

Asymmetric Dirhodium-Catalyzed Modification of Immunomodulatory Imide Drugs and their Biological Assessment

Supporting Information
(92 Pages)

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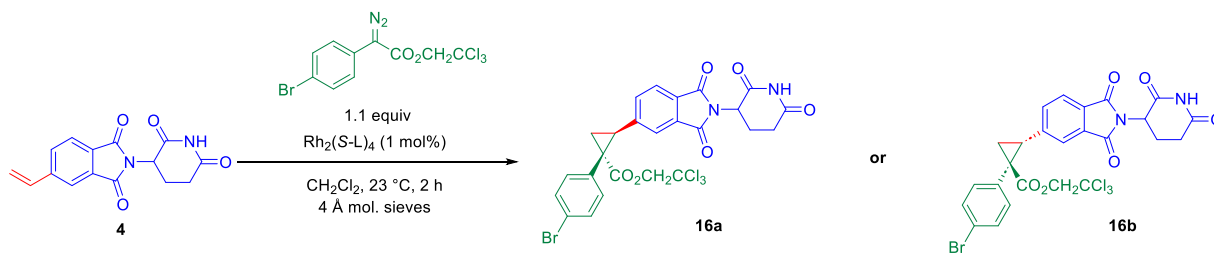
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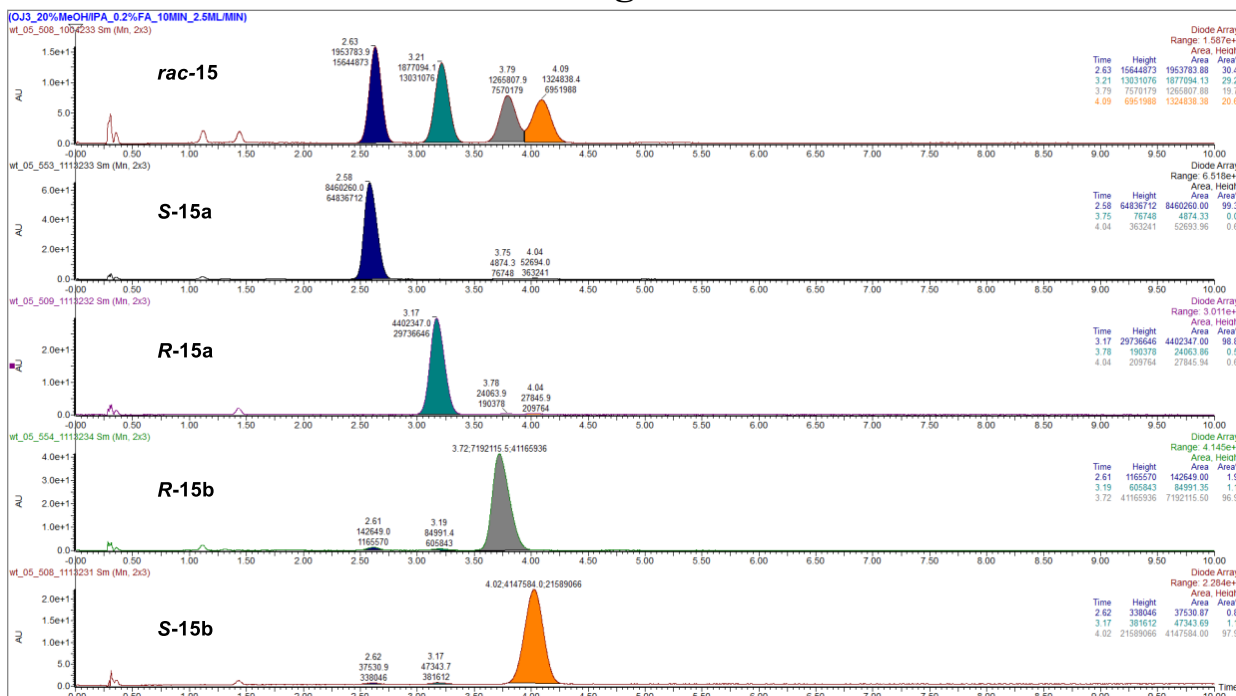
Scheme S1. Catalyst screen for the cyclopropanation of 4.



Entry	L	% yield	d.r. ^a	d.r. ^b (16x)
1	TPPTTL	77	>20:1	73:27 (16b)
2	PTAD	80	>20:1	87:13 (16a)
3 ^c	NTTL	82	>20:1	95:5 (16b)
4	<i>p</i> -Ph-TPCP	72	>20:1	99:1 (16a)

Reactions conducted on material in which the glutarimide stereocenter is racemic. Entries 1 and 3 gave the stereoisomers of product 16b, equivalent to those produced from the reaction with Rh₂(*R*-*p*-Ph-TPCP)₄. ^a Diastereomeric ratio for the relative configuration of the two new stereogenic centers, determined by ¹H NMR analysis. ^b Asymmetric induction arising from formation of the major relative diastereomer, determined by SFC analysis. ^c Reaction run with HFIP as solvent instead of CH₂Cl₂.

Scheme S2. Combined chromatograms of the stereoisomers of 15b



Section 1: General Information

All reactions were conducted in flame-dried glassware under an inert atmosphere of dry nitrogen or argon, unless otherwise noted. All reagents were purchased from Oakwood, CombiBlocks, Millipore Sigma, Ambeed, and WuXi and used as received. Anhydrous dichloromethane was obtained from a Grubbs-type solvent purification system and further dried under an argon atmosphere for 24 hours over 4 Å molecular sieves. Molecular sieves were activated by heating under vacuum (<1 torr) for three hours at 300 °C.

Proton (^1H) NMR spectra were recorded at 400 MHz on Varian and Bruker spectrometers, 500 MHz on an Inova-500 spectrometer, or 600 MHz on an Inova-600 spectrometer. Proton-decoupled carbon ($^{13}\text{C}\{^1\text{H}\}$) NMR spectra were recorded at 100 MHz on Varian and Bruker spectrometers or 151 MHz on an Inova-600 spectrometer. NMR spectra were recorded in deuterated solvents and were referenced to tetramethylsilane (in the case of CDCl_3) or the respective residual solvent signal. NMR chemical shifts were reported in parts per million (ppm). Abbreviations for signal couplings are as follows: s, singlet; d, doublet; t, triplet; q, quartet; quin, quintet; sex, sextet; sep, septet; and m, multiplet. The coupling constants were taken from the spectra directly and are uncorrected.

Mass spectrometric determinations were carried out on a Thermo Finnigan LTQ-FTMS spectrometer with atmospheric pressure chemical ionization (APCI), using a Fourier transform ion cyclotron resonance mass analyzer. Optical rotations were measured on a Rudolph Research Analytical Automatic Polarimeter, APIV-1W. Analytical thin layer chromatography was performed on silica gel plates using ultraviolet light to visualize analytes. Normal phase column chromatography was performed with SiliCycle silica gel 60 Å (50 μm) hand-packed in Biotage Sfaï columns on Biotage Isolera Four chromatographs, with SiliCycle silica gel 60 Å or Celite® as a dry-loading absorbent. Reverse phase chromatography was conducted using Teledyne ISCO RediSep Gold 18 reverse-phase columns using Teledyne ISCO CombiFlash or Biotage Isolera chromatographs. Supercritical fluid chromatography (SFC) analysis was performed on a Waters Acquity UPC2 instrument.

Purity of key compounds was assessed using SFC analysis (Waters Acquity UPC2 instrument), high-performance liquid chromatography analysis (HPLC) on an Agilent 1260 Infinity HPLC, or Waters AutoPurification System. All key compounds were $\geq 95\%$ pure by HPLC or SFC analysis.

Dirhodium catalysts $\text{Rh}_2(\text{S-p-Ph-TPCP})_4$,¹ $\text{Rh}_2(\text{R-p-Ph-TPCP})_4$,¹ $\text{Rh}_2(\text{S-tetra-p-Br-PPTL})_4$,² and $\text{Rh}_2(\text{R-tetra-p-Br-PPTL})_4$ ² were prepared according to their respective literature procedures. Racemic standards were prepared by using a 1:1 mixture of *R* and *S* catalyst under the same reaction conditions used to prepare enantioenriched compounds.

Caution! Glutarimide-containing compounds such as thalidomide are known reproductive, neurological, and hematological toxins. Care must be exercised to avoid direct contact with glutarimide-containing material; any glassware used with glutarimide-containing material should be treated with a 2 M aqueous solution of a strong base such as sodium hydroxide to destroy the material.

Section 2: Synthetic Procedures and Compound Characterization

Synthesis of Starting Materials

Preparation of Aryl Alkenes.

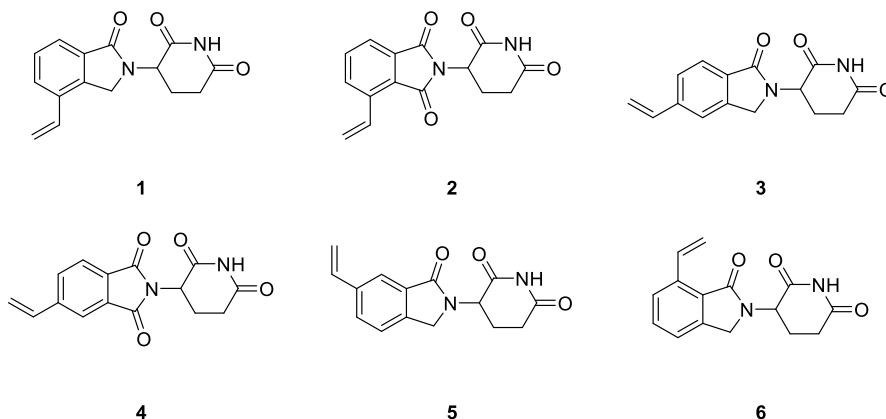


Figure S1. Aryl alkenes used in this study

Aryl alkenes **1-6** were prepared following the literature procedure,³ in 84-93% yield.

Preparation of Aryl Alkynes.

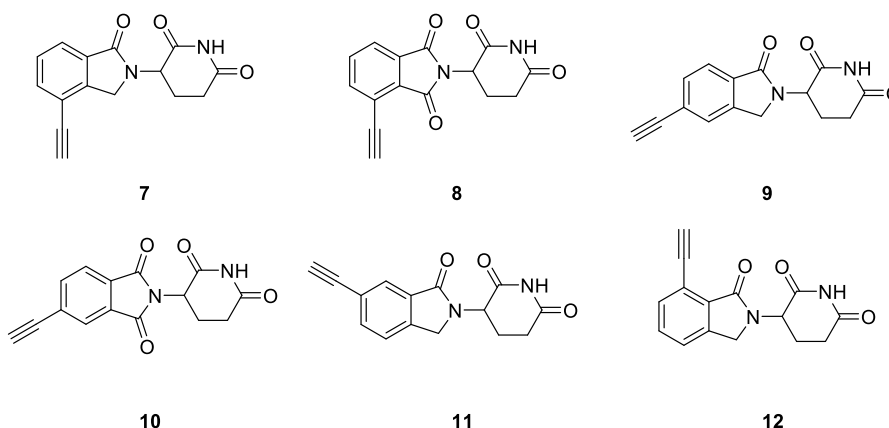


Figure S2. Aryl alkynes used in this study

Aryl alkynes **7-12** were prepared in 25-68% yield over two steps following an adapted patent procedure.⁴ A flame-dried round-bottom flask was charged with aryl bromide (1.0 equiv), Pd(PPh₃)₂Cl₂ (10 mol%), diisopropylethylamine (3.0 equiv), and CuI (10 mol%) under inert atmosphere. Dry *N,N*-dimethylformamide (0.20 M) and ethynyltrimethylsilane (5.0 equiv) were charged by syringe, and the reaction was heated to 65 °C overnight. The reaction was concentrated under vacuum and dry-mounted onto silica gel, then purified by flash column chromatography. The isolated material was carried on to the next step without further purification.

The trimethylsilyl-protected alkyne was charged to a flame-dried flask under inert atmosphere. Dry tetrahydrofuran (THF, 0.15 M) was charged via syringe, and the solution was cooled to 0 °C in an ice bath. 1.0 M tetrabutylammonium fluoride in THF (1.2 equiv) was added over thirty minutes via syringe pump to the cooled reaction. The reaction was stirred for thirty minutes longer, after which the reaction was concentrated and purified by flash column chromatography.

Preparation of Aryl Diazoacetates.

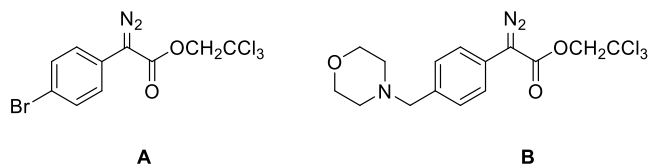


Figure S3. Aryl diazoacetates used in this study

2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (**A**) was prepared via the literature procedure, in 90% yield.⁵

2,2,2-trichloroethyl 2-diazo-2-(4-(morpholinomethyl)phenyl)acetate (**B**) was prepared from 2-(4-(bromomethyl)phenyl)acetic acid in three steps. Following an adaptation of the literature procedure,⁵ 2-(4-(bromomethyl)phenyl)acetic acid (1.0 g, 1.0 equiv, 4.4 mmol), 2,2,2-trichloroethanol (0.51 mL, 1.2 equiv, 5.2 mmol), and 4-(dimethylamino)pyridine (53 mg, 10 mol%, 0.44 mmol) were dissolved in dry CH₂Cl₂ (10 mL), and cooled to 0 °C. A solution of dicyclohexylcarbodiimide (1.0 g, 4.8 mmol, 1.1 equiv) in CH₂Cl₂ (5.7 mL) was added dropwise to the cooled reaction, which was allowed to stir and come to room temperature overnight. The reaction was filtered through Celite® to remove the white precipitate that had formed, rinsing with diethyl ether, and the filtrate was concentrated in vacuo. The material was filtered through a short plug of silica gel with diethyl ether, and concentrated in vacuo. The product obtained was used in the next step without further purification.

Following a procedure from the literature,⁶ potassium carbonate (359 mg, 1.3 equiv, 2.6 mmol), morpholine (0.17 mL, 1.0 equiv, 2.0 mmol), and 2,2,2-trichloroethyl 2-(4-(bromomethyl)phenyl)acetate (793 mg, 1.1 equiv, 2.2 mmol) were combined in a flame-dried vial under inert atmosphere, dissolved in dry acetonitrile (6.7 mL), and stirred at room temperature overnight. The acetonitrile was removed in vacuo and the residue was partitioned between water and ethyl acetate. The aqueous layer was extracted thrice with ethyl acetate, after which the organic layers were washed with brine, dried over magnesium sulfate, and concentrated in vacuo. The material was filtered through a silica plug with diethyl ether. The product obtained was used in the next step without further purification.

Following the literature procedure,⁵ 2,2,2-trichloroethyl 2-(4-(morpholinomethyl)phenyl)acetate (475 mg, 1 equiv, 1.3 mmol) and 2-nitrobenzenesulfonyl azide (443 mg, 1.5 equiv, 1.9 mmol) were taken up in dry acetonitrile (7.2 mL) and cooled to 0 °C. 2,3,4,6,7,8,9,10-octahydropyrimido[1,2-a]azepine (0.43 mL, 2.2 equiv, 2.9 mmol) was added dropwise to the cooled reaction. The reaction was allowed to stir for three hours at 0 °C, after which the reaction was quenched with a saturated solution of ammonium chloride in water. The reaction was extracted with three portions of diethyl ether. The combined organic layers were washed with brine, dried over magnesium sulfate, and concentrated in vacuo. The crude residue was purified via flash column chromatography (SiO₂, 10-30% EtOAc in hexanes), which gave 2,2,2-trichloroethyl 2-diazo-2-(4-(morpholinomethyl)phenyl)acetate (345 mg, 0.88 mmol, 68% yield) as a yellow-orange amorphous solid.

¹H NMR (400 MHz, CDCl₃) δ 7.45 (d, *J* = 8.4 Hz, 1H), 7.38 (d, *J* = 8.3 Hz, 1H), 4.92 (s, 1H), 3.71 (t, *J* = 4.7 Hz, 3H), 3.49 (s, 1H), 2.44 (t, *J* = 4.7 Hz, 2H).

Synthesis of Final Compounds

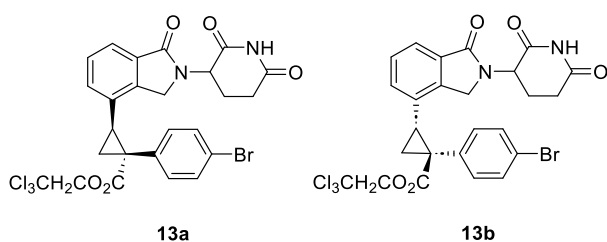
General Cyclopropanation Procedure (GP 1)

A 10 mL round-bottom flask was charged with a PTFE magnetic stir bar and ca. 200% w/w 4 Å molecular sieves, then flame dried under vacuum. The flask was then backfilled with dry nitrogen. Rh₂(*S*-p-Ph-TPCP)₄ (1 mol%) and the aryl alkene (1.0 equiv) were charged to the flask, and the flask was evacuated and backfilled with nitrogen. The flask was equipped with an argon balloon and rubber septum, and dry CH₂Cl₂

(0.2 M) was charged via syringe. A solution of 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (1.1 equiv) in dry CH₂Cl₂ (0.2 M, prepared in the same manner under inert atmosphere) was added dropwise to the stirred reaction at room temperature. The reaction was allowed to stir for two hours, after which the reaction was filtered through Celite, rinsing the filter pad with CH₂Cl₂. The crude residue was dry-mounted onto silica gel and was purified by flash column chromatography (SiO₂).

General Cyclopropanation Procedure (GP 2)

A 10 mL round-bottom flask was charged with a PTFE magnetic stir bar and ca. 200% w/w 4 Å molecular sieves, then flame dried under vacuum. The flask was then backfilled with dry nitrogen. Rh₂(*S*-p-Ph-TPCP)₄ (1 mol%) and the aryl alkyne (1.0 equiv) were charged to the flask, and the flask was evacuated and backfilled with nitrogen. The flask was equipped with an argon balloon and rubber septum, and dry CH₂Cl₂ (0.2 M) was charged via syringe. A solution of 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (1.1 equiv) in dry CH₂Cl₂ (0.2 M, prepared in the same manner under inert atmosphere) was added over 15 minutes to the stirred reaction at room temperature, via syringe pump. The reaction was allowed to stir for two hours, after which the reaction was filtered through Celite, rinsing the filter pad with CH₂Cl₂. The crude residue was dry-mounted onto silica gel and was purified by flash column chromatography (SiO₂).



2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)cyclopropane-1-carboxylate (13a).

2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)cyclopropane-1-carboxylate (13b).

Compound **13a** was prepared following **GP 1**, using Rh₂(*S*-p-Ph-TPCP)₄ (3.5 mg, 1 mol%, 2.0 μmol), 3-(1-oxo-4-vinylisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH₂Cl₂ (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO₂, gradient of 40% to 100% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)cyclopropane-1-carboxylate as an amorphous white solid. (78 mg, 0.12 mmol, 62% yield).

Compound **13b** was prepared in the same manner using Rh₂(*R*-p-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)cyclopropane-1-carboxylate as an amorphous white solid. (77 mg, 0.12 mmol, 61% yield).

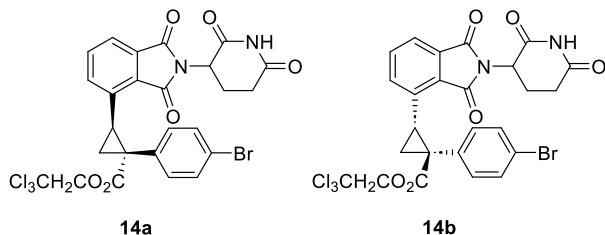
HRMS (APCI): *m/z* 612.9702 [(M+H)⁺ requires 612.9694], calc'd for C₂₅H₂₀⁷⁹Br³⁵Cl₃N₂O₅

¹H NMR (600 MHz, CDCl₃, reported as a mixture of diastereomers) δ 8.26 and 8.25 (s, 1H), 7.655 and 7.652 (d, *J* = 7.6 Hz, 1H), 7.28 and 7.26 (d, *J* = 8.5 Hz, 2H, partially obscured by solvent signal), 7.16-7.10 (m, 1H), 6.89 and 6.86 (d, *J* = 8.5 Hz, 2H), 6.49 and 6.43 (d, *J* = 7.7 Hz, 1H), 5.29 and 5.26 (dd, *J* = 13.2, 5.2 Hz, 1H), 4.88 and 4.87 (d, *J* = 12.0 Hz, 1H), 4.66 and 4.65 (d, *J* = 12.0 Hz, 1H), 4.59 and 4.54 (d, *J* = 16.0 Hz, 1H), 4.39 and 4.37 (d, *J* = 16.0 Hz, 1H), 3.07 (dd, *J* = 9.3, 7.3 Hz, 1H), 2.98-2.81 (m, 2H), 2.40-2.20 (m, 3H), 2.12-2.06 (m, 1H).

¹³C{¹H} NMR (126 MHz, CDCl₃, reported as a mixture of diastereomers) δ 171.4, 171.1, 169.63, 169.59, 169.42, 169.37, 141.6, 133.3, 133.2, 132.3, 131.5, 131.4, 130.7, 129.0, 128.9, 128.4, 122.93, 122.91, 122.2, 74.4, 74.3, 52.1, 52.0, 46.4, 36.8, 36.7, 31.7, 29.9, 29.7, 23.7, 23.6, 19.8, 19.7.

SFC analysis: **13a** (Trefoil® AMY1, 25% 1:1 MeOH:PrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 99:1 d.r.: *t_R* (major diastereomers) = 2.89 and 4.82 min, *t_R* (minor diastereomers) = 3.24 and 3.96 min. **13b** indicated the opposite diastereomers in 99:1 d.r.

Purity (SFC): **13a** – 99%; **13b** – 99%



14a

14b

2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-4-yl)cyclopropane-1-carboxylate (14a).

2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-4-yl)cyclopropane-1-carboxylate (14b).

Compound **14a** was prepared following **GP 1**, using $\text{Rh}_2(\text{S-p-Ph-TPCP})_4$ (3.5 mg, 1 mol%, 2.0 μmol), 2-(2,6-dioxopiperidin-3-yl)-4-vinylisindoline-1,3-dione (57 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH_2Cl_2 (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO_2 , gradient of 40% to 100% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-4-yl)cyclopropane-1-carboxylate as an amorphous white solid. (40 mg, 62 μmol , 31% yield).

Compound **14b** was prepared in the same manner using $\text{Rh}_2(\text{R-p-Ph-TPCP})_4$, which afforded 2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-4-yl)cyclopropane-1-carboxylate as an amorphous white solid. (38 mg, 59 μmol , 30% yield).

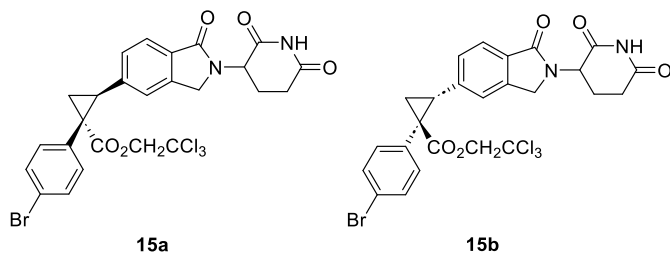
HRMS (APCI): m/z 625.9417 ($[\text{M}]^+$ requires 625.9422), calc'd for $\text{C}_{25}\text{H}_{18}\text{BrCl}_3\text{N}_2\text{O}_6$.

^1H NMR (500 MHz, CDCl_3 , reported as a mixture of diastereomers) δ 8.22 and 8.21 (s, 1H), 7.624 and 7.617 (dd, $J = 7.40, 0.70$ Hz, 1H), 7.38-7.33 (m, 1H), 7.27 (d, $J = 7.9$ Hz, 2H, partially obscured by solvent signal), 6.95 (t, $J = 8.2$ Hz, 2H), 6.77 and 6.68 (d, $J = 8.0$ Hz, 1H), 5.00 (dt, $J = 12.4, 6.1$ Hz, 1H), 4.95 and 4.93 (d, $J = 11.9$ Hz, 1H), 4.64 and 4.61 (d, $J = 11.2$ Hz, 1H), 4.21-4.09 (m, 1H) 2.95-2.72 (m, 3H), 2.39 and 2.35 (dd, $J = 9.1, 5.4$ Hz, 1H), 2.20-2.13 (m, 2H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3 , reported as a mixture of diastereomers) δ 171.1, 171.0, 170.9, 168.1, 168.0, 167.9, 166.95, 166.93, 136.3, 136.1, 134.0, 133.9, 133.2, 133.1, 132.4, 132.37, 132.32, 132.19, 132.17, 131.40, 131.37, 130.0, 129.8, 122.4, 122.3, 122.1, 122.0, 94.8, 74.81, 74.76, 49.5, 37.1, 36.8, 31.5, 29.5, 29.0, 22.8, 22.7, 19.7, 19.3.

SFC analysis: **14a** (Chiralcel® OX-3, 35% 1:1 MeOH:PrOH with 0.2% formic acid in CO_2 , 2.5 mL/min, diode array) indicated >99:1 d.r.: t_R (major diastereomers) = 2.16 and 2.36 min, t_R (minor diastereomers) = 4.09 and 7.32 min. **14b** indicated the opposite diastereomers in 99:1 d.r.

Purity (SFC): **14a** – 99%; **14b** – 96%



15a

15b

2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate (15a).

2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate (15b).

Compound **15a** was prepared following **GP 1**, using $\text{Rh}_2(\text{S-p-Ph-TPCP})_4$ (3.5 mg, 1 mol%, 2.0 μmol), 3-(1-oxo-5-vinylisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH_2Cl_2 (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO_2 , gradient of 25% to 75% EtOAc in hexanes)

afforded 2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (95 mg, 0.16 mmol, 77% yield).

Compound **15b** was prepared in the same manner using Rh₂(*R*-p-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (97 mg, 0.16 mmol, 79% yield).

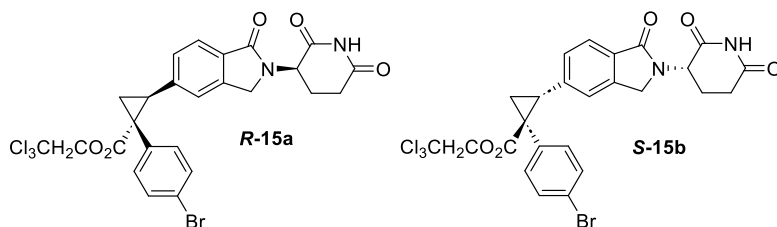
HRMS (APCI): *m/z* 612.9702 [(M+H)⁺ requires 612.9686], calc'd for C₂₅H₂₀BrCl₃N₂O₅

¹H NMR (500 MHz, CDCl₃, reported as a mixture of diastereomers) 8.42 (s, 1H), 7.60 (12.0, 8.0 Hz, 1H), 7.28 and 7.27 (d, *J*=8.5 Hz, 2H), 6.96-6.93 (m, 3H), 6.86-6.85 (m, 1H), 5.16 (ddd, *J*=18.0, 13.3, 5.2 Hz), 4.83 and 4.81 (d, *J*=2.8 Hz, 1H), 4.66 and 4.64 (d, *J*=3.2 Hz, 1H), 4.34 and 4.31 (d, *J*=16.0 Hz, 1H), 4.21 and 4.10 (d, 16.0 Hz, 1H), 3.29 (dd, *J*=9.4, 7.2 Hz, 1H), 2.89-2.76 (m, 2H), 2.36-2.23 (m, 2H), 2.23-2.18 (m, 1H), 2.03-2.00 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl₃, reported as a mixture of diastereomers) δ 171.33, 171.30, 171.3, 169.72, 169.66, 169.0, 141.50, 141.48, 140.4, 133.7, 133.6, 132.5, 132.4, 131.46, 131.38, 130.33, 130.27, 128.7, 128.3, 123.8, 122.9, 122.4, 122.1, 122.0, 94.9, 74.6, 52.0, 51.8, 47.0, 46.8, 37.4, 37.3, 33.9, 33.8, 31.6, 23.54, 23.47, 20.91, 20.86.

SFC analysis: **15a** (Chiralcel® OJ-3, 30% 1:1 MeOH:ⁱPrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated >99:1 d.r.: *t*_R (major diastereomers) = 2.72 and 4.60 min, *t*_R (minor diastereomers) = 3.38 and 3.98 min. **15b** indicated the opposite diastereomers in >99:1 d.r.

Purity (SFC): **15a** – 99%; **15b** – 99%



2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-((*R*)-2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate (R-15a**).**

2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-((*S*)-2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate (S-15b**).**

Compound **S-15a** was prepared following **GP 1**, using Rh₂(*S*-p-Ph-TPCP)₄ (3.5 mg, 1 mol%, 2.0 μmol), (*S*)-3-(1-oxo-5-vinylisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH₂Cl₂ (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO₂, gradient of 25% to 75% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-((*S*)-2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (115 mg, 0.19 mmol, 94% yield).

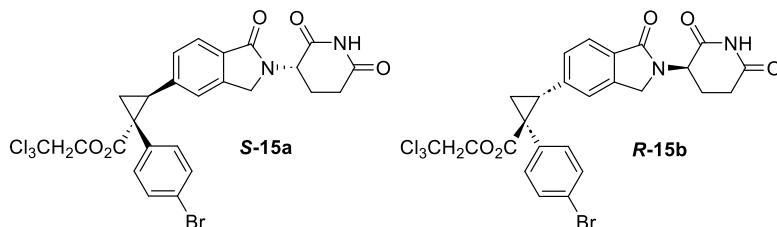
Compound **R-15b** was prepared in the same manner using (*R*)-3-(1-oxo-5-vinylisindolin-2-yl)piperidine-2,6-dione and Rh₂(*R*-p-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-((*R*)-2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (117 mg, 0.19 mmol, 95% yield).

¹H NMR (400 MHz, CDCl₃) δ 8.00 (s, 1H), 7.61 (d, *J* = 7.9 Hz, 1H), 7.29 (d, *J* = 8.5 Hz, 2H), 7.00 – 6.91 (m, 3H), 6.87 (dd, *J* = 7.9, 1.5 Hz, 1H), 5.15 (dd, *J* = 13.2, 5.2 Hz, 1H), 4.83 (d, *J* = 11.9 Hz, 1H), 4.65 (d, *J* = 11.9 Hz, 1H), 4.32 (d, *J* = 16.0 Hz, 1H), 4.22 (d, *J* = 16.0 Hz, 1H), 3.30 (dd, *J* = 9.3, 7.4 Hz, 1H), 2.94 – 2.87 (m, 1H), 2.87-2.73 (m, 1H), 2.41 – 2.26 (m, 2H), 2.25-2.13 (m, 1H), 2.06 – 1.95 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl₃) δ 171.5, 171.3, 169.8, 168.9, 141.5, 140.4, 133.6, 132.3, 131.3, 130.3, 128.2, 123.7, 122.9, 122.0, 94.9, 74.5, 51.9, 47.0, 37.3, 33.8, 31.6, 23.4, 20.9.

SFC analysis: **S-15a** (Chiralcel® OJ-3, 30% 1:1 MeOH:ⁱPrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 98:2 d.r.: *t*_R (major diastereomers) = 4.08 min (peak representing minor enantiomer of **S-15a** n.d.), *t*_R (minor diastereomers) = 2.64 and 3.22 min. **R-15b** indicated the opposite diastereomers in 96:4 d.r.

Specific rotation: **S-15a** $[\alpha]_D^{22}$ 12.7 (c 1, CHCl₃) **R-15b** $[\alpha]_D^{22}$ -17.6 (c 1.9, CHCl₃)
Purity (SFC): **S-15a** – 97%; **R-15b** – 98%



2,2,2-trichloroethyl (1R,2S)-1-(4-bromophenyl)-2-(2-((S)-2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate (S-15a).

2,2,2-trichloroethyl (1S,2R)-1-(4-bromophenyl)-2-(2-((R)-2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate (R-15b).

Compound **R-15a** was prepared following **GP 1**, using Rh₂(*S*-p-Ph-TPCP)₄ (3.5 mg, 1 mol%, 2.0 μmol), (*R*)-3-(1-oxo-5-vinylisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH₂Cl₂ (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO₂, gradient of 25% to 75% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-((*S*)-2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (118 mg, 0.19 mmol, 96% yield).

Compound **S-15b** was prepared in the same manner using (*S*)-3-(1-oxo-5-vinylisindolin-2-yl)piperidine-2,6-dione and Rh₂(*R*-p-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-((*S*)-2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (117 mg, 0.19 mmol, 95% yield).

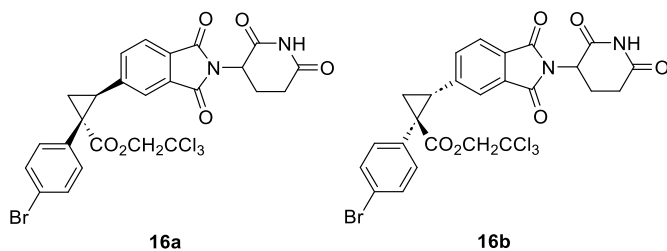
¹H NMR (400 MHz, CDCl₃) δ 7.97 (s, 1H), 7.63 (d, *J* = 7.9 Hz, 1H), 7.29 (d, *J* = 8.5 Hz, 2H), 6.98 – 6.93 (m, 3H), 6.86 (s, 1H), 5.19 (dd, *J* = 13.2, 5.3 Hz, 1H), 4.83 (d, *J* = 11.9 Hz, 1H), 4.66 (d, *J* = 11.9 Hz, 1H), 4.36 (d, *J* = 16.0 Hz, 1H), 4.12 (d, *J* = 16.1 Hz, 1H), 3.30 (dd, *J* = 9.3, 7.4 Hz, 1H), 2.97-2.87, (m, 1H), 2.87-2.72 (m, 1H), 2.41 – 2.27 (m, 2H), 2.27 – 2.15 (m, 1H), 2.08 – 1.94 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 171.5, 171.3, 169.8, 169.0, 141.5, 140.4, 133.6, 132.4, 131.3, 130.2, 128.6, 123.7, 122.4, 121.9, 94.9, 74.6, 51.8, 46.8, 37.4, 33.8, 31.6, 23.5, 20.8.

SFC analysis: **R-15a** (Chiralcel® OJ-3, 30% 1:1 MeOH:PrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 99:1 d.r.: *t_R* (major diastereomers) = 2.53 min (peak representing minor enantiomer of **R-15a** n.d.), *t_R* (minor diastereomers) = 3.71 and 3.98 min. **S-15b** indicated the opposite diastereomers in 98:2 d.r.

Specific rotation: **R-15a** $[\alpha]_D^{22}$ -4.8 (c 1, CHCl₃) **S-15b** $[\alpha]_D^{22}$ 5.5 (c 1.9, CHCl₃)

Purity (SFC): **R-15a** – 96%; **S-15b** – 97%



2,2,2-trichloroethyl (1R,2S)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-5-yl)cyclopropane-1-carboxylate (16a).

2,2,2-trichloroethyl (1S,2R)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-5-yl)cyclopropane-1-carboxylate (16b).

Compound **16a** was prepared following **GP 1**, using Rh₂(*S*-p-Ph-TPCP)₄ (3.5 mg, 1 mol%, 2.0 μmol), 2-(2,6-dioxopiperidin-3-yl)-5-vinylisindoline-1,3-dione (57 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH₂Cl₂ (2 mL) at room temperature for two

hours. Purification via flash column chromatography (SiO₂, gradient of 20% to 60% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (90 mg, 0.14 mmol, 72% yield). Compound **16b** was prepared in the same manner using Rh₂(*R*-*p*-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (90 mg, 0.14 mmol, 72% yield).

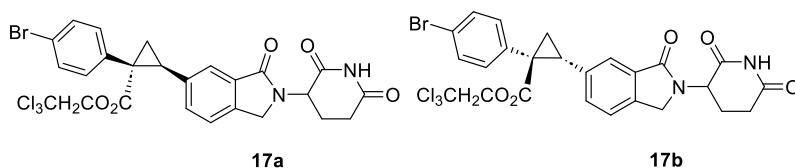
HRMS (APCI): *m/z* 626.9492 [(M+H)⁺ requires 625.9487], calc'd for C₂₅H₁₈⁷⁹Br³⁵Cl₃N₂O₆

¹H NMR (500 MHz, CDCl₃, reported as a mixture of diastereomers) δ 8.49 (s, 1H), 7.59 and 7.58 (d, *J*=2.0 Hz, 1H), 7.40 (d, 10.2 Hz, 1H), 7.29 (d, 8.0 Hz, 2H), 7.12-7.10 (m, 1H), 6.95 (d, *J*=8.0 Hz, 2H), 4.93 (dd, *J*=12.2, 5.3 Hz, 1H), 4.83 (d, 12.0 Hz, 1H), 4.65 (d, 12.0 Hz, 1H), 3.37-3.33 (m, 1H), 2.88-2.71 (m, 3H), 2.37 (dd, 9.2, 5.5 Hz, 1H), 2.13-2.08 (m, 2H).

¹³C{¹H} NMR (126 MHz, CDCl₃, reported as a mixture of diastereomers) δ 171.2, 170.9, 168.2, 167.04, 167.02, 166.90, 166.88, 143.72, 143.70, 133.91, 133.88, 133.47, 133.46, 131.9, 131.8, 131.6, 130.1, 129.1, 128.3, 125.4, 123.4, 123.47, 123.44, 122.3, 94.8, 74.7, 49.42, 49.39, 37.76, 37.74, 33.4, 31.42, 31.40, 22.69, 22.64, 20.68, 20.64.

SFC analysis: **16a** (Trefoil® CEL1, 20% 1:1 MeOH:PrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 99:1 d.r.: *t_R* (major diastereomers) = 5.17 and 5.62 min, *t_R* (minor diastereomers) = 4.31 and 4.92 min. **16b** indicated the opposite diastereomers in 98:2 d.r.

Purity (SFC): **16a** – 98%; **16b** – 96%



2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-5-yl)cyclopropane-1-carboxylate (17a**).**

2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-5-yl)cyclopropane-1-carboxylate (17b**).**

Compound **17a** was prepared following **GP 1** using Rh₂(*S*-*p*-Ph-TPCP)₄ (3.5 mg, 1 mol%, 2.0 μmol), 3-(1-oxo-6-vinylisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH₂Cl₂ (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO₂, gradient of 20% to 75% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (96 mg, 0.16 mmol, 78% yield).

Compound **17b** was prepared in the same manner using Rh₂(*R*-*p*-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-5-yl)cyclopropane-1-carboxylate as an amorphous white solid. (95 mg, 0.15 mmol, 77% yield).

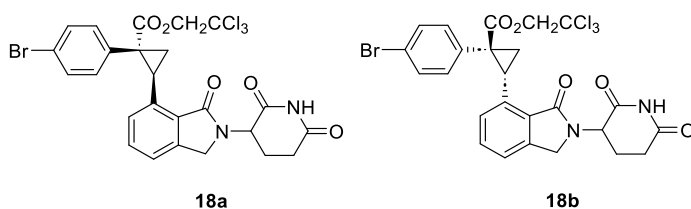
HRMS (APCI): *m/z* 612.9698 [(M+H)⁺ requires 612.9694], calc'd for C₂₅H₂₁O₅N₂⁷⁹Br³⁵Cl₃

¹H NMR (600 MHz, CDCl₃, reported as a mixture of diastereomers) δ 7.94 (s, 1H), 7.53 and 7.43 (s, 1H), 7.26 (d, *J* = 8.6 Hz, 2H, partially obscured by solvent signal), 7.21 and 7.19 (d, *J* = 7.9 Hz, 1H), 7.02 and 6.92 (dd, *J* = 8.0, 1.7 Hz, 1H), 6.96 (d, *J* = 8.6 Hz, 2H), 5.20 and 5.17 (dd, *J* = 15.6, 5.0 Hz, 1H), 4.843 and 4.841 (d, *J* = 12.0 Hz, 1H), 4.65 and 4.648 (d, *J* = 11.9 Hz, 1H), 4.41 and 4.39 (d, *J* = 16.0 Hz, 1H), 4.25 and 4.24 (d, *J* = 15.9 Hz, 0H), 3.34 (t, *J* = 8.3 Hz, 1H), 2.97 – 2.88 (m, 1H), 2.86 – 2.77 (m, 1H), 2.45 – 2.26 (m, 2H), 2.24-2.18 (m, 1H), 2.10 – 1.97 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl₃, reported as a mixture of diastereomers) δ 171.4, 171.3, 169.72, 169.69, 169.1, 140.20, 140.18, 136.23, 136.21, 133.7, 133.6, 132.5, 132.2, 131.62, 131.58, 131.28, 131.26, 124.4, 123.7, 122.7, 121.9, 121.8, 95.0, 74.6, 51.98, 51.85, 47.0, 46.8, 36.94, 36.87, 33.57, 31.64, 31.62, 23.5, 20.1, 20.0.

SFC analysis: **17a** (Trefoil® CEL1, 25% 1:1 EtOH:PrOH with 0.2% formic acid in CO₂, 2.0 mL/min, diode array) indicated 98:2 d.r.: *t_R* (major diastereomers) = 4.85 and 5.60 min, *t_R* (minor diastereomers) = 4.34 and 4.72min. **17b** indicated the opposite diastereomers in 97:3 d.r.

Purity (SFC): 17a – 99%; 17b – 95%



2,2,2-trichloroethyl (1R,2S)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-4-yl)cyclopropane-1-carboxylate (18a).

2,2,2-trichloroethyl (1S,2R)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-4-yl)cyclopropane-1-carboxylate (18b).

Compound **18a** was prepared following **GP 1**, using $\text{Rh}_2(\text{S-p-Ph-TPCP})_4$ (3.5 mg, 1 mol%, 2.0 μmol), 3-(1-oxo-7-vinylisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH_2Cl_2 (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO_2 , gradient of 50% to 60% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1R,2S)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-4-yl)cyclopropane-1-carboxylate as an amorphous white solid. (35 mg, 56 μmol , 28% yield).

Compound **18b** was prepared in the same manner using $\text{Rh}_2(\text{R-p-Ph-TPCP})_4$, which afforded 2,2,2-trichloroethyl (1S,2R)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-4-yl)cyclopropane-1-carboxylate as an amorphous white solid. (36 mg, 57 μmol , 29% yield).

