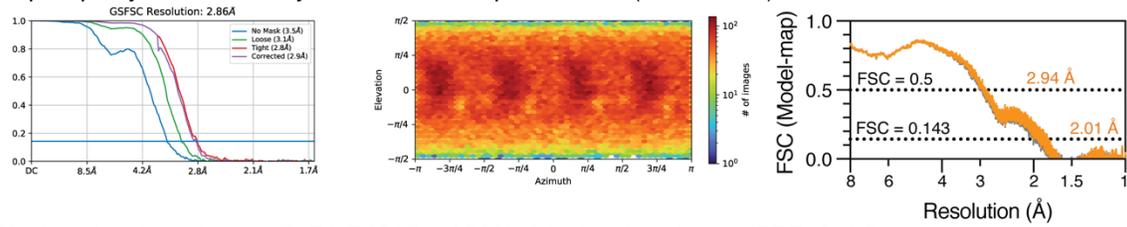


human RyR2-S2808D (H_2O_2) + H_2O_2 /NOC-12/GSH + Rapamycin in the primed state (PDB: 8UQ5)

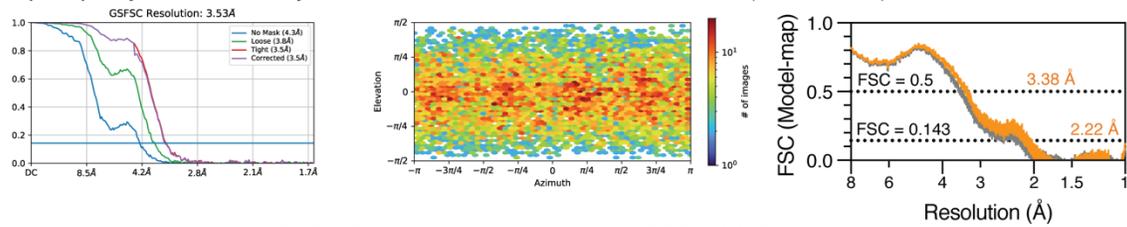


b

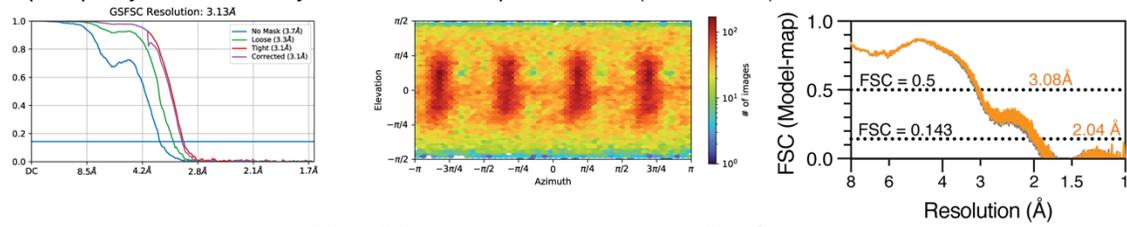
PKA-phosphorylated human RyR2-R420Q in the primed state (PDB: 8UXC)



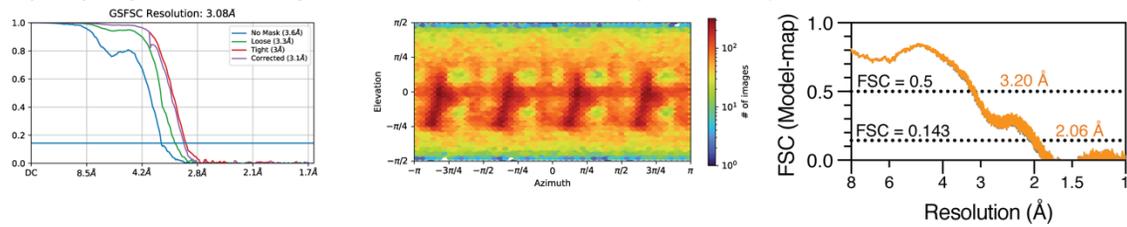
PKA-phosphorylated human RyR2-R420Q + ARM210 in the closed state (PDB: 8UXE)



PKA-phosphorylated human RyR2-R420W in the primed state (PDB: 8UXF)

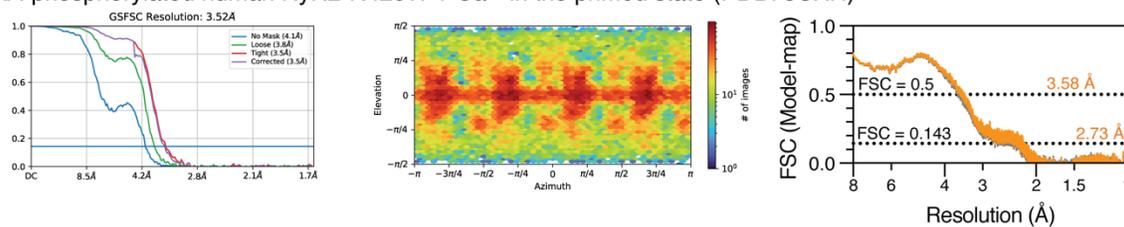


PKA-phosphorylated human RyR2-R420W in the closed state (PDB: 8UXG)

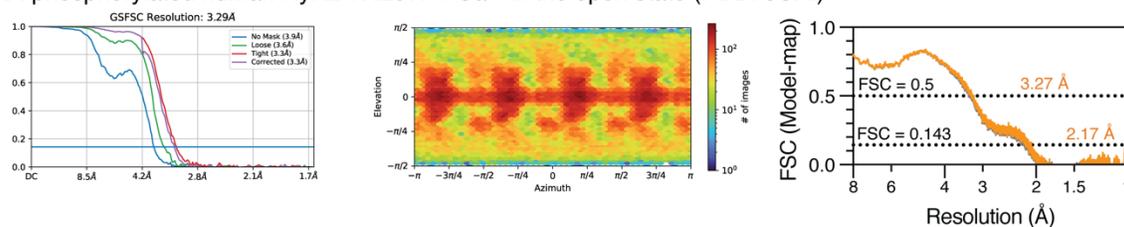


c

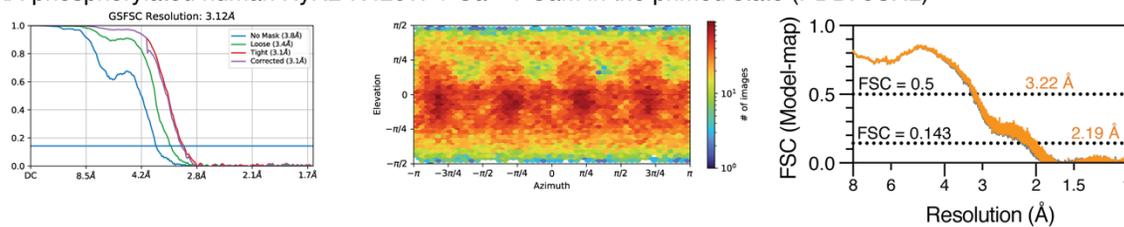
PKA-phosphorylated human RyR2-R420W + Ca²⁺ in the primed state (PDB: 8UXH)



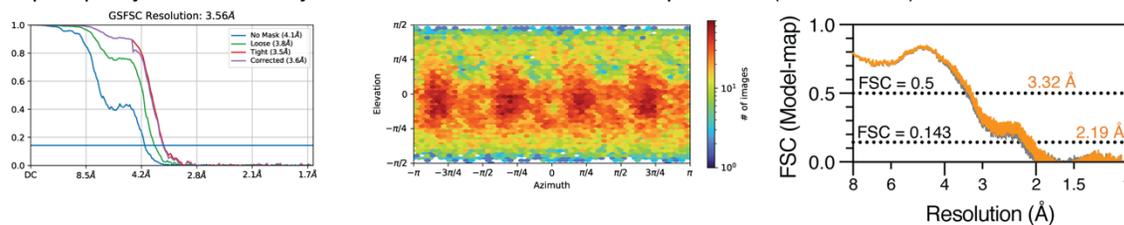
PKA-phosphorylated human RyR2-R420W + Ca²⁺ in the open state (PDB: 8UXI)