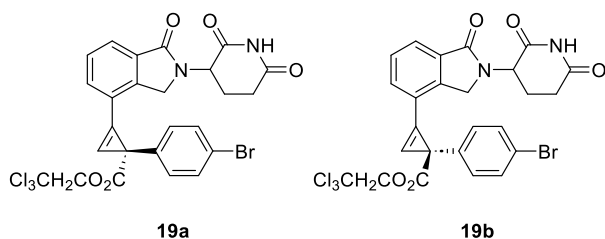
HRMS (APCI): m/z 612.9698 [$\text{M}+\text{H}$]⁺ requires 612.9694], calc'd for $\text{C}_{25}\text{H}_{21}\text{O}_5\text{N}_2^{79}\text{Br}^{35}\text{Cl}_3$

¹H NMR (400 MHz, CDCl_3 , reported as a mixture of diastereomers) δ 8.30 and 8.23 (s, 1H), 7.24 – 7.14 (m, 4H), 7.00 and 6.97 (d, $J = 8.5$ Hz, 2H), 6.60-6.52 and 6.52-6.45 (m, 1H), 5.27 and 5.19 (dd, $J = 13.3, 5.1$ Hz, 1H), 4.91 and 4.89, (d, $J = 11.9$ Hz, 1H), 4.67 and 4.65 (d, $J = 11.9$ Hz, 1H), 4.47-4.22 (m, 3H), 3.00 – 2.75 (m, 2H), 2.50 – 2.18 (m, 3H), 2.15-2.10 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl_3 , reported as a mixture of diastereomers) δ 171.5 and 171.4, 171.34 and 171.30, 170.0 and 169.73, 169.72 and 169.61, 142.14 and 142.12, 135.47 and 135.45, 133.45 and 133.28, 133.42 and 133.38, 131.6 and 131.5, 131.0 and 130.9, 130.1 and 129.9, 126.1 and 126.0, 121.7 and 121.6, 121.49, 121.46, 95.1, 74.8 and 74.7, 52.1 and 51.9, 46.9 and 46.6, 36.8 and 36.5, 31.8 and 31.7, 29.6 and 29.3, 23.6 and 23.4, 19.2 and 19.0.

SFC analysis: **18a** (Trefoil® AMY1, 30% 1:1 MeOH:^tPrOH with 0.2% formic acid in CO_2 , 2.5 mL/min, diode array) indicated 99:1 d.r.: t_R (major diastereomers) = 3.65 and 5.13 min, t_R (minor diastereomers) = 3.17 and 4.24 min. **18b** indicated the opposite diastereomers in 99:1 d.r.

Purity (SFC): 18a – 98%; 18b – 99%



2,2,2-trichloroethyl (1S)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate (19a).

2,2,2-trichloroethyl (1R)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate (19b).

Compound **19a** was prepared following **GP 2**, using $\text{Rh}_2(\text{S-p-Ph-TPCP})_4$ (3.5 mg, 1 mol%, 2.0 μmol), 3-(4-ethynyl-1-oxoisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH_2Cl_2 (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO_2 , gradient of 30% to 90% EtOAc in hexanes)

afforded 2,2,2-trichloroethyl (1*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (76 mg, 0.12 mmol, 62% yield).

Compound **19b** was prepared in the same manner using Rh₂(*R*-p-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (79 mg, 0.13 mmol, 64% yield).

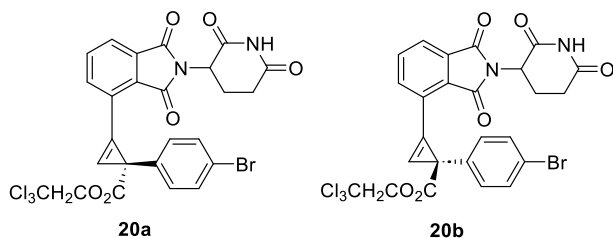
HRMS (APCI): *m/z* 610.9536 [(M+H)⁺ requires 610.9537], calc'd for C₂₅H₁₉O₅N₂⁷⁹Br³⁵Cl₃

¹H NMR (600 MHz, CDCl₃, reported as a mixture of diastereomers) δ 8.27 (s, 1H), 7.96 (d, *J* = 7.6 Hz, 1H), 7.74 (t, *J* = 7.5 Hz, 1H), 7.58 (td, *J* = 7.6, 2.7 Hz, 1H), 7.444 and 7.437 (d, *J* = 8.5 Hz, 2H), 7.39 and 7.38 (s, 1H), 7.29 and 7.28 (d, *J* = 8.5 Hz, 2H), 5.25 and 5.22 (dd, *J* = 13.4, 5.2 Hz, 1H), 4.81 and 4.80 (d, *J* = 12 Hz, 1H), 4.762 and 4.760 (d, *J* = 12 Hz, 1H), 4.60 and 4.58 (d, *J* = 15.7 Hz, 1H), 4.43 and 4.40 (d, 16.5 Hz, 1H), 2.95-2.89 (m, 1H), 2.88-2.81 (m, 1H), 2.40-2.30 (m, 1H), 2.26 – 2.18 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl₃, reported as a mixture of diastereomers) δ 171.92, 171.90, 171.0, 169.5, 169.4, 168.60, 168.58, 142.41, 142.40, 138.6, 138.5, 133.02, 133.97, 132.60, 132.57, 131.6, 129.9, 129.32, 126.29, 121.3, 120.32, 120.29, 113.7, 113.3, 102.41, 102.37, 95.0, 74.5, 74.4, 52.1, 52.0, 46.8, 46.7, 32.5, 32.4, 31.6, 23.6.

SFC analysis: **19a** (Trefoil® AMY1, 30% 1:1 MeOH:^tPrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 98:2 d.r.: *t*_R (major diastereomers) = 3.00 and 3.55 min, *t*_R (minor diastereomers) = 4.41 and 5.81 min. **19b** indicated the opposite diastereomers in 98:2 d.r.

Purity (SFC): **19a** – 98%; **19b** – 95%



2,2,2-trichloroethyl (1*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate (20a).

2,2,2-trichloroethyl (1*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate (20b).

Compound **20a** was prepared following GP 2, using Rh₂(*S*-p-Ph-TPCP)₄ (3.5 mg, 1 mol%, 2.0 μmol), 2-(2,6-dioxopiperidin-3-yl)-4-ethynylisindoline-1,3-dione (56 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH₂Cl₂ (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO₂, gradient of 30% to 55% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (93 mg, 0.15 mmol, 76% yield).

Compound **20b** was prepared in the same manner using Rh₂(*R*-p-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (99 mg, 0.16 mmol, 79% yield).

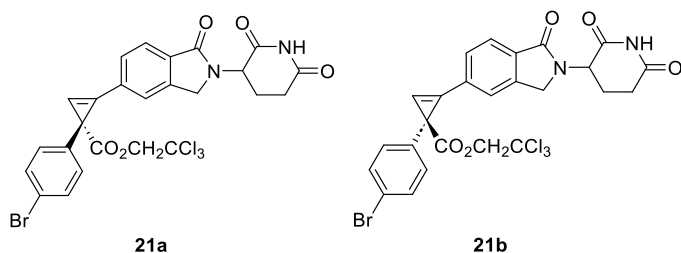
HRMS (APCI): *m/z* 624.9330 [(M+H)⁺ requires 624.9330], calc'd for C₂₅H₁₇O₆N₂⁷⁹Br³⁵Cl₃

¹H NMR (600 MHz, CDCl₃, reported as a mixture of diastereomers) δ 8.29 (s, 1H), 7.95 (d, *J* = 7.7 Hz, 1H), 7.74 (t, *J* = 7.6 Hz, 1H), 7.584 and 7.58 (t, *J* = 7.7 Hz, 1H), 7.443 and 7.436 (d, *J* = 8.6 Hz, 2H), 7.39 and 7.38 (s, 1H), 7.29 and 7.28 (d, *J* = 8.6 Hz, 2H), 5.26 and 5.22 (dd, *J* = 13.4, 5.0 Hz, 1H), 4.81 and 4.79 (d, *J* = 12.0 Hz, 1H), 4.761 and 4.760 (d, *J* = 11.9 Hz, 1H), 4.61 and 4.56 (d, *J* = 16.6 Hz, 1H), 4.43 and 4.40 (d, *J* = 16.5 Hz, 1H), 2.97-2.87 (m, 1H), 2.87-2.79 (m, 1H), 2.41-2.29 (m, 1H), 2.27 – 2.11 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl₃, reported as a mixture of diastereomers) δ 171.8, 170.9, 168.0, 166.59, 166.57, 166.24, 166.22, 138.3, 138.2, 135.8, 135.7, 134.9, 133.1, 131.5, 130.1, 129.91, 129.86, 125.2, 122.70, 122.68, 121.2, 112.48, 112.45, 108.11, 108.06, 95.11, 95.08, 74.49, 74.47, 49.6, 33.92, 33.91, 31.5, 22.7.

SFC analysis: **20a** (Chiralcel® OX-3, 30% 1:1 MeOH:^tPrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 80:20 d.r.: *t*_R (major diastereomers) = 6.92 and 12.88 min, *t*_R (minor diastereomers) = 4.57 and 4.97 min. **20b** indicated the opposite diastereomers in 81:19 d.r.

Purity (SFC): **20a** – 99%; **20b** – 98%



2,2,2-trichloroethyl (1S)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate (21a).

2,2,2-trichloroethyl (1R)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate (21b).

Compound **21a** was prepared following **GP 2**, using $\text{Rh}_2(\text{S-p-Ph-TPCP})_4$ (3.5 mg, 1 mol%, 2.0 μmol), 3-(5-ethynyl-1-oxoisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH_2Cl_2 (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO_2 , gradient of 35% to 55% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (S)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (48 mg, 78 μmol , 39% yield).

Compound **21b** was prepared in the same manner using $\text{Rh}_2(\text{R-p-Ph-TPCP})_4$, which afforded 2,2,2-trichloroethyl (R)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (46 mg, 74 μmol , 37% yield).

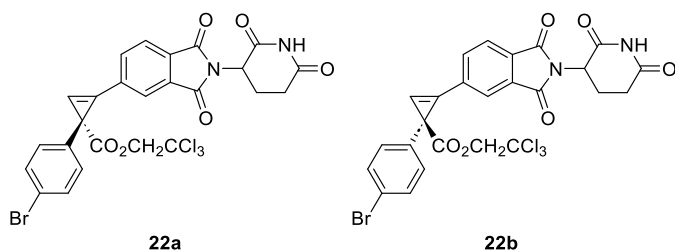
HRMS (APCI): m/z 610.9536 [(M+H)⁺ requires 610.9537], calc'd for $\text{C}_{25}\text{H}_{19}\text{O}_5\text{N}_2^+\text{Br}^-\text{Cl}_3$

¹H NMR (400 MHz, CDCl_3 , reported as a mixture of diastereomers) δ 8.27 (s, 1H), 7.952 and 7.950 (d, J = 7.9 Hz, 1H), 7.79 – 7.71 (m, 1H), 7.69 – 7.64 (m, 1H), 7.43 (d, J = 8.5 Hz, 2H), 7.39 (s, 1H), 7.29 (d, J = 8.6 Hz, 2H), 5.23 (dd, J = 13.3, 5.2 Hz, 1H), 4.823 and 4.816 (d, J = 12.0 Hz, 1H), 4.764 and 4.760 (d, J = 12.0 Hz, 1H), 4.52 and 4.51 (d, J = 16.3 Hz, 1H), 4.37 and 4.35 (d, J = 16.3 Hz, 1H), 2.98 – 2.88 (m, 2H), 2.88 – 2.76 (m, 1H), 2.35 (qd, J = 13.1, 5.1 Hz, 1H), 2.26 – 2.15 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl_3 , reported as a mixture of diastereomers) δ ¹³C NMR (101 MHz, CDCl_3) δ 172.08, 172.06, 171.1, 169.5, 168.5, 142.1, 138.5, 133.17, 133.15, 131.50, 131.48, 130.32, 130.30, 129.95, 128.4, 125.0, 124.5, 121.13, 121.10, 116.2, 102.2, 102.1, 95.1, 74.42, 74.39, 52.1, 47.1, 33.3, 31.6, 23.5.

SFC analysis: **21a** (Chiralpak® AS3, 20% 1:1:1 EtOH:PrOH:MeCN with 20 mM NH_4HCO_2 in CO_2 , 2.5 mL/min, diode array) indicated >99:1 d.r.: t_R (major diastereomers) = 3.94 and 6.41 min, t_R (minor diastereomers) = 3.41 and 4.88 min. **21b** indicated the opposite diastereomers in 99:1 d.r.

Purity (SFC): **21a** – 97%; **21b** – 95%



2,2,2-trichloroethyl (1S)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate (22a).

2,2,2-trichloroethyl (1R)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate (22b).

Compound **22a** was prepared following **GP 2**, using $\text{Rh}_2(\text{S-p-Ph-TPCP})_4$ (3.5 mg, 1 mol%, 2.0 μmol), 2-(2,6-dioxopiperidin-3-yl)-5-ethynylisindoline-1,3-dione (56 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH_2Cl_2 (2 mL) at room temperature for two

hours. Purification via flash column chromatography (SiO₂, gradient of 20% to 50% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (87 mg, 0.14 mmol, 69% yield). Compound **22b** was prepared in the same manner using Rh₂(*R*-p-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1,3-dioxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (88 mg, 0.14 mmol, 70% yield).

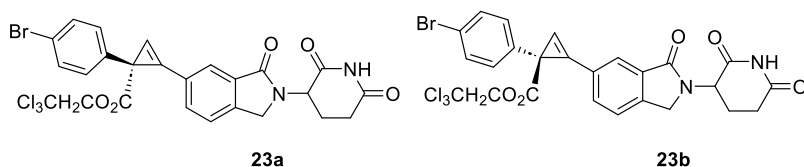
HRMS (APCI): *m/z* 624.9341 [(M+H)⁺ requires 624.9344], calc'd for C₂₅H₁₆⁷⁹Br₃Cl₃N₂O₆

¹H NMR (500 MHz, CDCl₃, reported as a mixture of diastereomers) δ 8.26 (s, 1H), 8.11-8.10 (m, 1H), 7.98-7.94 (m, 2H), 7.54 (s, 1H), 7.45 (d, *J*=8.5 Hz, 2H), 7.28 (d, *J*=8.5 Hz, 2H), 5.01 (dd, *J*=12.5, 5.1 Hz, 1H), 4.84 and 4.83 (d, *J*=12.0 Hz, 1H), 4.76 (d, *J*=12.0 Hz, 1H), 2.94-2.72 (m, 3H), 2.19-2.14 (m, 1H).

¹³C{¹H} NMR (126 MHz, CDCl₃, reported as a mixture of diastereomers) δ 171.5, 171.0, 167.9, 166.5, 166.4, 137.9, 135.7, 132.7, 132.6, 131.6, 131.2, 129.90, 129.89, 124.89, 124.88, 124.6, 121.4, 116.1, 116.0, 104.5, 95.0, 74.5, 60.5, 49.7, 33.7, 31.48, 31.46, 22.7, 22.6, 14.3.

SFC analysis: **22a** (Chiralcel® OJ-3, 20% 1:1 MeOH:ⁱPrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 98:2 d.r.: *t*_R (major diastereomers) = 7.10 and 8.24 min, *t*_R (minor diastereomers) = 9.20 and 11.52 min. **22b** indicated the opposite diastereomers in 99:1 d.r.

Purity (SFC): **22a** – 98%; **22b** – 95%



2,2,2-trichloroethyl (1*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate (23a**).**

2,2,2-trichloroethyl (1*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate (23b**).**

Compound **23a** was prepared following **GP 2**, using Rh₂(*S*-p-Ph-TPCP)₄ (3.5 mg, 1 mol%, 2.0 μmol), 3-(6-ethynyl-1-oxoisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH₂Cl₂ (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO₂, gradient of 60% to 75% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (76 mg, 12 μmol, 62% yield).

Compound **23b** was prepared in the same manner using Rh₂(*R*-p-Ph-TPCP)₄, which afforded 2,2,2-trichloroethyl (1*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-5-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (77 mg, 0.13 mmol, 63% yield).

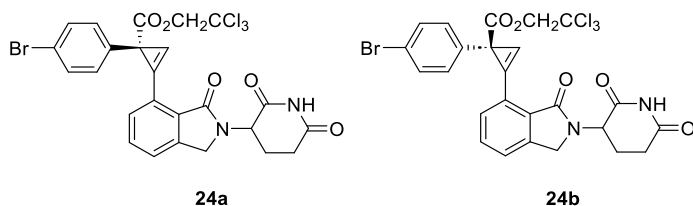
HRMS (APCI): *m/z* 610.9540 [(M+H)⁺ requires 610.9537], calc'd for C₂₅H₁₉O₅N₂⁷⁹Br³⁵Cl₃

¹H NMR (600 MHz, CDCl₃, reported as a mixture of diastereomers) δ 8.17 (s, 1H), 8.13 (d, *J* = 6.2 Hz, 1H), 7.81 (t, *J* = 7.9 Hz, 1H), 7.54 (d, *J* = 7.9 Hz, 1H), 7.42 (d, *J* = 8.6 Hz, 2H), 7.32 (s, 1H), 7.31 (d, *J* = 8.4 Hz, 2H), 5.29 – 5.17 (m, 1H), 4.82 and 4.81 (d, *J* = 11.9 Hz, 1H), 4.76 and 4.75 Hz (d, *J* = 11.9 Hz, 1H), 4.54 (d, *J* = 16.7 Hz, 1H), 4.39 (d, *J* = 16.5 Hz, 1H), 2.97 – 2.90 (m, 1H), 2.88 – 2.77 (m, 1H), 2.41 – 2.30 (m, 1H), 2.25 – 2.18 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl₃, reported as a mixture of diastereomers) δ 172.2, 169.5, 171.3, 168.5, 143.3, 138.6, 133.61, 133.57, 132.54, 132.52, 131.4, 130.1, 125.65, 125.61, 125.4, 123.9, 121.02, 121.00, 116.3, 116.2, 100.7, 95.1, 74.4, 52.03, 52.00, 47.3, 33.3, 31.6, 23.5.

SFC analysis: **23a** (Chiralcel® OJ-3, 30% 1:1 MeOH:ⁱPrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 98:2 d.r.: *t*_R (major diastereomers) = 1.81 and 2.40 min, *t*_R (minor diastereomers) = 2.12 and 3.16 min. **23b** indicated the opposite diastereomers in 98:2 d.r.

Purity (SFC): **23a** – 99%; **23b** – 99%



2,2,2-trichloroethyl (1*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate (24a).

2,2,2-trichloroethyl (1*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate (24b).

Compound **24a** was prepared following **GP 2**, using $\text{Rh}_2(\text{S-p-Ph-TPCP})_4$ (3.5 mg, 1 mol%, 2.0 μmol), 3-(7-ethynyl-1-oxoisindolin-2-yl)piperidine-2,6-dione (54 mg, 1.0 equiv, 0.20 mmol), 2,2,2-trichloroethyl 2-(4-bromophenyl)-2-diazoacetate (82 mg, 1.1 equiv, 0.22 mmol), and CH_2Cl_2 (2 mL) at room temperature for two hours. Purification via flash column chromatography (SiO_2 , gradient of 45% to 70% EtOAc in hexanes) afforded 2,2,2-trichloroethyl (1*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (99 mg, 0.16 mmol, 81% yield).

Compound **24b** was prepared in the same manner using $\text{Rh}_2(\text{R-p-Ph-TPCP})_4$, which afforded 2,2,2-trichloroethyl (1*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-3-oxoisindolin-4-yl)cycloprop-2-ene-1-carboxylate as an amorphous white solid. (97 mg, 0.16 mmol, 79% yield).

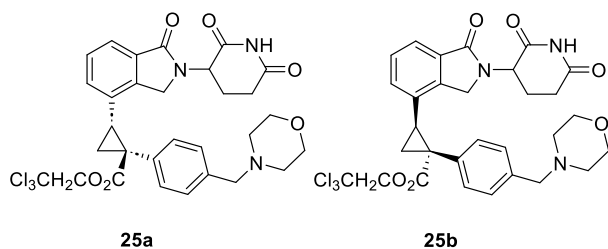
HRMS (APCI): m/z 610.9545 [($\text{M}+\text{H}$)⁺ requires 610.9537], calc'd for $\text{C}_{25}\text{H}_{19}\text{O}_5\text{N}_2^{79}\text{Br}^{35}\text{Cl}_3$

¹H NMR (600 MHz, CDCl_3) δ 8.17 (s, 1H), 7.66 (d, $J = 3.6$ Hz, 1H), 7.63 – 7.54 (m, 2H), 7.51 and 7.50 (s, 1H), 7.41 and 7.40 (d, $J = 8.6$ Hz, 2H), 7.34 and 7.33 (d, $J = 8.6$ Hz, 2H), 5.25 and 5.23 (dd, $J = 13.0, 5.1$ Hz, 1H), 4.85 and 4.83 (d, $J = 11.9$ Hz, 1H), 4.74 and 4.73 (d, $J = 12.0$ Hz, 1H), 2.95–2.89 (m, 1H), 2.89–2.84 (m, 1H), 2.43–2.33 (m, 1H), 2.29–2.14 (m, 1H).

¹³C{¹H} NMR (101 MHz, CDCl_3 , reported as a mixture of diastereomers) δ 172.5, 172.4, 171.3, 169.7, 168.4, 143.1, 139.1, 139.0, 132.2, 131.29, 131.28, 130.45, 130.43, 130.37, 130.26, 130.24, 124.8, 122.5, 120.80, 120.77, 112.7, 112.5, 106.0, 105.9, 95.3, 74.4, 74.3, 52.1, 52.0, 47.1, 47.0, 33.27, 33.26, 31.7, 23.5.

SFC analysis: **24a** (Chiralpak® AS3, 20% 1:1 MeOH:PrOH with 0.2% formic acid in CO_2 , 2.5 mL/min, diode array) indicated 61:39 d.r.: t_R (major diastereomers) = 4.05 and 5.21 min, t_R (minor diastereomers) = 4.49 and 7.67 min. **24b** indicated the opposite diastereomers in 60:40 d.r.

Purity (SFC): **24a** – 95%; **24b** – 95%



2,2,2-trichloroethyl (1*S*,2*R*)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)-1-(4-(morpholinomethyl)phenyl)cyclopropane-1-carboxylate (25a).

2,2,2-trichloroethyl (1*R*,2*S*)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-4-yl)-1-(4-(morpholinomethyl)phenyl)cyclopropane-1-carboxylate (25b).

A 10 mL round-bottom flask was charged with a PTFE magnetic stir bar and ca. 200% w/w 4 Å molecular sieves, then flame dried under vacuum. The flask was then backfilled with dry nitrogen. $\text{Rh}_2(\text{S-tetra-p-BrPPTTL})_4$ (3.7 mg, 1 mol%, 1.0 μmol) and 3-(1-oxo-4-vinylisindolin-2-yl)piperidine-2,6-dione (27 mg, 1.0 equiv, 0.10 mmol) were charged to the flask, and the flask was evacuated and backfilled with nitrogen. The flask was equipped with an argon balloon and rubber septum, and dry CH_2Cl_2 (0.5 mL) along with 0.10 mL 1,1,1,3,3,3-hexafluoroisopropanol (10 equiv, 1.0 mmol) were charged via syringe. A solution of 2,2,2-trichloroethyl 2-diazo-2-(4-(morpholinomethyl)phenyl)acetate (43 mg, 1.1 equiv, 0.11 mmol) in dry CH_2Cl_2 (0.5 mL, prepared in the same manner under inert atmosphere) was added dropwise to the stirred reaction at

room temperature. The reaction was allowed to stir for two hours, after which the reaction was filtered through Celite®, rinsing the filter pad with CH₂Cl₂. The crude residue was dry-mounted onto silica gel and was purified by flash column chromatography (SiO₂, 0-5% methanol in CH₂Cl₂), which afforded 2,2,2-trichloroethyl (1*S*,2*R*)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisoindolin-4-yl)-1-(4-(morpholinomethyl)phenyl)cyclopropane-1-carboxylate as an amorphous white solid (50 mg, 79 μmol, 79% yield).

Compound **25b** was prepared in the same manner using Rh₂(*R*-tetra-*p*-BrPPTTL)₄, which afforded 2,2,2-trichloroethyl (1*R*,2*S*)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisoindolin-4-yl)-1-(4-(morpholinomethyl)phenyl)cyclopropane-1-carboxylate as an amorphous white solid as an amorphous white solid. (49 mg, 77 μmol, 77% yield).

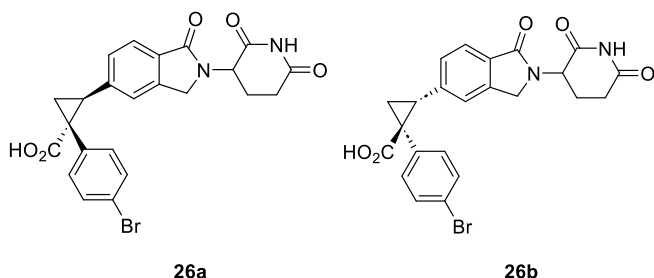
HRMS (APCI): *m/z* 634.1279 ([*M*+*H*]⁺ requires 634.1273), calc'd for C₃₀H₃₁O₆N₃³⁵Cl₃

¹H NMR (400 MHz, CDCl₃, reported as a mixture of diastereomers) δ 8.54 (s, 1H), 7.595 and 7.593 (d, *J* = 7.6 Hz, 1H), 7.10 – 7.00 (m, 3H), 6.96 and 6.93 (d, *J* = 12.3 Hz, 2H), 5.29 and 5.25 (dd, *J* = 9.6, 5.2 Hz, 1H), 4.86 and 4.85 (d, *J* = 12.0 Hz, 1H), 4.664 and 4.658 (d, *J* = 12.0 Hz, 1H), 4.58 and 4.56 (d, *J* = 16.0 Hz, 1H), 4.42 and 4.38 (d, *J* = 16.2 Hz, 1H), 3.69 – 3.58 (m, 4H), 3.43 – 3.36 (m, 2H), 3.07 (ddd, *J* = 9.4, 7.4, 2.1 Hz, 1H), 3.01 – 2.77 (m, 2H), 2.42 – 2.19 (m, 7H), 2.15 (ddd, *J* = 7.5, 5.3, 2.2 Hz, 1H).

¹³C{¹H} NMR (101 MHz, CDCl₃, reported as a mixture of diastereomers) δ 171.9, 171.8, 171.29, 171.27, 169.7, 169.53, 169.48, 141.7, 141.6, 137.3, 137.2, 132.0, 131.9, 131.5, 131.4, 131.30, 131.27, 131.19, 131.18, 129.12, 129.06, 129.05, 128.0, 122.6, 95.3, 95.2, 74.4, 74.3, 67.0, 62.9, 53.50, 53.46, 52.03, 51.96, 46.61, 46.50, 37.1, 37.0, 31.7, 29.8, 29.7, 29.6, 23.63, 23.60, 19.7, 19.6.

SFC analysis: **25a** (Trefoil® CEL2, 35% 1:1 MeOH:PrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 83:17 d.r.: *t*_R (major diastereomers) = 6.04 and 8.83 min, *t*_R (minor diastereomers) = 6.80 and 13.50 min. **25b** indicated the opposite diastereomers in 85:15 d.r.

Purity (SFC): 25a – 99%; 25b – 99%



26a

26b

(1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisoindolin-5-yl)cyclopropane-1-carboxylic acid (26a).

(1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisoindolin-5-yl)cyclopropane-1-carboxylic acid (26b).

A 100 mL round-bottom flask was charged with a PTFE magnetic stir bar and zinc dust (754 mg, 16 equiv, 11.5 mmol) under ambient conditions, followed by 2,2,2-trichloroethyl (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisoindolin-5-yl)cyclopropane-1-carboxylate (443 mg, 1.0 equiv, 0.72 mmol). Tetrahydrofuran (16.6 mL) was charged via syringe followed by acetate buffer (10 mL, pH = 3.7, 1.2 M in H₂O, NaOAc/AcOH). The reaction was stirred at room temperature for 18 hours, after which the reaction was filtered through Celite®, rinsing with ethyl acetate. The aqueous layer was extracted thrice with 50 mL portions of ethyl acetate, after which the combined organics were washed with 150 mL brine. The combined organics were dried over magnesium sulfate, then filtered and concentrated in vacuo. The crude material was purified via flash column chromatography (C18, 10-95% MeCN in H₂O, 0.1% trifluoroacetic acid buffer), which gave (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisoindolin-5-yl)cyclopropane-1-carboxylic acid as an amorphous white solid (280 mg, 0.58 mmol, 83% yield).

Compound **26b** was prepared in the same manner using zinc dust (727 mg, 16 equiv, 11.1 mmol), 2,2,2-trichloroethyl (1*R*,2*S*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisoindolin-5-yl)cyclopropane-

1-carboxylate (427 mg, 1.0 equiv, 0.70 mmol), tetrahydrofuran (16.0 mL), and acetate buffer (9.6 mL), which gave (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylic acid (286 mg, 0.59 mmol, 82% yield) as an amorphous white solid.

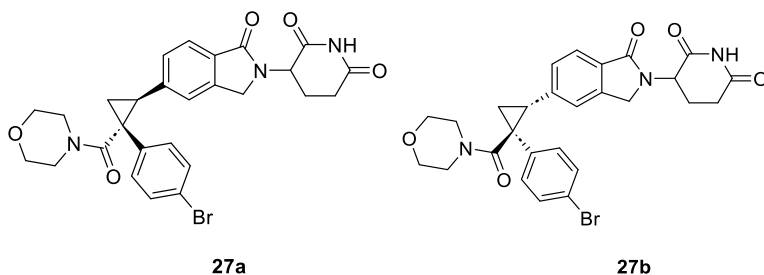
HRMS (APCI): m/z 483.0559 ($[M+H]^+$ requires 483.0550), calc'd for C₂₃H₂₀O₅N₂⁷⁹Br

¹H NMR (600 MHz, CD₃OD, reported as a mixture of diastereomers) δ 7.55 – 7.42 (m, 1H), 7.26 (d, J = 8.1 Hz, 2H), 7.14 and 7.10 (s, 1H), 7.05-6.98 (m, 1H, partially obscured by signal at 7.00), 7.00 (d, J = 8.0 Hz, 3H), 5.10 and 5.07 (dd, J = 10.9, 5.2 Hz, 1H), 4.39 and 4.34 (d, J = 17.0 Hz, 1H), 4.27 and 4.26 (d, J = 17.0 Hz, 1H), 3.27 (t, J = 8.2 Hz, 1H), 2.87 (ddd, J = 18.5, 13.6, 5.4 Hz, 1H), 2.81 – 2.70 (m, 1H), 2.43 (pd, J = 13.5, 4.5 Hz, 1H), 2.17-2.08 (m, 3H).

¹³C NMR (101 MHz, CD₃OD) δ 176.2, 174.7, 172.22, 172.20, 171.19, 171.17, 143.5, 143.01, 143.29, 135.7, 135.6, 135.0, 134.9, 131.9, 131.0, 130.9, 129.7, 129.3, 124.3, 124.0, 123.73, 123.71, 122.0, 53.6, 53.5, 38.6, 38.5, 34.0, 32.3, 24.0, 20.5, 20.4.

SFC analysis: **26a** (Chiralpak® AS3, 20% 1:1 EtOH:PrOH with 0.2% formic acid in CO₂, 2.5 mL/min, diode array) indicated 99:1 d.r.: t_R (major diastereomers) = 4.56 and 6.30 min, t_R (minor diastereomers) = 5.73 and 8.81 min. **26b** indicated the opposite diastereomers in 99:1 d.r.

Purity (SFC): **26a** – 98%; **26b** – 98%



3-(5-((1*S*,2*R*)-2-(4-bromophenyl)-2-(morpholine-4-carbonyl)cyclopropyl)-1-oxoisindolin-2-yl)piperidine-2,6-dione (27a).

3-(5-((1*R*,2*S*)-2-(4-bromophenyl)-2-(morpholine-4-carbonyl)cyclopropyl)-1-oxoisindolin-2-yl)piperidine-2,6-dione (27b).

A 4 mL scintillation vial equipped with a PTFE magnkjetic stir bar was flame-dried under vacuum and backfilled with dry nitrogen. The vial was charged with (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylic acid (50 mg, 1.0 equiv, 0.10 mmol) and N-(chloro(dimethylamino)methylene)-N-methylmethanaminium hexafluorophosphate(V) (34 mg, 1.2 equiv, 0.12 mmol). Morpholine (11 μ L, 1.3 equiv, 0.13 mmol), 1-methyl-1H-imidazole (28 μ L, 3.5 equiv, 0.35 mmol), and acetonitrile (0.25 mL) were charged successively by syringe, and the reaction was stirred at room temperature for 18 hours. The reaction was stirred at room temperature for 18 hours, after which the reaction was partitioned between water (50 mL) and ethyl acetate (50 mL). The aqueous layer was extracted thrice with 50 mL portions of ethyl acetate, after which the combined organics were washed with 150 mL of brine. The combined organics were dried over magnesium sulfate, then filtered and concentrated in vacuo. The crude material was purified via flash column chromatography (SiO₂, 0-10% methanol in CH₂Cl₂), which gave 3-(5-((1*S*,2*R*)-2-(4-bromophenyl)-2-(morpholine-4-carbonyl)cyclopropyl)-1-oxoisindolin-2-yl)piperidine-2,6-dione as an amorphous white solid (31 mg, 55 μ mol, 55% yield).

Compound **27b** was prepared in the same manner from (1*S*,2*R*)-1-(4-bromophenyl)-2-(2-(2,6-dioxopiperidin-3-yl)-1-oxoisindolin-5-yl)cyclopropane-1-carboxylic acid, which gave 3-(5-((1*R*,2*S*)-2-(4-bromophenyl)-2-(morpholine-4-carbonyl)cyclopropyl)-1-oxoisindolin-2-yl)piperidine-2,6-dione as an amorphous white solid (34 mg, 60 μ mol, 60% yield).

HRMS (APCI): m/z 552.1139 ($[M+H]^+$ requires 552.1129), calc'd for C₂₇H₂₇O₅N₃⁷⁹Br

¹H NMR (600 MHz, CDCl₃, reported as a mixture of diastereomers) δ 7.99 and 7.98 (s, 1H), 7.60 (d, J and 7.58 (d, J = 5.9 Hz, 1H), 7.29 – 7.26 (m, 2H, partially obscured by solvent signal), 7.17 and 7.10 (s, 1H), 7.07

and 7.02 (dd, $J = 7.9, 1.4$ Hz, 1H), 6.99 – 6.92 (m, 2H), 5.160 and 5.156 (dd, $J = 13.3, 5.1$ Hz, 1H), 4.343 and 4.338 (d, $J = 15.9$ Hz, 1H), 4.17 and 4.16 (d, $J = 15.9$ Hz, 1H), 3.74 – 3.37 (m, 7H), 3.30 (dt, $J = 9.2, 6.9$ Hz, 1H), 3.18 (s, 1H), 2.92 – 2.87 (m, 1H), 2.80 (ddd, $J = 18.1, 13.4, 5.4$ Hz, 1H), 2.37 – 2.25 (m, 1H), 2.23 – 2.13 (m, 1H), 2.08 (ddd, $J = 7.0, 5.8, 3.2$ Hz, 1H), 1.68 (ddd, $J = 9.1, 5.8, 4.5$ Hz, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 171.74, 171.72, 170.27, 170.23, 170.0, 169.1, 141.4, 141.25, 141.21, 134.26, 134.21, 131.74, 131.71, 129.87, 129.7, 128.6, 128.3, 123.8, 123.51, 123.48, 123.2, 121.2, 121.1, 66.4, 51.8, 51.7, 46.80, 46.76, 38.1, 37.9, 31.6, 30.2, 30.1, 23.4, 16.1, 16.0.

SFC analysis: **27a** (Chiralcel® OJ-3, 20% 1:1 MeOH:PrOH with 0.2% formic acid in CO_2 , 2.5 mL/min, diode array) indicated 99:1 d.r.: t_{R} (major diastereomers) = 2.80 and 3.65 min, t_{R} (minor diastereomers) = 3.21 and 4.77 min. **27b** indicated the opposite diastereomers in 99:1 d.r.

Purity (SFC): **27a** – 98%; **27b** – 98%

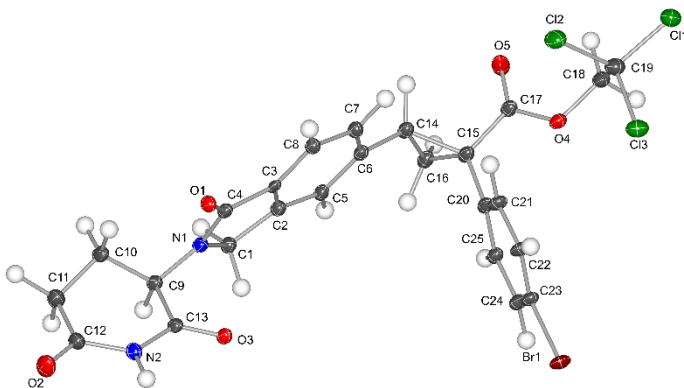
Section 3: X-ray Crystallographic Data for *R*-15a

Submitted by: **William Tracy, Huw Davies Lab**

Solved by: **John Bacsá, Mackenzie Young**

$R_1 = 5.45\%$

Crystal Data and Experimental



matrix least squares minimisation on F^2 .

Crystal Data. $\text{C}_{25}\text{H}_{20}\text{BrCl}_3\text{N}_2\text{O}_5$, $M_r = 614.709$, monoclinic, $P2_1$ (No. 4), $a = 6.1901(4)$ Å, $b = 9.2069(6)$ Å, $c = 22.1261(17)$ Å, $\beta = 90.543(7)^\circ$, $\alpha = \gamma = 90^\circ$, $V = 1260.94(16)$ Å³, $T = 104(6)$ K, $Z = 2$, $Z' = 1$, $\mu(\text{Mo } K\alpha) = 1.988$, 15376 reflections measured, 5610 unique ($R_{\text{int}} = 0.0782$) which were used in all calculations. The final wR_2 was 0.1153 (all data) and R_1 was 0.0545 ($I \geq 2 \sigma(I)$).

Experimental. Single colourless plate-shaped crystals of WT-05-553 were crystallized from methanol by slow evaporation. A suitable crystal with dimensions $0.13 \times 0.10 \times 0.05$ mm³ was selected and mounted on a loop with paratone on a XtaLAB Synergy, Dualflex, HyPix diffractometer. The crystal was kept at a steady $T = 104(6)$ K during data collection. The structure was solved with the ShelXT (Sheldrick, 2015) solution program using dual methods and by using Olex2 1.5-alpha (Dolomanov et al., 2009) as the graphical interface. The model was refined with olex2.refine 1.5-alpha (Bourhis et al., 2015) using full

Compound	WT-05-553
Formula	C ₂₅ H ₂₀ BrCl ₃ N ₂ O ₅
<i>D</i> _{calc.} / g cm ⁻³	1.619
<i>μ</i> /mm ⁻¹	1.988
Formula Weight	614.709
Color	colorless
Shape	plate-shaped
Size/mm ³	0.13×0.10×0.05
<i>T</i> /K	104(6)
Crystal System	monoclinic
Flack Parameter	-0.006(8)
Hooft Parameter	-0.006(8)
Space Group	<i>P</i> 2 ₁
<i>a</i> /Å	6.1901(4)
<i>b</i> /Å	9.2069(6)
<i>c</i> /Å	22.1261(17)
<i>α</i> /°	90
<i>β</i> /°	90.543(7)
<i>γ</i> /°	90
<i>V</i> /Å ³	1260.94(16)
<i>Z</i>	2
<i>Z</i> '	1
Wavelength/Å	0.71073
Radiation type	Mo K _α
<i>θ</i> _{min} /°	3.41
<i>θ</i> _{max} /°	27.57
Measured Refl's.	15376
Indep't Refl's	5610
Refl's I≥2 σ(I)	4416
<i>R</i> _{int}	0.0782
Parameters	451
Restraints	823
Largest Peak	1.5961
Deepest Hole	-0.6735
GooF	1.0057
<i>wR</i> ₂ (all data)	0.1153
<i>wR</i> ₂	0.1076
<i>R</i> ₁ (all data)	0.0786
<i>R</i> ₁	0.0545

Structure Quality Indicators

Reflections:	d min (MoK α) 2 Θ =55.1°	0.77	I/ σ (I)	10.2	R _{int} m=2.69	7.82%	Full 50.5°	95.6		
Refinement:	Shift	-0.001	Max Peak	1.6	Min Peak	-0.7	Goof	1.006	Hoof	-0.006(8)

A colorless plate-shaped crystal with dimensions 0.13 × 0.10 × 0.05 mm³ was mounted on a loop with paratone. Data were collected using a XtaLAB Synergy, Dualflex, HyPix diffractometer equipped with an Oxford Cryosystems low-temperature device operating at $T = 104(6)$ K.

Data were measured using ω scans with Mo K α radiation. The diffraction pattern was indexed and the total number of runs and images was based on the strategy calculation from the program CrysAlisPro system (CCD 43.92a 64-bit (release 05-10-2023)). The maximum resolution that was achieved was $\Theta = 27.57^\circ$ (0.77 Å).

The unit cell was refined using CrysAlisPro 1.171.43.121a (Rigaku OD, 2024) on 3358 reflections, 22% of the observed reflections.

Data reduction, scaling and absorption corrections were performed using CrysAlisPro 1.171.43.121a (Rigaku OD, 2024). The final completeness is 98.65 % out to 27.57° in Θ . An analytical numeric absorption correction using a multifaceted crystal model based on expressions derived by R.C. Clark & J.S. Reid. (Clark, R. C. & Reid, J. S. (1995). Acta Cryst. A51, 887-897) was performed using CrysAlisPro 1.171.43.121a (Rigaku Oxford Diffraction, 2024). An empirical absorption correction using spherical harmonics, implemented in SCALE3 ABSPACK scaling algorithm was also applied. The absorption coefficient μ of this material is 1.988 mm⁻¹ at this wavelength ($\lambda = 0.71073\text{Å}$) and the minimum and maximum transmissions are 0.821 and 0.924.

The structure was solved and the space group $P2_1$ (# 4) determined by the ShelXT (Sheldrick, 2015) structure solution program using dual methods and refined by full matrix least squares minimisation on F^2 using version of olex2.refine 1.5-alpha (Bourhis et al., 2015). All atoms, even hydrogen atoms, were refined anisotropically. Hydrogen atom positions were located from the electron densities and freely refined using Hirshfeld scattering factors. Refinement was by using NoSpherA2, an implementation of non-spherical atom-form-factors (F. Kleemiss, H. Puschmann, O. Dolomanov, S.Grabowsky - <https://doi.org/10.1039/D0SC05526C> – 2020). NoSpherA2 implementation of HAR makes use of tailor-made aspherical atomic form factors calculated from a Hirshfeld-partitioned electron density (ED) not from spherical-atom form factors. The ED was calculated from a Gaussian basis set single determinant SCF wavefunction from DFT using selected functionals for a fragment of this crystal. This fragment was embedded in an electrostatic crystal field by employing cluster charges. The following options were used: software: SOFTWARE: ORCA PARTITIONING: NoSpherA2 INT ACCURACY: Normal METHOD: PBE BASIS SET: x2c-SVP CHARGE: 0 MULTIPLICITY: 1 SOLVATION: Methanol RELATIVISTIC: DKH2 DATE: 2024-05-14_13-44-20

There is a single formula unit in the asymmetric unit, which is represented by the reported sum formula. In other words: Z is 2 and Z' is 1. The moiety formula is C₂₅ H₂₀ Br Cl₃ N₂ O₅.

The Flack parameter was refined to -0.006(8). Determination of absolute structure using Bayesian statistics on Bijvoet differences using the Olex2 results in -0.006(8). The chiral atoms in this structure are: C9(R), C14(S), C15(R). Note: The Flack parameter is used to determine chirality of the crystal studied, the value should be near 0, a value of 1 means that the stereochemistry is wrong and the model should be inverted. A value of 0.5 means that the crystal consists of a racemic mixture of the two enantiomers.

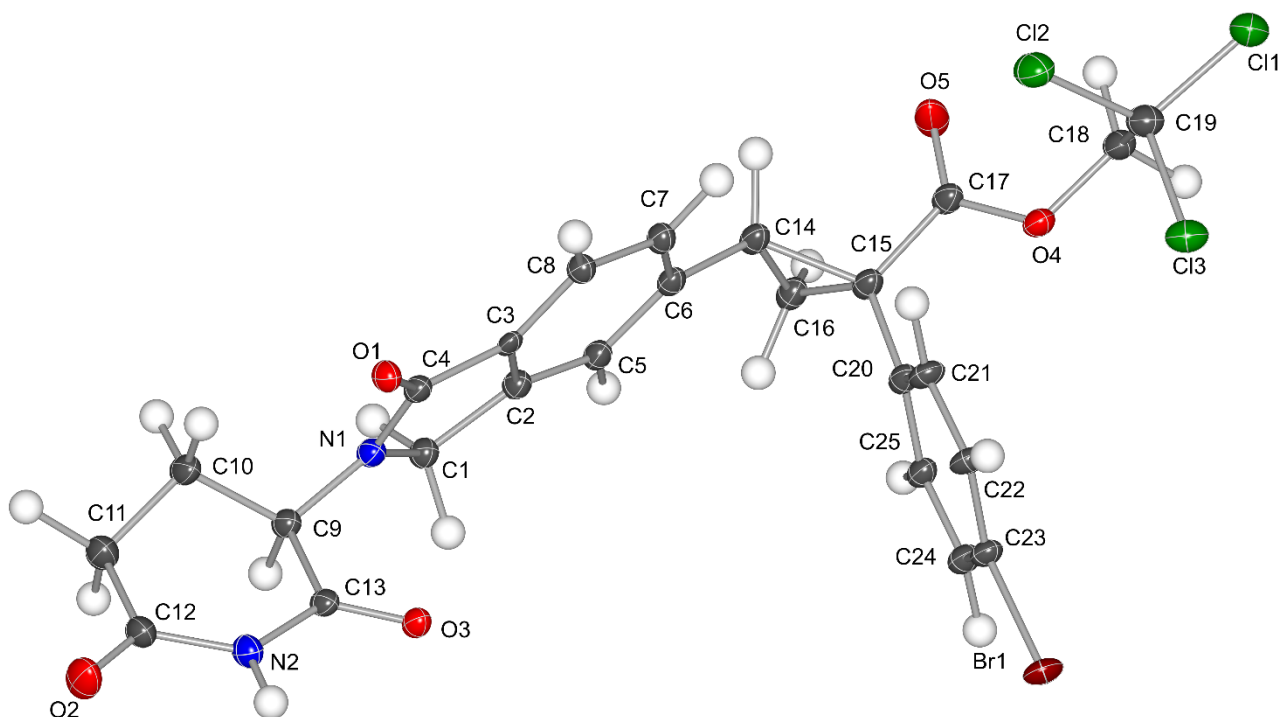
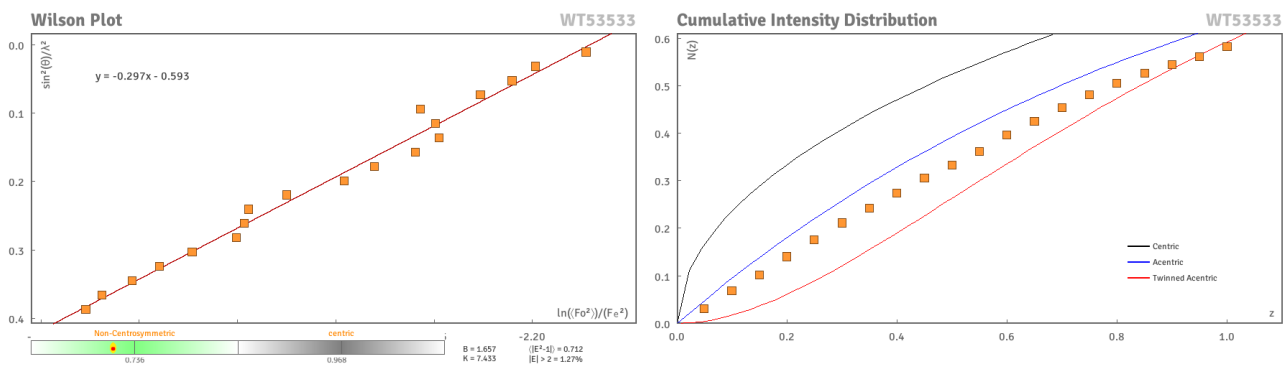
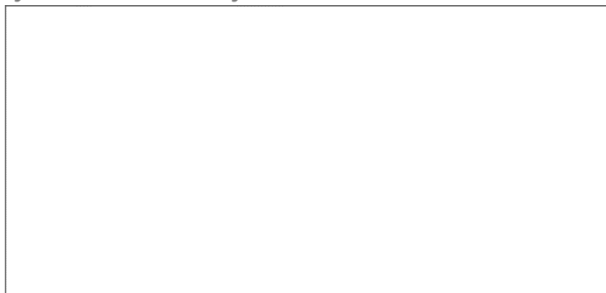


Figure 1 A thermal ellipsoidal representation of the asymmetric unit in the crystal structure (50% probability) which consists of one whole molecule. The chiral atoms in this structure are: C9(R), C14(S), and C15(R).

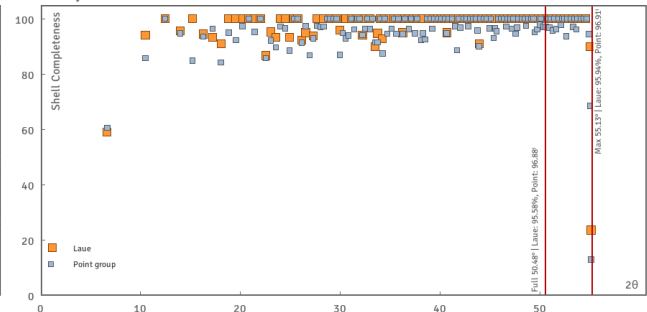
Data Plots: Diffraction Data



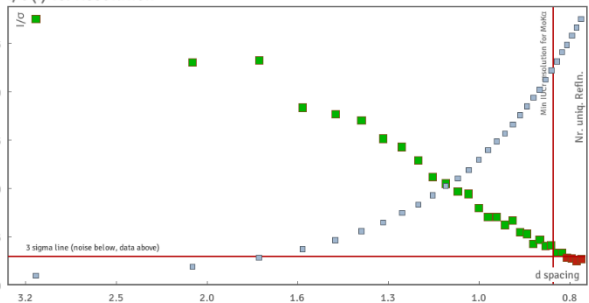
Systematic Absences Intensity Distribution WT53533



Completeness Plot WT53533

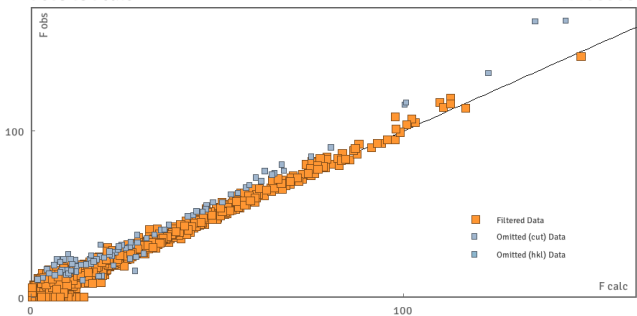


I/σ(I) vs. Resolution WT53533

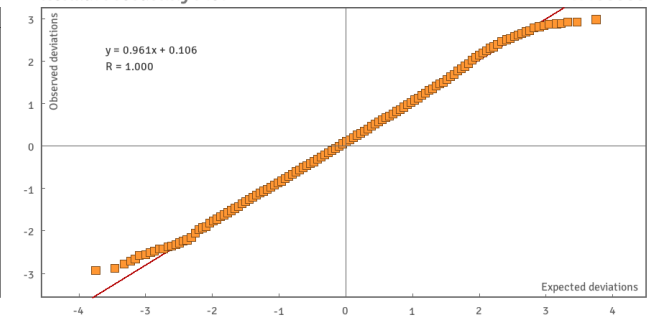


Data Plots: Refinement and Data

Fobs vs Fcalc WT53533



Normal Probability Plot WT53533



Reflection Statistics

Total reflections (after filtering)	15067	Unique reflections	5610
Completeness	0.969	Mean I/σ	9.61
hkl _{max} collected	(8, 11, 28)	hkl _{min} collected	(-8, -11, -28)
hkl _{max} used	(8, 11, 28)	hkl _{min} used	(-8, -11, 0)
Lim d _{max} collected	100.0	Lim d _{min} collected	0.36
d _{max} used	5.98	d _{min} used	0.77
Friedel pairs	3967	Friedel pairs merged	0
Inconsistent equivalents	5	R _{int}	0.079
R _{sigma}	0.0982	Intensity transformed	0
Omitted reflections	309	Omitted by user (OMIT hkl)	3
Multiplicity	(5283, 3149, 948, 182, 41, 3)	Maximum multiplicity	9
Removed systematic absences	0	Filtered off (Shel/OMIT)	0

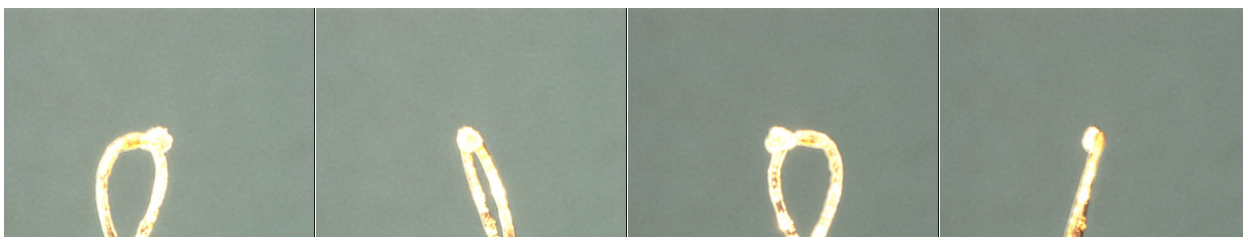


Table 1: Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for WT-05-553. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} .

Atom	x	y	z	U_{eq}
Br1	5371.5(7)	1137.5(3)	1682.0(2)	21.17(12)
Cl3	3100.3(17)	5997.6(11)	86.8(5)	26.1(3)
Cl1	761(2)	8186.8(11)	-588.1(6)	28.2(3)
Cl2	2633(2)	8864.4(11)	589.3(6)	29.2(3)
O1	7753(5)	8378(3)	4542.3(14)	17.2(6)
O2	10793(5)	6114(5)	6695.3(14)	32.8(8)
O3	7875(5)	4979(3)	4887.5(14)	17.5(7)
O4	-410(5)	6480(3)	1003.0(14)	19.3(5)
O5	-2542(6)	8132(4)	1452.0(16)	33.7(9)
N1	4848(6)	7116(4)	4900.2(18)	16.2(6)
N2	9217(6)	5538(4)	5810.9(18)	18.4(8)
C1	2699(7)	6635(5)	4700(2)	16.9(7)
C2	2630(7)	7101(4)	4056(2)	16.0(7)
C3	4518(7)	7884(4)	3935(2)	13.4(7)
C4	5927(7)	7865(4)	4472(2)	15.2(7)
C5	1035(7)	6939(4)	3619(2)	16.3(8)
C6	1334(7)	7600(5)	3054(2)	15.8(6)
C7	3204(7)	8426(4)	2945(2)	16.9(8)
C8	4842(7)	8574(4)	3376(2)	16.2(8)
C9	5785(7)	6673(4)	5473(2)	17.0(6)
C10	6482(8)	7933(5)	5882(2)	22.2(9)
C11	7441(9)	7325(5)	6465(2)	25.4(9)
C12	9287(7)	6297(5)	6349(2)	21.8(9)
C13	7694(7)	5653(4)	5353(2)	15.7(8)
C14	-410(8)	7585(5)	2582(2)	17.9(5)
C15	-528(7)	6447(4)	2067(2)	17.8(4)
C16	-2184(7)	6491(4)	2567(2)	19.1(5)
C17	-1285(7)	7124(5)	1485(2)	18.5(6)
C18	-833(8)	7092(5)	422(2)	21.0(5)
C19	1315(9)	7524(5)	144(2)	23.8(4)
C20	963(7)	5191(5)	2009(2)	16.9(5)
C21	3039(7)	5365(5)	1791(2)	18.8(8)
C22	4392(8)	4176(5)	1687(2)	20.1(9)
C23	3612(8)	2779(5)	1814(2)	17.2(8)
C24	1527(8)	2574(5)	2030(2)	18.4(8)
C25	228(8)	3780(5)	2133(2)	17.6(8)

Table 2: Anisotropic Displacement Parameters ($\times 10^4$) for WT-05-553. The anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^{*2} \times U_{11} + \dots + 2hka^* \times b^* \times U_{12}]$

Atom	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
Br1	22.6(2)	11.99(18)	28.9(3)	5.0(2)	1.08(16)	-0.4(2)

Atom	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
Cl3	27.2(5)	19.5(5)	31.5(6)	1.8(4)	0.1(4)	3.2(4)
Cl1	36.4(7)	22.4(6)	25.6(5)	4.3(4)	-3.2(3)	2.4(3)
Cl2	35.7(7)	20.7(5)	31.0(6)	-1.6(4)	-7.2(4)	0.8(3)
O1	18.1(8)	17.2(14)	16.3(12)	-0.9(5)	-4.0(5)	-0.6(7)
O2	38.0(15)	32.9(17)	27.2(15)	10.1(13)	-15.7(8)	-6.6(14)
O3	18.8(17)	16.0(14)	17.6(11)	1.9(11)	-5.1(8)	-2.3(7)
O4	23.5(11)	13.6(10)	20.9(6)	3.4(5)	-3.4(3)	-2.2(3)
O5	45.3(17)	33.9(13)	21.7(12)	25.3(8)	-7.5(6)	-4.9(5)
N1	17.9(8)	13.8(13)	16.9(7)	0.8(5)	-3.2(4)	-1.3(5)
N2	18.5(15)	19.0(16)	17.6(11)	-0.5(8)	-4.7(7)	-0.3(8)
H2	17(8)	10(20)	18(9)	-3(6)	-2(4)	3(6)
C1	18.1(8)	15.1(16)	17.3(7)	0.4(6)	-3.2(4)	-1.6(5)
H1a	19(5)	15(2)	17(5)	0.3(10)	-2.3(19)	-1.9(10)
H1b	18(3)	16(5)	19(4)	0.1(15)	-3.0(14)	-2.9(17)
C2	17.2(8)	13.7(16)	17.2(7)	1.2(6)	-3.0(4)	-2.1(5)
C3	15.8(8)	9.0(15)	15.4(7)	3.7(6)	-2.3(4)	-3.9(5)
C4	17.1(8)	12.2(16)	16.2(7)	1.3(6)	-3.3(4)	-2.0(5)
C5	17.0(9)	14.3(16)	17.5(7)	2.2(6)	-3.1(4)	-3.2(5)
H5	27(8)	50(30)	23(6)	-15(7)	-10(3)	9(5)
C6	16.4(9)	13.5(12)	17.5(7)	3.4(5)	-2.6(4)	-3.4(4)
C7	17.3(9)	16.6(17)	16.5(8)	1.7(6)	-3.6(4)	-2.1(5)
H7	26(7)	50(20)	23(4)	-16(6)	-12(3)	13(5)
C8	17.3(9)	14.9(16)	16.4(8)	1.2(6)	-3.4(4)	-1.3(5)
H8	26(7)	50(20)	23(4)	-16(6)	-12(3)	13(5)
C9	18.9(10)	14.7(12)	17.2(8)	0.8(6)	-3.6(4)	-0.9(4)
H9	19(4)	15(4)	18(4)	1.6(16)	-3.1(15)	-0.8(16)
C10	30(2)	15.6(12)	20.6(13)	0.5(7)	-8.2(8)	-1.9(6)
H10a	30(4)	18(4)	20(5)	-0.4(16)	-9.0(17)	-1.6(17)
H10b	31(4)	17(4)	22(5)	1.8(16)	-7.6(16)	-2.0(18)
C11	32.4(16)	23.4(17)	20.3(14)	6.0(10)	-6.7(7)	-1.1(7)
H11a	32(5)	24(4)	21(4)	5.9(17)	-7.4(18)	-2.1(16)
H11b	32(4)	24(5)	22(5)	6.0(18)	-6.3(16)	0.5(18)
C12	29.2(15)	18.6(17)	17.6(12)	2.2(10)	-6.6(7)	-0.1(8)
C13	17.5(12)	12.9(15)	16.7(11)	-0.7(7)	-3.7(5)	-0.7(7)
C14	18.1(8)	15.7(8)	19.7(7)	3.7(4)	-4.6(3)	-3.8(3)
H14	19(6)	16(2)	22(6)	3.7(11)	-6(2)	-3.5(11)
C15	18.1(8)	15.3(7)	20.0(6)	3.9(3)	-4.8(3)	-3.7(3)
C16	18.7(8)	17.3(11)	21.1(9)	2.8(4)	-4.0(4)	-4.8(4)
H16a	19(5)	19(5)	22(5)	2.6(17)	-4.1(18)	-4(2)
H16b	20(2)	22(6)	26(7)	3.9(12)	-4.4(11)	-3(2)
C17	19.1(12)	15.8(10)	20.5(6)	3.4(5)	-5.5(3)	-3.3(3)
C18	26.4(7)	15.2(11)	21.4(7)	1.6(4)	-4.5(3)	-1.2(4)
H18a	27(5)	16(3)	21(4)	1.2(15)	-4.2(16)	-1.1(15)
H18b	26(4)	16(3)	23(6)	1.6(15)	-4.7(17)	-1.0(15)
C19	26.9(7)	18.8(6)	25.5(6)	1.4(3)	-2.9(3)	1.4(3)
C20	17.7(8)	14.8(7)	18.3(12)	3.4(3)	-4.9(4)	-3.4(4)
C21	18.6(8)	12.3(6)	25(2)	4.2(3)	-2.4(6)	-2.1(4)
H21	30(5)	11(6)	100(30)	6.7(15)	28(6)	5(3)
C22	19.2(9)	12.0(6)	29(2)	4.1(3)	-0.4(6)	-1.3(4)
H22	30(5)	11(6)	100(30)	6.7(15)	28(6)	5(3)
C23	19.1(8)	11.8(5)	21(2)	3.7(3)	-3.2(6)	-2.0(4)
C24	19.2(8)	14.4(7)	22(2)	3.2(3)	-2.7(6)	-2.6(4)
H24	29(8)	15(2)	90(40)	4.1(14)	22(9)	1.6(19)
C25	18.7(8)	14.8(7)	19(2)	3.3(3)	-4.3(6)	-3.3(4)
H25	38(9)	17(6)	140(50)	7.1(16)	44(11)	7(3)

Table 3: Bond Lengths in Å for WT-05-553.

Atom	Atom	Length/Å	Atom	Atom	Length/Å
Br1	C23	1.887(4)	C9	H9	1.17(5)
Cl3	C19	1.793(5)	C9	C10	1.531(6)
Cl1	C19	1.762(5)	C9	C13	1.534(6)
Cl2	C19	1.773(5)	C10	H10a	1.08(3)
O1	C4	1.234(5)	C10	H10b	1.08(3)
O2	C12	1.213(5)	C10	C11	1.523(6)
O3	C13	1.208(5)	C11	H11a	1.12(4)
O4	C17	1.340(6)	C11	H11b	1.12(4)
O4	C18	1.426(6)	C11	C12	1.508(7)
O5	C17	1.213(5)	C14	H14	1.070(3)
N1	C1	1.467(5)	C14	C15	1.550(6)
N1	C4	1.353(6)	C14	C16	1.490(6)
N1	C9	1.447(6)	C15	C16	1.516(7)
N2	H2	0.97(5)	C15	C17	1.500(6)
N2	C12	1.381(6)	C15	C20	1.486(6)
N2	C13	1.382(5)	C16	H16a	1.0702(18)
C1	H1a	1.04(3)	C16	H16b	1.0702(18)
C1	H1b	1.04(3)	C18	H18a	1.098(9)
C1	C2	1.489(6)	C18	H18b	1.097(9)
C2	C3	1.402(6)	C18	C19	1.522(7)
C2	C5	1.383(6)	C20	C21	1.386(6)
C3	C4	1.469(6)	C20	C25	1.405(6)
C3	C8	1.405(6)	C21	H21	1.0780
C5	H5	1.06(5)	C21	C22	1.399(6)
C5	C6	1.405(6)	C22	H22	1.0780
C6	C7	1.407(6)	C22	C23	1.403(6)
C6	C14	1.494(6)	C23	C24	1.394(7)
C7	H7	1.0780	C24	H24	1.0780
C7	C8	1.393(6)	C24	C25	1.391(6)
C8	H8	1.0780	C25	H25	1.0780

Table 4: Bond Angles in ° for WT-05-553.

Atom	Atom	Atom	Angle/°	Atom	Atom	Atom	Angle/°
C18	O4	C17	118.1(3)	C5	C2	C3	120.9(4)
C4	N1	C1	113.3(4)	C4	C3	C2	109.2(4)
C9	N1	C1	122.3(4)	C8	C3	C2	121.8(4)
C9	N1	C4	124.1(4)	C8	C3	C4	129.0(4)
C12	N2	H2	117(3)	N1	C4	O1	124.4(4)
C13	N2	H2	115(3)	C3	C4	O1	129.3(4)
C13	N2	C12	127.5(4)	C3	C4	N1	106.3(4)
H1a	C1	N1	111.2(2)	H5	C5	C2	120.8(3)
H1b	C1	N1	111.2(2)	C6	C5	C2	118.4(4)
H1b	C1	H1a	109.1	C6	C5	H5	120.8(3)
C2	C1	N1	102.7(4)	C7	C6	C5	120.1(4)
C2	C1	H1a	111.2(2)	C14	C6	C5	121.1(4)
C2	C1	H1b	111.2(2)	C14	C6	C7	118.4(4)
C3	C2	C1	108.4(4)	H7	C7	C6	119.0(3)
C5	C2	C1	130.7(4)	C8	C7	C6	122.0(4)

Atom	Atom	Atom	Angle/°	Atom	Atom	Atom	Angle/°
C8	C7	H7	119.0(3)	C20	C15	C17	116.0(4)
C7	C8	C3	116.7(4)	C15	C16	C14	62.1(3)
H8	C8	C3	121.6(3)	H16a	C16	C14	117.6(2)
H8	C8	C7	121.6(3)	H16a	C16	C15	117.6(2)
H9	C9	N1	107.5(2)	H16b	C16	C14	117.6(2)
C10	C9	N1	114.4(4)	H16b	C16	C15	117.6(2)
C10	C9	H9	107.5(3)	H16b	C16	H16a	114.7
C13	C9	N1	108.9(4)	O5	C17	O4	123.6(4)
C13	C9	H9	107.5(2)	C15	C17	O4	112.0(4)
C13	C9	C10	110.7(4)	C15	C17	O5	124.4(4)
H10a	C10	C9	109.8(3)	H18a	C18	O4	105(2)
H10b	C10	C9	109.8(3)	H18b	C18	O4	113(2)
H10b	C10	H10a	108.3	H18b	C18	H18a	107.6(11)
C11	C10	C9	109.2(4)	C19	C18	O4	108.3(4)
C11	C10	H10a	109.8(3)	C19	C18	H18a	111(2)
C11	C10	H10b	109.8(3)	C19	C18	H18b	111(2)
H11a	C11	C10	109.2(3)	Cl1	C19	Cl3	108.7(3)
H11b	C11	C10	109.2(3)	Cl2	C19	Cl3	107.7(3)
H11b	C11	H11a	107.892608686(15)	Cl2	C19	Cl1	110.7(3)
C12	C11	C10	112.1(4)	C18	C19	Cl3	111.4(3)
C12	C11	H11a	109.2(3)	C18	C19	Cl1	107.3(3)
C12	C11	H11b	109.2(3)	C18	C19	Cl2	110.9(4)
N2	C12	O2	119.5(4)	C21	C20	C15	121.3(4)
C11	C12	O2	124.0(4)	C25	C20	C15	120.0(4)
C11	C12	N2	116.6(4)	C25	C20	C21	118.6(4)
N2	C13	O3	121.1(4)	H21	C21	C20	119.1(2)
C9	C13	O3	122.7(4)	C22	C21	C20	121.7(4)
C9	C13	N2	116.2(4)	C22	C21	H21	119.1(3)
H14	C14	C6	115(2)	H22	C22	C21	120.8(3)
C15	C14	C6	123.3(4)	C23	C22	C21	118.4(4)
C15	C14	H14	111(2)	C23	C22	H22	120.8(3)
C16	C14	C6	123.4(4)	C22	C23	Br1	120.2(4)
C16	C14	H14	114(2)	C24	C23	Br1	118.8(3)
C16	C14	C15	59.8(3)	C24	C23	C22	121.0(4)
C16	C15	C14	58.2(3)	H24	C24	C23	120.4(3)
C17	C15	C14	111.3(3)	C25	C24	C23	119.1(4)
C17	C15	C16	114.0(4)	C25	C24	H24	120.4(3)
C20	C15	C14	124.4(4)	C24	C25	C20	121.1(4)
C20	C15	C16	120.6(4)	H25	C25	C20	119.4(3)
				H25	C25	C24	119.4(3)

Table 5: Torsion Angles in ° for WT-05-553.

Atom	Atom	Atom	Atom	Angle/°
Br1	C23	C22	C21	-179.9(3)
Br1	C23	C24	C25	179.5(3)
Cl3	C19	C18	O4	58.1(3)
Cl1	C19	C18	O4	177.0(3)
Cl2	C19	C18	O4	-61.9(3)
O1	C4	N1	C1	179.6(4)
O1	C4	N1	C9	-7.4(5)
O1	C4	C3	C2	177.0(5)
O1	C4	C3	C8	-4.6(6)

Atom	Atom	Atom	Atom	Angle/°
O2	C12	N2	C13	-176.8(4)
O2	C12	C11	C10	150.3(5)
O3	C13	N2	C12	176.1(4)
O3	C13	C9	N1	-22.4(5)
O3	C13	C9	C10	-148.9(5)
O4	C17	C15	C14	148.0(4)
O4	C17	C15	C16	-148.5(4)
O4	C17	C15	C20	-1.7(4)
O5	C17	C15	C14	-32.1(5)
O5	C17	C15	C16	31.4(5)
O5	C17	C15	C20	178.2(5)
N1	C1	C2	C3	-4.8(4)
N1	C1	C2	C5	179.0(3)
N1	C4	C3	C2	-1.5(4)
N1	C4	C3	C8	176.9(3)
N1	C9	C10	C11	-179.9(4)
N1	C9	C13	N2	157.8(3)
N2	C12	C11	C10	-29.4(5)
N2	C13	C9	C10	31.2(4)
C1	C2	C3	C4	4.0(4)
C1	C2	C3	C8	-174.5(3)
C1	C2	C5	C6	174.7(5)
C2	C3	C8	C7	-0.9(5)
C2	C5	C6	C7	-1.1(5)
C2	C5	C6	C14	-174.3(4)
C3	C8	C7	C6	-1.4(5)
C5	C6	C7	C8	2.4(5)
C5	C6	C14	C15	-95.8(4)
C5	C6	C14	C16	-22.6(5)
C6	C14	C15	C16	112.3(6)
C6	C14	C15	C17	-141.9(5)
C6	C14	C15	C20	4.8(6)
C6	C14	C16	C15	-112.2(6)
C9	C10	C11	C12	55.9(4)
C14	C15	C20	C21	-77.5(5)
C14	C15	C20	C25	107.2(5)
C14	C16	C15	C17	-101.0(3)
C14	C16	C15	C20	113.9(3)
C15	C20	C21	C22	-174.5(4)
C15	C20	C25	C24	174.1(4)
C20	C21	C22	C23	-0.6(5)
C20	C25	C24	C23	1.6(5)
C21	C22	C23	C24	0.9(5)
C22	C23	C24	C25	-1.4(5)

Table 6: Hydrogen Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for WT-05-553. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} .

Atom	x	y	z	U_{eq}
H2	10440(80)	4920(60)	5720(20)	16(11)
H1a	2539(9)	5520(40)	4738(2)	17(3)
H1b	1490(40)	7135(16)	4947(8)	17(3)
H5	-380(60)	6330(30)	3709(5)	32(13)
H7	3374(7)	8960(4)	2514(2)	33(10)
H8	6286(7)	9189(4)	3287(2)	33(10)
H9	4480(50)	6010(30)	5731(10)	17(3)
H10a	7660(40)	8590(20)	5656(8)	23(3)
H10b	5110(50)	8610(20)	5982(4)	23(3)
H11a	8030(20)	8240(30)	6755(10)	26(3)
H11b	6150(40)	6738(19)	6720(9)	26(3)
H14	-890(70)	8640(20)	2420(20)	19(3)
H16a	-2135(7)	5640(5)	2896(2)	20(3)
H16b	-3773(8)	6866(4)	2452(2)	22(3)
H18a	-1610(50)	6220(30)	163(17)	22(3)
H18b	-1960(50)	8010(30)	430(20)	22(3)
H21	3626(7)	6444(5)	1700(2)	48(13)
H22	5997(8)	4329(5)	1513(2)	48(13)
H24	929(8)	1495(5)	2117(2)	45(17)
H25	-1371(8)	3629(5)	2312(2)	70(20)

Citations

CrysAlisPro (ROD), Rigaku Oxford Diffraction, Poland (?).

CrysAlisPro Software System, Rigaku Oxford Diffraction, (2024).

L.J. Bourhis and O.V. Dolomanov and R.J. Gildea and J.A.K. Howard and H. Puschmann, The Anatomy of a Comprehensive Constrained, Restrained, Refinement Program for the Modern Computing Environment - Olex2 Disected, *Acta Cryst. A*, (2015), **A71**, 59-71.

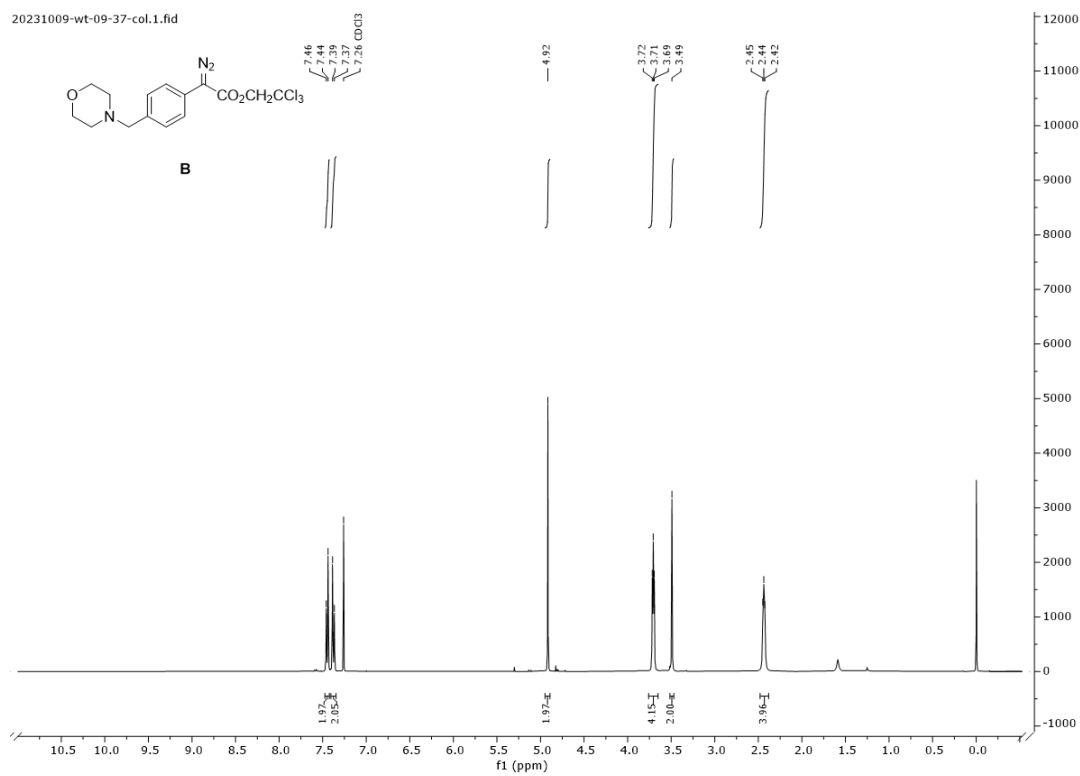
O.V. Dolomanov and L.J. Bourhis and R.J. Gildea and J.A.K. Howard and H. Puschmann, Olex2: A complete structure solution, refinement and analysis program, *J. Appl. Cryst.*, (2009), **42**, 339-341.

Sheldrick, G.M., ShelXT-Integrated space-group and crystal-structure determination, *Acta Cryst.*, (2015), **A71**, 3-8.

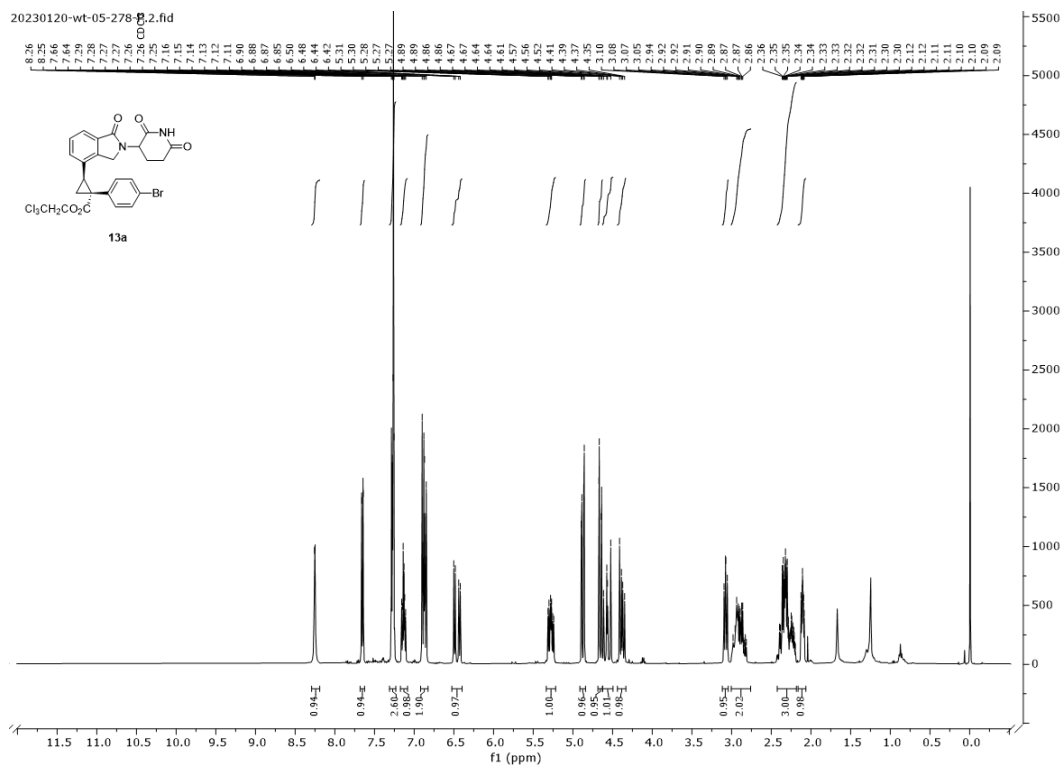
Section 4: Spectroscopic Data

¹H NMR Spectra

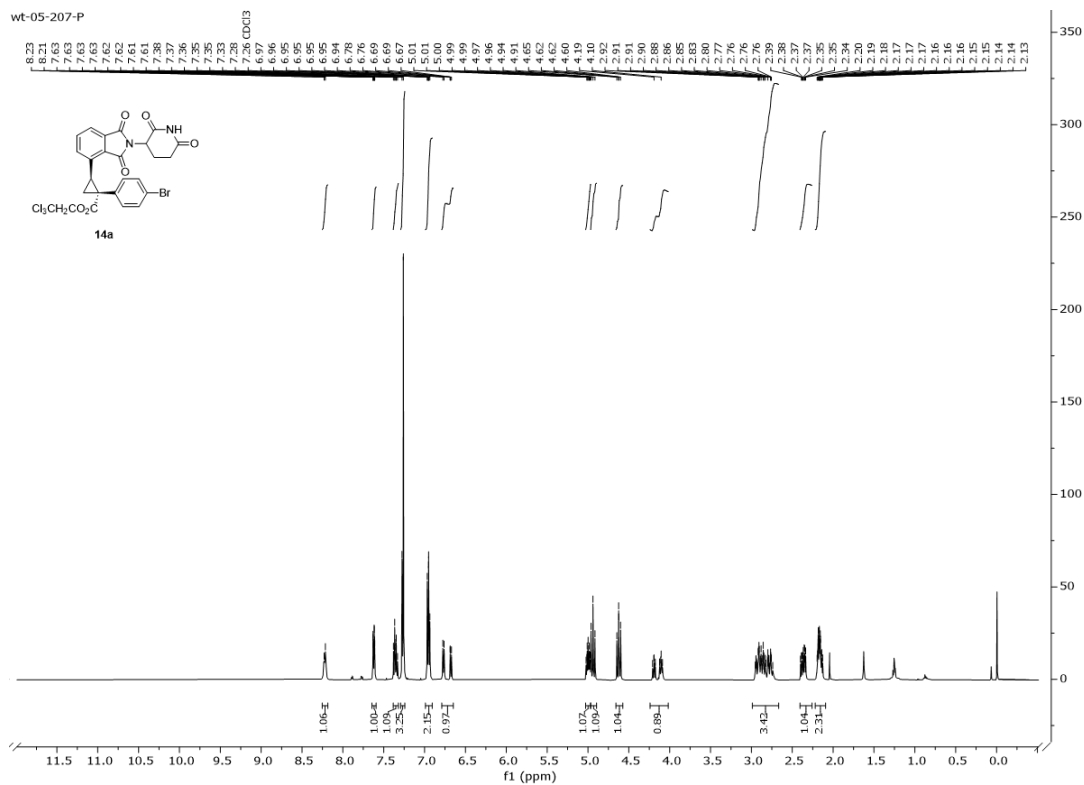
¹H NMR Spectrum for Compound B



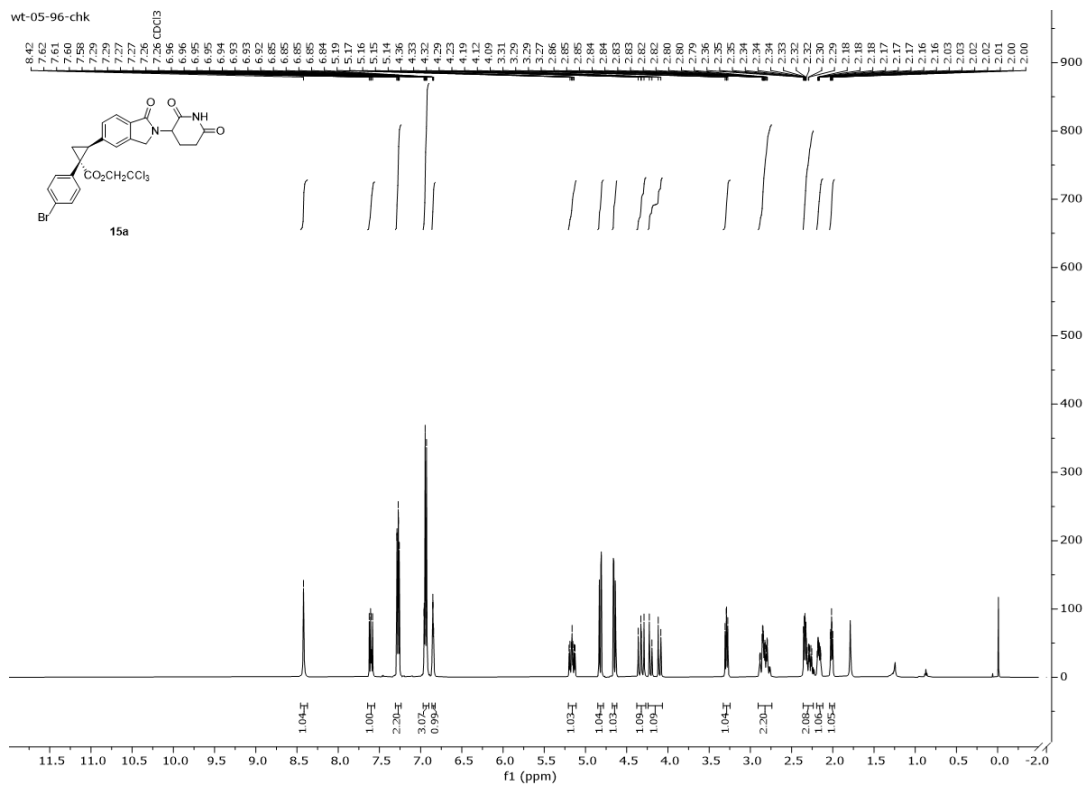
¹H NMR Spectrum for Compounds 13a and 13b