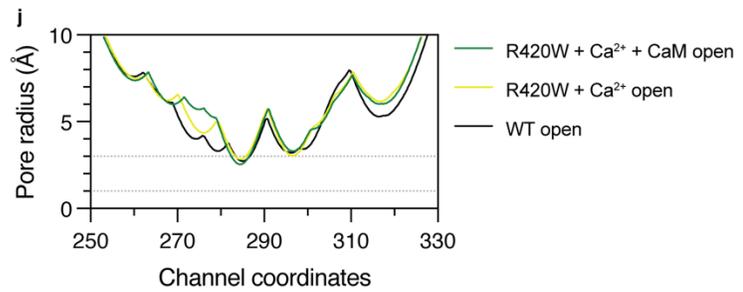
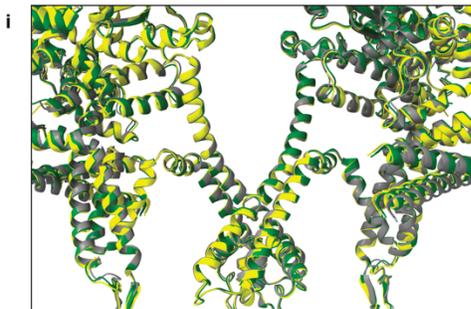
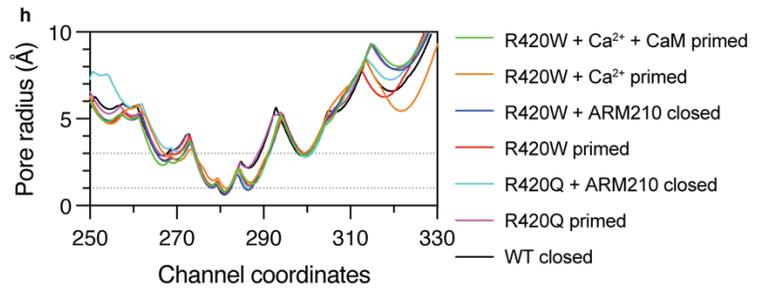
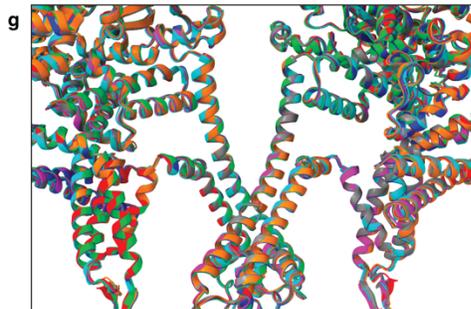
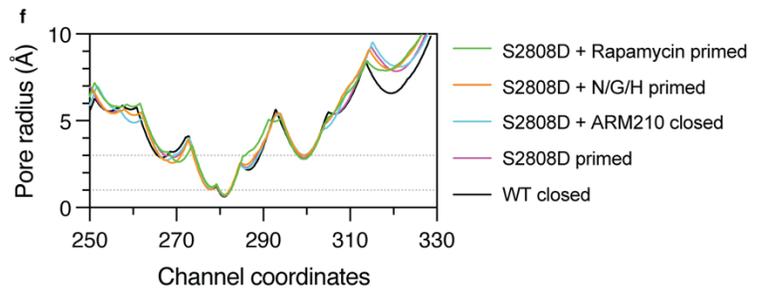
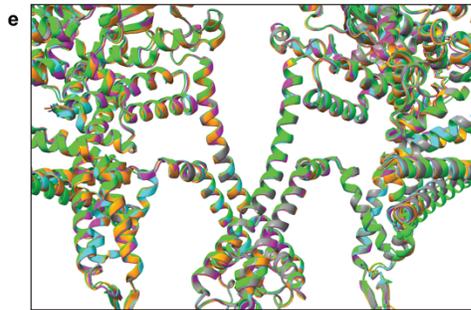
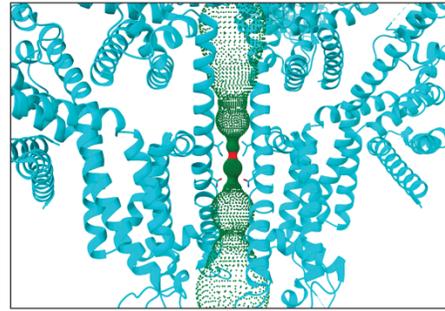
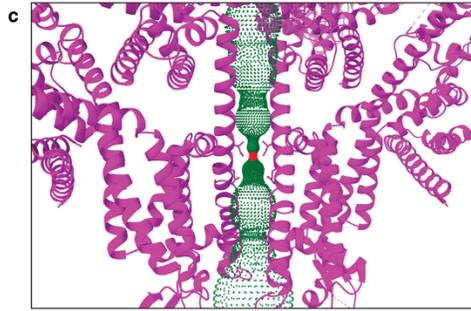
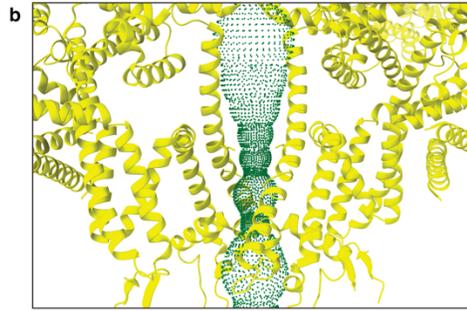
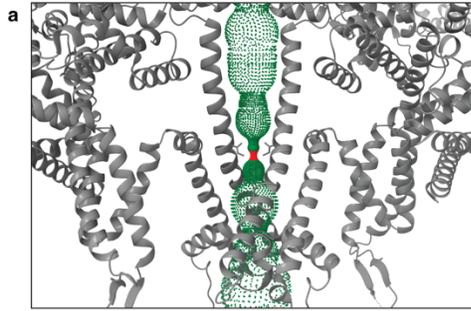
PKA-phosphorylated human RyR2-R420W + Ca²⁺ + CaM in the primed state (PDB: 8UXL)



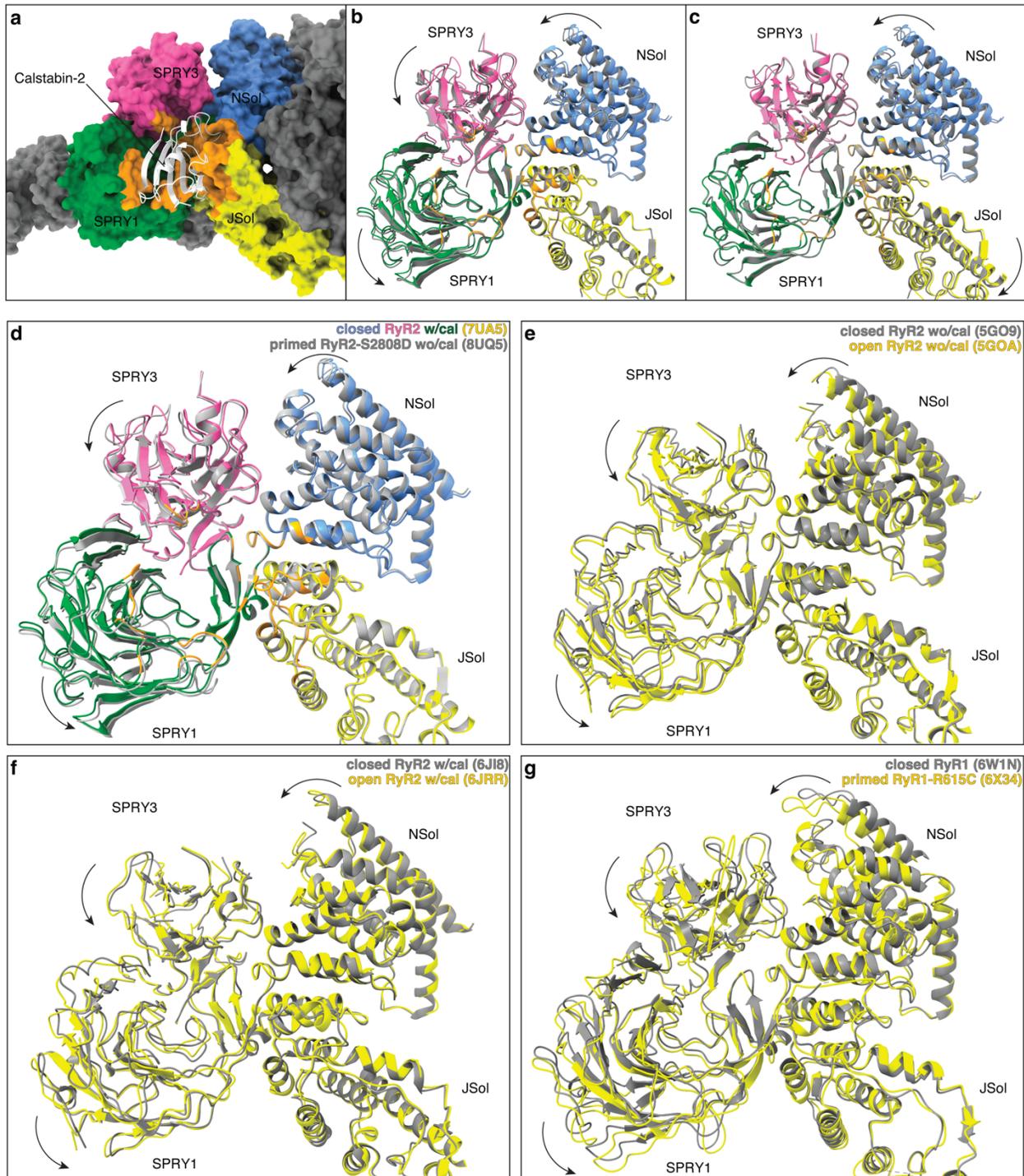
PKA-phosphorylated human RyR2-R420W + Ca²⁺ + CaM in the open state (PDB: 8UXM)



Supplementary Fig. 3. FSC curves. a-c The GSFSCs (left) and the viewing angle distributions (middle) of the global non-uniform refinement performed in cryoSPARC before any local refinement, and the FSC model-map performed in PHENIX (right) for each structure are shown.