¹H NMR Spectrum for Compounds 14a and 14b

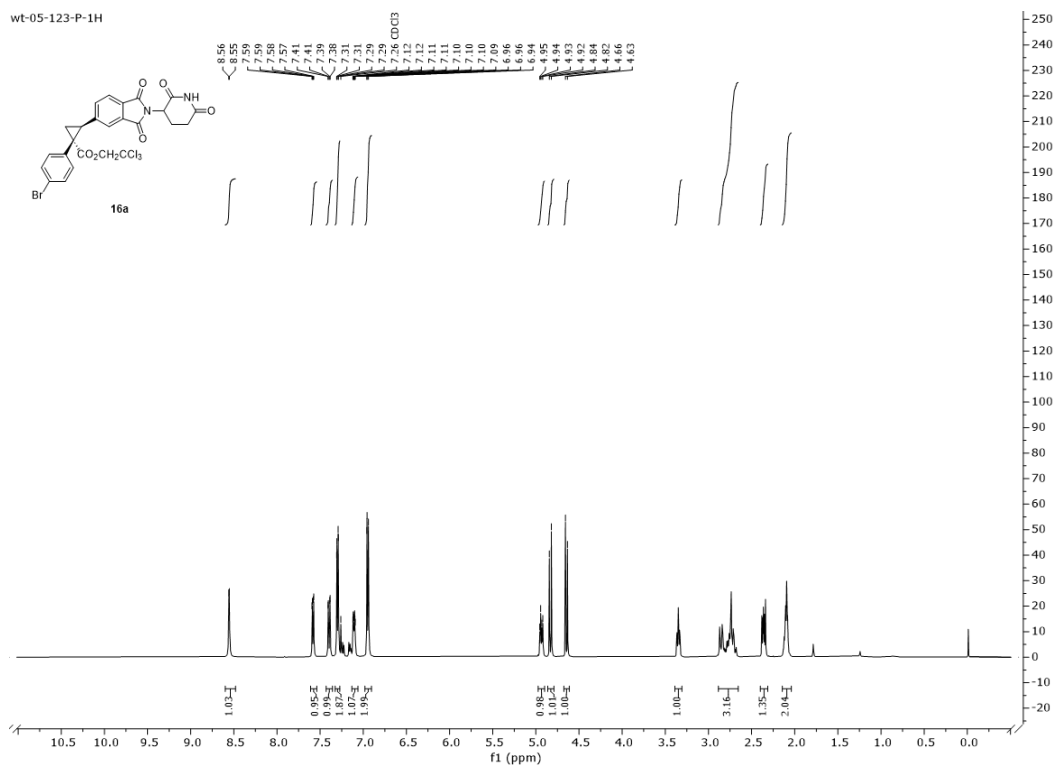


¹H NMR Spectrum for Compounds 15a and 15b

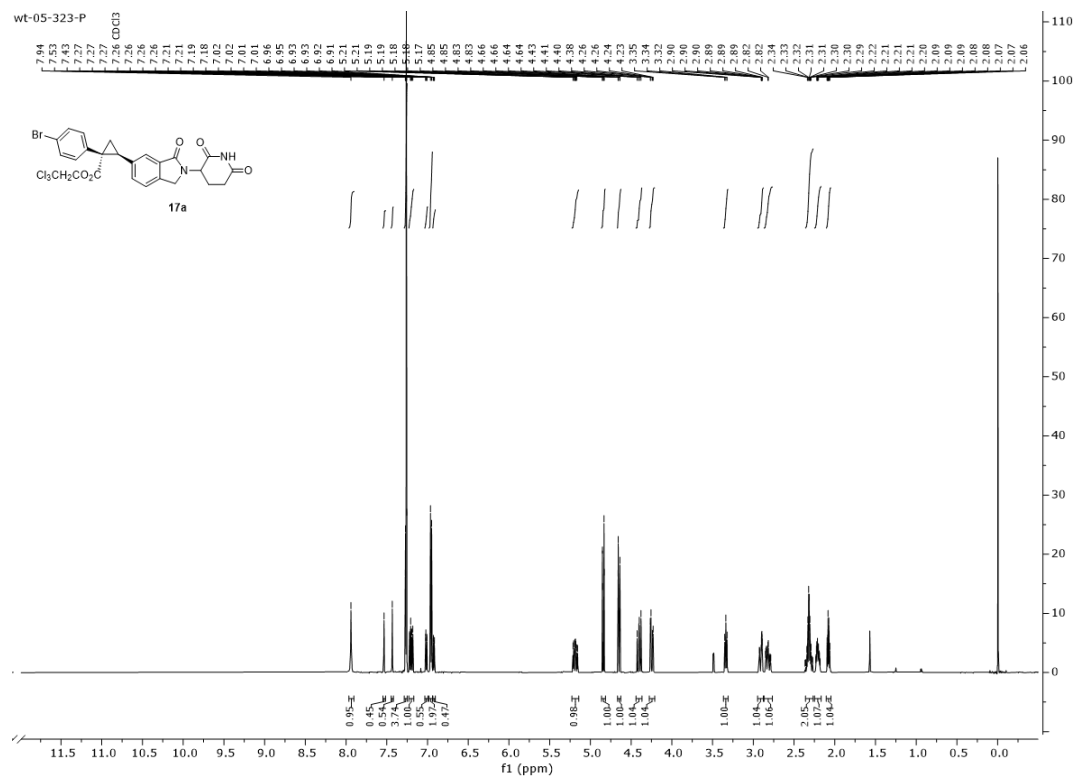


¹H NMR Spectrum for Compounds S-15a and R-15b

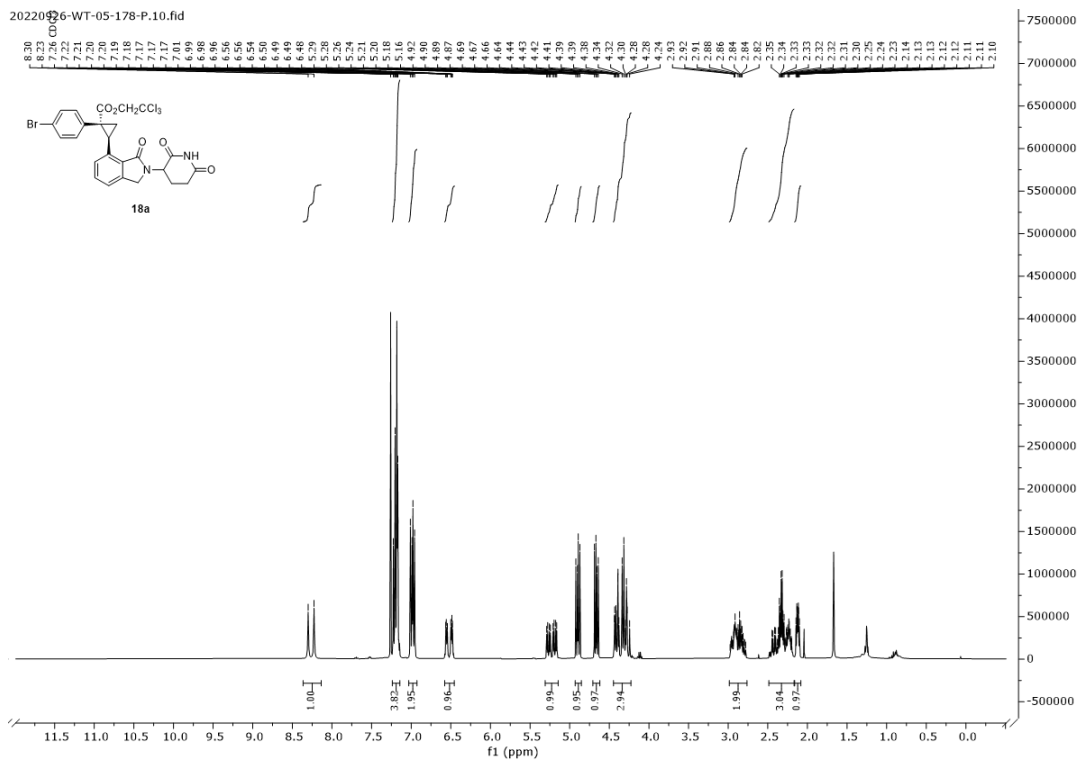
¹H NMR Spectrum for Compounds 16a and 16b



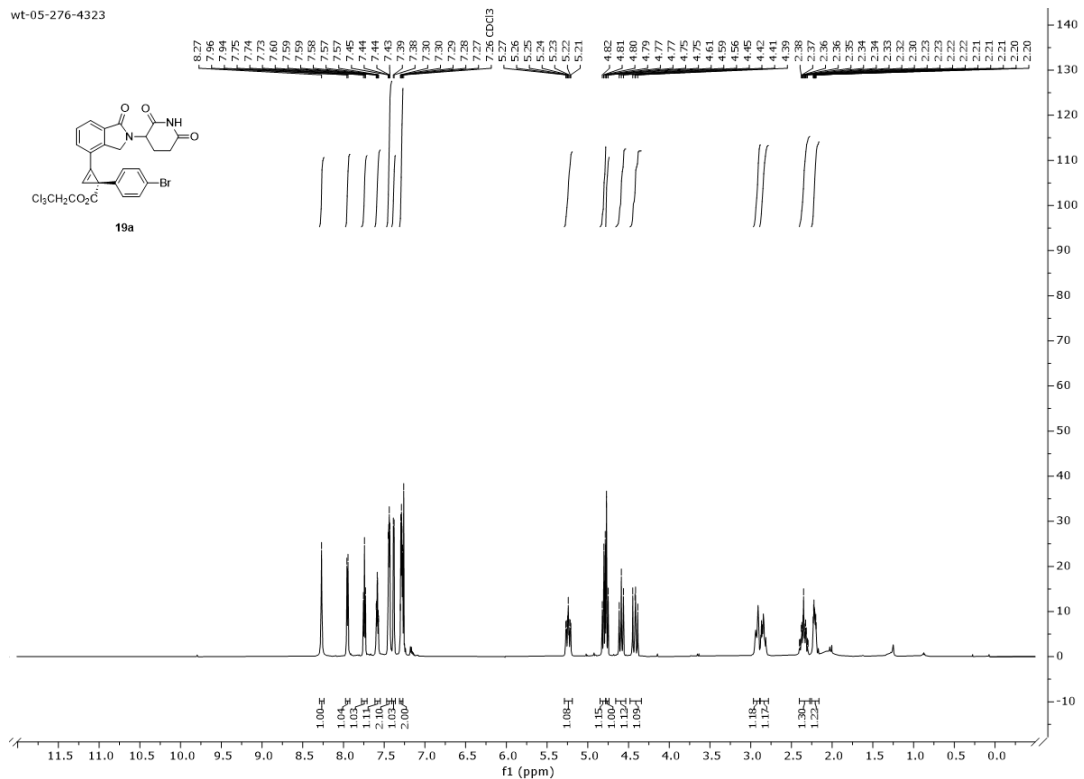
¹H NMR Spectrum for Compounds 17a and 17b



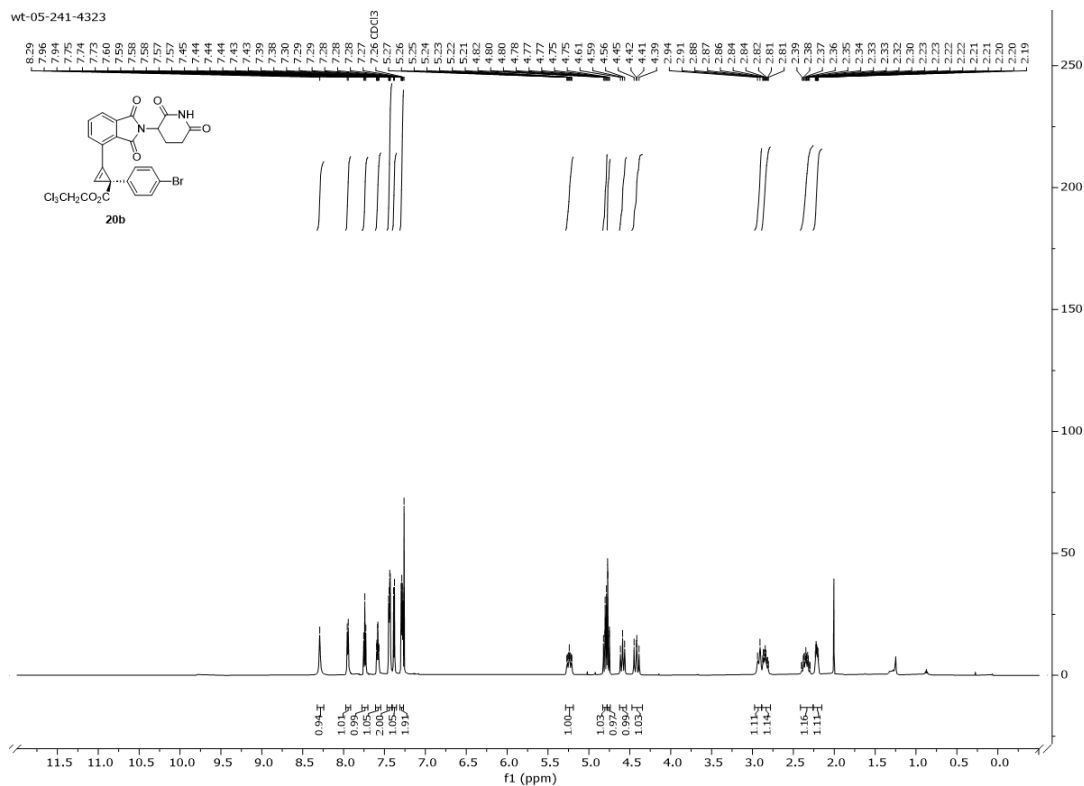
¹H NMR Spectrum for Compounds 18a and 18b



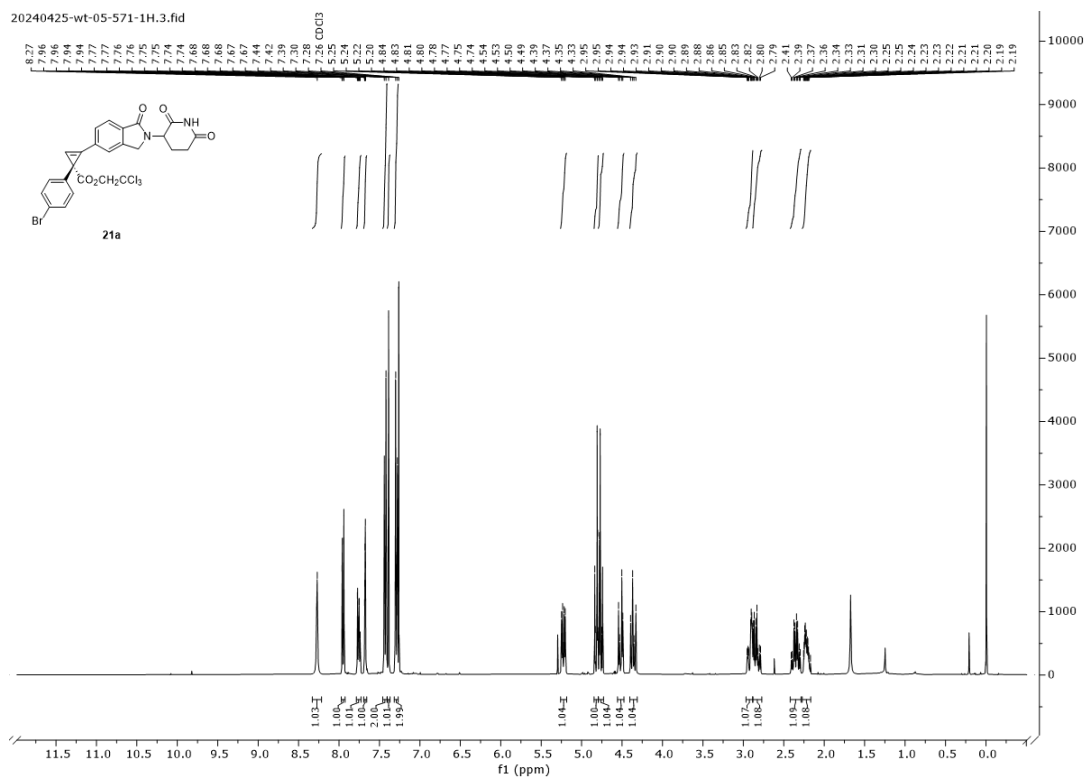
¹H NMR Spectrum for Compounds 19a and 19b



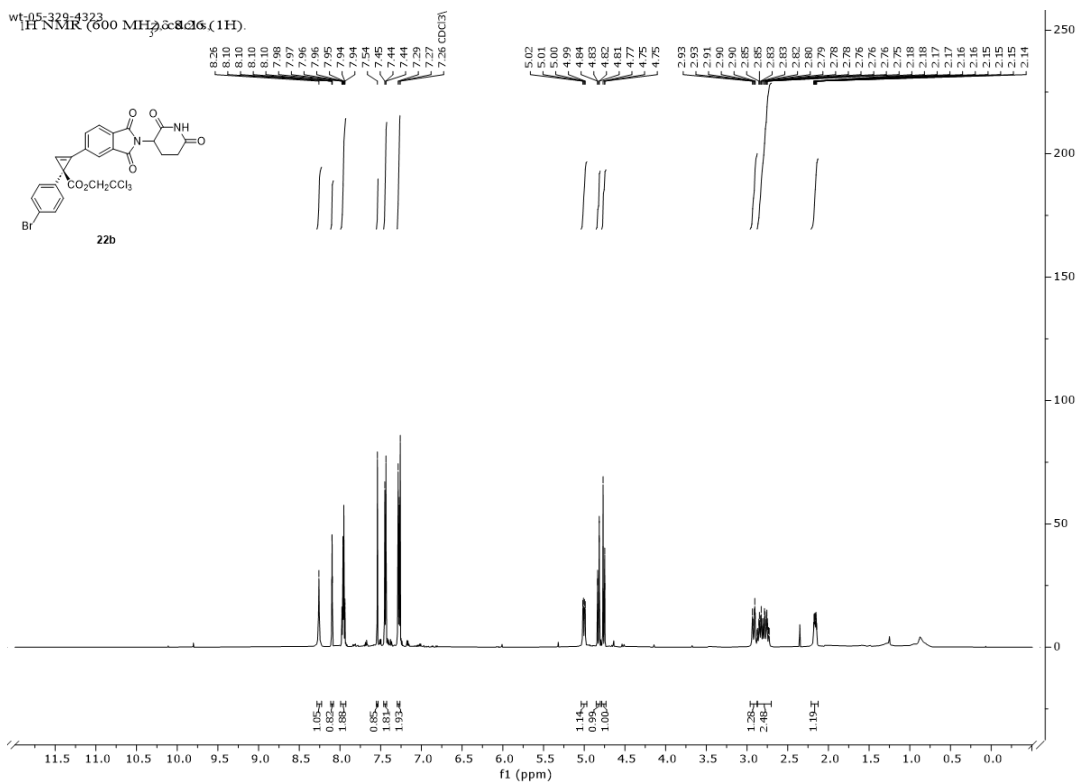
¹H NMR Spectrum for Compounds 20a and 20b



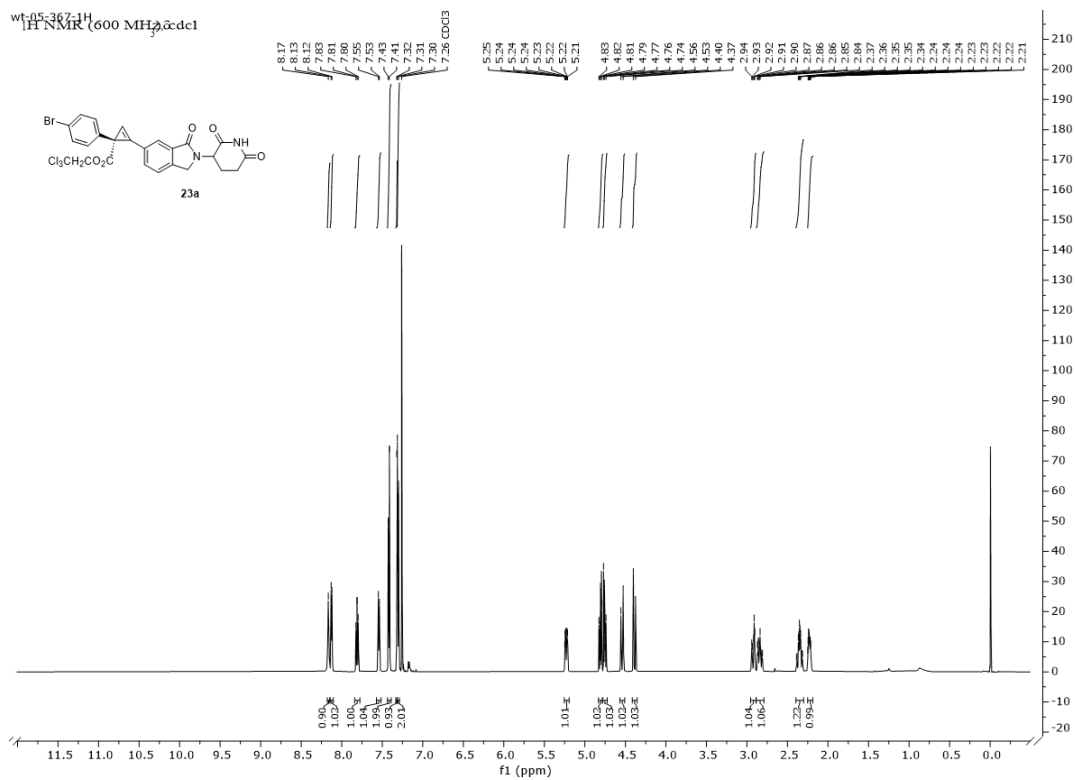
¹H NMR Spectrum for Compounds 21a and 21b



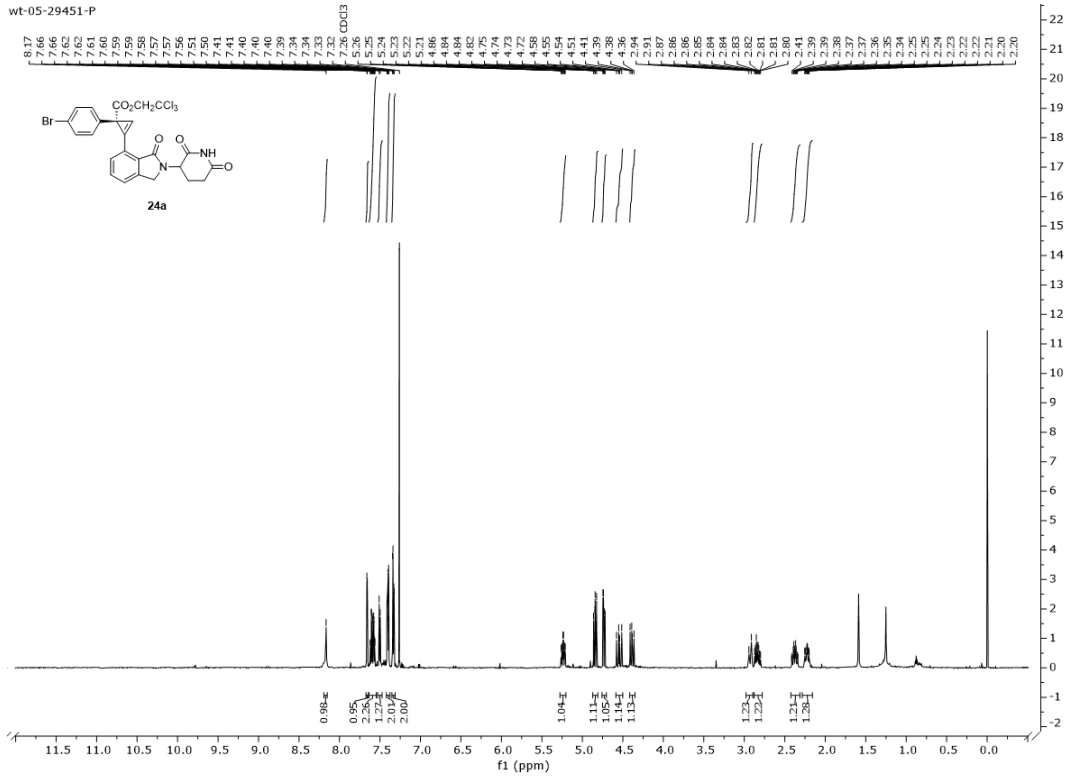
¹H NMR Spectrum for Compounds 22a and 22b



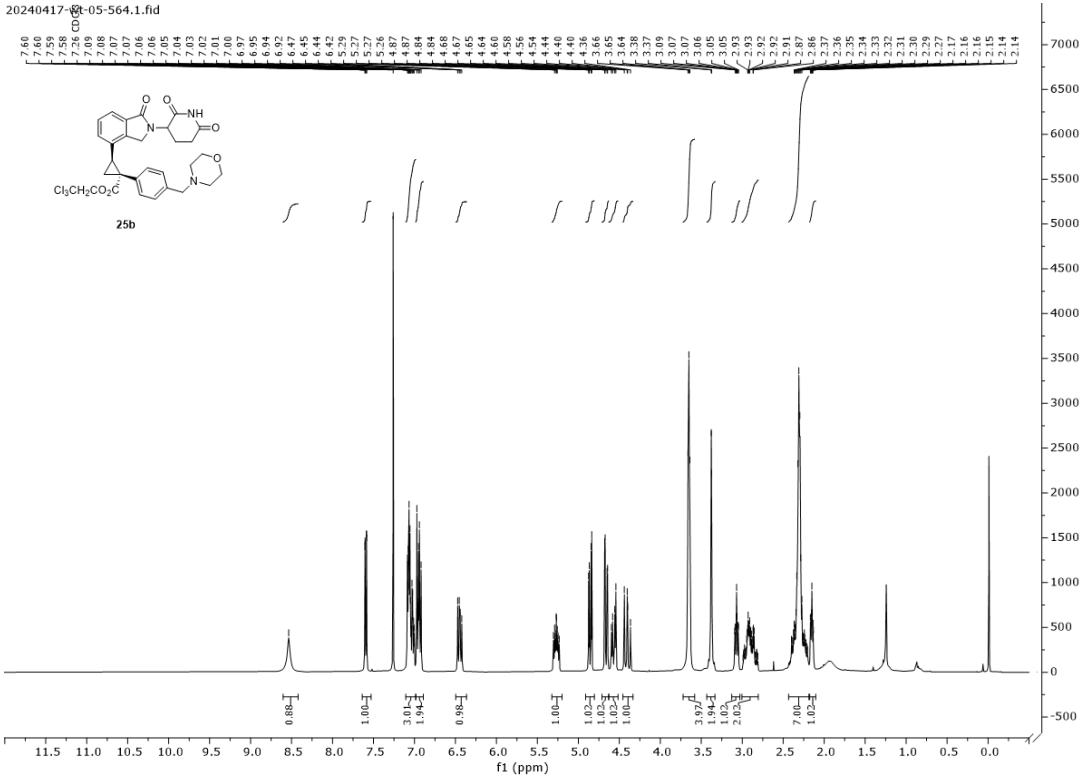
¹H NMR Spectrum for Compounds 23a and 23b



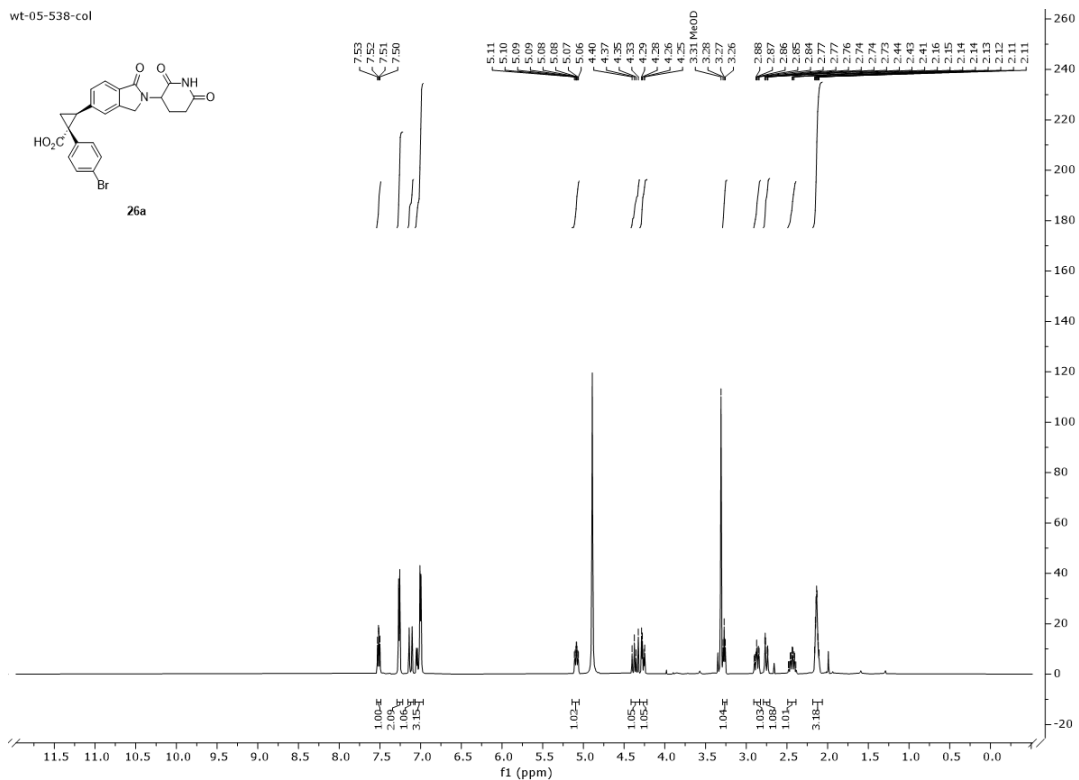
¹H NMR Spectrum for Compounds 24a and 24b



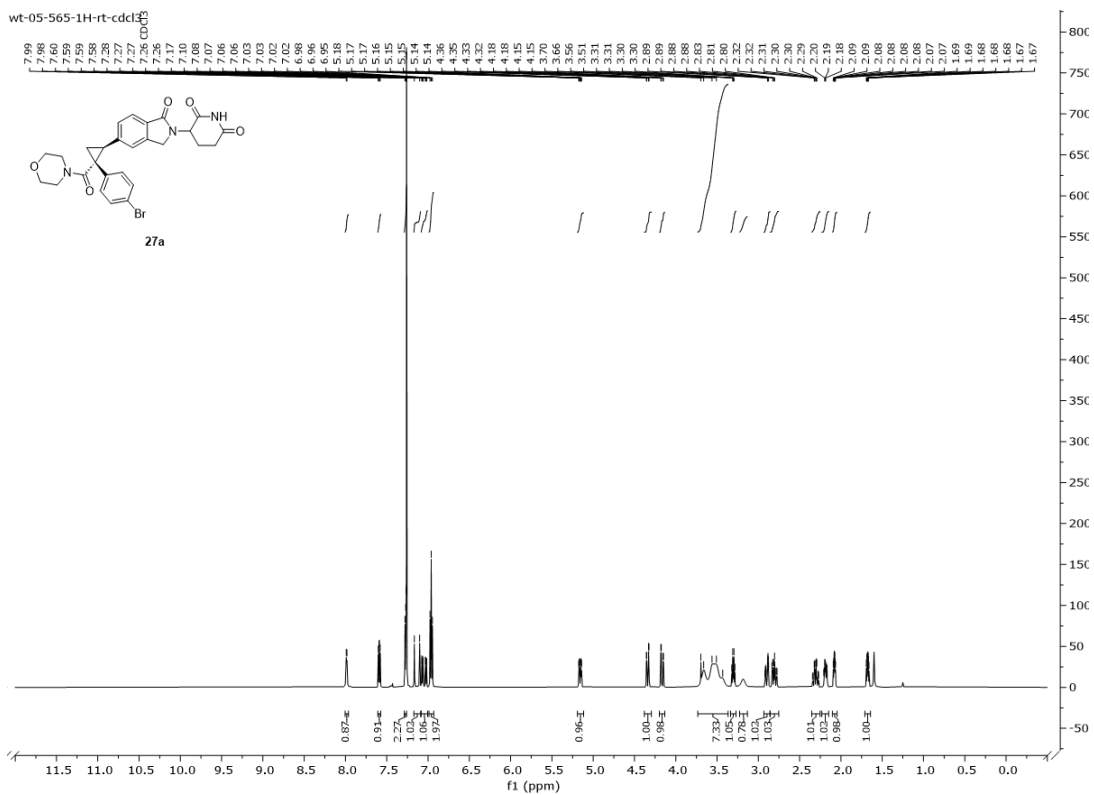
¹H NMR Spectrum for Compounds 25a and 25b



¹H NMR Spectrum for Compounds 26a and 26b



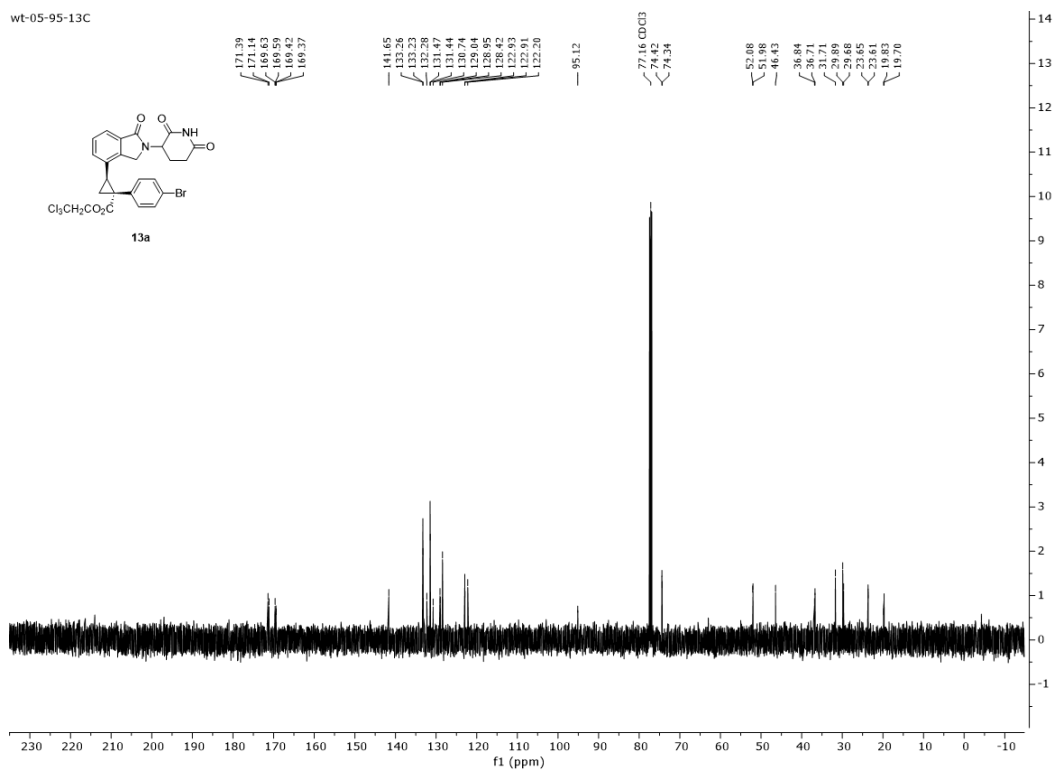
¹H NMR Spectrum for Compounds 27a and 27b



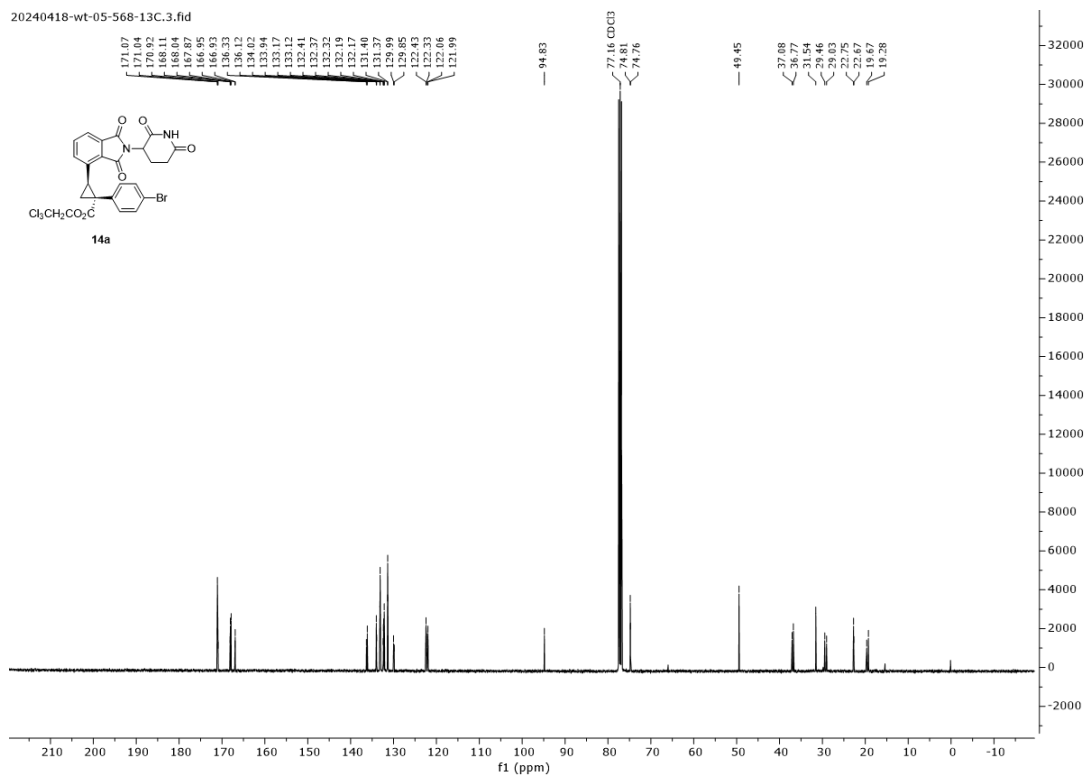
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectra

$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 13a and 13b.

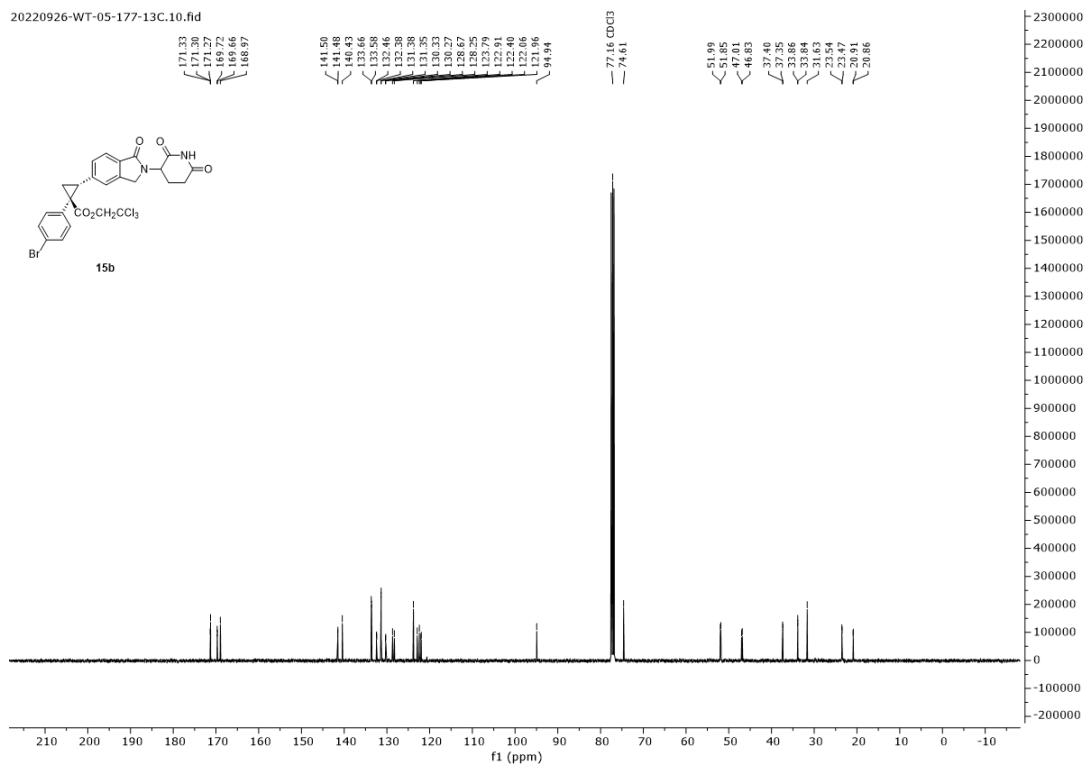
wt-05-95-13C



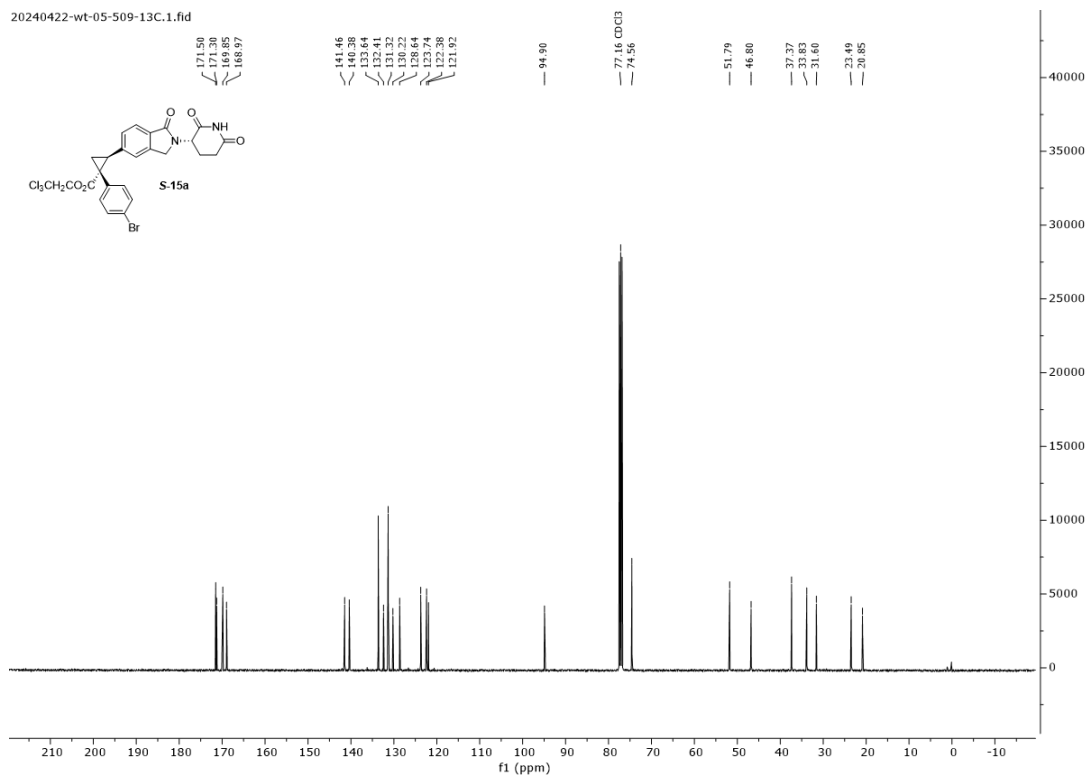
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 14a and 14b.



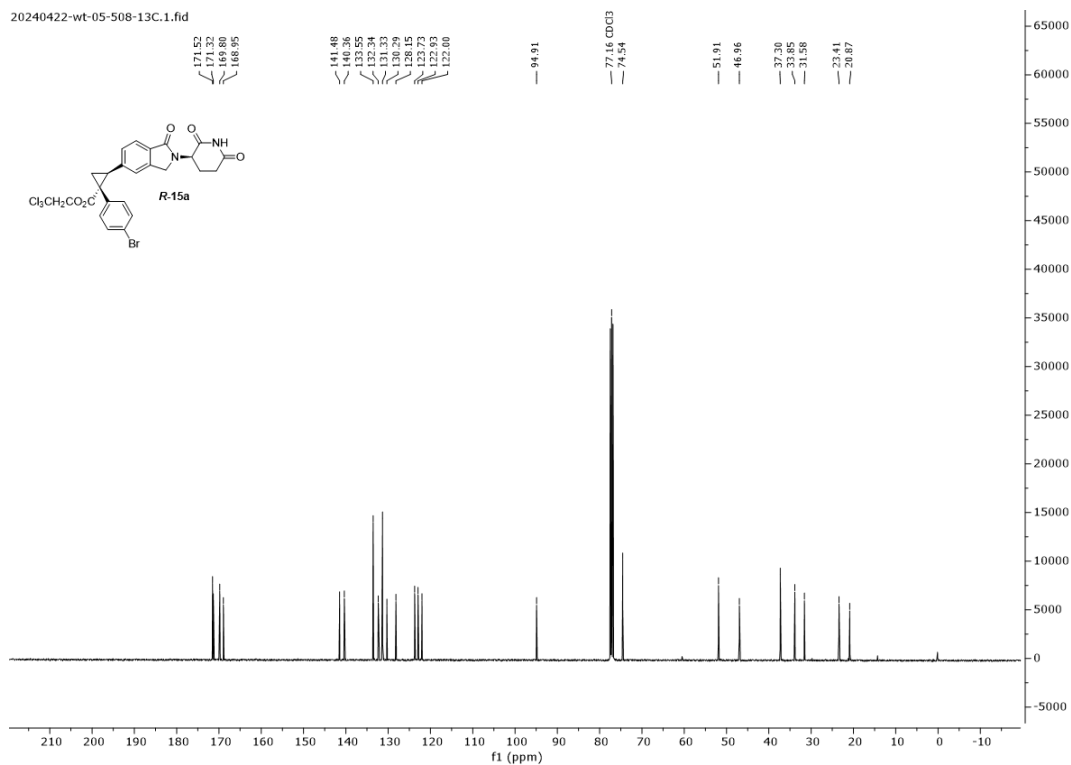
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 15a and 15b.



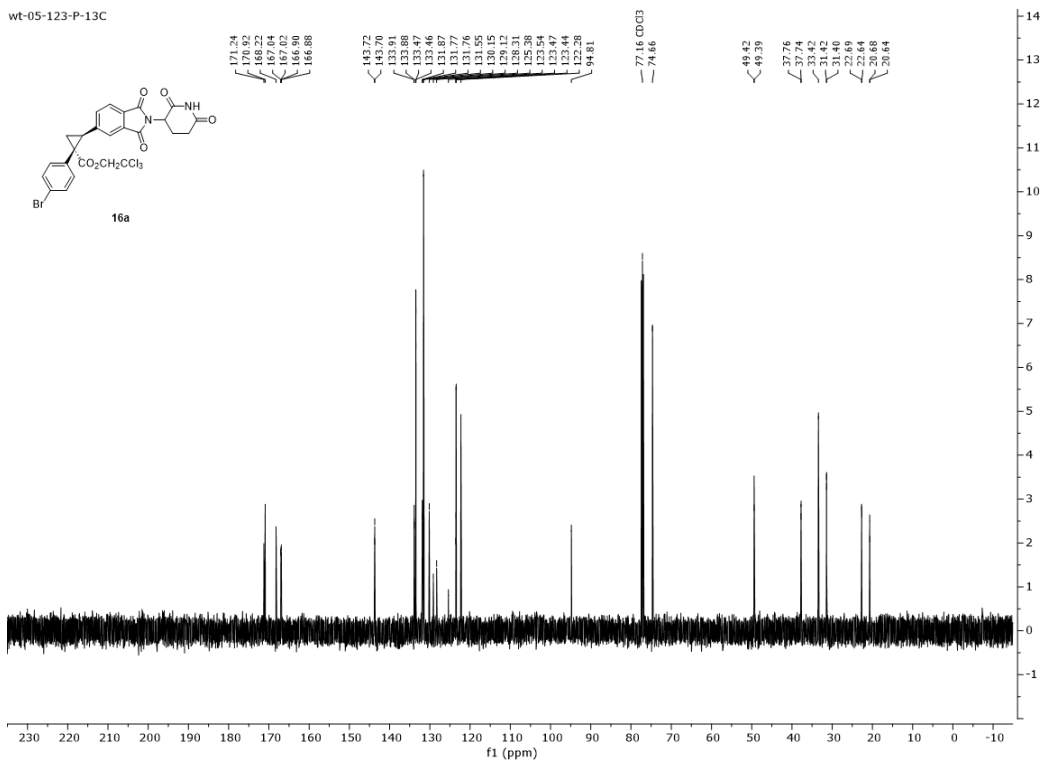
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds *S*-15a and *R*-15b.



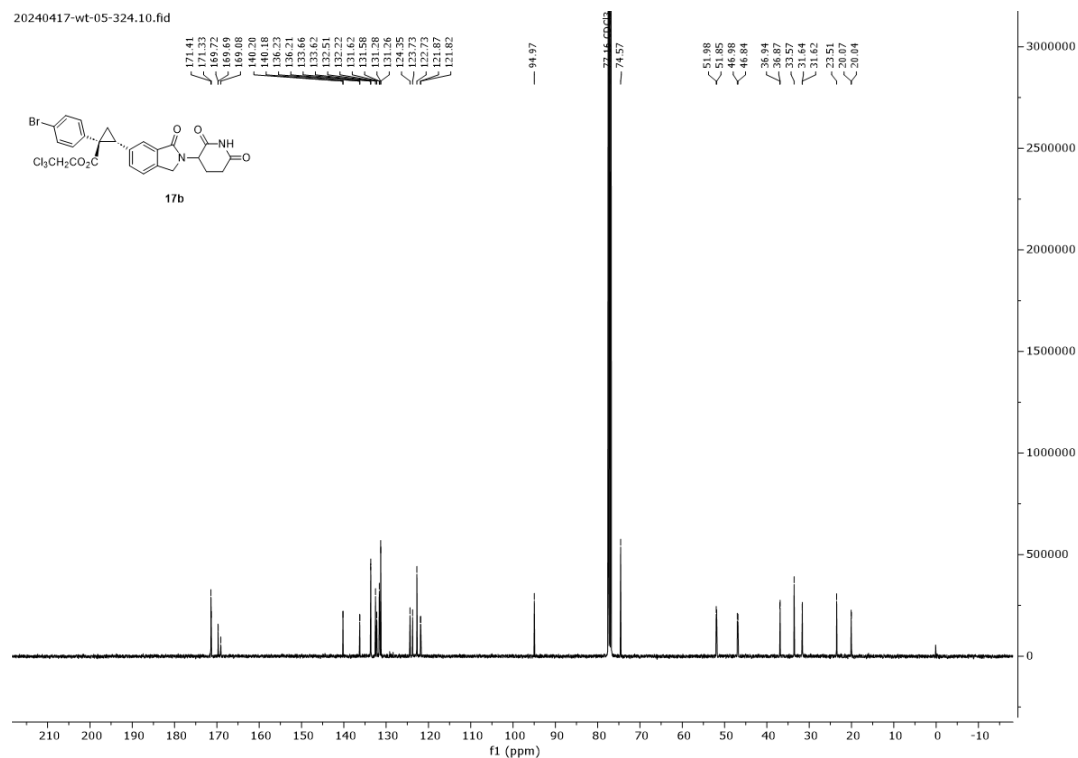
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds *R*-15a and *S*-15b.



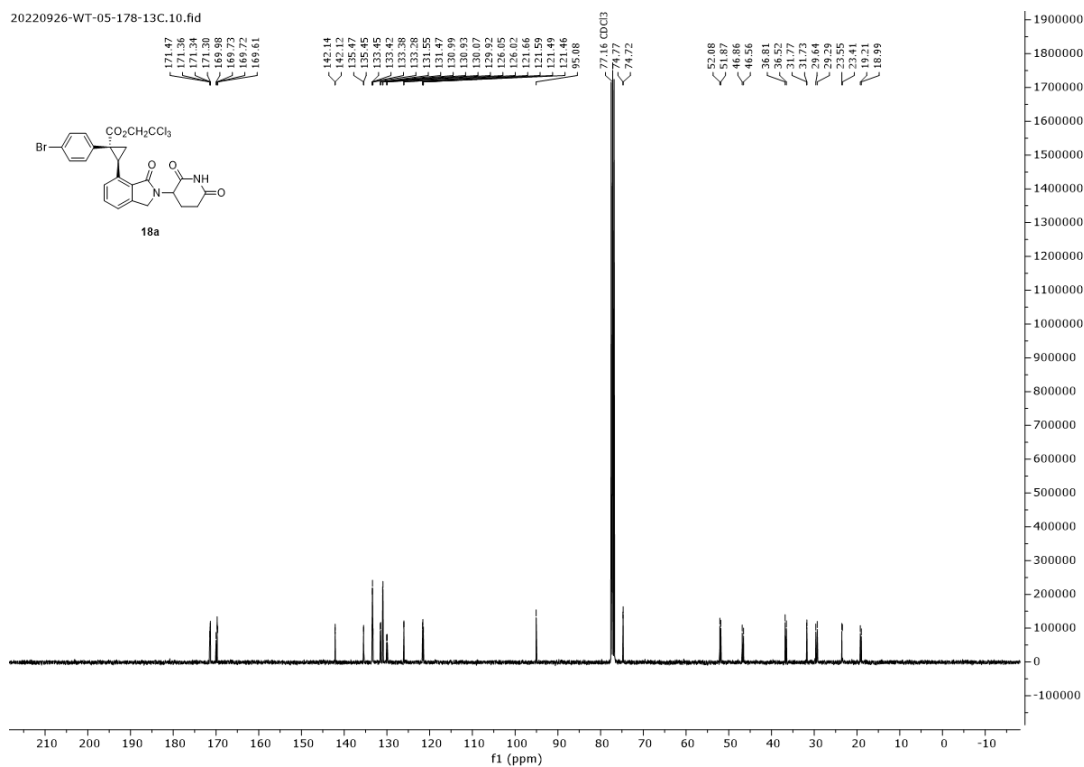
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 16a and 16b.



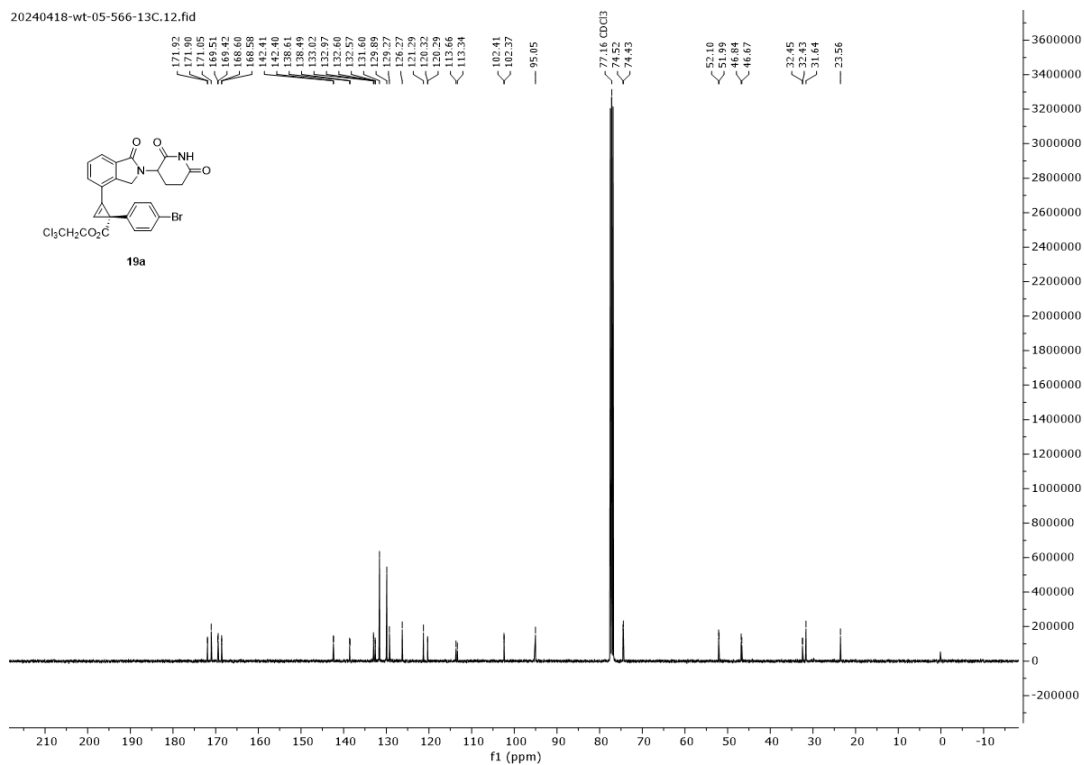
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 17a and 17b.



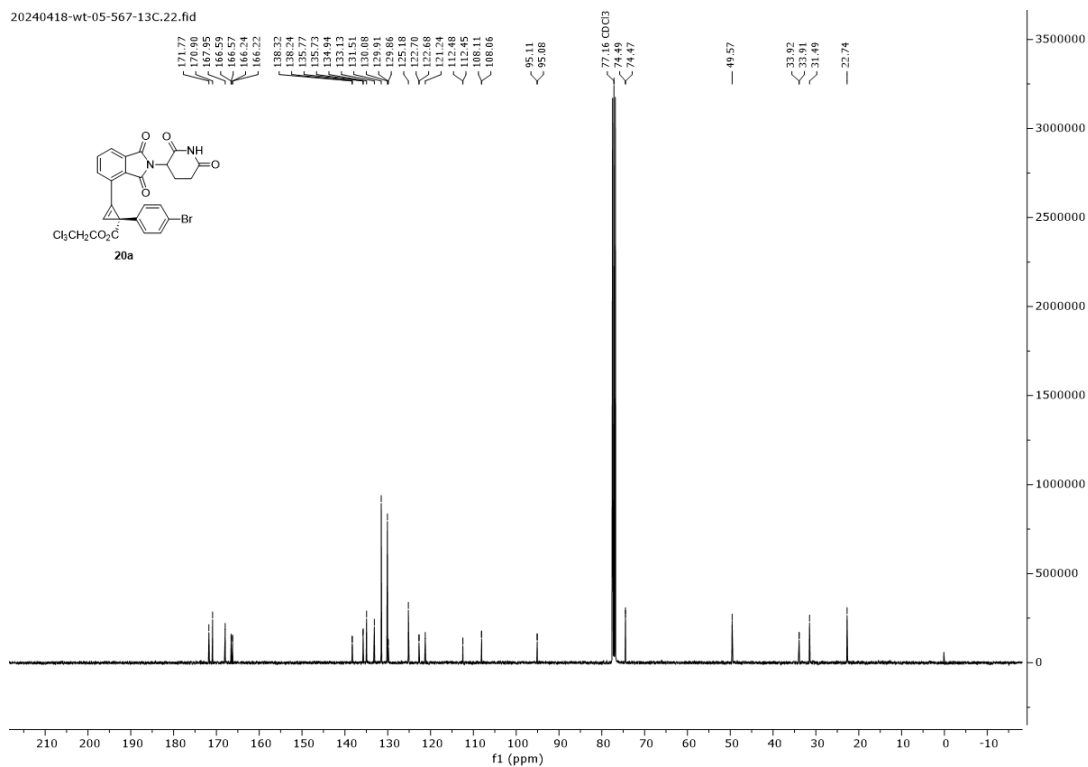
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 18a and 18b.



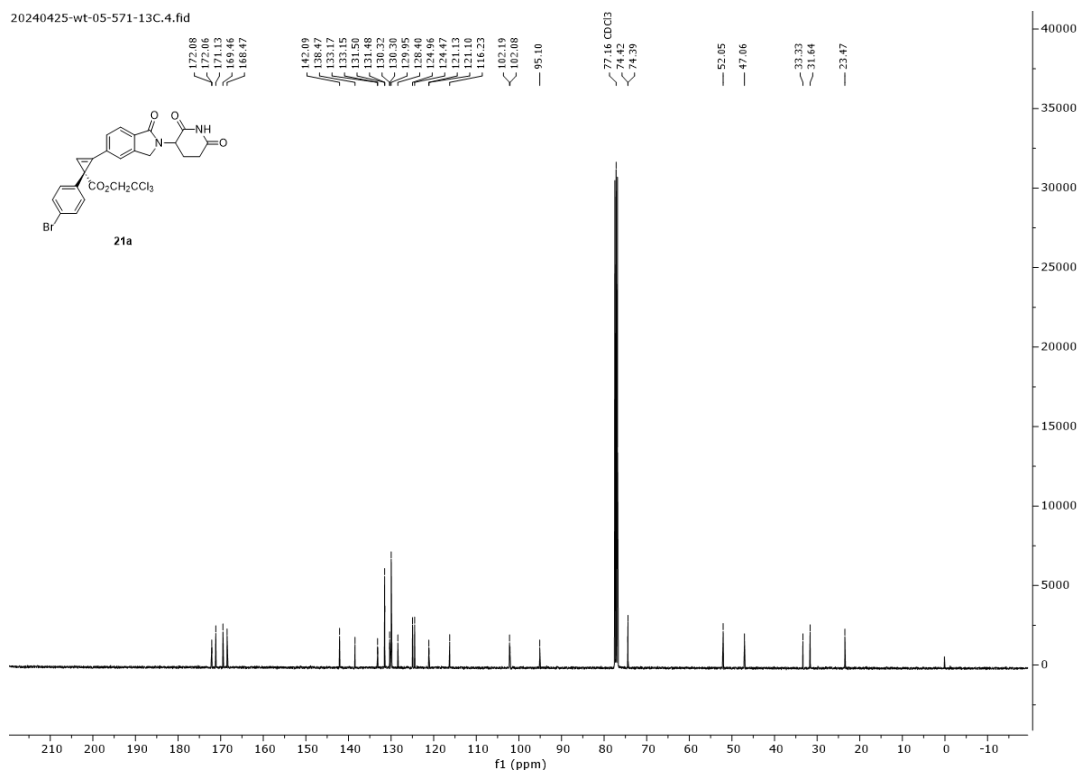
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 19a and 19b.



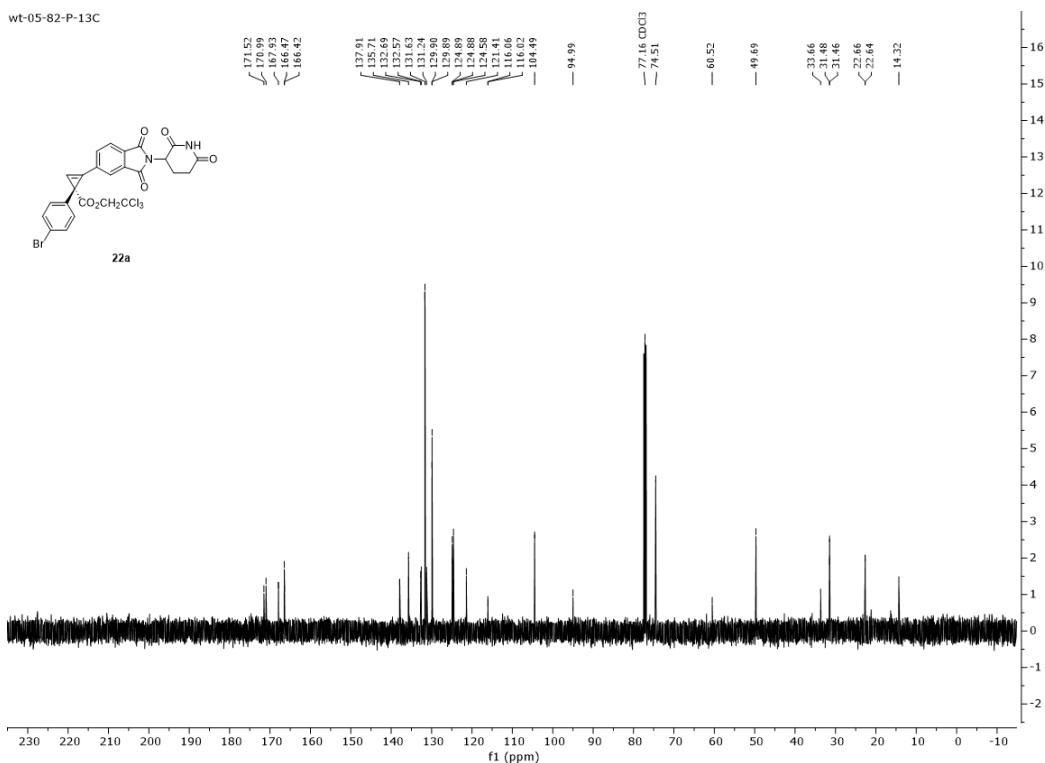
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 20a and 20b.



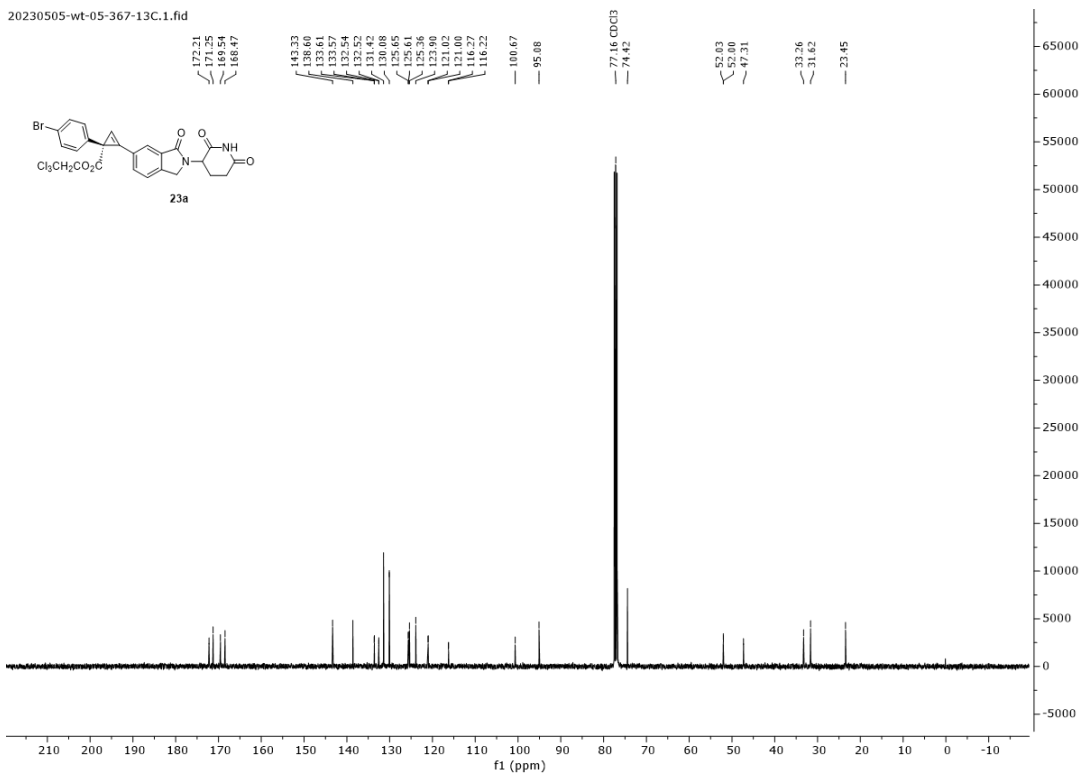
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 21a and 21b.



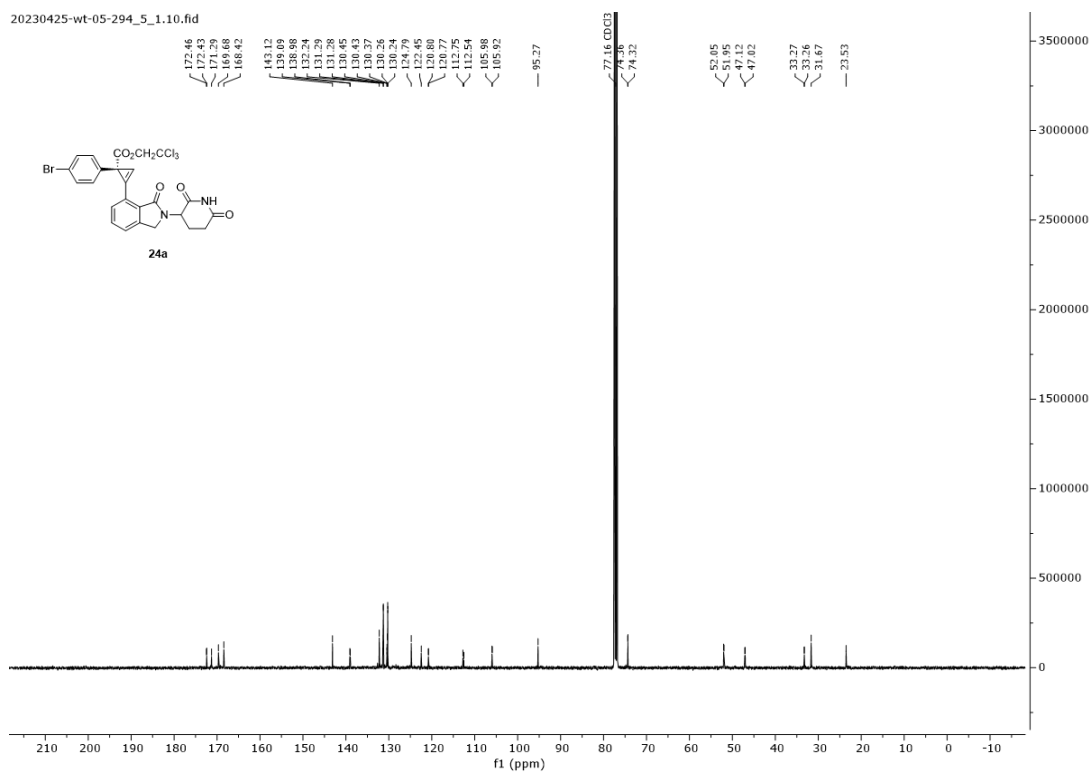
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 22a and 22b.



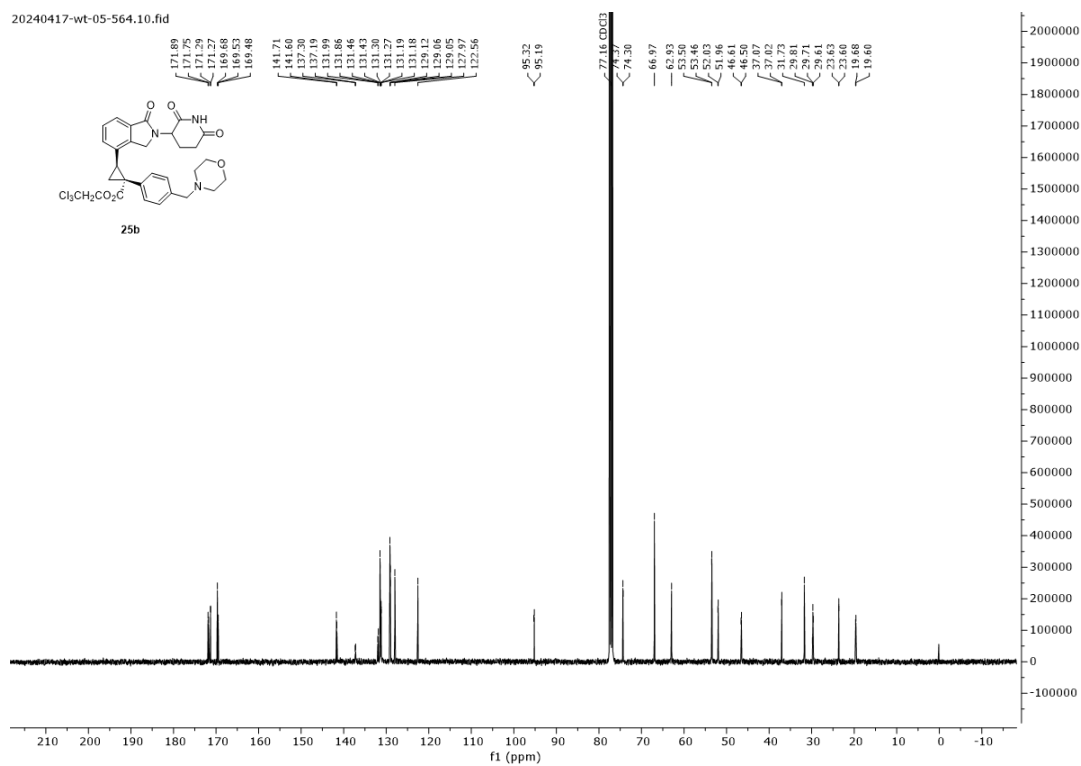
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 23a and 23b.



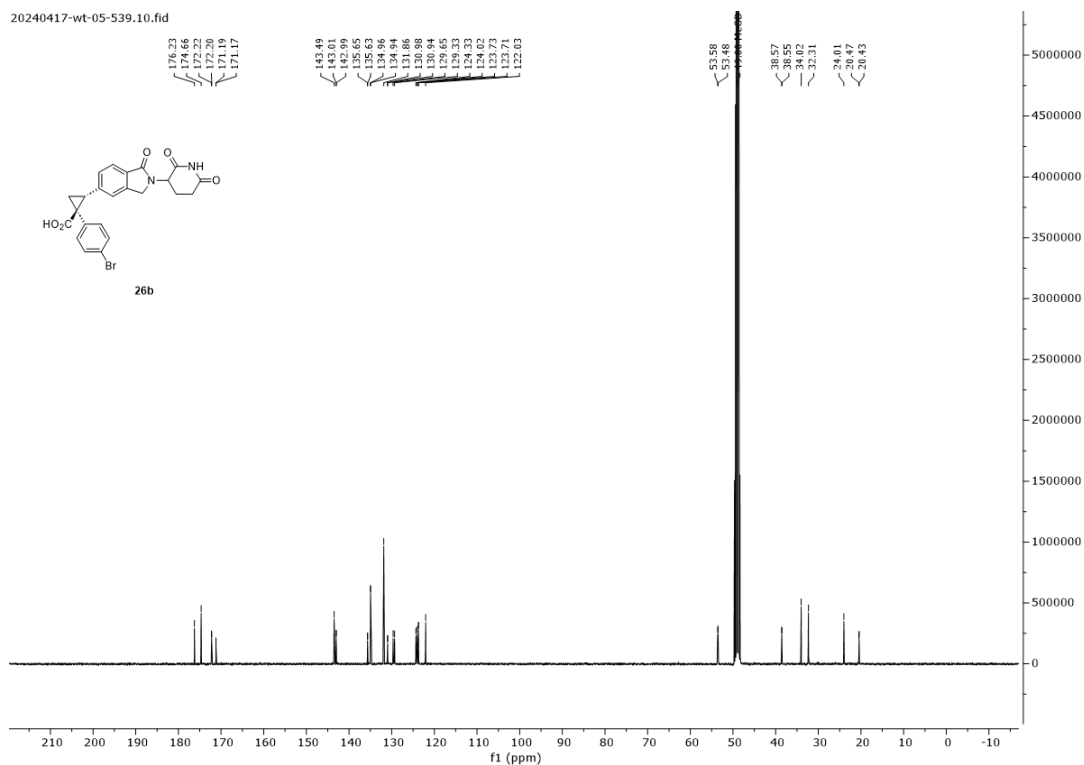
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 24a and 24b.



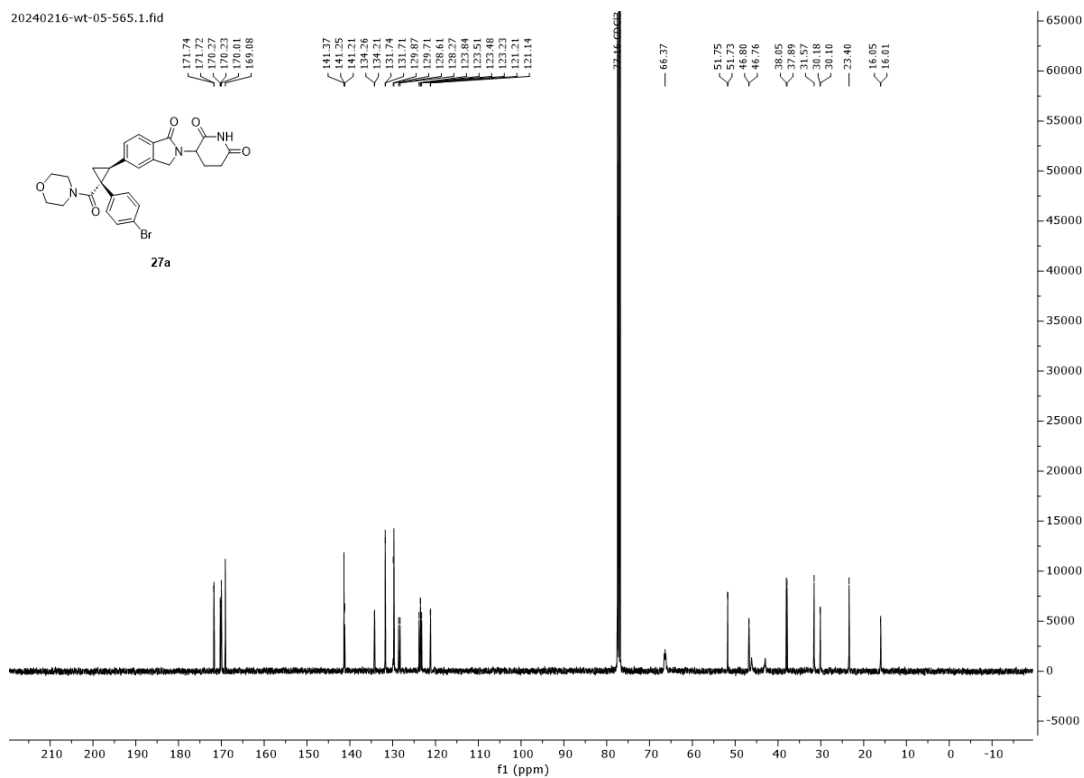
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 25a and 25b.



$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 26a and 26b.



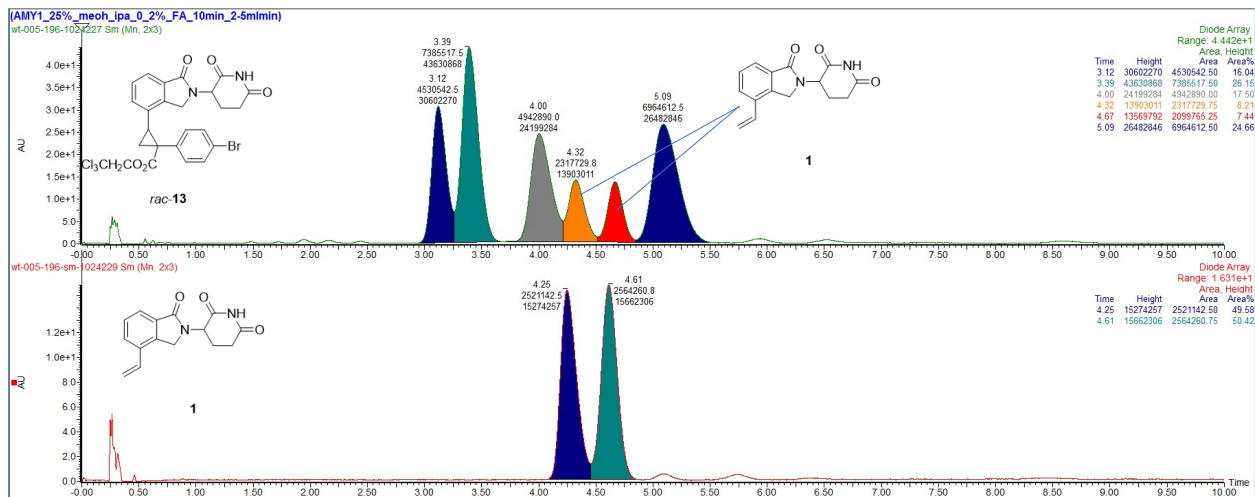
$^{13}\text{C}\{^1\text{H}\}$ NMR Spectrum for Compounds 27a and 27b.