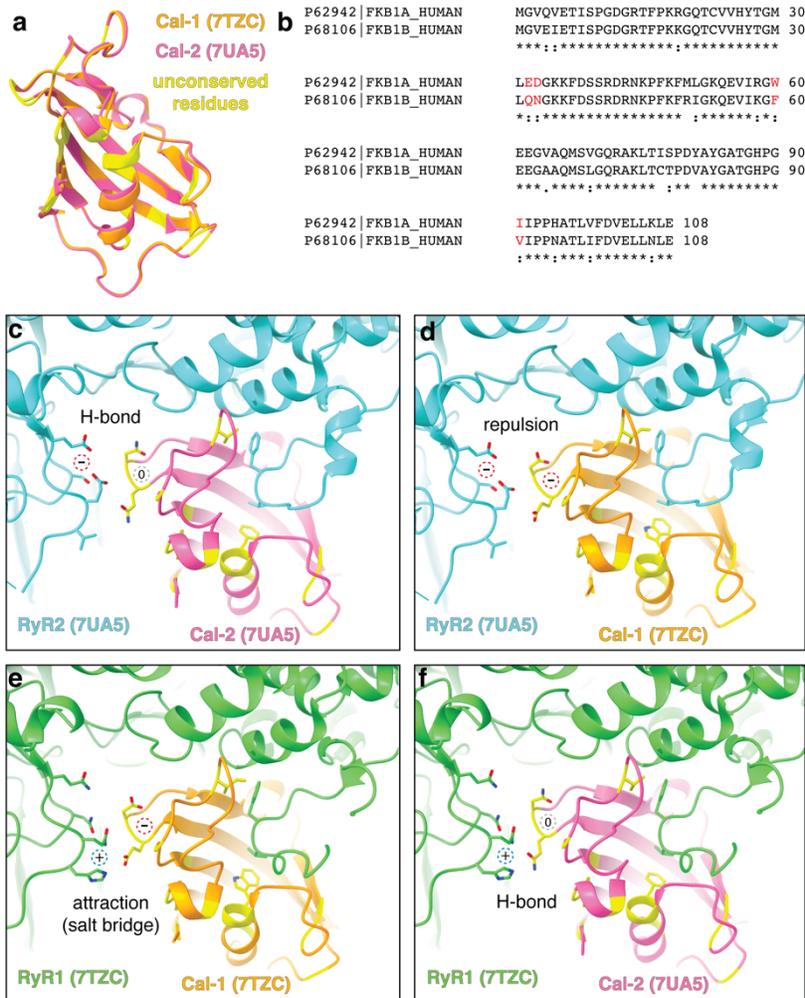


Supplementary Fig. 4. Pore analyses of all structures. a-d Atomic models overlapped with the permeation path calculated using HOLE (illustrated as green dots for diameter greater than 2 Å and red dots for diameter less than 2 Å) for closed PKA-phosphorylated WT RyR2 (**a**), open PKA-phosphorylated WT RyR2 (**b**), primed PKA-phosphorylated RyR2-R420Q (**c**), and closed PKA-phosphorylated RyR2-R420Q + ARM210 (**d**). **e** Aligned atomic models of all closed/primed structures focusing on the pore and TM domain. **f** Pore radii estimation for each structure calculated with HOLE. Channel coordinates are arbitrary. Atomic model alignment and HOLE analyses suggest that no significant differences are found in the pore of the closed states. Only the sidechain of residue Q4863 seems to fluctuate, but probably because of the weak cryo-EM density resulting from the expected dynamic behavior of the solvent-exposed polar sidechain. **g-j** same as **e,f** but with the RyR2-S2808D (**g,h**) and the RyR2-R420W open states (**i,j**).

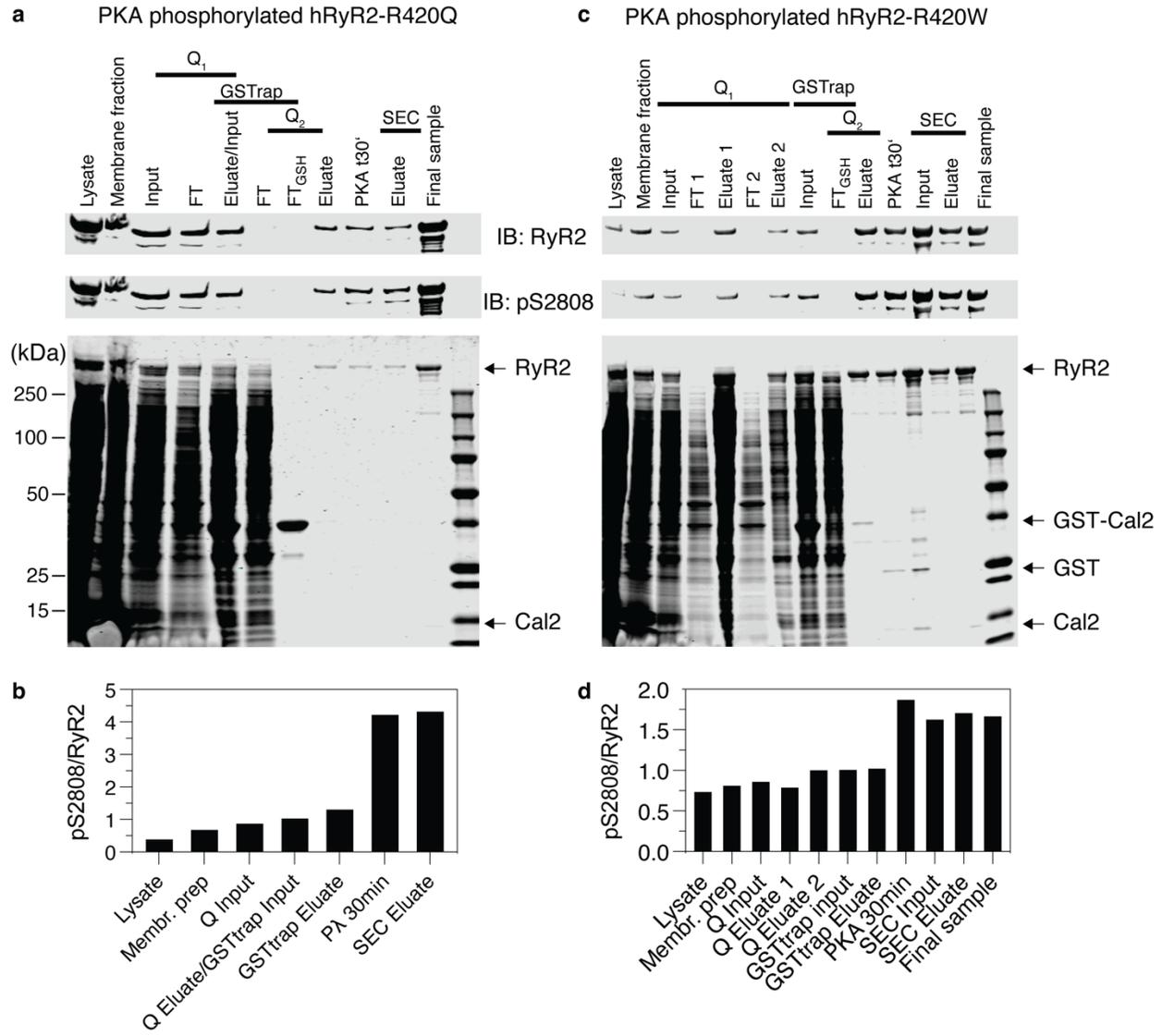


Supplementary Fig. 5. Calstabin binding site in different models. **a** Atomic model of RyR2 showing calstabin-2 (white ribbons), calstabin-2 interacting RyR2 domains (NSol in light blue, SPRY1 in green, SPRY3 in pink, and JSol in yellow), and RyR2-calstabin-2 interacting surface (orange). **b,c** Atomic models of the calstabin-2 interacting RyR2 domains in the closed state