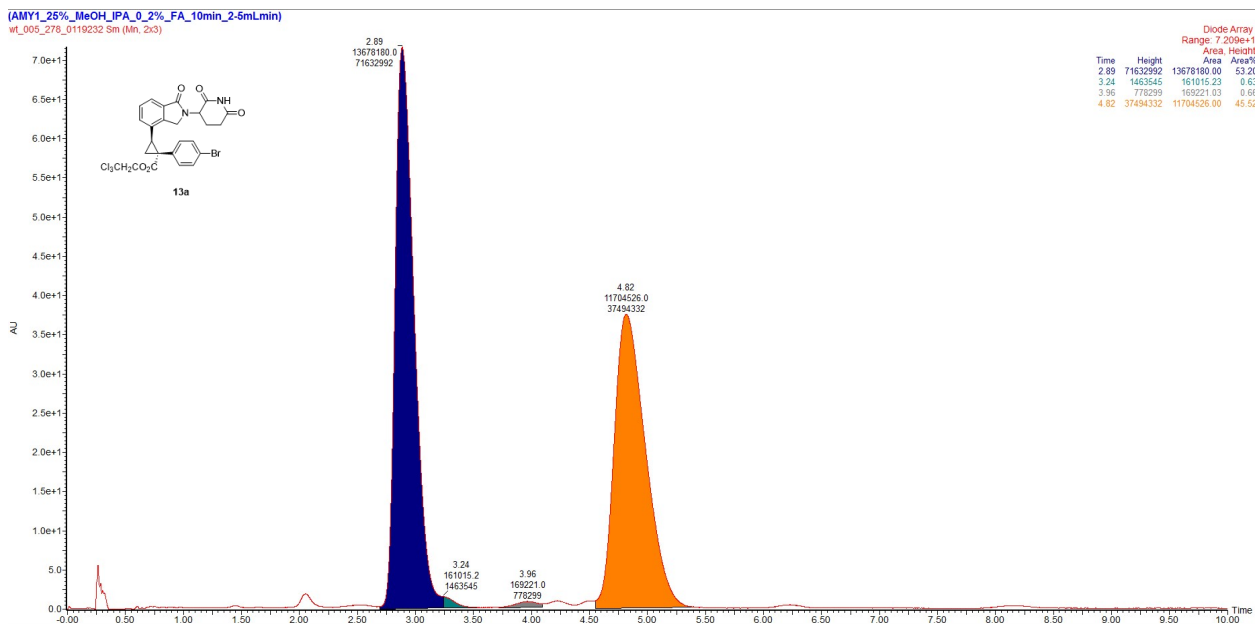
Section 5: Chromatographic Data

Chiral SFC Chromatograms

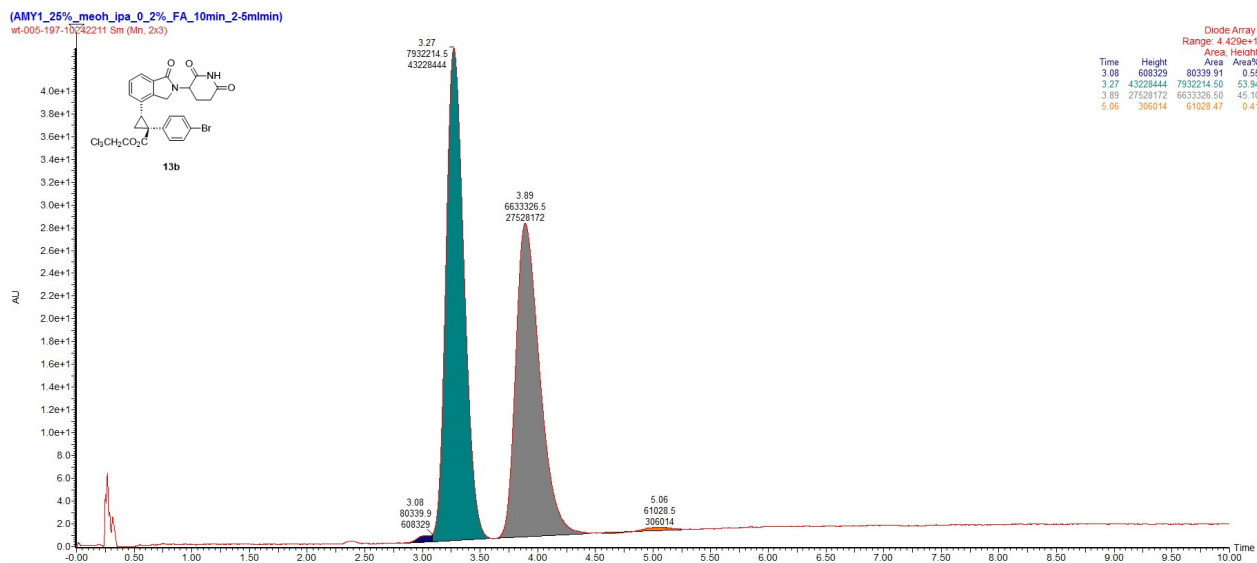
Racemic Chromatogram, 13



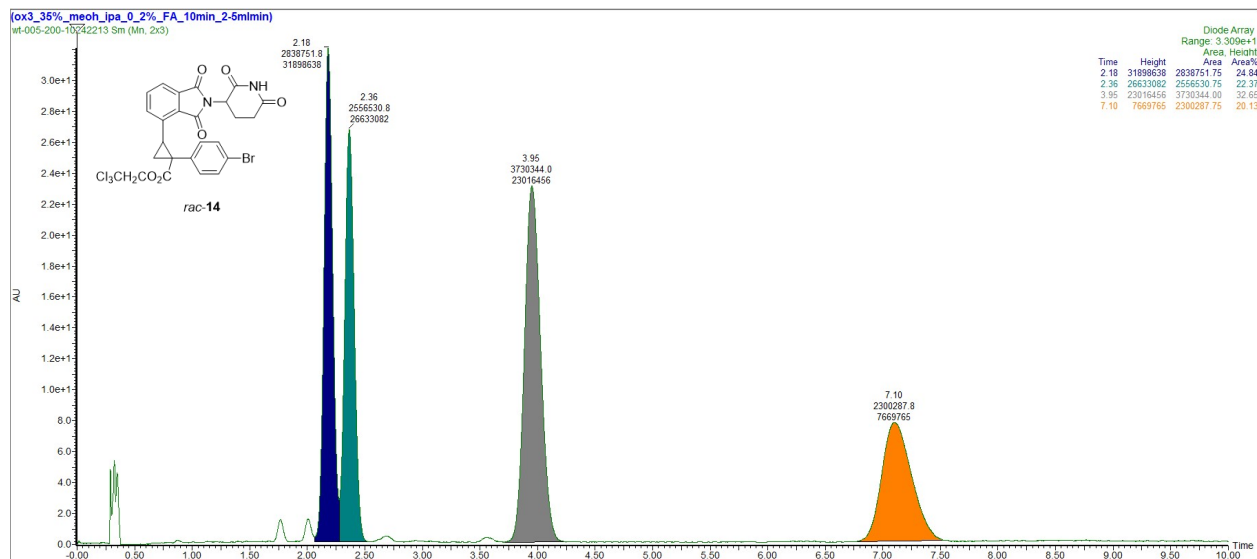
13a



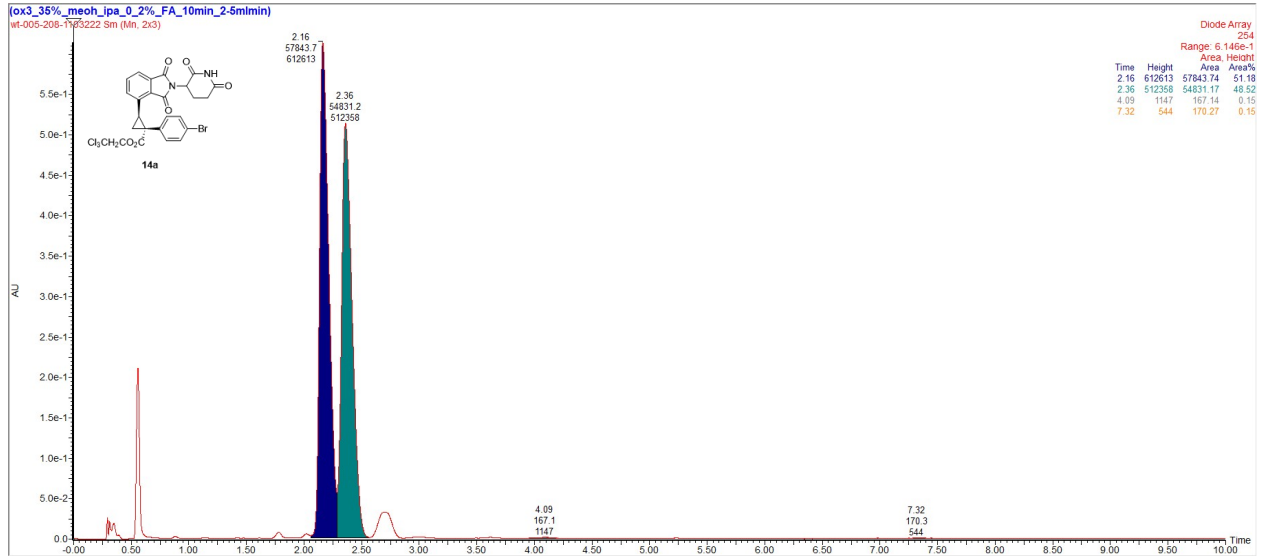
13b



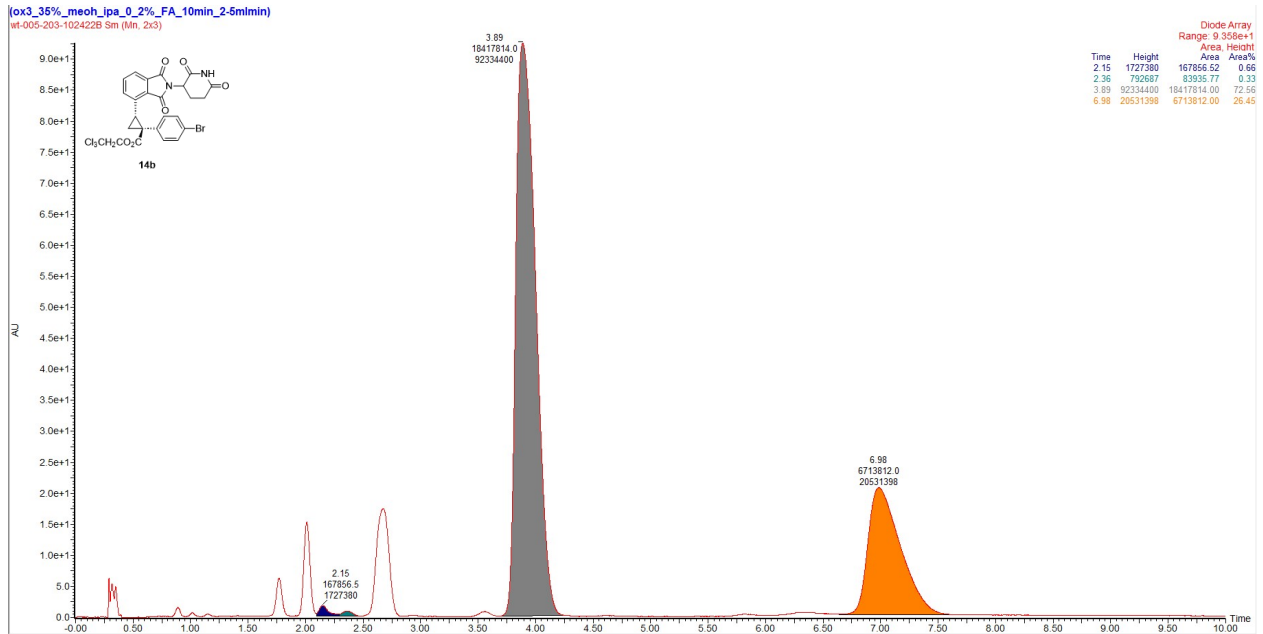
Racemic Chromatogram, 14



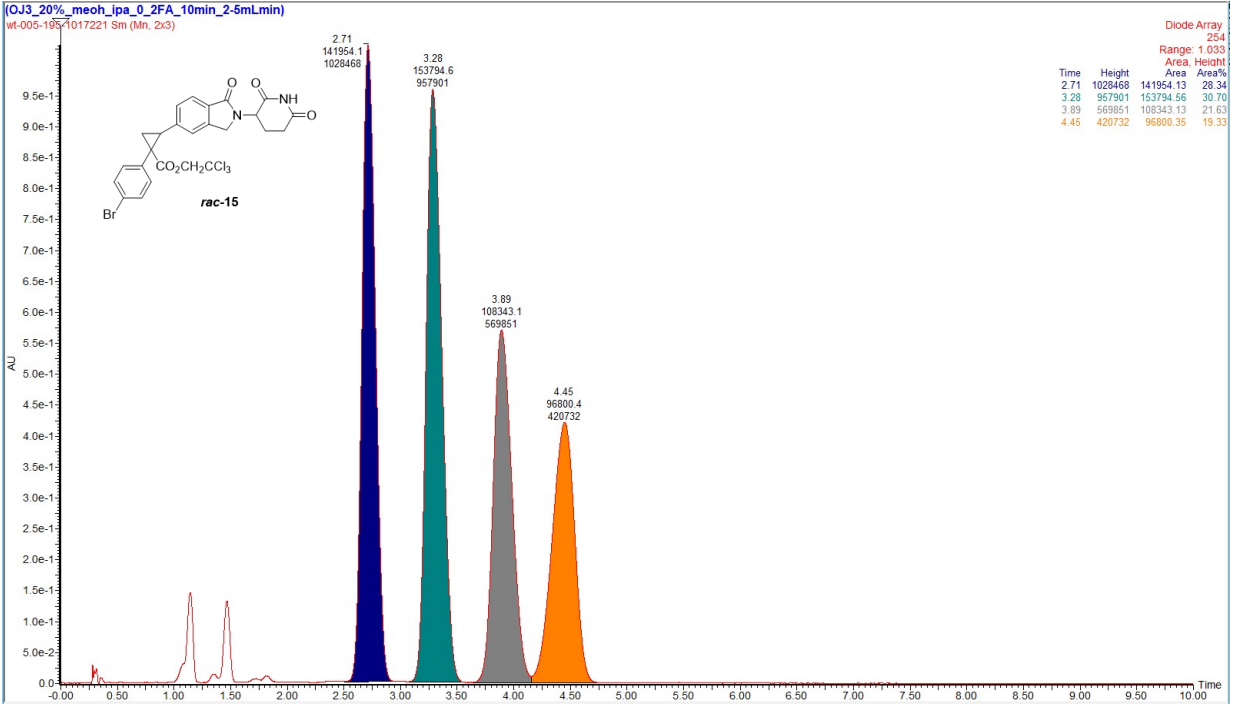
14a



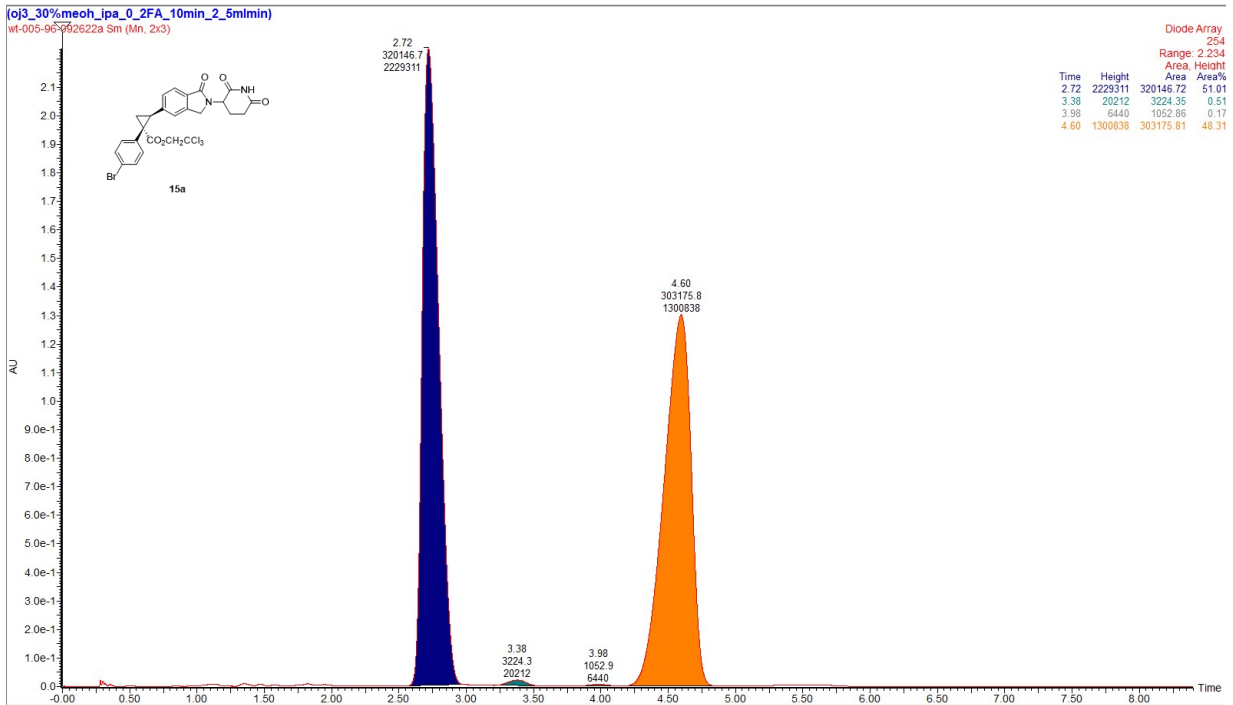
14b



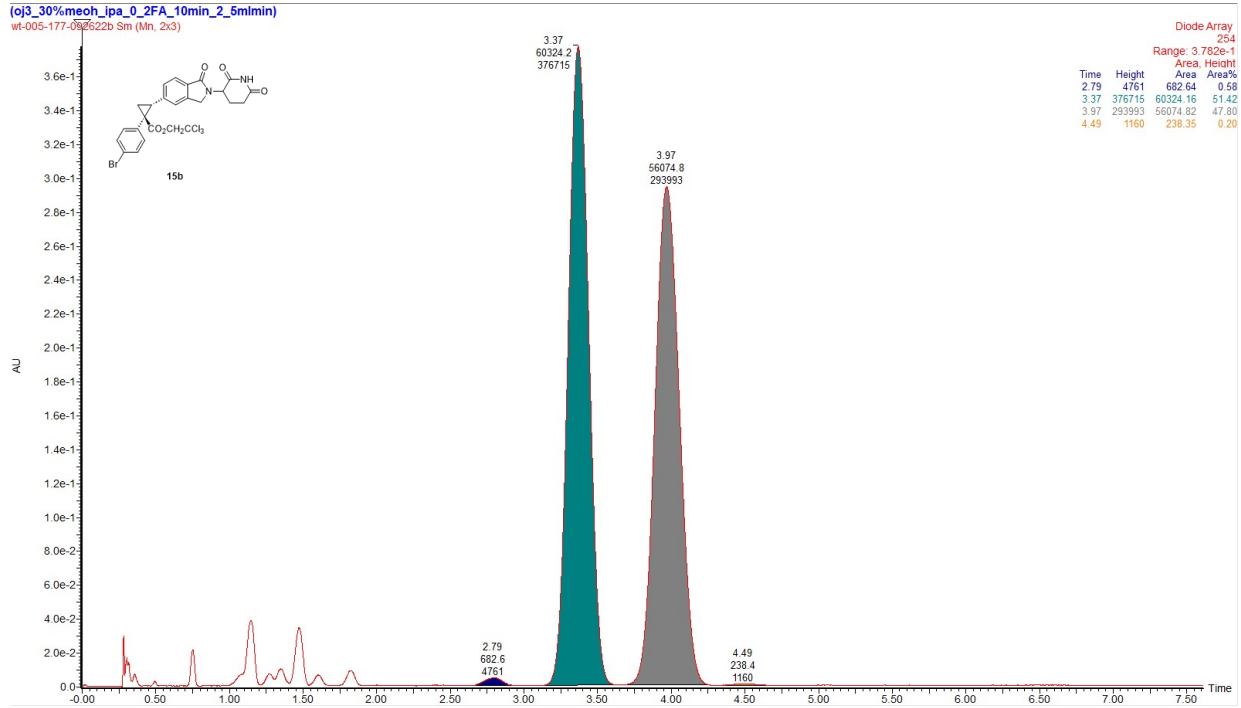
Racemic Chromatogram, 15



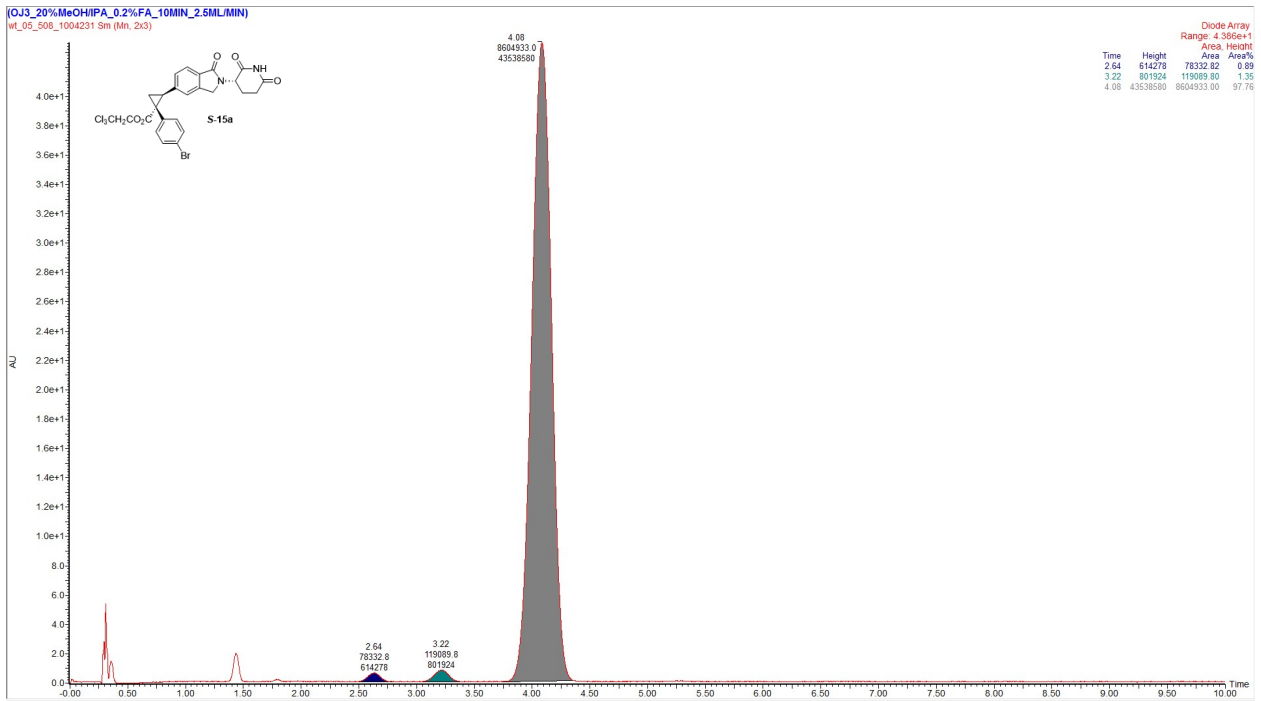
15a



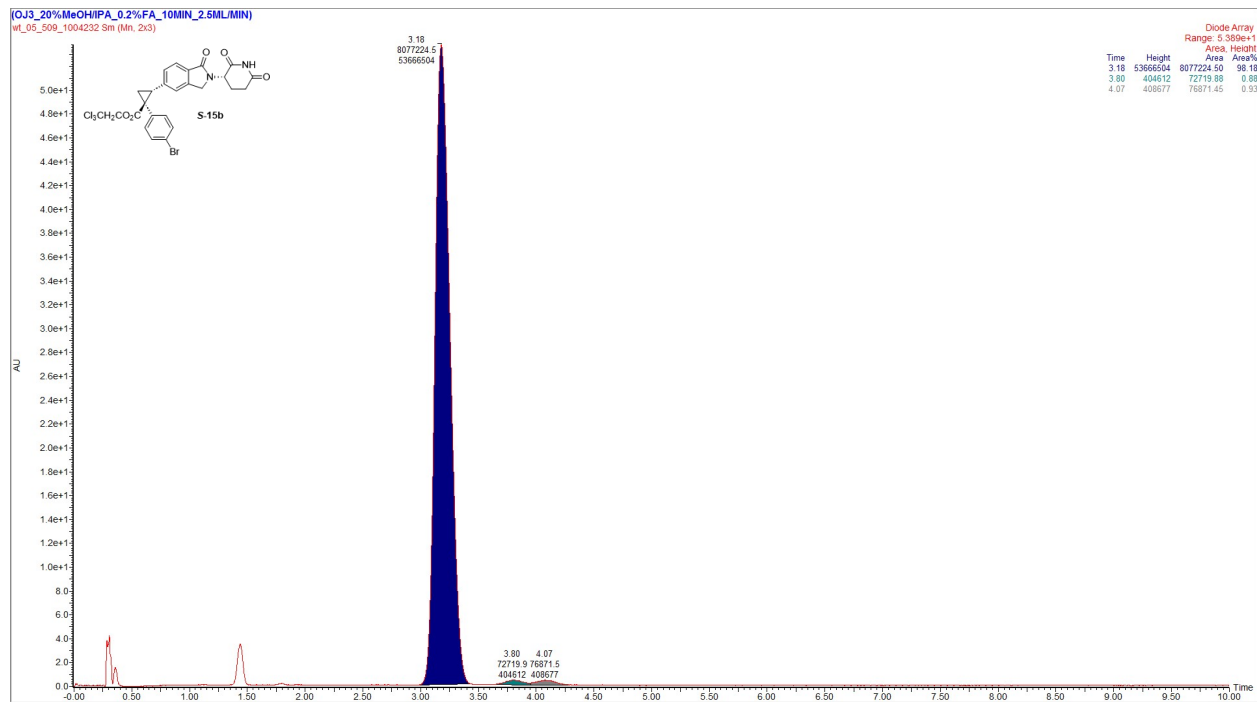
15b



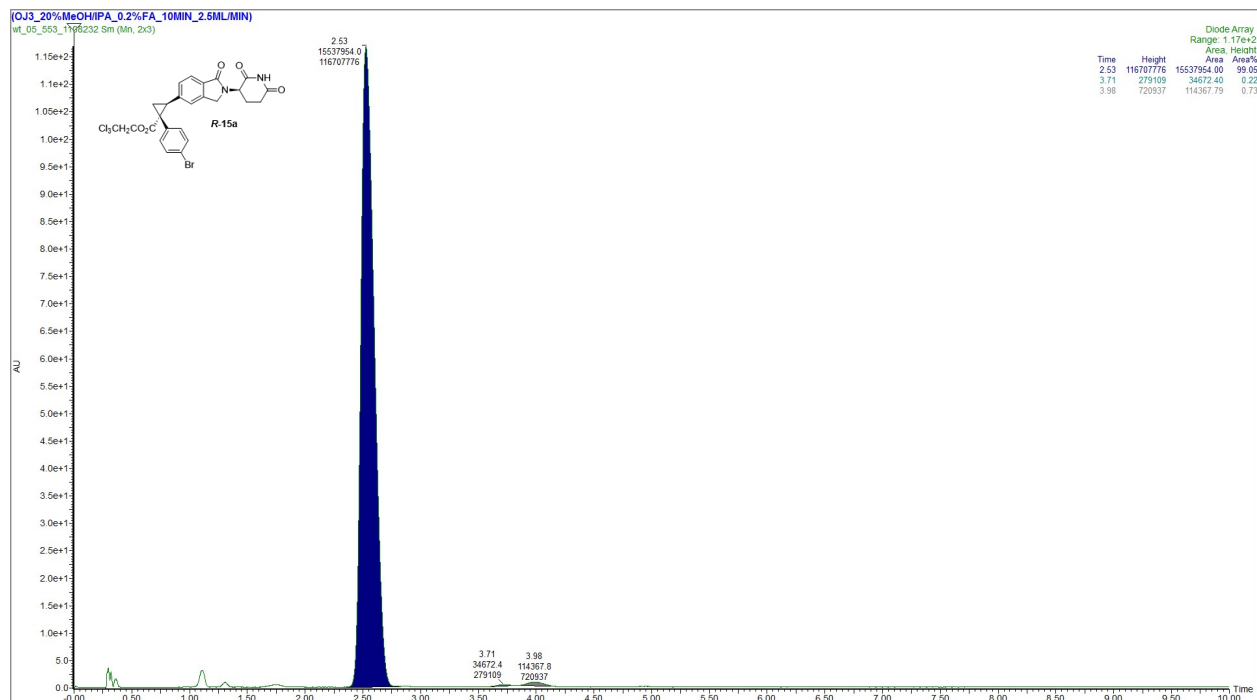
S-15a



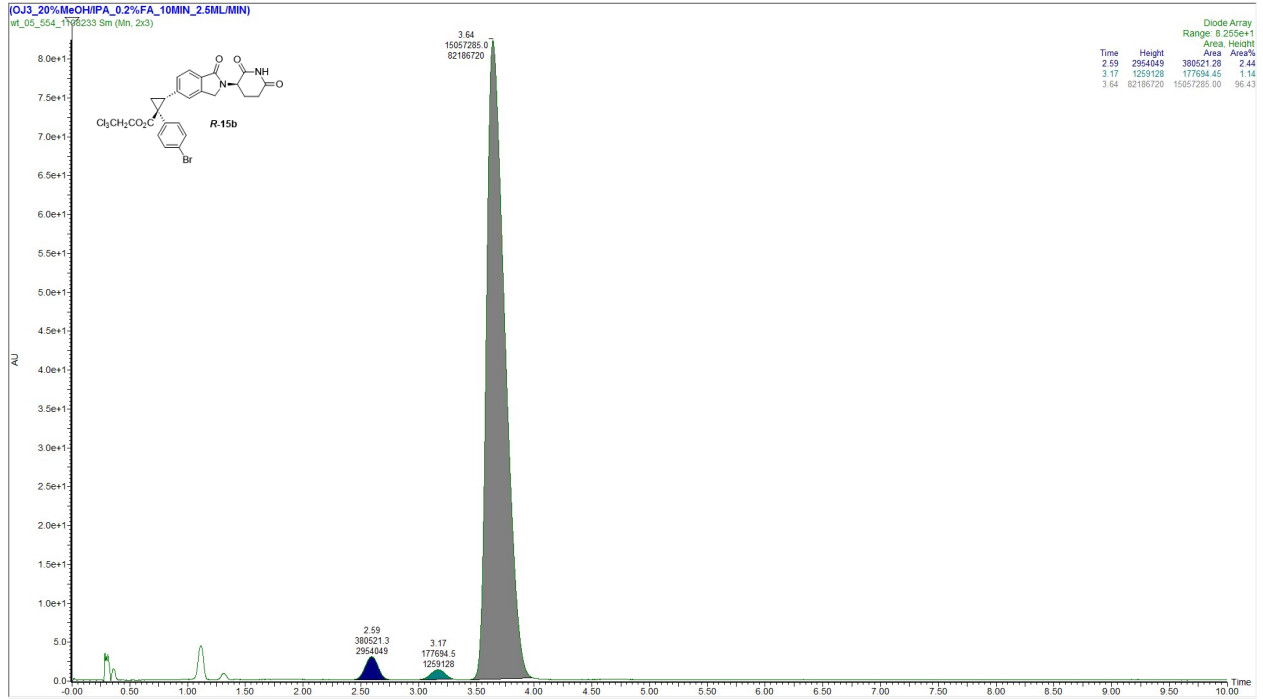
S-15b



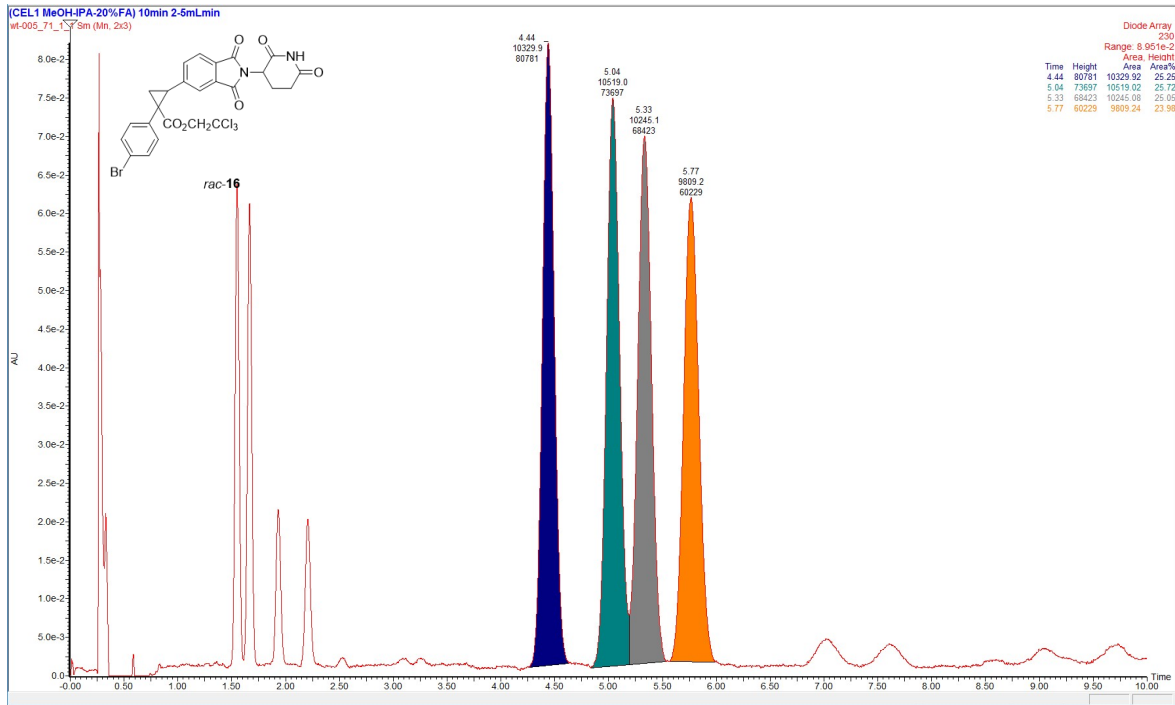
R-15a



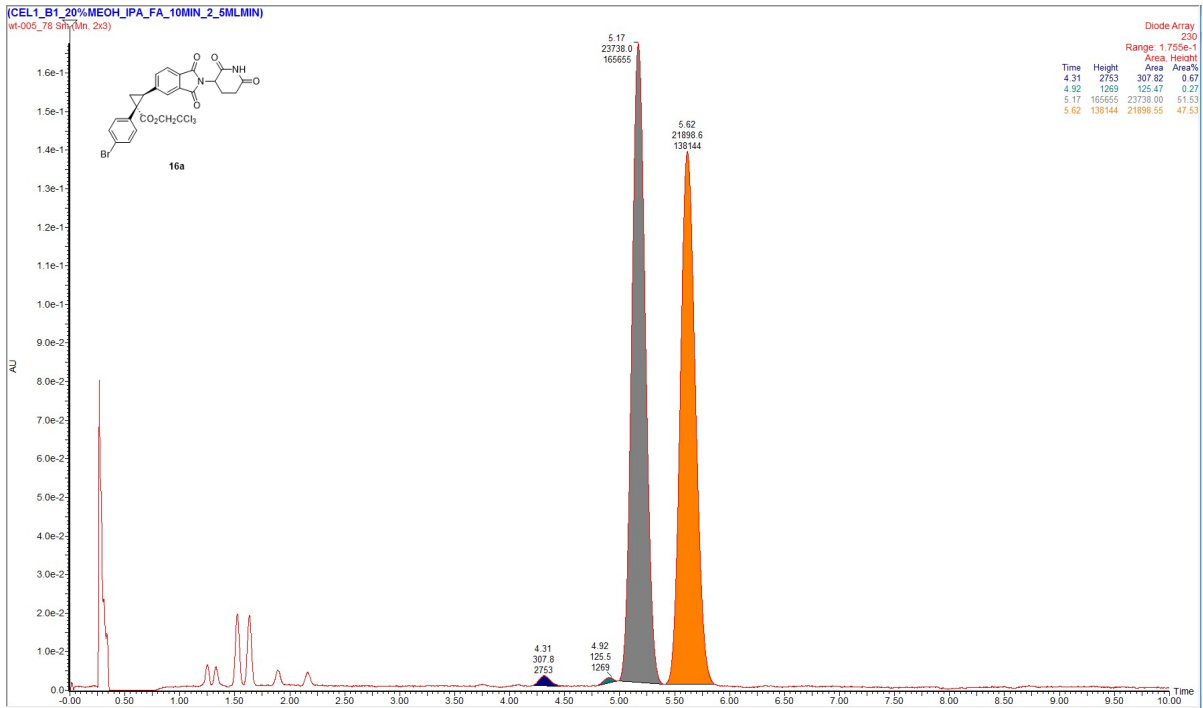
R-15b



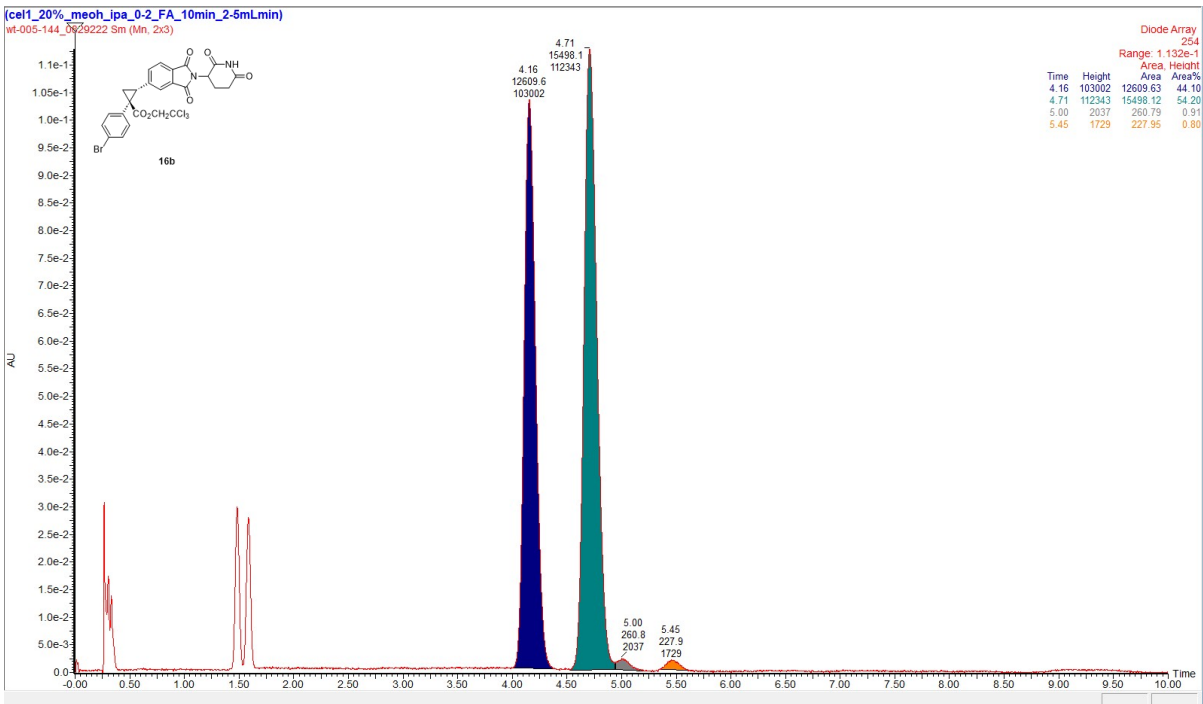
Racemic Chromatogram, 16



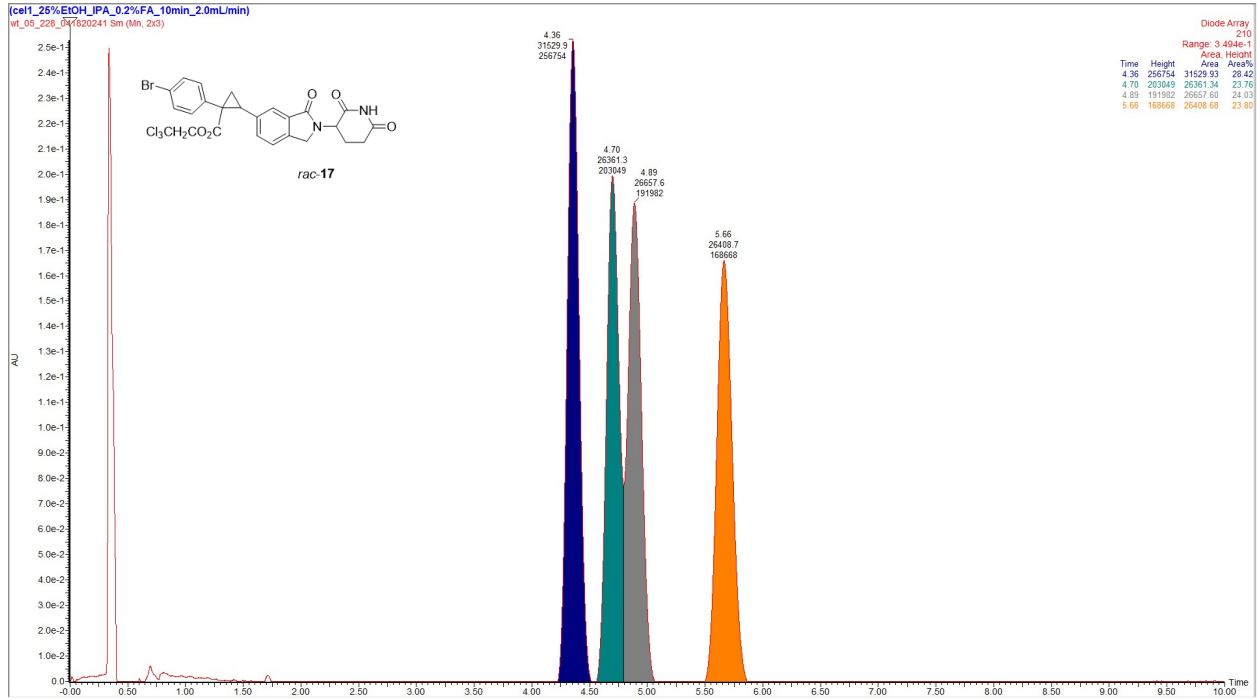
16a



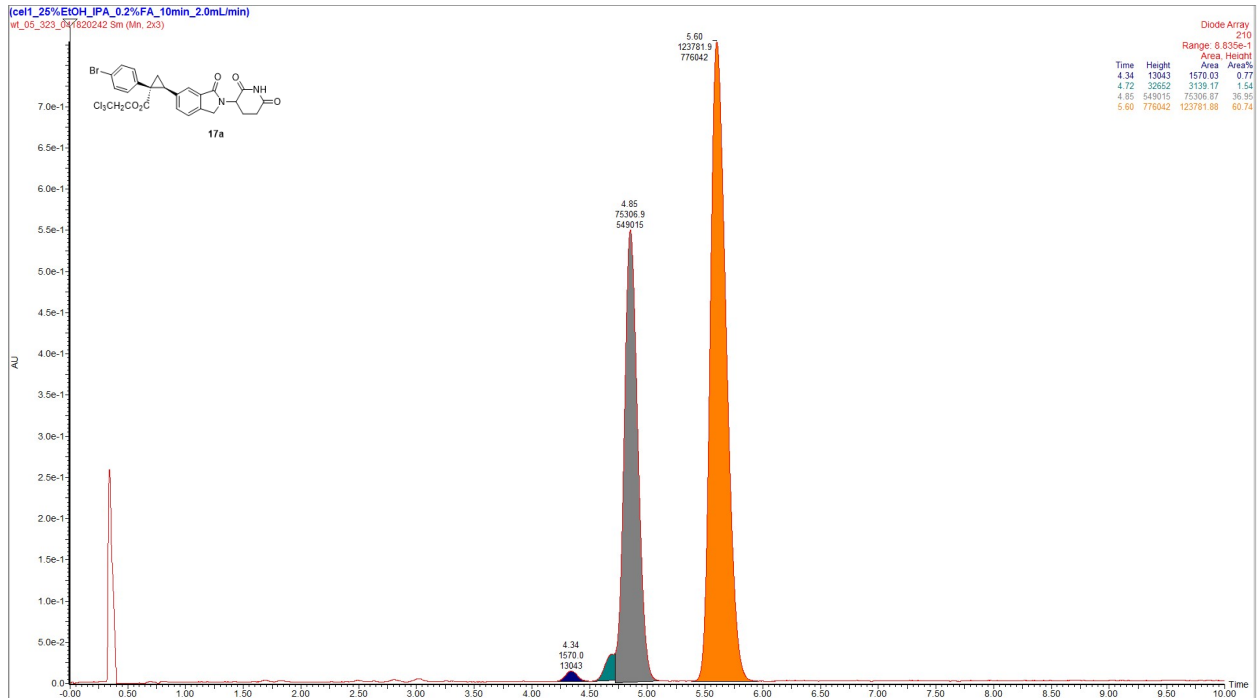
16b



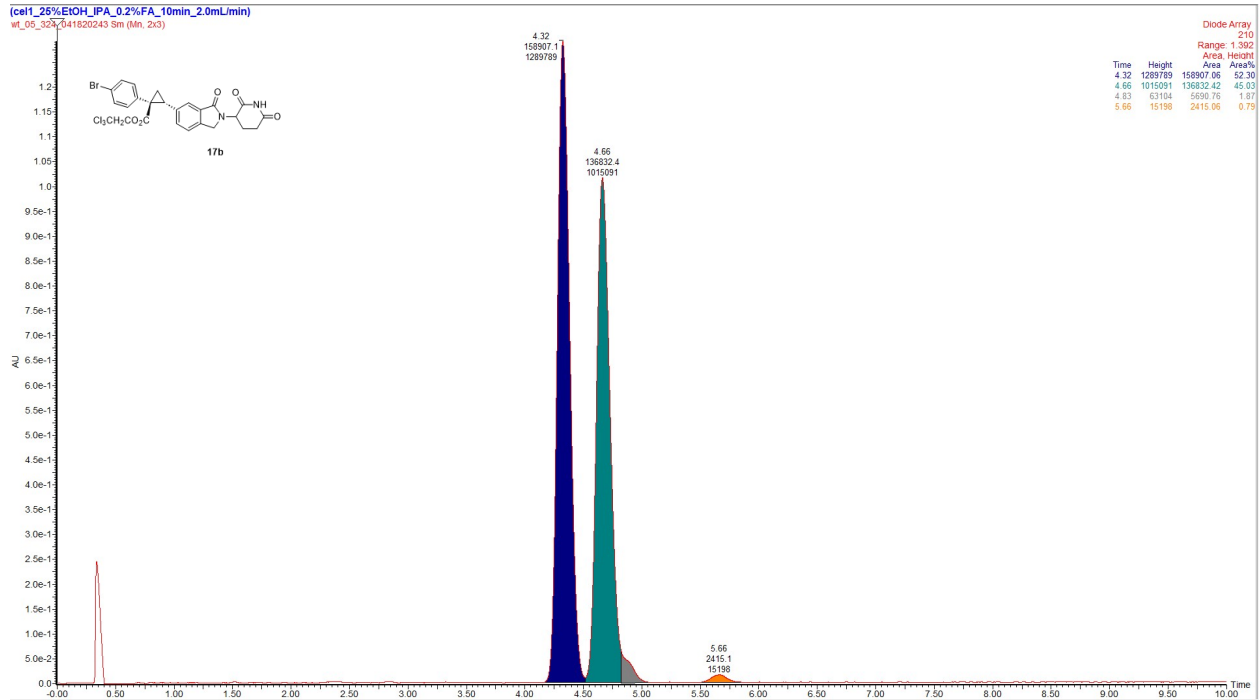
Racemic Chromatogram, 17



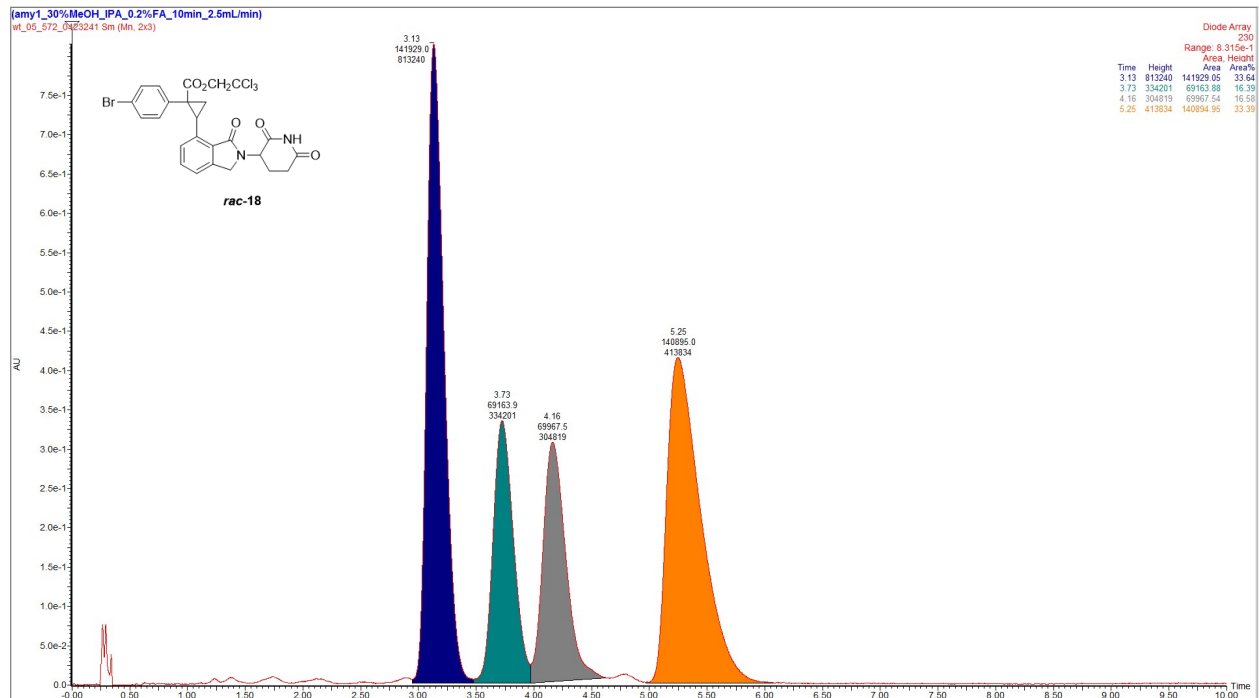
17a



17b

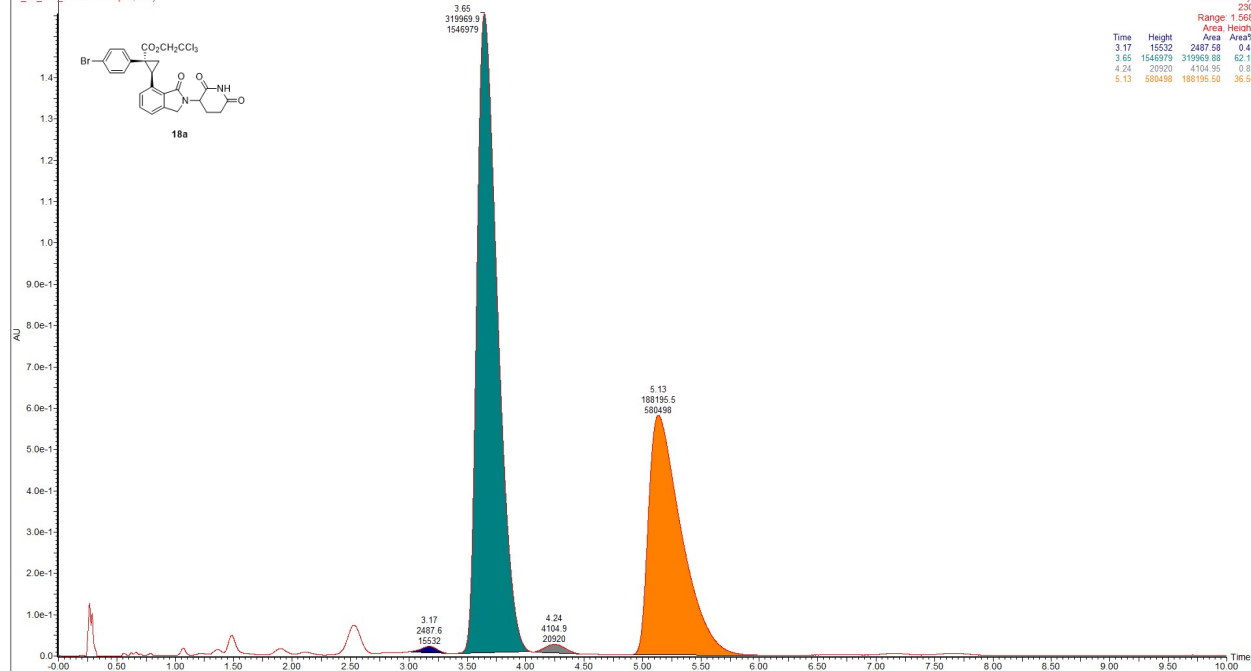


Racemic Chromatogram, 18



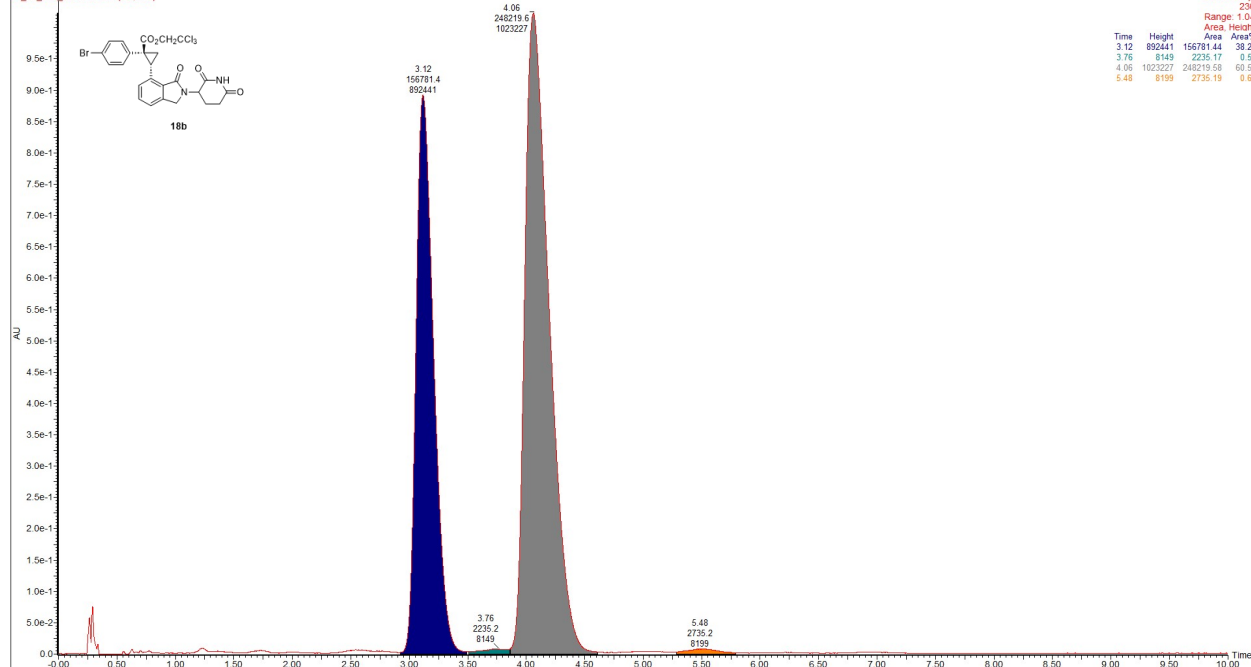
18a

[amy1_30%MeOH_IPA_0.2%FA_10min_2.6mL/min]
 wt_05_2035_0422243 Sm (Min, 2x3)

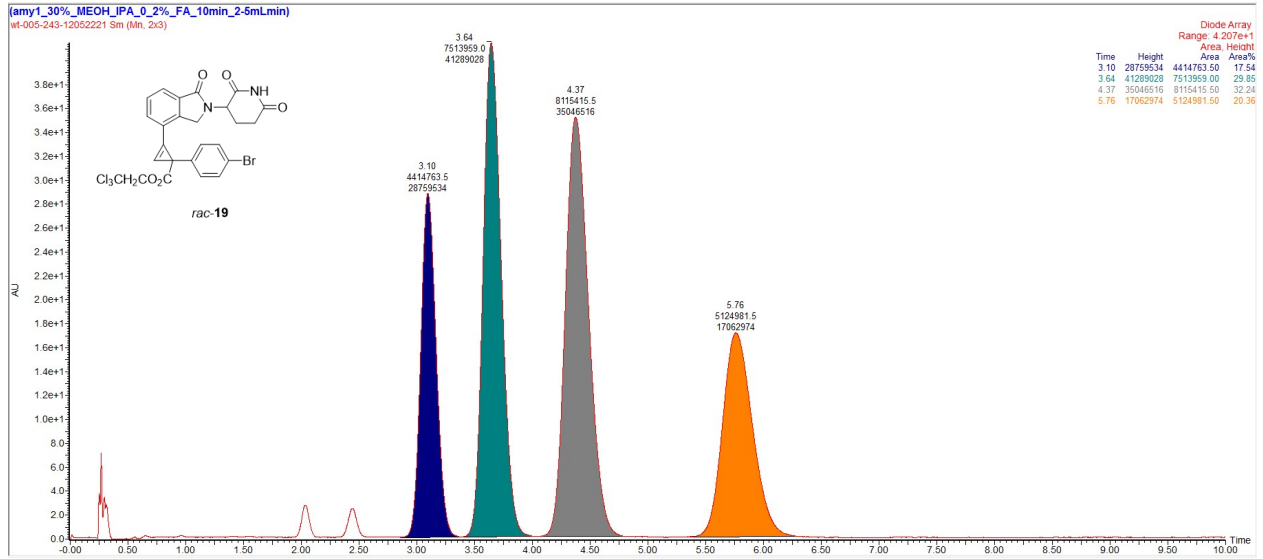


18b

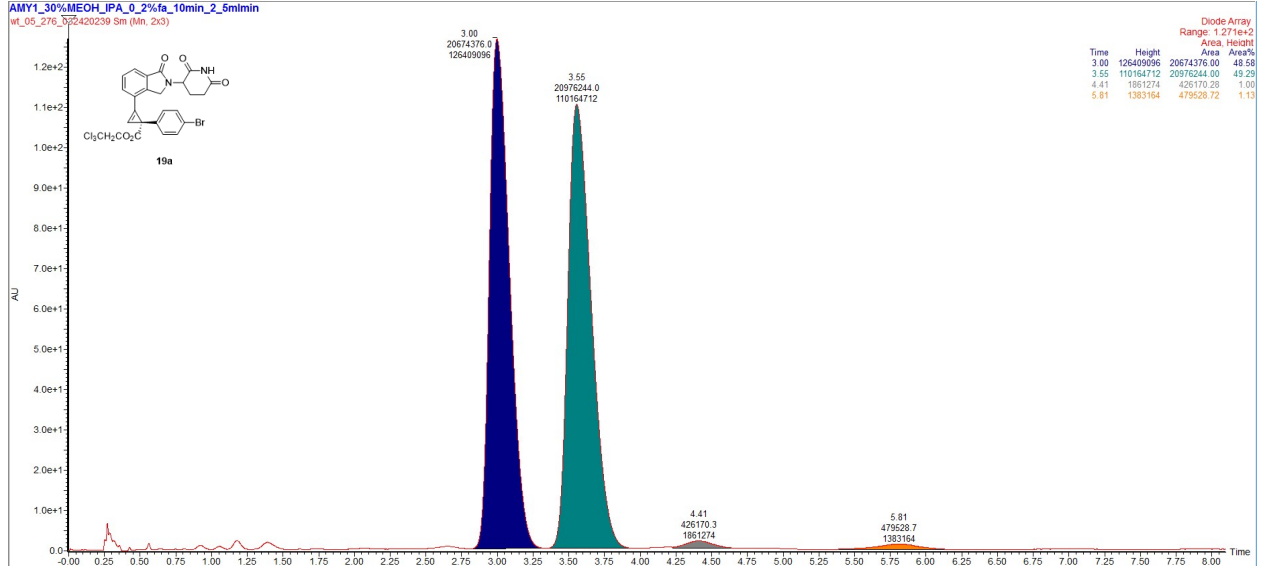
[amy1_30%MeOH_IPA_0.2%FA_10min_2.6mL/min]
 wt_05_5735_0423242 Sm (Min, 2x3)



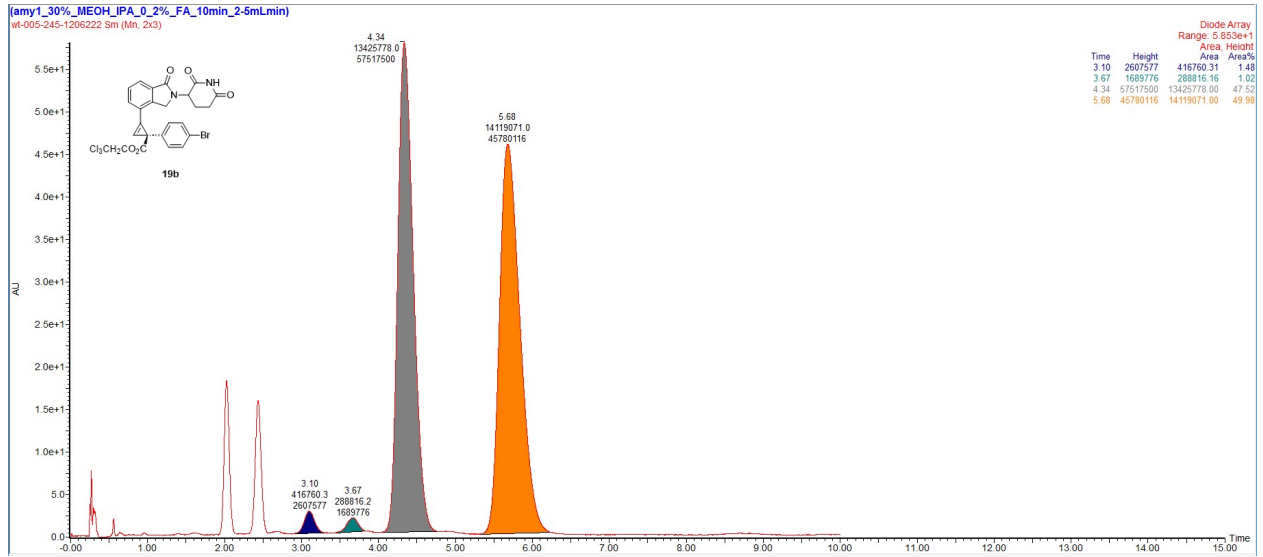
Racemic Chromatogram, 19



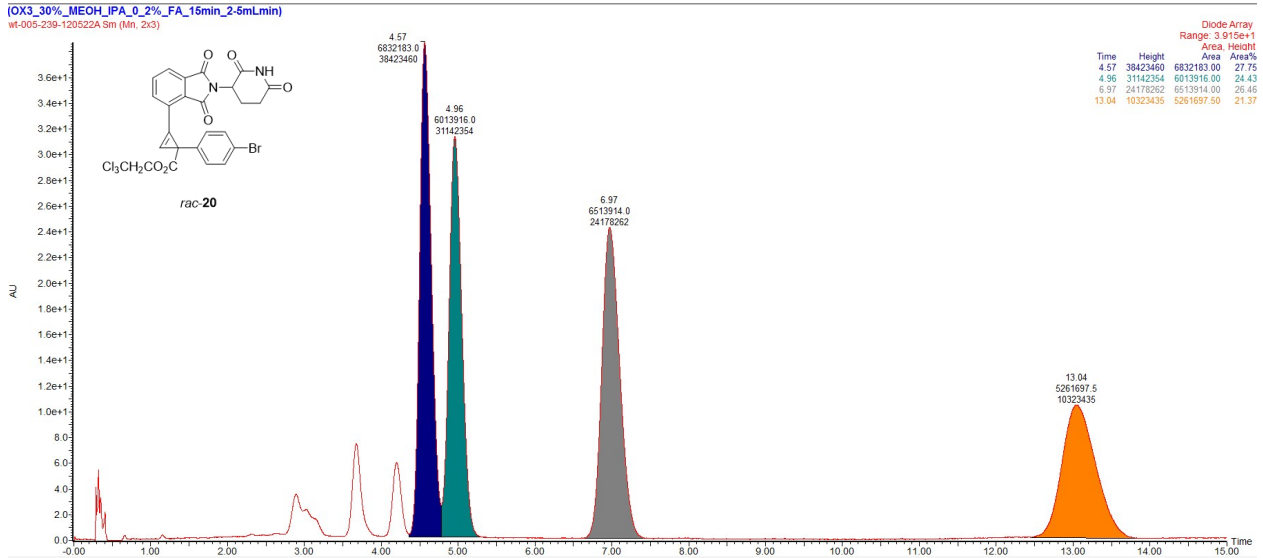
19a



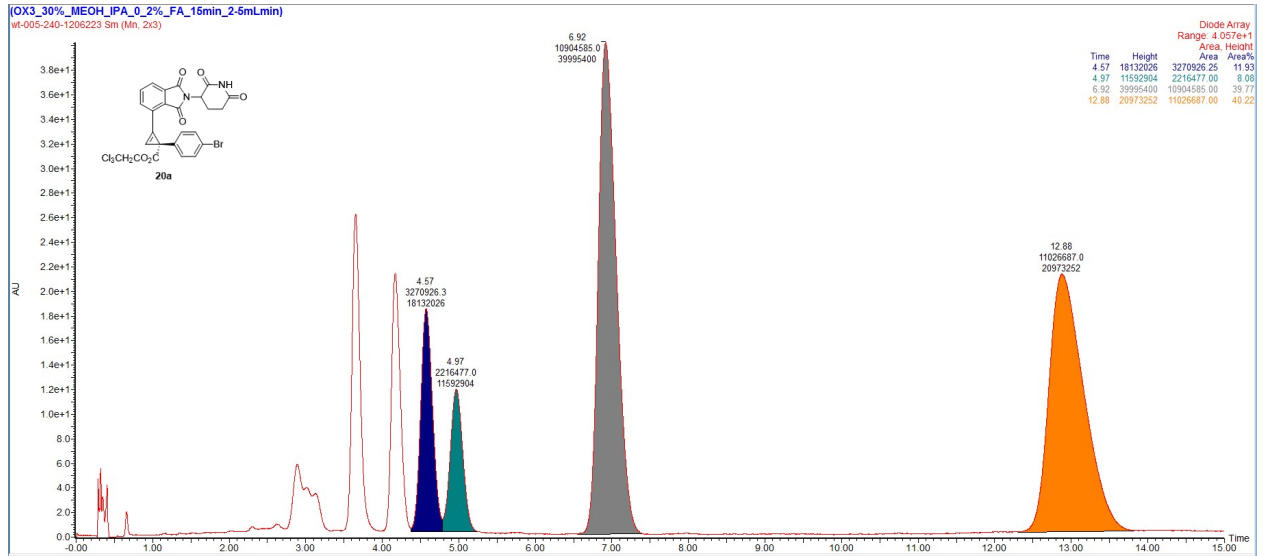
19b



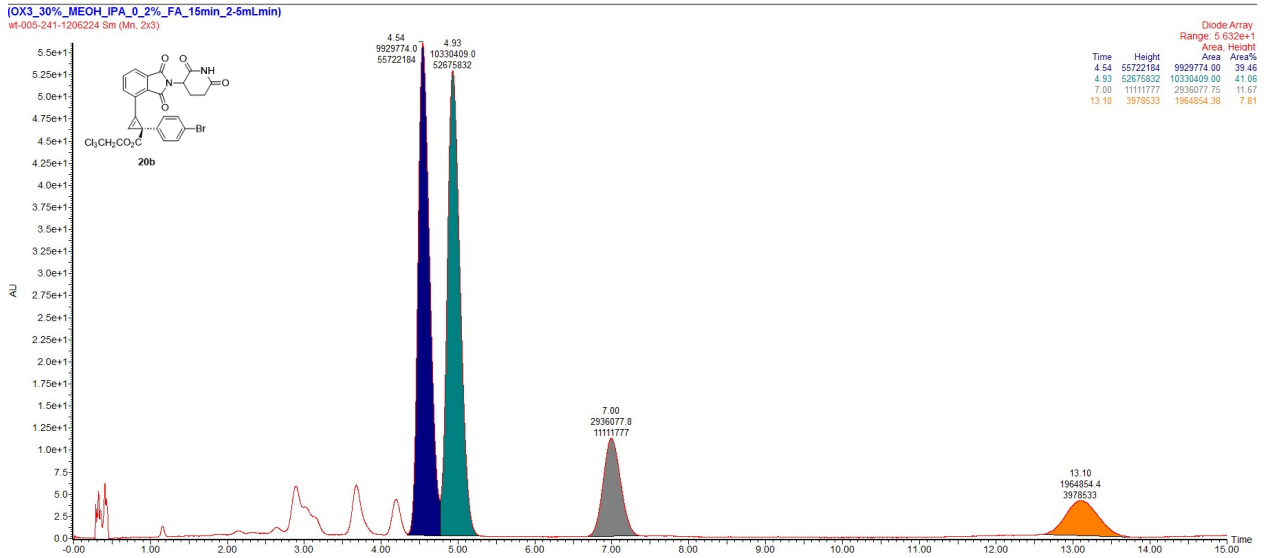
Racemic Chromatogram, 20



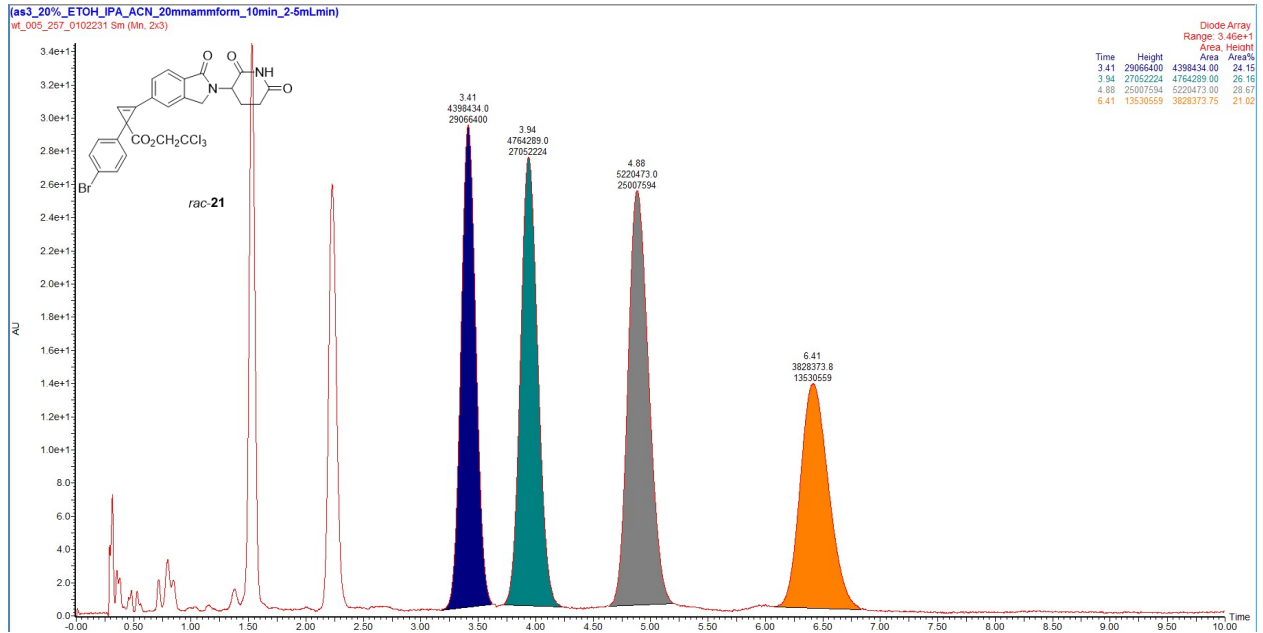
20a



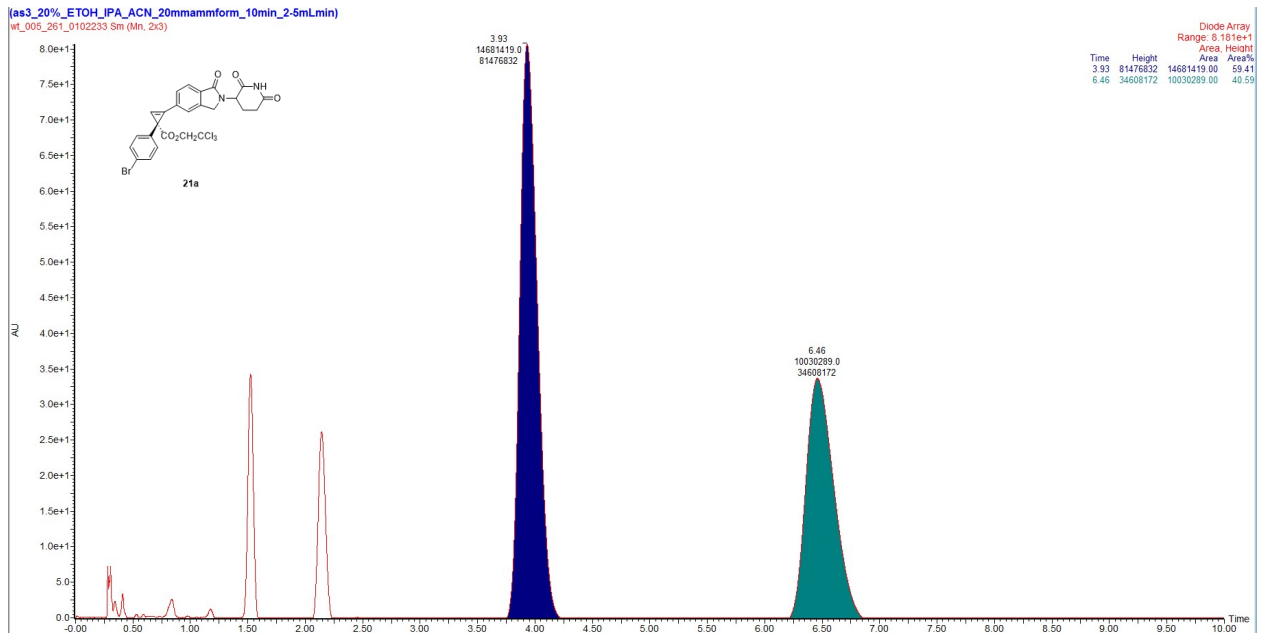
20b



Racemic Chromatogram, 21

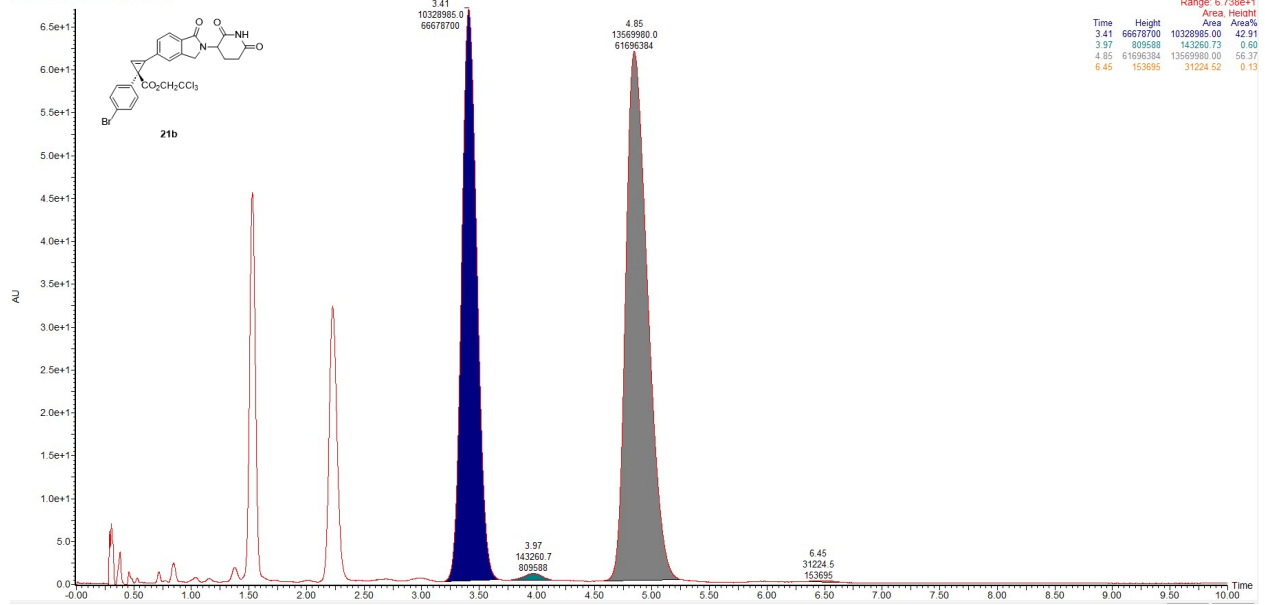


21a



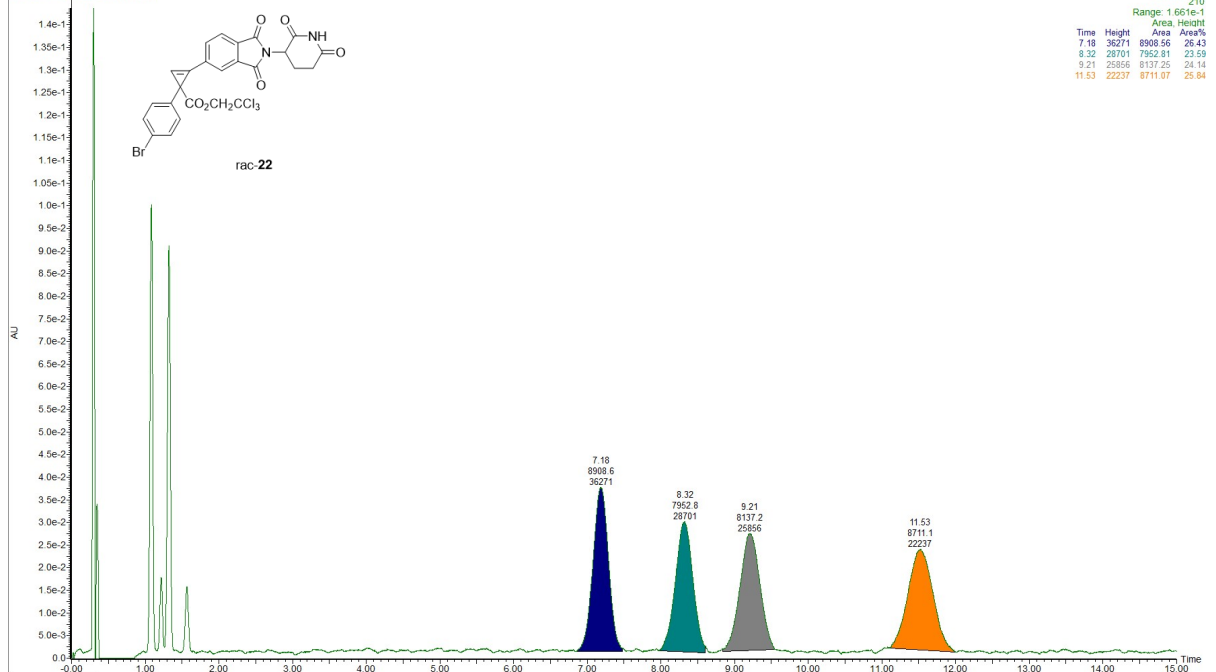
21b

(as3_20%_ETOH_IPA_ACN_20mmmmform_10min_2-5mLmin)
wt_005_262_0102234 Sm (Mn, 2x3)

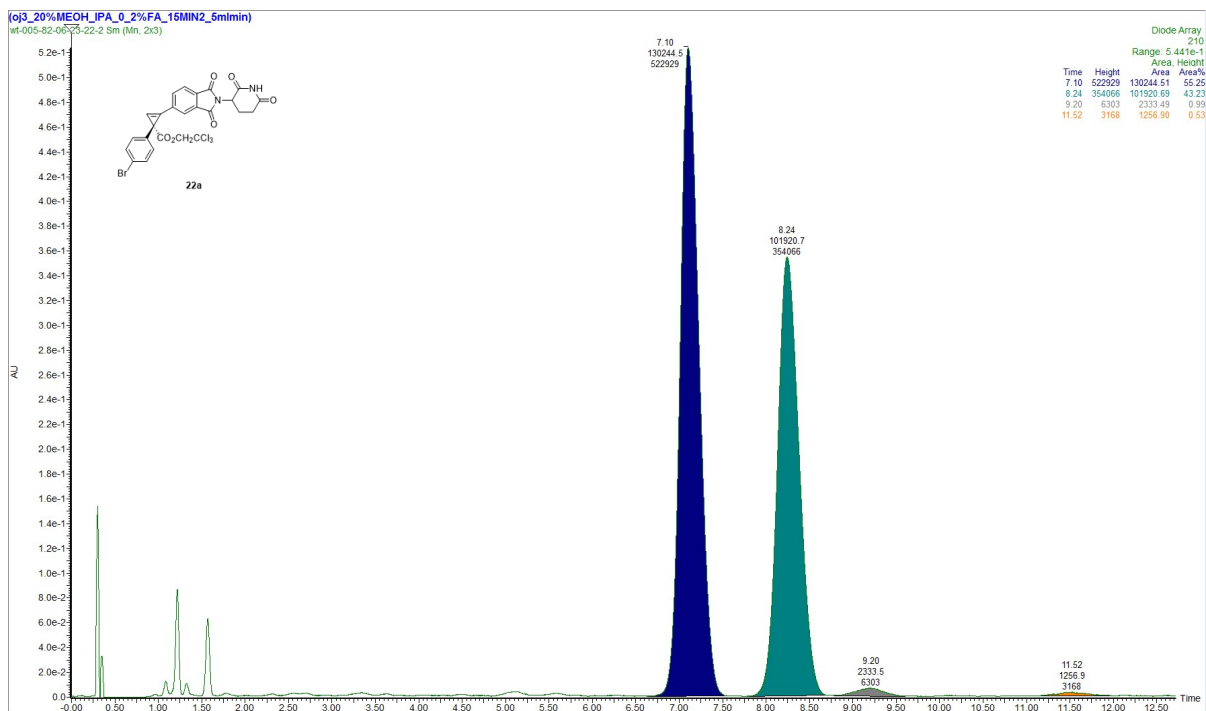


Racemic Chromatogram, 22

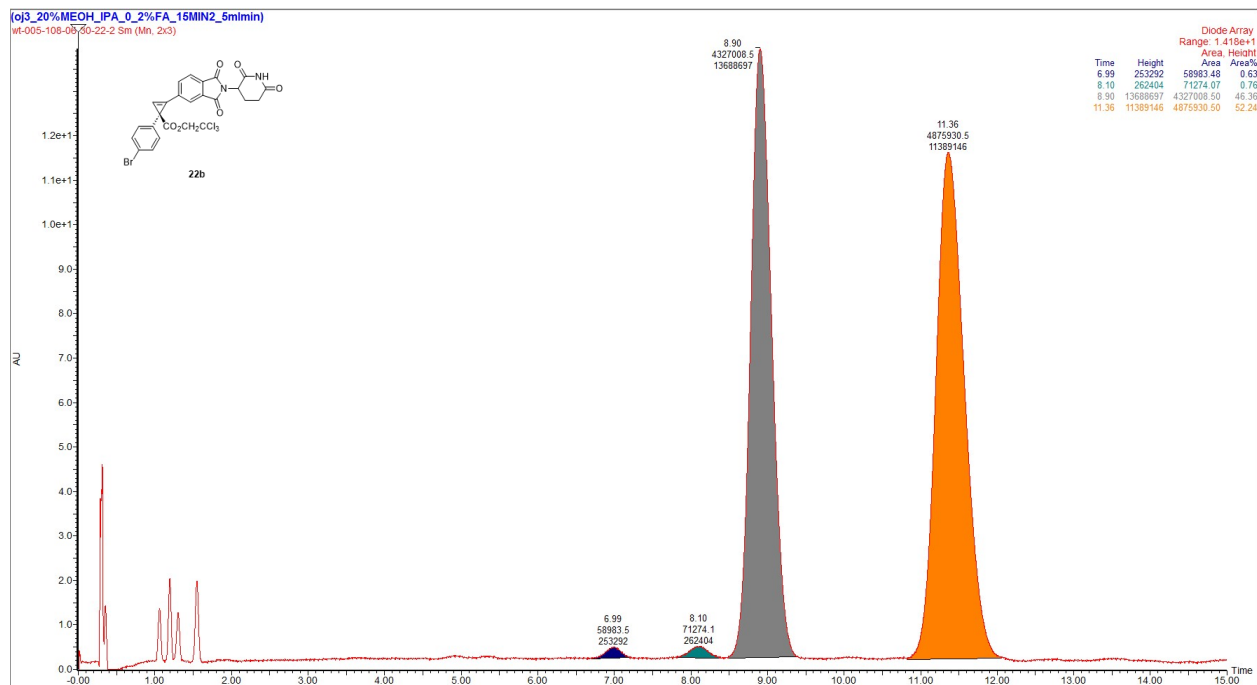
(cj3_20%_MEOH_IPA_0_2%_FA_15MIN2_5minin)
wt-005-101-0523228 Sm (Mn, 2x3)