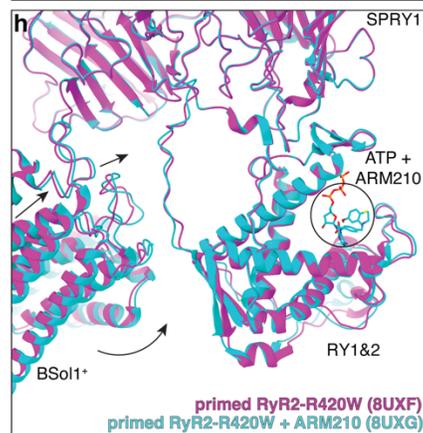
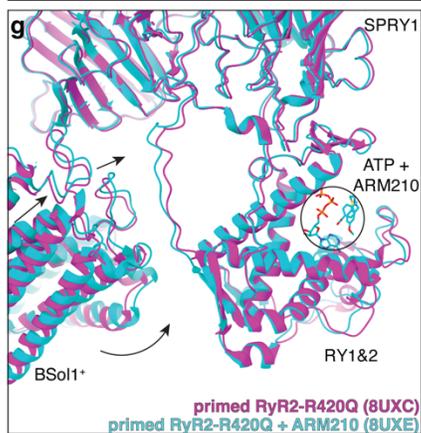
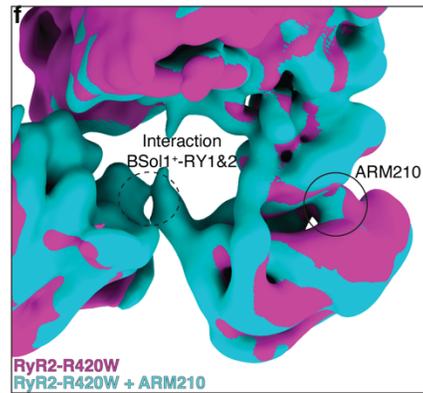
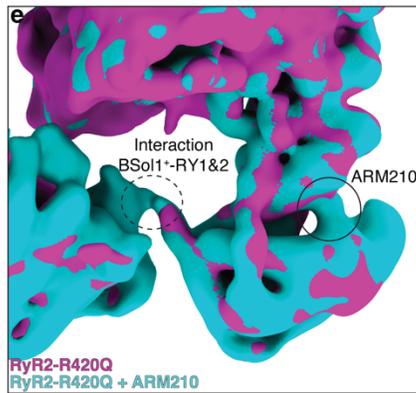
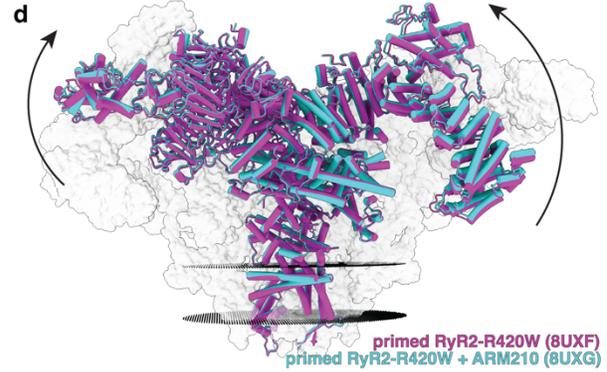
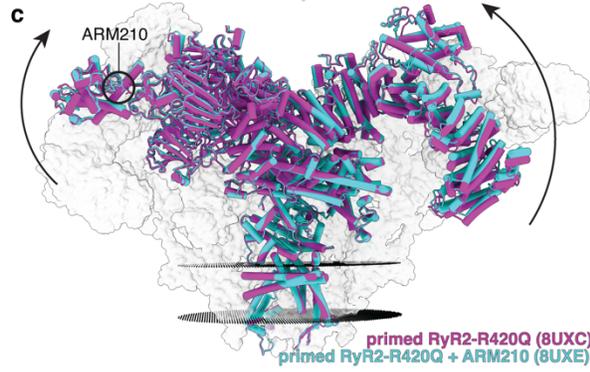
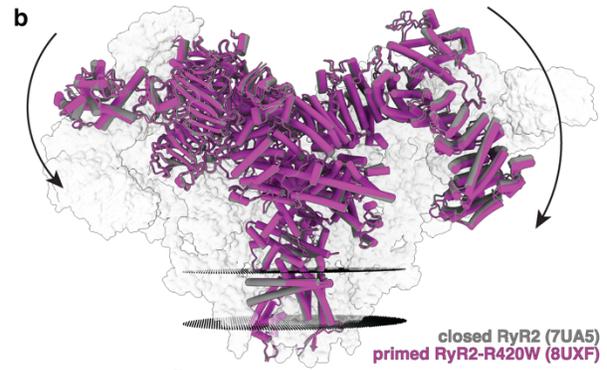
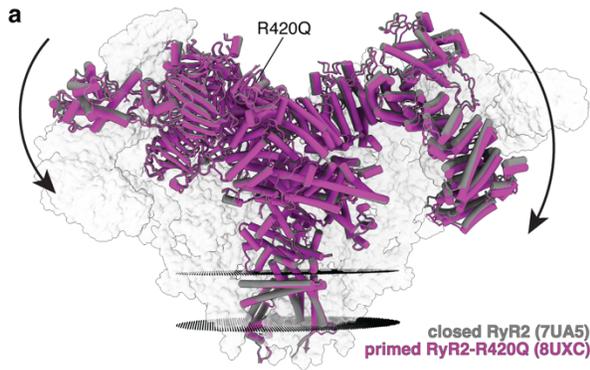
(PDB:7UA5, colors) and in the open state (PDB: 7UA9, grey). Models are aligned on the JSol (**b**) and SPRY1-3 (**c**) domains. Relative inter-domain motions are shown with arrows. **d** Atomic models of the calstabin-2 interacting RyR2 domains of human WT RyR2 in the closed state (PDB:7UA5, colors) and RyR2-S2808D in the primed state without calstabin-2 (PDB:8UQ5, grey). Models are aligned in the JSol. Relative inter-domain motions are shown with arrows. **e** Same as **d** but of pig RyR2 in the closed state without calstabin-2 (PDB:5GO9, grey) and pig RyR2 in the open state without calstabin-2 (PDB:5GOA, yellow). **f** Same as **d** but of pig RyR2 in the closed state with calstabin-2 (PDB:6JI8, grey) and pig RyR2 in the open state with calstabin-2 (PDB:6JRR, yellow). **g** Same as **d** but of pig RyR1 in the closed state (PDB:6W1N, grey) and pig RyR1-R615C in the primed state (PDB:6X34, yellow).



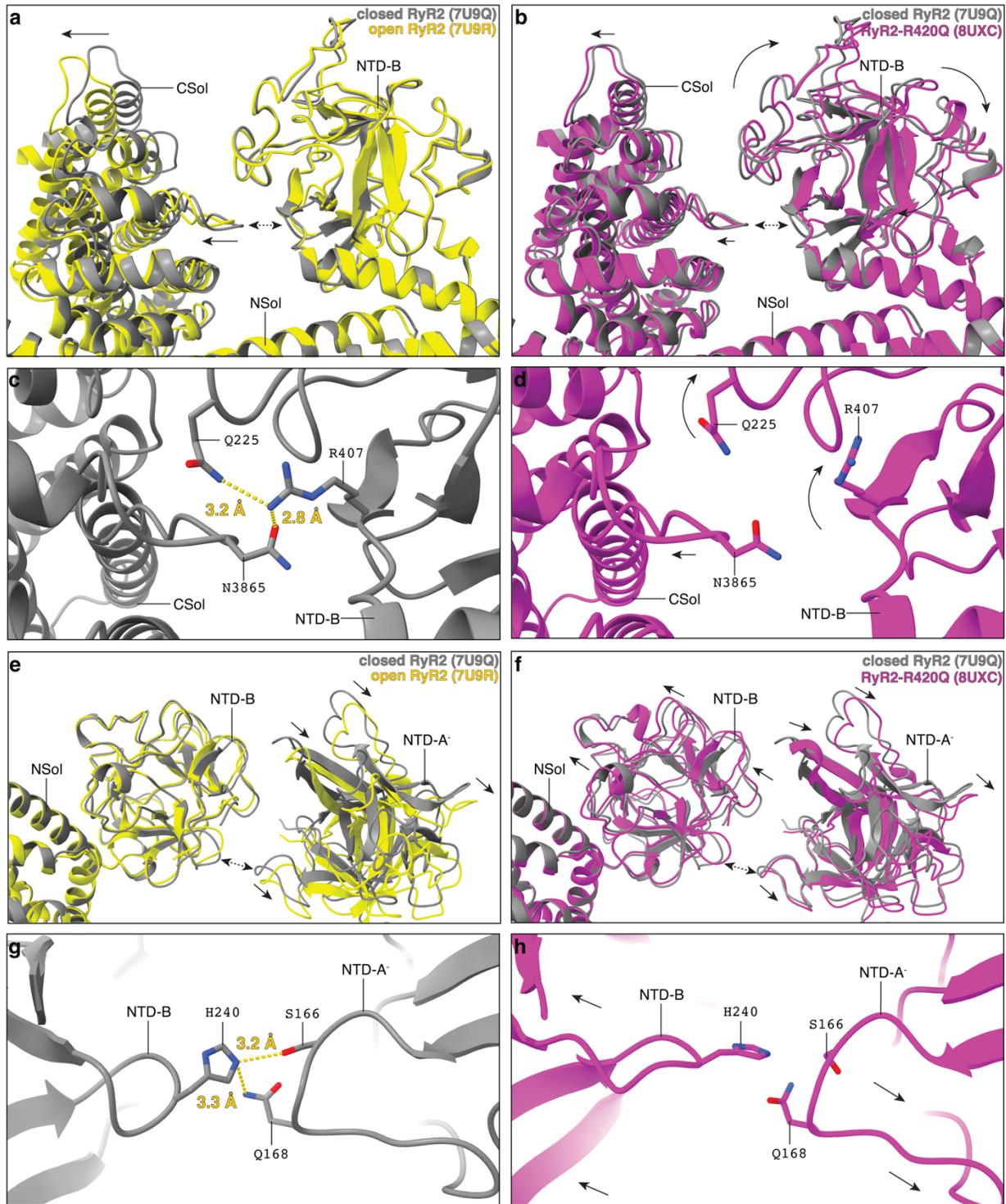
Supplementary Fig. 6. Structures of the complexes between RyRs and calstabins. a Overlapped atomic models of calstabin-1 (PDB: 7TZC, orange) and calstabin-2 (PDB: 7UA5, magenta) showing their almost identical backbone structure. Unconserved residues are colored (yellow). **b** Sequence alignment of human calstabin-1 (FKBP1A) and calstabin-2 (FKBP1B). Only the four unconserved residues that interact with RyRs are colored (red). **c-f** Atomic models of RyR2 and calstabin-2 (**c**), RyR2 and calstabin-1 (**d**), RyR1 and calstabin-1 (**e**), and RyR1 and calstabin-2 (**f**). Models are aligned on the calstabins models. Potential interaction (H-bond, repulsion, and attraction) and potential interacting residues between RyRs and calstabins are shown for RyR1 (N1296, Q1299, H1300, and N1545), RyR2 (E1309 and E1312), calstabin-1 (E32 and D33), and calstabin-2 (Q32 and N33).



Supplementary Fig. 7. Purification of recombinant human RyR2. **a,c** Immunoblot (top) and SDS-PAGE (bottom) of purification of human recombinant RyR2-R420Q (**a**) and RyR2-R420W (**c**) expressed in HEK293 cells. **b,d** Ratio of normalized intensities of the pS2808 and total RyR2 bands. After PKA phosphorylation, the intensity increases ~3-fold (**b**) and ~2-fold (**d**), reaching a saturation point.



Supplementary Fig. 8. Cryo-EM reconstructions of human RyR2 shows that the mutation-induced “primed” state is reversed by treatment with the Rycal ARM210. **a** Overlapped models of closed PKA-phosphorylated RyR2 (grey) and primed PKA-phosphorylated RyR2-R420Q (magenta). The arrows show that the cytoplasmic shell of the channel shifts downward-outward when going from the closed to the primed state. **b** Same as **a** but with primed PKA-phosphorylated RyR2-R420W (magenta). **c** Overlapped models of primed PKA-phosphorylated RyR2-R420Q (magenta) and closed PKA-phosphorylated RyR2-R420Q in the presence of ARM210 (cyan). The arrows show that the cytoplasmic shell of the channel shifts upward-inward when going from the primed to the closed state. **d** Same as **c** but with primed PKA-phosphorylated RyR2-R420W (magenta) and closed PKA-phosphorylated RyR2-R420W + ARM210 (cyan). **e** Cryo-EM maps of local refinement cryoSPRAC jobs of primed PKA-phosphorylated RyR2-R420Q (magenta), and closed PKA-phosphorylated RyR2-R420Q + ARM210 (cyan). For better interpretation, a softening gaussian was applied to the cryo-EM maps. Interaction between BSol1 and RY1&2, and ARM210 densities are labelled. **f** Same as **e** but with primed PKA-phosphorylated RyR2-R420W (magenta), and closed PKA-phosphorylated RyR2-R420W + ARM210 (cyan). **g** Aligned models of primed PKA-phosphorylated RyR2-R420Q (magenta), and closed PKA-phosphorylated RyR2-R420Q + ARM210 (cyan). Conformational changes of the RY1&2 and BSol domains are shown with arrows. ATP + ARM210 are labelled. **h** Same as **g** but with primed PKA-phosphorylated RyR2-R420W (magenta), and closed PKA-phosphorylated RyR2-R420W + ARM210 (cyan).

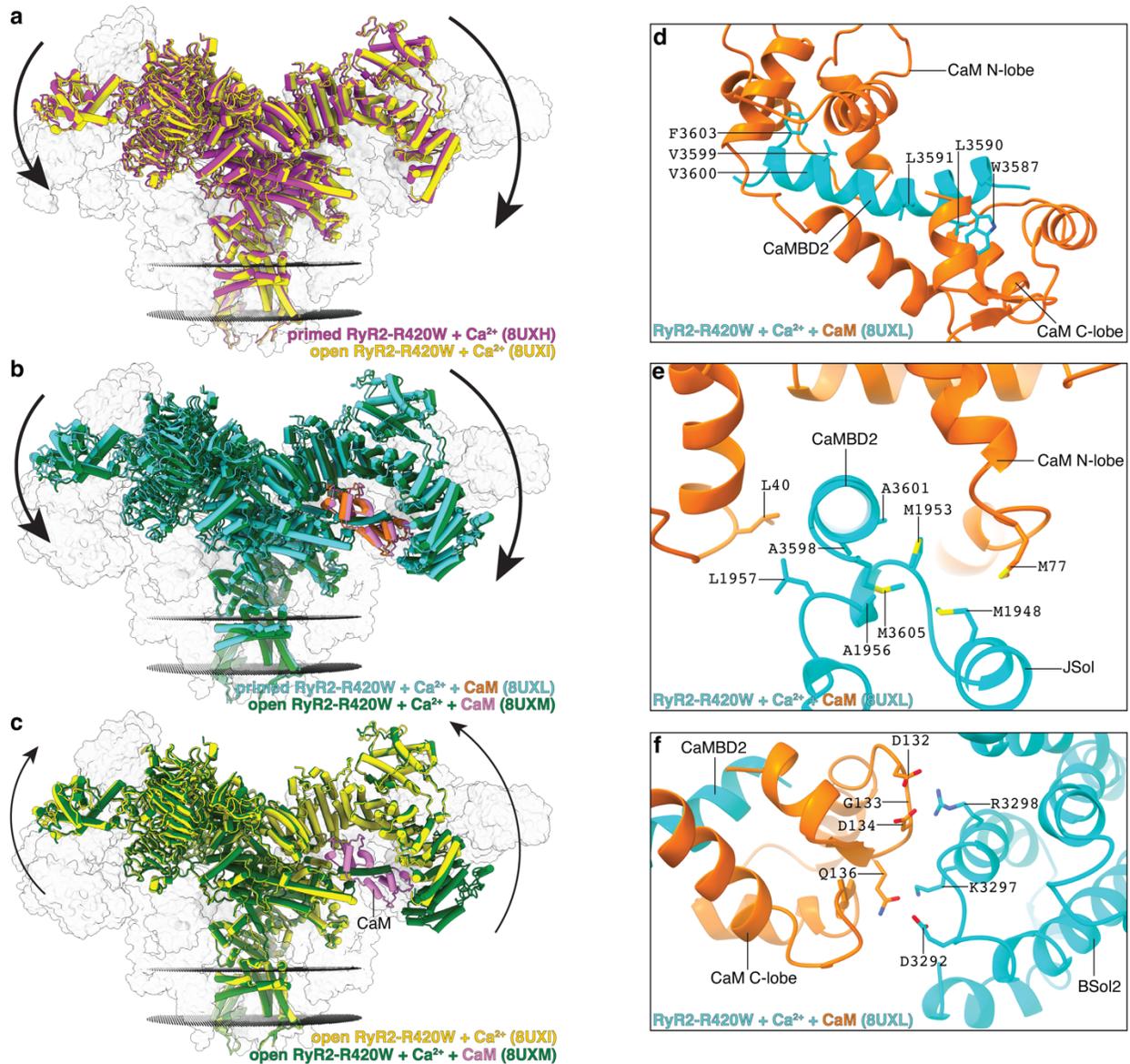


Supplementary Fig. 9. R420Q-induced rotation of the NTD-B domain disrupts interaction with the CSol and NTD-A* domains. a-d Aligned models focused on the NTD-B and CSol domains of closed PKA-phosphorylated WT RyR2 (PDB:7U9Q, grey), open PKA-phosphorylated WT RyR2 (PDB:7U9R, yellow), and primed PKA-phosphorylated RyR2-R420Q (PDB:8UXC,

magenta). Models were aligned at the NSol domain. Interdomain interactions are shown (dashed double arrow). Conformational changes are shown with arrows. Residues and distances stabilizing interdomain interactions are labelled. **e-h** Same as **a-d** but focusing on the interaction between the NTD-B and NTD-A⁻ domains. NTD-A⁻ stands for the NTD-A domain of the neighbor protomer.

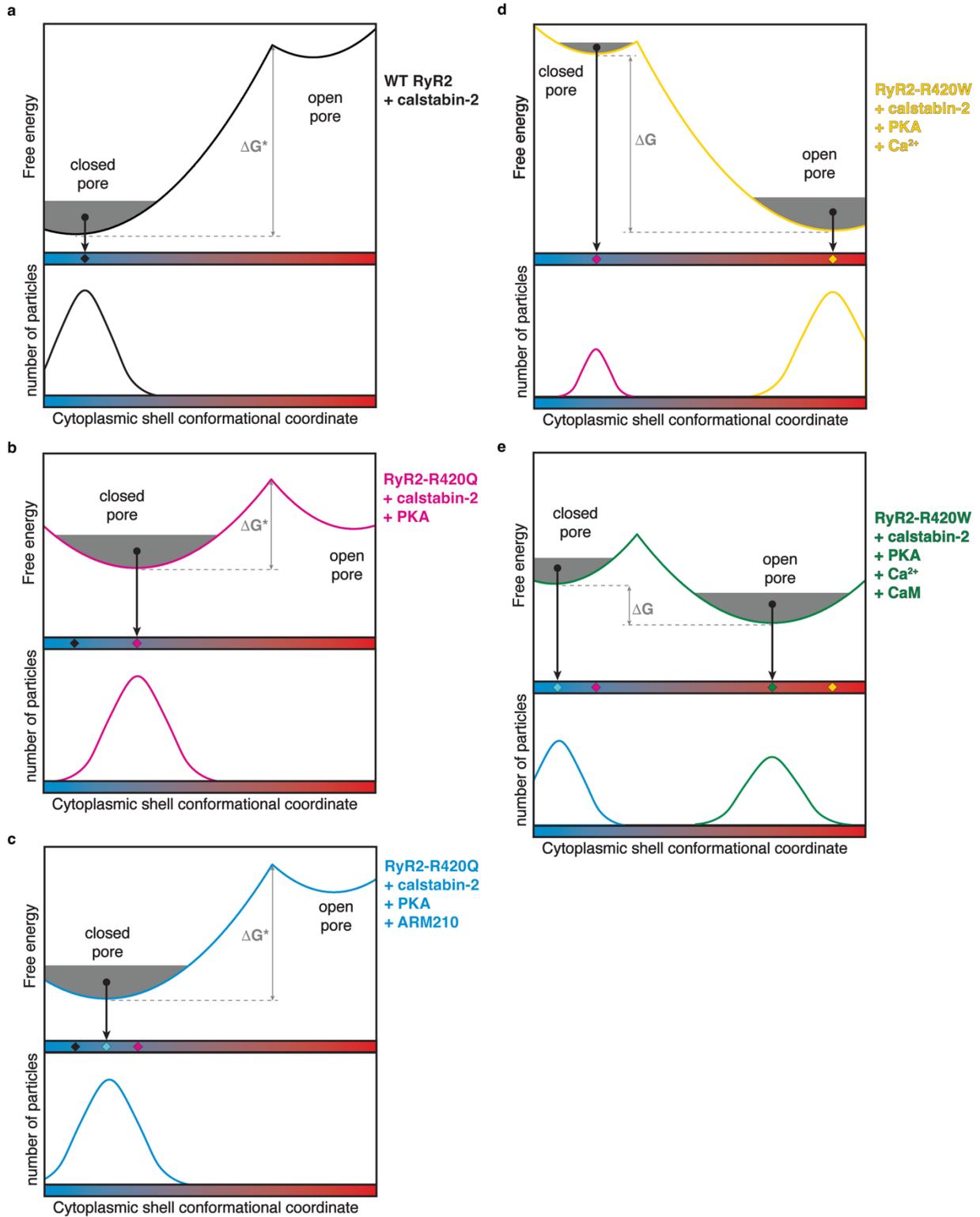
Variant	Onset age			koff	Kd	kon
	Average	SD	Count			
Arg2474Ser/Gly	7.4	4.5	8	0.095	232	407370
Ser2246Leu	8.7	4.8	18	0.076	185	410636
Arg420Gln	14.1	11.1	27	0.075	160	469200
Arg4959Gln	14.9	10.0	12	0.079	183	430215
Arg420Trp	20.1	7.3	12	0.071	168	407370
Glu4076Lys	29.7	20.6	3	0.061	151	400362
Arg3570Trp	36.0	26.9	2	0.064	155	415632
WT	60.0	*	*	0.043	105	406991

Supplementary Fig. 10. Calstabin binding site in different models. Table of the analyzed variants from the database,⁵³ showing average, standard deviation (SD), and number of patients (count) of onset age and diagnosis age. Also, included the k_{on} ($M^{-1} \cdot min^{-1}$), k_{off} (min^{-1}), and K_d (nM) of the recombinant RyR2 variants expressed in HEK293 cells.



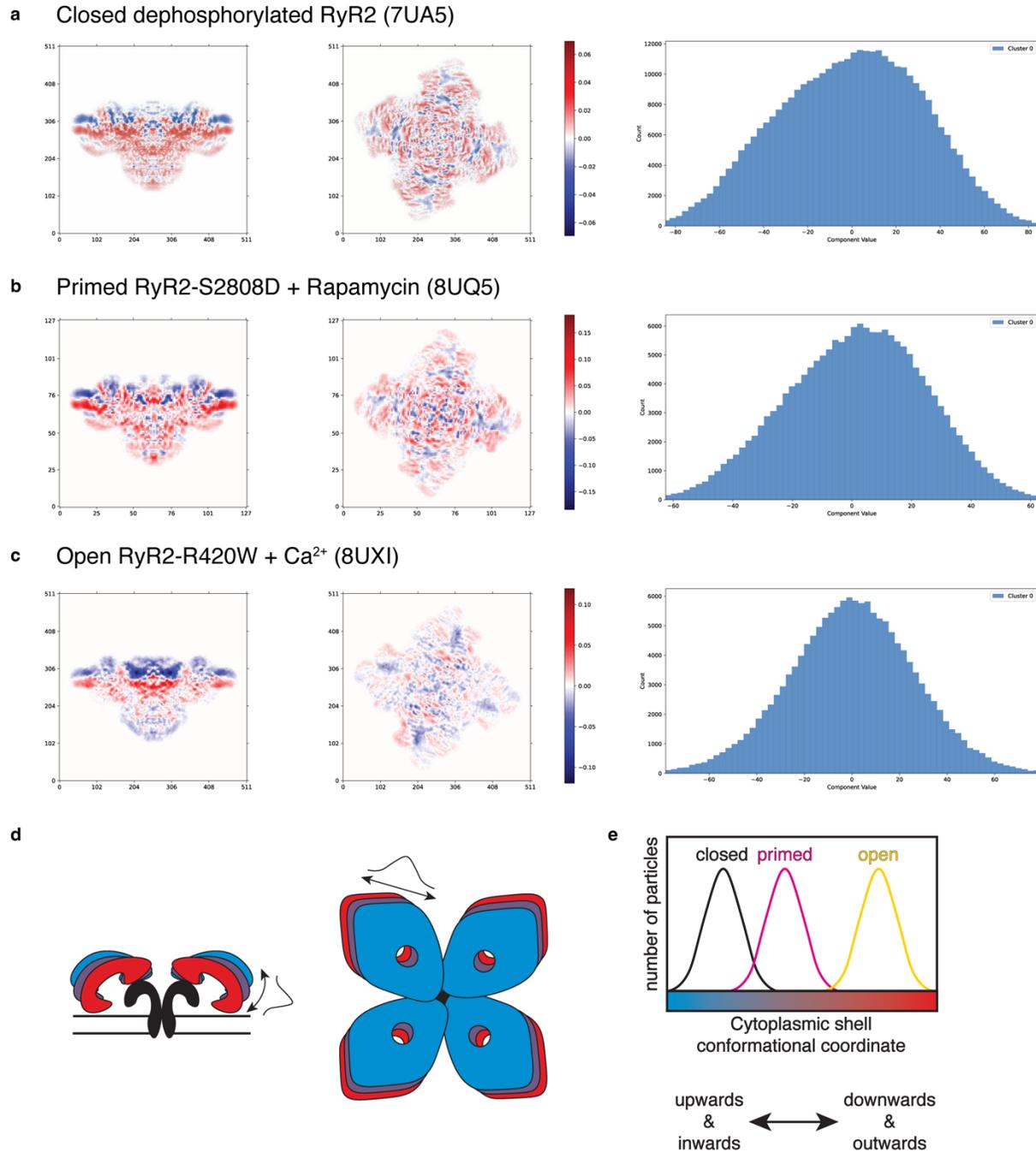
Supplementary Fig. 11. RyR2-R420W increased sensitivity to Ca²⁺ is partially reversed by accessory protein CaM. **a** Overlapped models of primed PKA-phosphorylated RyR2-R420W + Ca²⁺ (PDB:8UXH, magenta) and open PKA-phosphorylated RyR2-R420W + Ca²⁺ (PDB:8UXI, yellow). The arrows show that the cytoplasmic shell of the channel shifts downward-outward when going from the primed to the open state. To facilitate visualization only the front protomer is shown in colors, while the other three protomers are shown as grey transparent volumes. The positions of the sarcoplasmic reticulum membranes are shown as black discs. **b** Overlapped models of primed PKA-phosphorylated RyR2-R420W + Ca²⁺ + CaM (PDB:8UXL, cyan) and open PKA-phosphorylated RyR2-R420W + Ca²⁺ + CaM (PDB:8UXM, green). The arrows show that the

cytoplasmic shell of the channel shifts downward-outward when going from the primed to the open state, even in the presence of CaM. **c** Overlapped models of open PKA-phosphorylated RyR2-R420W + Ca²⁺ (PDB:8UXI, yellow) and open PKA-phosphorylated RyR2-R420W + Ca²⁺ + CaM (PDB:8UXM, green). The arrows show that the cytoplasmic shell of the channel shifts upward-inward in the presence of CaM, suggesting CaM has a stabilizing effect even in the open state. **d-f** Different views of the model of primed PKA-phosphorylated RyR2-R420W + Ca²⁺ + CaM (PDB:8UXL) focused on the CaM binding site. Residues involved in protein-protein interactions and interdomain interactions are labelled.

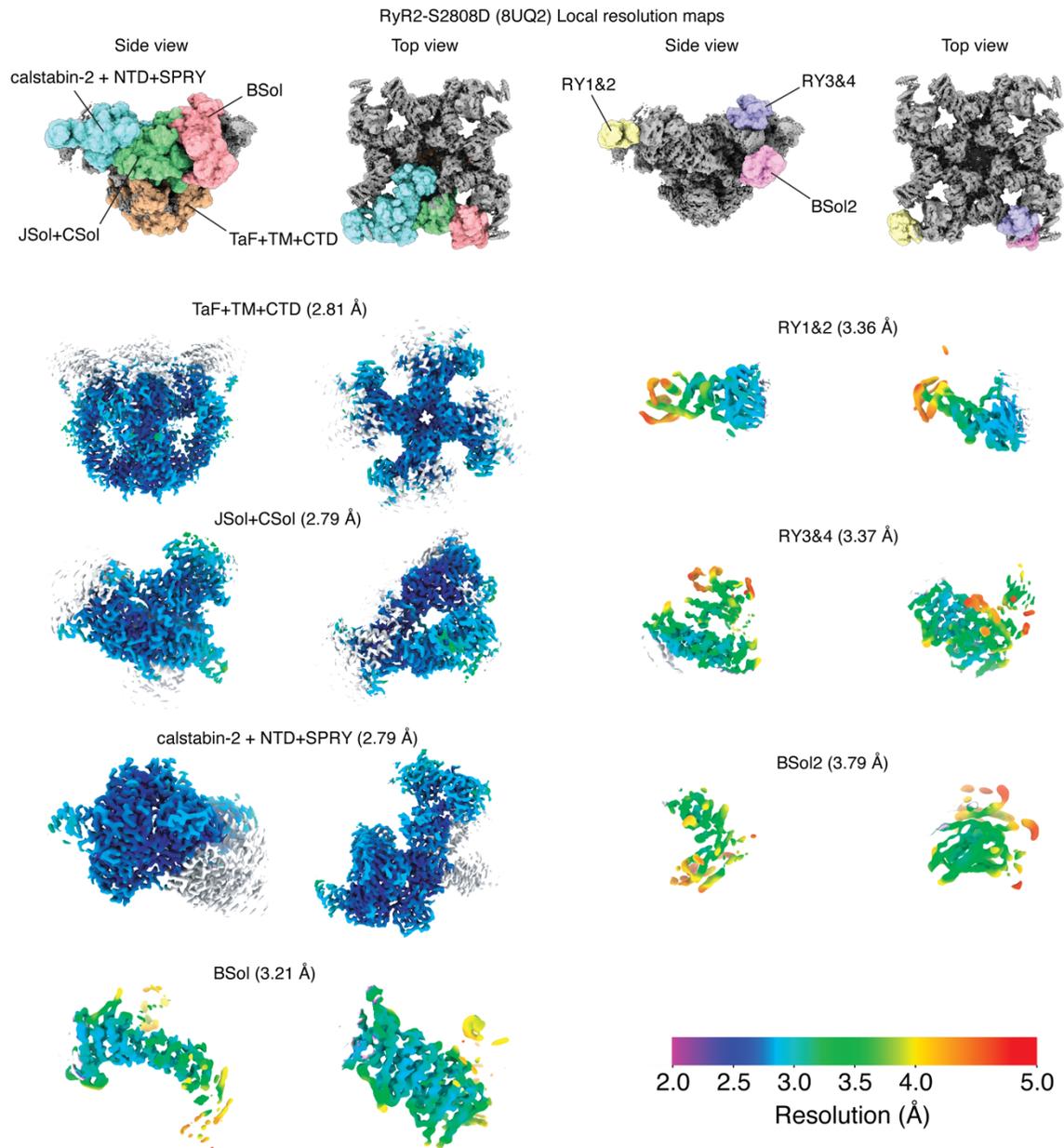


Supplementary Fig. 12. The proposed energy landscape explains the conformations and populations observed by cryo-EM. a-c Schematic view of the dynamic energy landscape of conformational space of the cytoplasmic shell for WT RyR2 + calstabin-2 (black, **a**), PKA

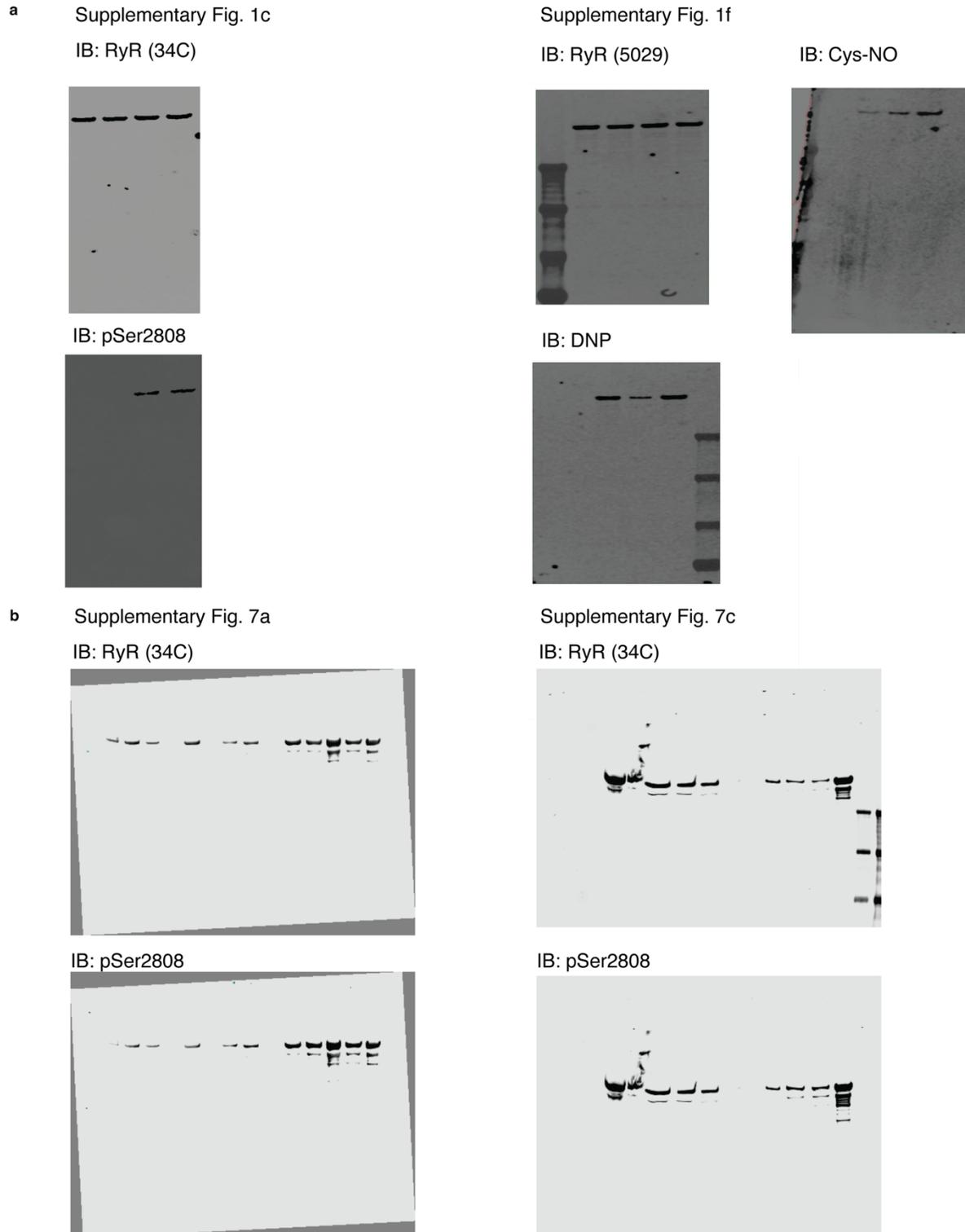
phosphorylated RyR2-R420Q + calstabin-2 (magenta, **b**), and PKA phosphorylated RyR2-R420Q + calstabin-2 + ARM210 (cyan, **c**). Energy landscapes were estimated using the approach from Figure 5. The grey area represents the conformations populated at a level of energy set by conditions and temperature. The average conformation is showed as a dot and projected to the conformational axis, which is observed in cryo-EM. The relative position of the average conformations correlates to the average conformations observed in cryo-EM (black > cyan > magenta). The population in the open state is very low and undetectable by cryo-EM. However, bilayer experiments suggest the channels visit the open state, with an increased open probability in the CPVT channels. The increased open probability is explained here by a decreased of the activation energy (ΔG^*) compared to the WT and the ARM210 treated channels. **d,e** Same as **a** but with PKA phosphorylated RyR2-R420W + calstabin-2 + Ca^{2+} (yellow, **d**) and PKA phosphorylated RyR2-R420W + calstabin-2 + Ca^{2+} + CaM (green, **e**). In this case both the open and closed states are present in cryo-EM. The change in the energy landscape explains the upward-inward movements observed for the cryo-EM structures. In addition, the lower net energy difference (ΔG) explains the shift in population observed in the cryo-EM particles.



Supplementary Fig. 13. 3D Variability Analysis of RyR2 variants. **a-c** Principal mode of variability and particle count showing normal distribution for closed dephosphorylated RyR2 (**a**), primed RyR2-S2808D + Rapamycin (**b**), and open RyR2-R420W + Ca²⁺ (**c**). **d** Schematic model of normal dynamics of RyR2 from the side and top views. **e** Representation of the cryo-EM particle count and the conformation of the cytoplasmic shell. Supplementary Figure 13d adapted from Marco C. Miotto et al., Structural analyses of human ryanodine receptor type 2 channels reveal the mechanisms for sudden cardiac death and treatment. *Sci. Adv.* 8,eabo1272(2022).DOI:10.1126/sciadv.abo1272 under a CC BY license: <https://creativecommons.org/licenses/by/4.0/>.¹⁰



Supplementary Fig. 14. Representative local resolution maps. Local refinement cryo-EM maps colored by local resolution shown for the RyR2-R2808D (8UQ2).



Supplementary Fig. 15. Uncropped western blots. a Uncropped western blots for supplementary figure 1. **b** Uncropped western blots for supplementary figure 7.

Supplementary Table 1. Domain span for human RyR2.

Identification	Symbol	Residue Span
Cytoplasmic shell		(1-3633)
N-terminal domain	NTD	(1-639)
N-terminal domain A	NTD-A	(1-219)
N-terminal domain B	NTD-B	(220-408)
N-terminal solenoid	NSol	(409-639)
SPRY domain	SPRY	(640-1646)
SP1a/ryanodine receptor domain 1	SPRY1	(640-861,1463-1483,1595-1646)
SP1a/ryanodine receptor domain 2	SPRY2	(1076-1255)
SP1a/ryanodine receptor domain 3	SPRY3	(1256-1462,1484-1594)
RYR repeats 1 & 2	RY1&2	(862-1076)
Junctional solenoid	JSol	(1647-2108)
Bridging solenoid	BSol	(2109-3564)
Bridging solenoid 1	BSol1	(2109-2681,2916-3042)
Bridging solenoid 2	BSol2	(3042-3344)
Bridging solenoid 3	BSol3	(3345-3564)
RYR repeats 3 & 4	RY3&4	(2682-2915)
Shell-core linker peptide	SCLP	(3565-3633)
Activation core and pore		(3634-4967)
Core solenoid	CSol	(3634-4130)
EF-hand pair	EF1&2	(4016-4090)
Thumb and forefingers domain	TaF	(4131-4209)
Transmembrane domain	TM	(4237-4886)
Auxiliary intramembrane helices	Sx	(4237-4310)
Pseudo voltage sensor domain	pVSD	(4480-4750)
Channel pore domain	Pore	(4751-4886)
C-terminal domain	CTD	(4887-4967)
Zn-finger domain	ZnF	(4887-4914)

Supplementary Table 2. Cryo-EM Statistics.

Sample	RyR2-S2808D	RyR2- S2808D + ARM210	RyR2- S2808D + NOC-12/GSH	RyR2- S2808D + Rapamycin
State	primed	closed	primed	primed
<i>PDB ID</i>	8UQ2	8UQ3	8UQ4	8UQ5
<i>EMDB ID</i>	EMD-42458	EMD-42459	EMD-42460	EMD-42461
Data collection				
Microscope	FEI Titan Krios	FEI Titan Krios	FEI Titan Krios	FEI Titan Krios
Detector	Gatan K3	Gatan K3	Gatan K3	Gatan K3
Voltage (kV)	300	300	300	300
Magnification	105,000	105,000	105,000	105,000
Exposure (e ⁻ /Å ²)	58	58	58	58
Defocus range (μm)	-0.4 to -1.2	-0.4 to -1.2	-0.4 to -1.2	-0.4 to -1.2
Pixel size (Å)	0.83	0.83	0.83	0.83
Processing				
Software	cryoSPARC	cryoSPARC	cryoSPARC	cryoSPARC
Symmetry	C4	C4	C4	C4
Initial particles (N)	267,550	137,303	83,074	320,951
Final particles (N)	191,898	83,460	19,753	18,232
Global map resolution (Å)*	2.98	3.18	3.64	3.96
Local maps resolution range (Å) [‡]	2.79-3.79	2.92-4.16	3.22-5.63	3.11-5.62
Model Composition				
Peptide chains	8	8	8	4
Nonhydrogen	138,608	138,700	138,608	135,336
Protein residues	17,324	17,324	17,324	16,896
Ligands	12	16	12	12
Mean B factors (Å²)				
Protein	112.09	120.82	151.90	141.97
Ligands	161.25	179.93	208.42	192.77
RMSD				
Bond length (Å)	0.003	0.003	0.003	0.003
Bond angles (°)	0.746	0.732	0.752	0.720
Ramachandran				
Favored (%)	97.16	97.04	96.58	97.31
Allowed (%)	2.77	2.92	3.33	2.60
Disallowed (%)	0.07	0.05	0.09	0.10
Validation				
MolProbity score	1.72	1.77	1.90	1.72
Clashscore	11.30	12.18	15.03	11.81
Rotamer outliers (%)	0.50	0.18	0.24	0.30
FSC model 0.5 (Å) [#]	2.99	3.13	3.53	3.43

Supplementary Table 2 continuation.

Sample	RyR2-R420Q	RyR2-R420Q +ARM210	RyR2-R420W	RyR2-R420W + ARM210
State	primed	closed	primed	closed
<i>PDB ID</i>	8UXC	8UXE	8UXF	8UXG
<i>EMDB ID</i>	EMD-42759	EMD-42761	EMD-42762	EMD-42763
Data collection				
Microscope	FEI Titan Krios	FEI Titan Krios	FEI Titan Krios	FEI Titan Krios
Detector	Gatan K3	Gatan K3	Gatan K3	Gatan K3
Voltage (kV)	300	300	300	300
Magnification	105,000	105,000	105,000	105,000
Exposure (e ⁻ /Å ²)	58	58	58	58
Defocus range (μm)	-0.4 to -1.2	-0.4 to -1.2	-0.4 to -1.2	-0.4 to -1.2
Pixel size (Å)	0.83	0.83	0.83	0.83
Processing				
Software	cryoSPARC	cryoSPARC	cryoSPARC	cryoSPARC
Symmetry	C4	C4	C4	C4
Initial particles (N)	187,859	20,900	115,505	206,663
Final particles (N)	143,933	14,924	102,478	181,724
Global map resolution (Å)*	2.86	3.53	3.13	3.08
Local maps resolution range (Å)†	2.64-3.35	3.10-5.02	2.90-3.74	2.91-3.38
Model Composition				
Peptide chains	8	8	8	8
Nonhydrogen	138,600	138,692	138,620	138,712
Protein residues	17,324	17,324	17,324	17,324
Ligands	12	16	12	16
Mean B factors (Å²)				
Protein	110.19	129.90	105.51	110.57
Ligands	147.54	258.08	171.76	165.53
RMSD				
Bond length (Å)	0.003	0.004	0.004	0.003
Bond angles (°)	0.672	0.713	0.697	0.720
Ramachandran				
Favored (%)	97.38	96.83	97.62	97.67
Allowed (%)	2.55	3.05	2.34	2.28
Disallowed (%)	0.07	0.12	0.05	0.05
Validation				
MolProbity score	1.61	1.81	1.62	1.58
Clashscore	9.29	12.83	10.46	9.75
Rotamer outliers (%)	0.03	0.37	0.58	0.74
FSC model 0.5 (Å)#	2.94	3.38	3.08	3.20

Supplementary Table 2 continuation.

Sample	RyR2-R420W + Ca ²⁺		RyR2-R420W + Ca ²⁺ + CaM	
	primed	open	primed	open
State	primed	open	primed	open
PDB ID	8UXH	8UXI	8UXL	8UXM
EMDB ID	EMD-42764	EMD-42765	EMD-42768	EMD-42769
Data collection				
Microscope	FEI Titan Krios		FEI Titan Krios	
Detector	Gatan K3		Gatan K3	
Voltage (kV)	300		300	
Magnification	105,000		105,000	
Exposure (e ⁻ /Å ²)	58		58	
Defocus range (µm)	-0.4 to -1.2		-0.4 to -1.2	
Pixel size (Å)	0.83		0.83	
Processing				
Software	cryoSPARC		cryoSPARC	
Symmetry	C4		C4	
Initial particles (N)	254,828		134,776	
Final particles (N)	55,886	149,448	77,625	49,606
Global map resolution (Å)*	3.52	3.29	3.12	3.56
Local maps resolution range (Å)‡	3.15-4.11	3.01-3.82	2.88-5.03	3.02-6.01
Model Composition				
Peptide chains	8	8	12	12
Nonhydrogen	131,656	131,656	143,500	143,500
Protein residues	16,444	16,444	17,928	17,928
Ligands	16	16	32	32
Mean B factors (Å²)				
Protein	158.66	130.95	129.59	134.13
Ligands	338.76	234.58	197.73	205.78
R.m.s. deviations				
Bond length (Å)	0.004	0.003	0.004	0.011
Bond angles (°)	0.854	0.825	0.881	0.751
Ramachandran				
Favored (%)	97.05	96.77	96.82	97.09
Allowed (%)	2.85	3.15	3.09	2.82
Disallowed (%)	0.10	0.07	0.09	0.09
Validation				
MolProbity score	1.94	1.90	1.92	1.75
Clashscore	13.01	12.67	13.79	11.89
Rotamer outliers (%)	1.58	1.28	1.25	0.74
FSC model 0.5 (Å)#	3.58	3.27	3.22	3.32

* Map resolution of the non-uniform refinement determined by CryoSPARC before local masks.

‡ Map resolution range represents the range determined by local refinements in cryoSPARC using the masks described in Methods.

Value obtained from Phenix validation tool.