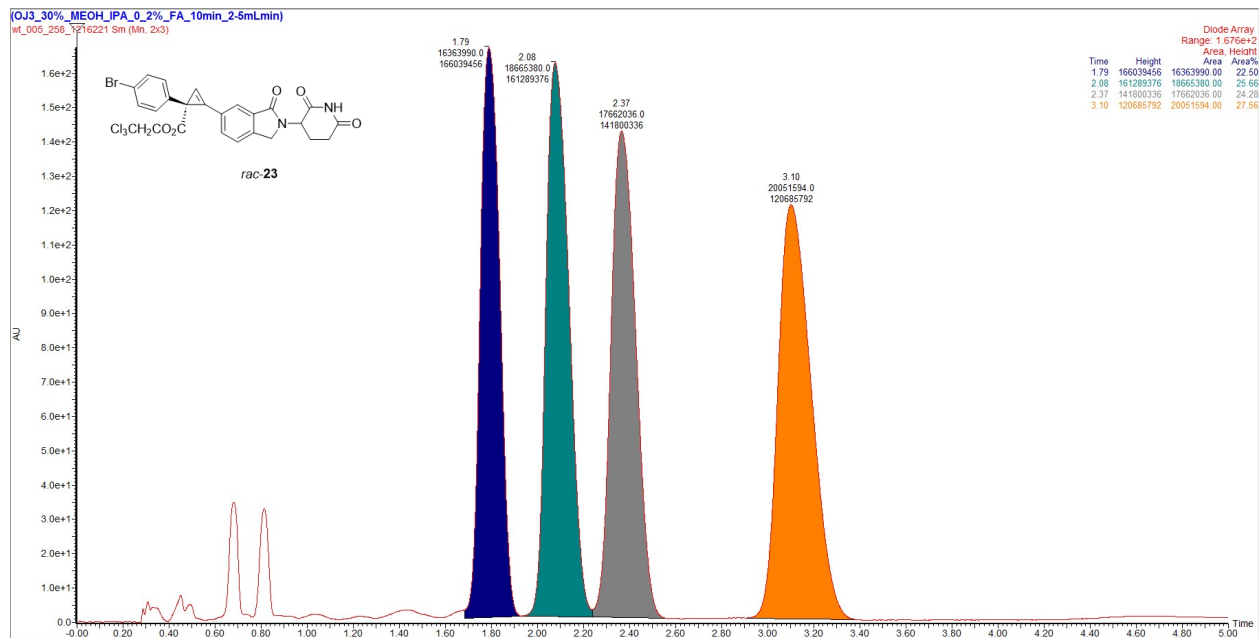
22a



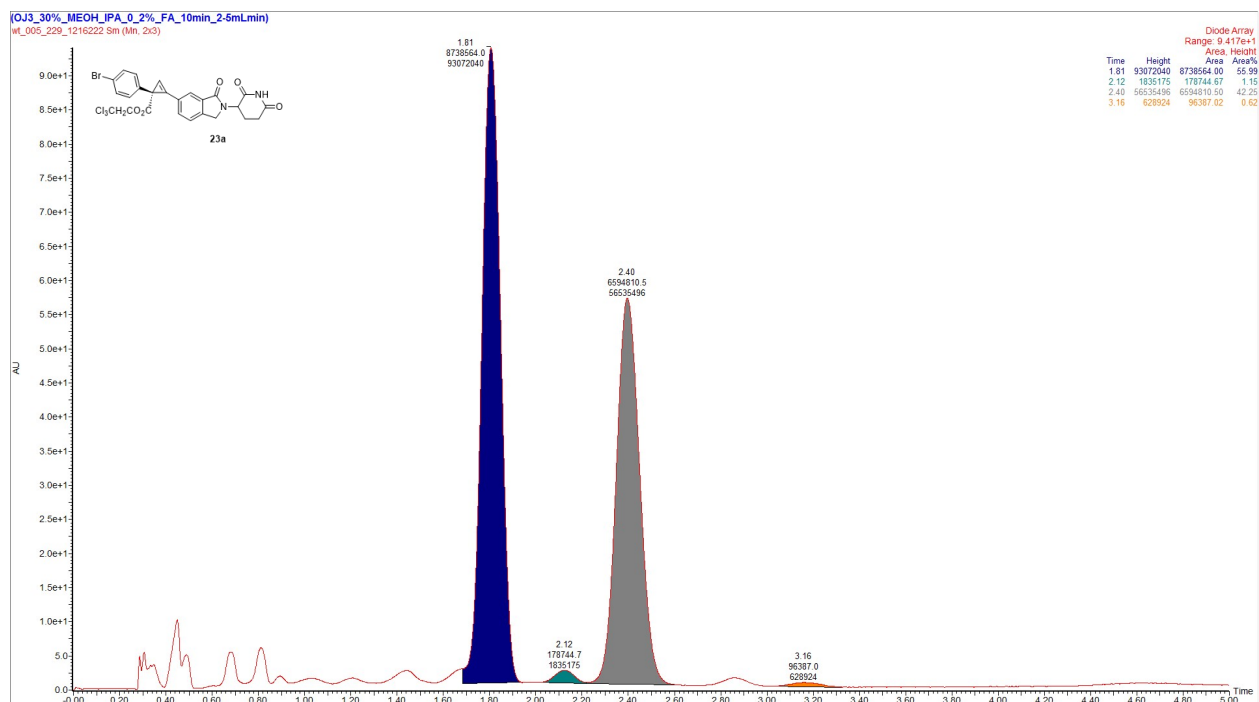
22b



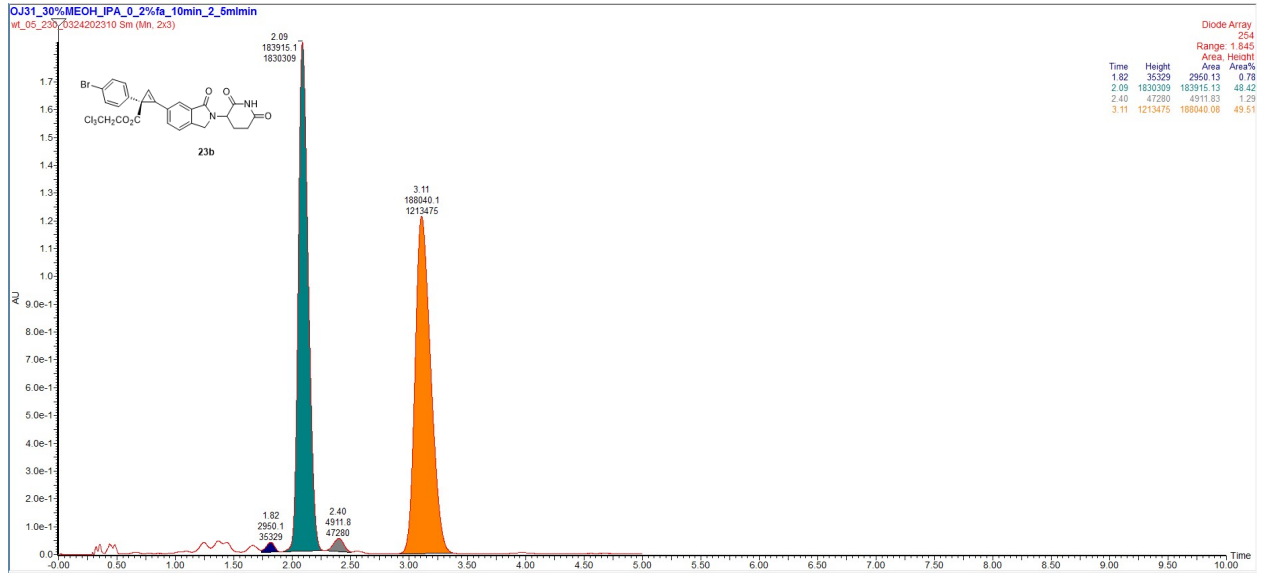
Racemic Chromatogram, 23



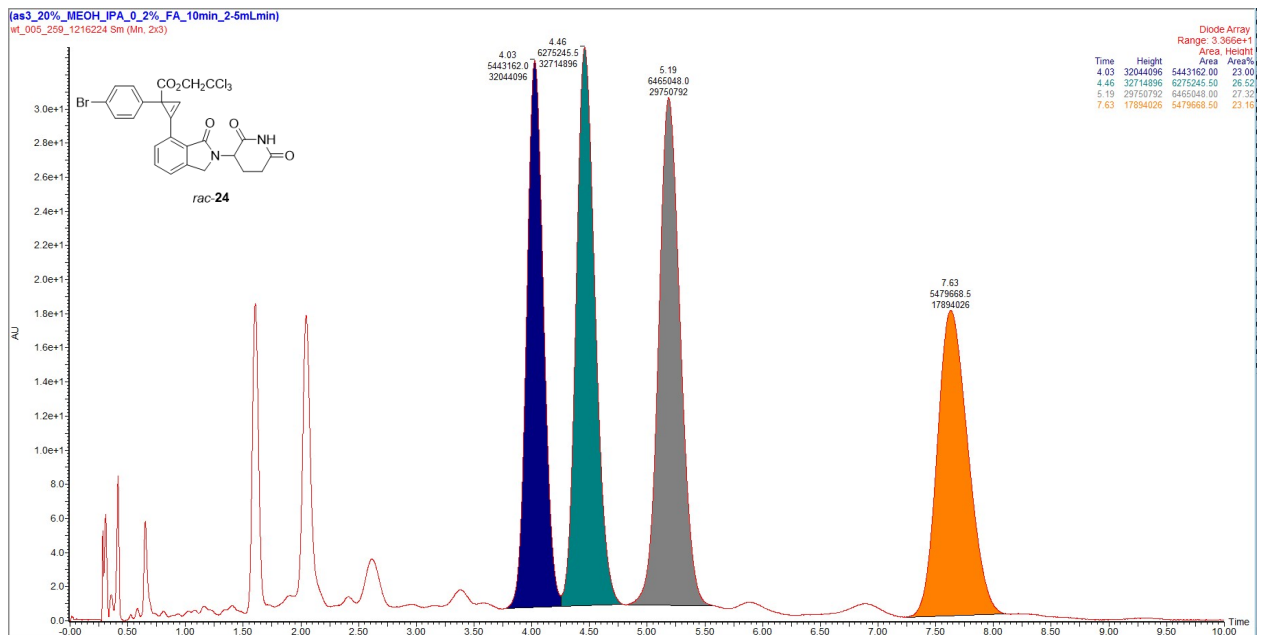
23a



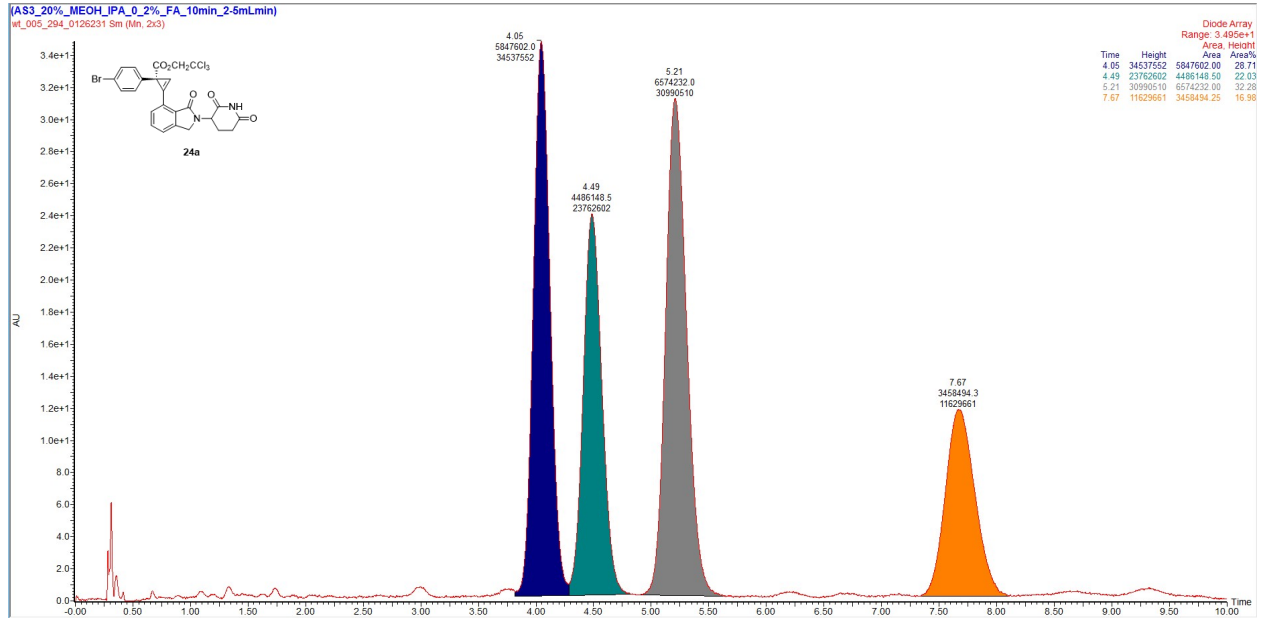
23b



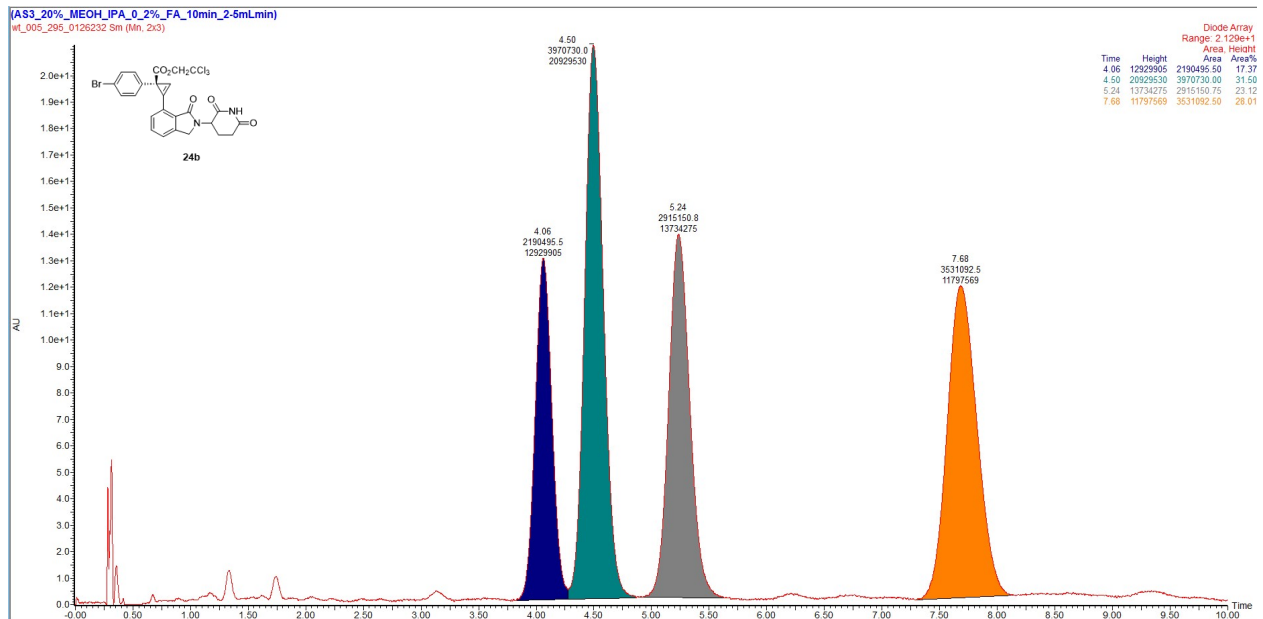
Racemic Chromatogram, 24



24a

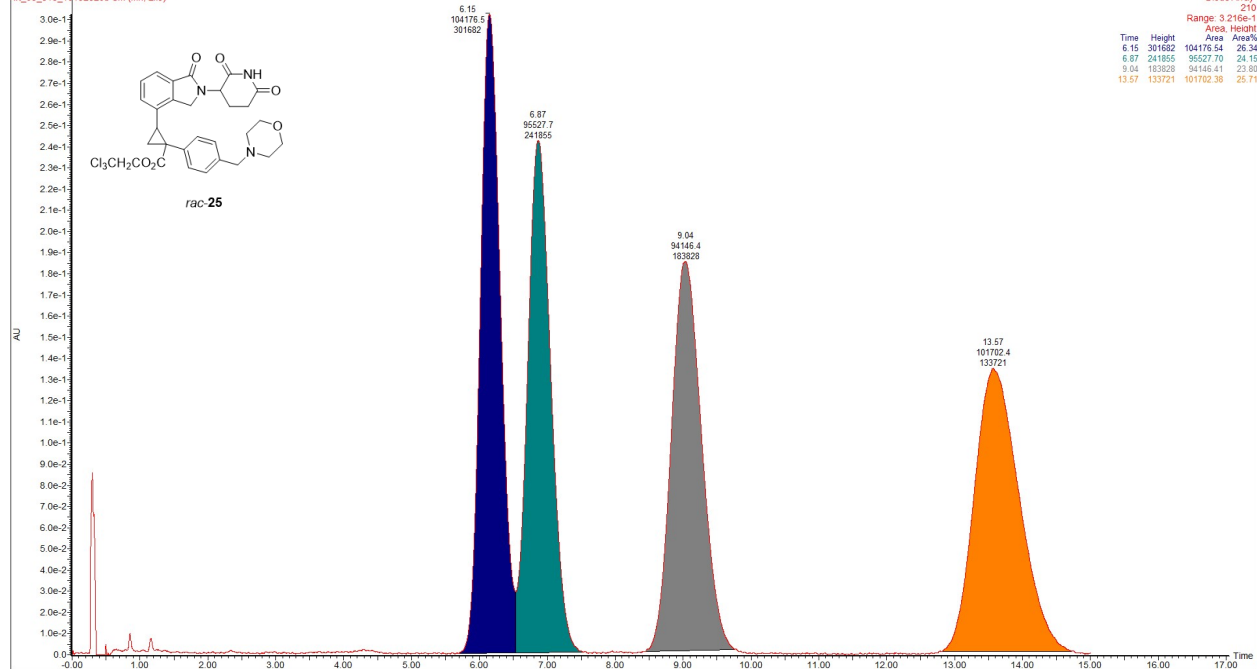


24b



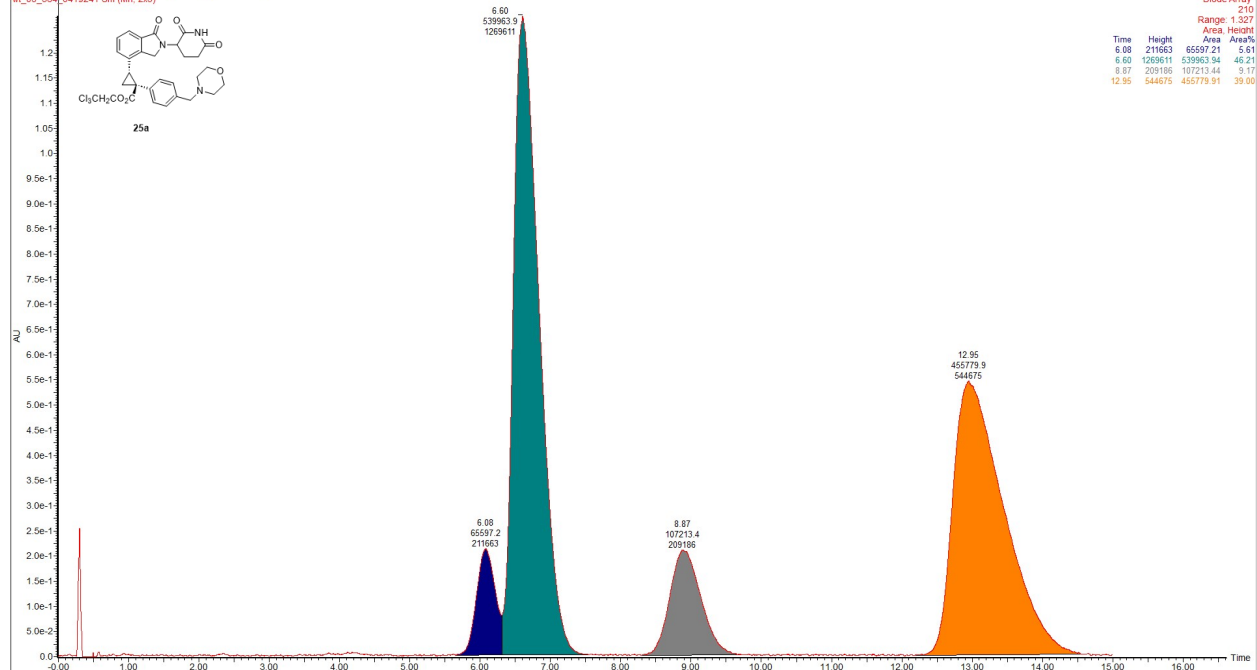
Racemic Chromatogram, 25

[CEL2_35%MeOH_IPA_0.2%FA_16min_2.5mL/min]
 wt_05_519_151923209 Sm (Mn, 2x3)

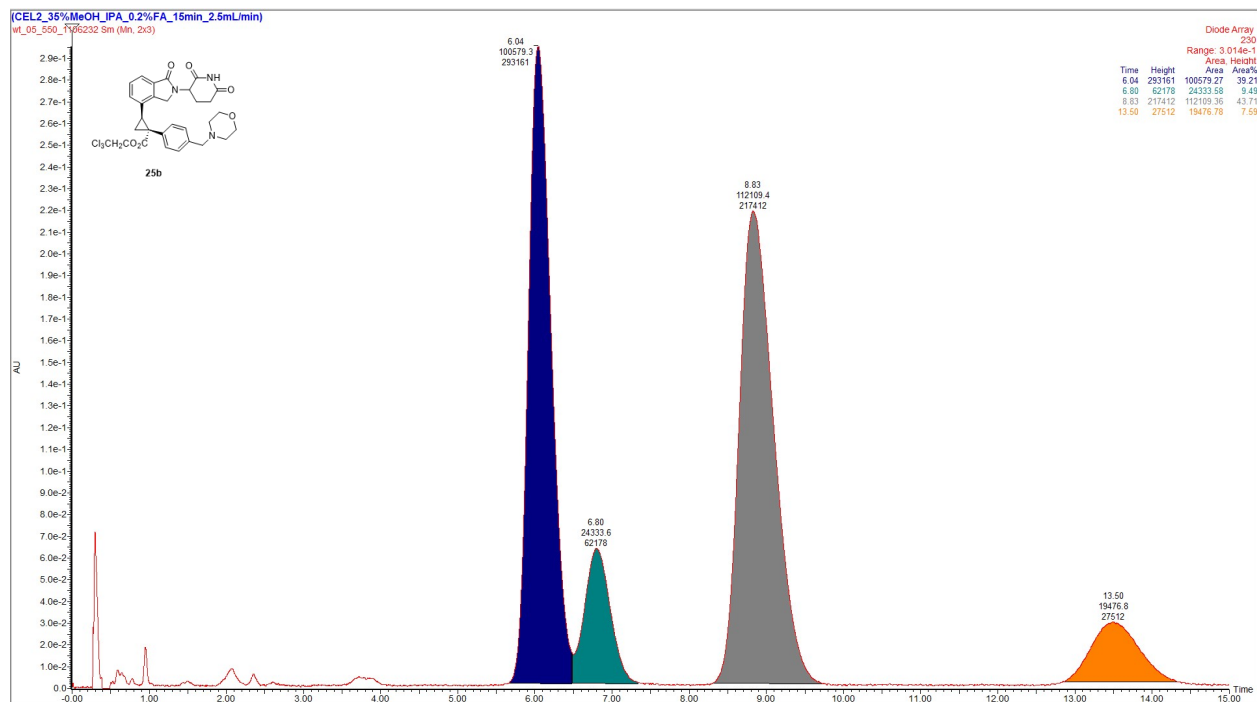


25a

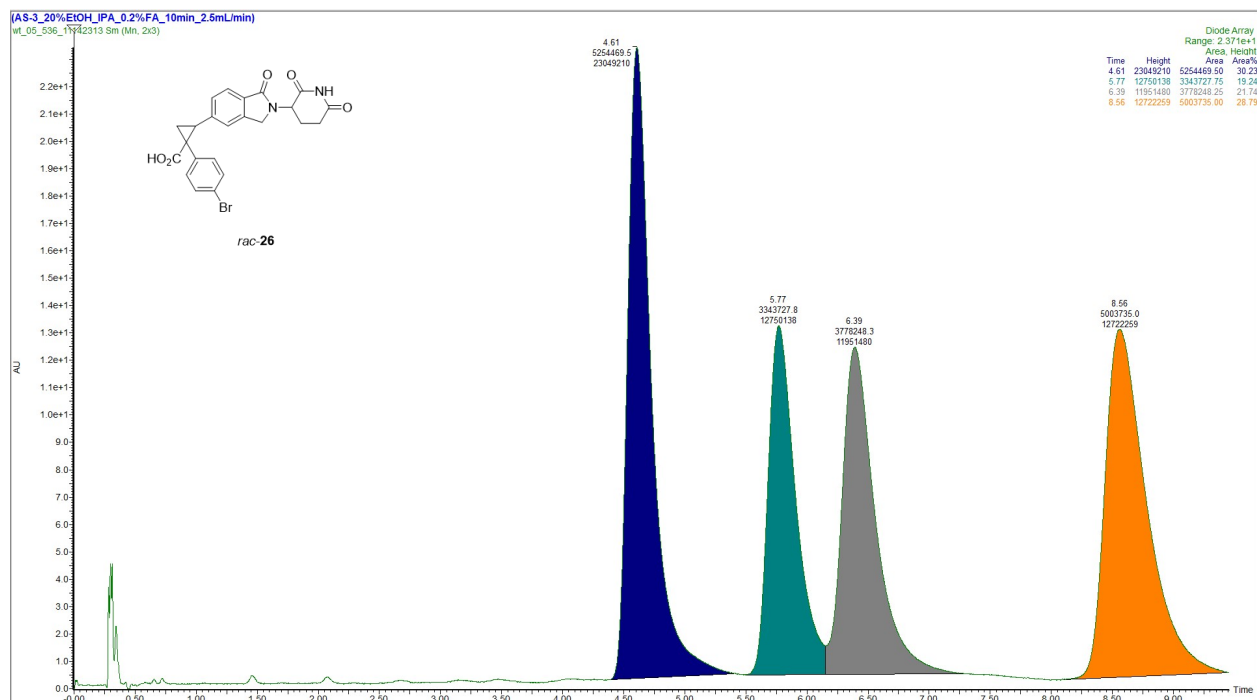
[CEL2_35%MeOH_IPA_0.2%FA_16min_2.5mL/min]
 wt_05_563_0419241 Sm (Mn, 2x3)



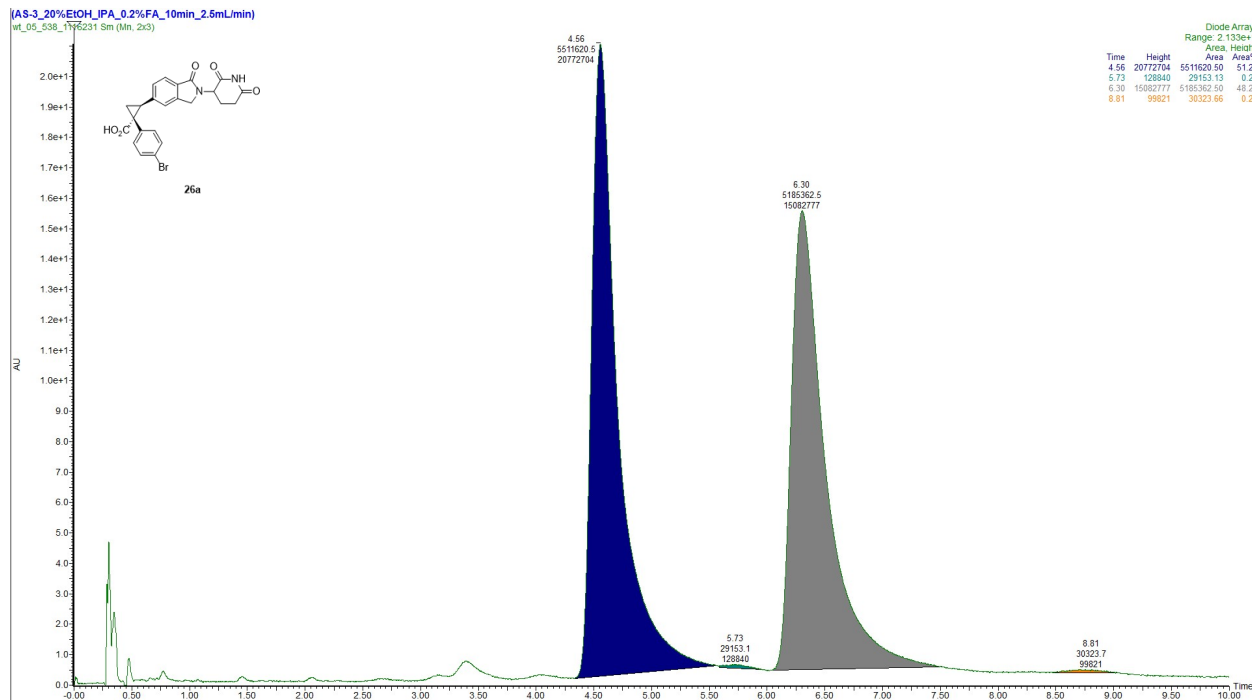
25b



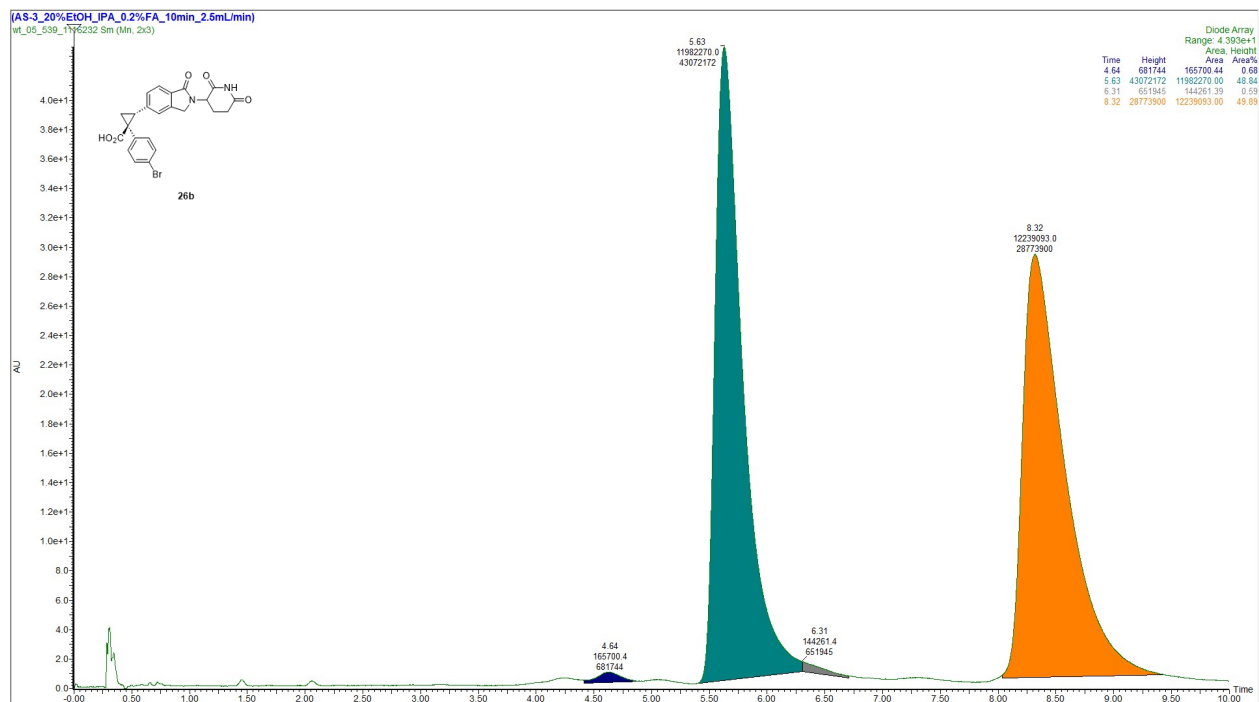
Racemic Chromatogram, 26



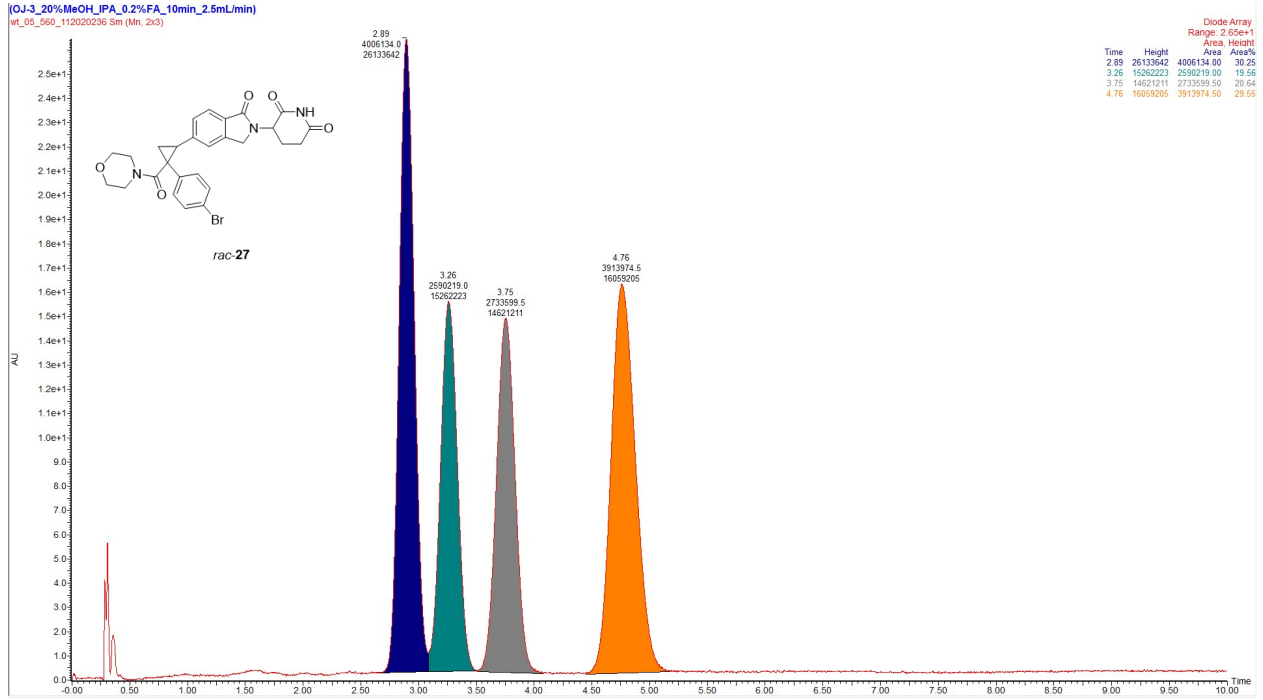
26a



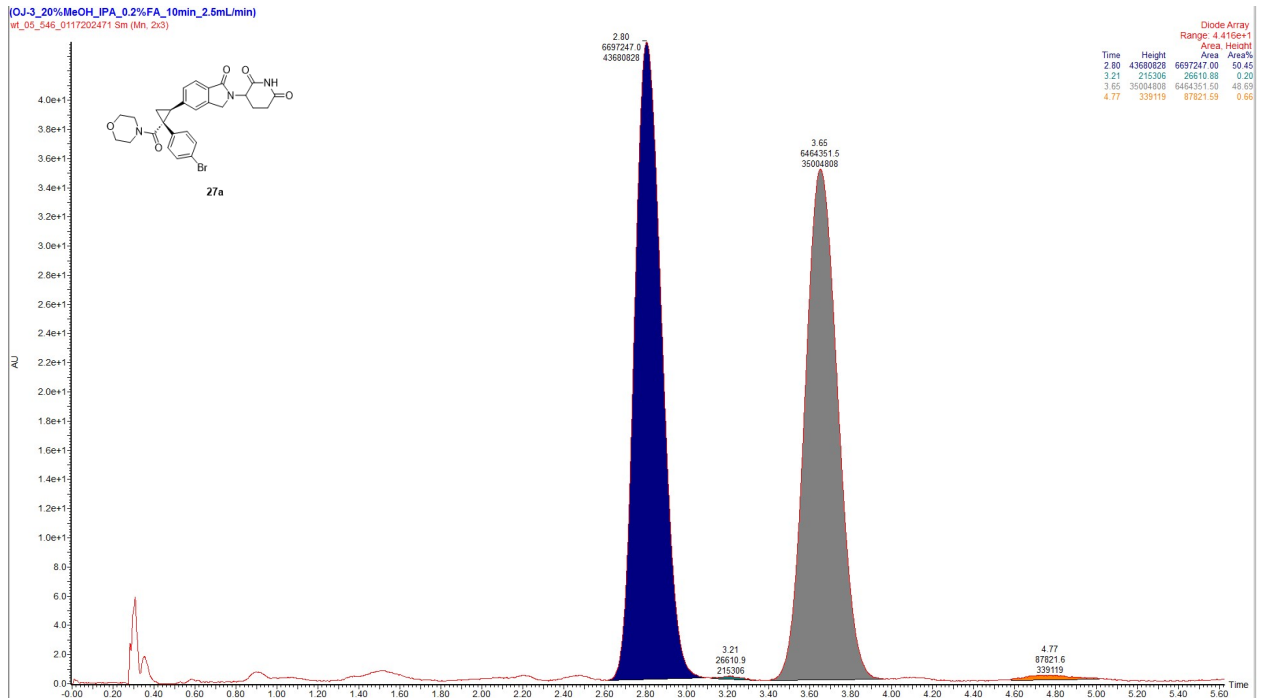
26b



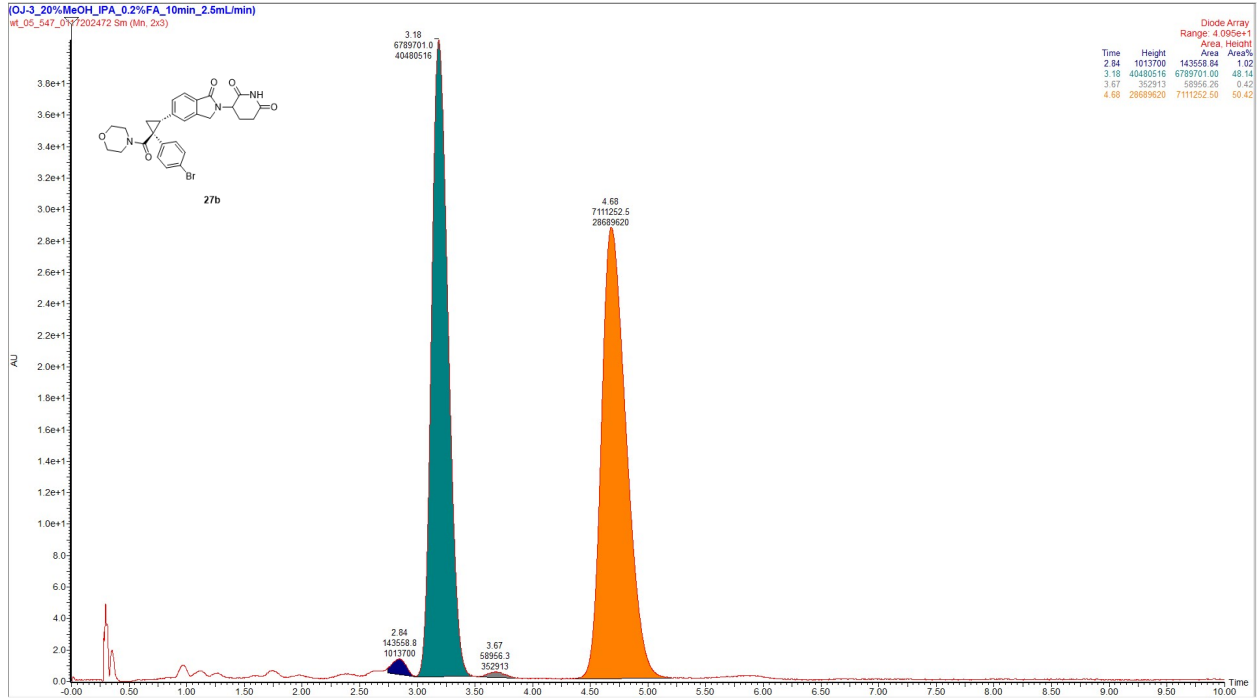
Racemic Chromatogram, 27



27a



27b



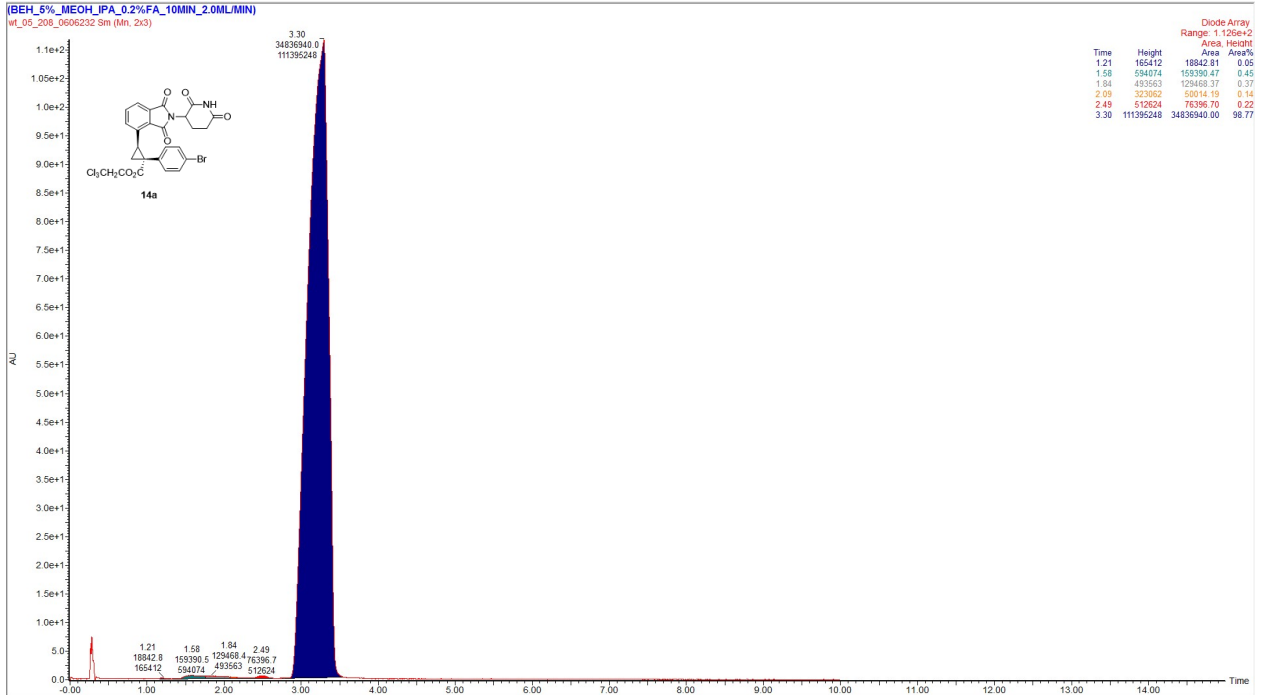
Purity Chromatograms 13a



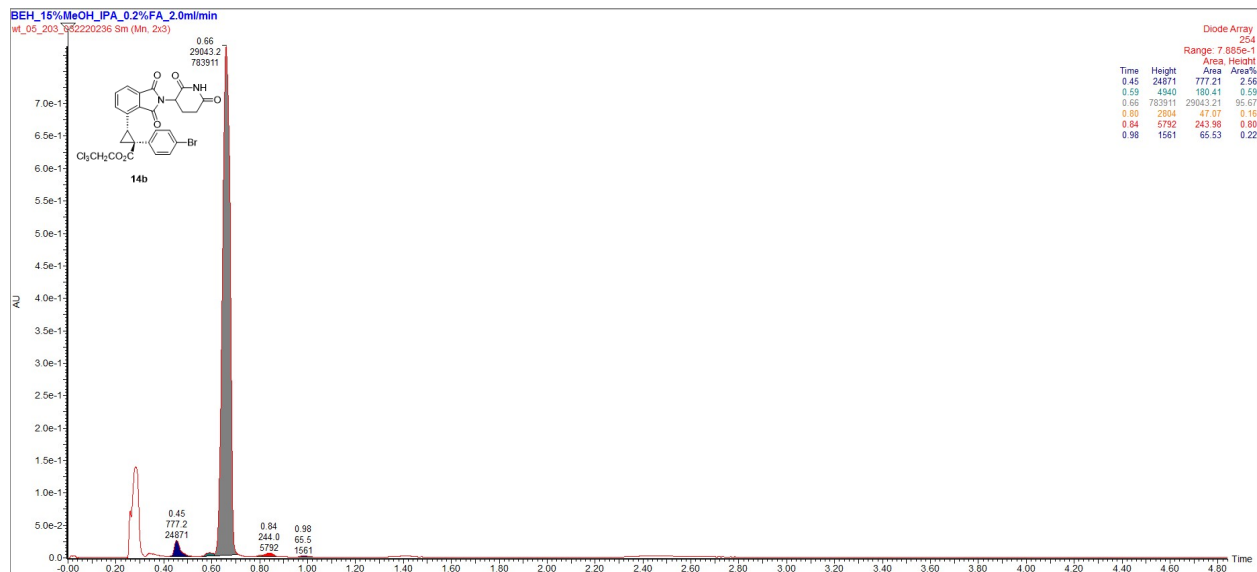
13b



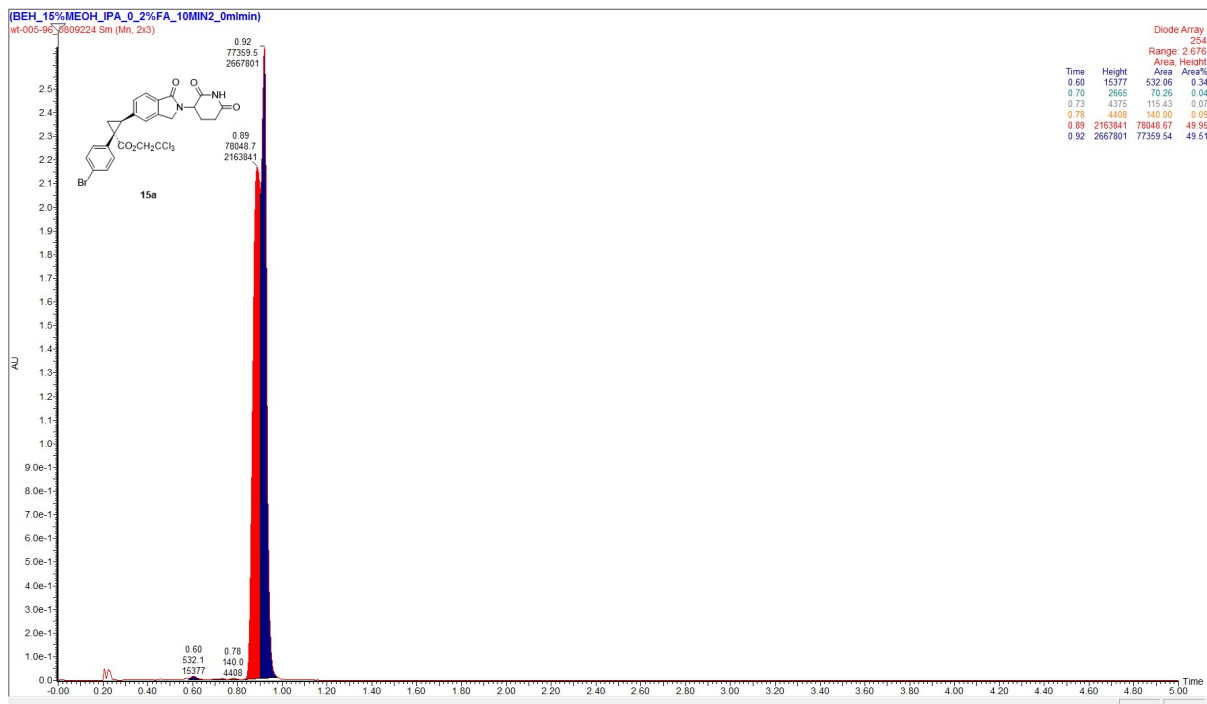
14a



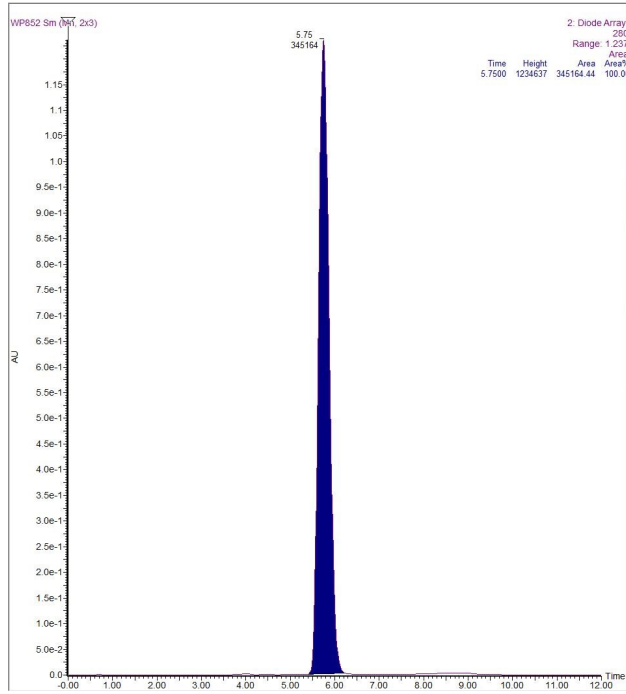
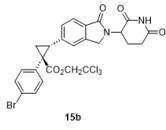
14b



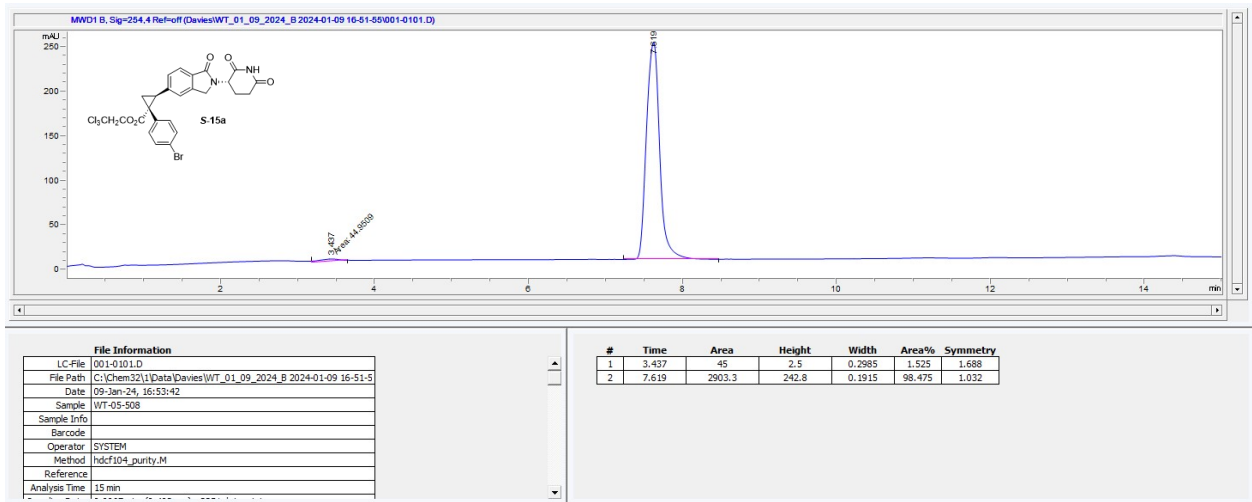
15a



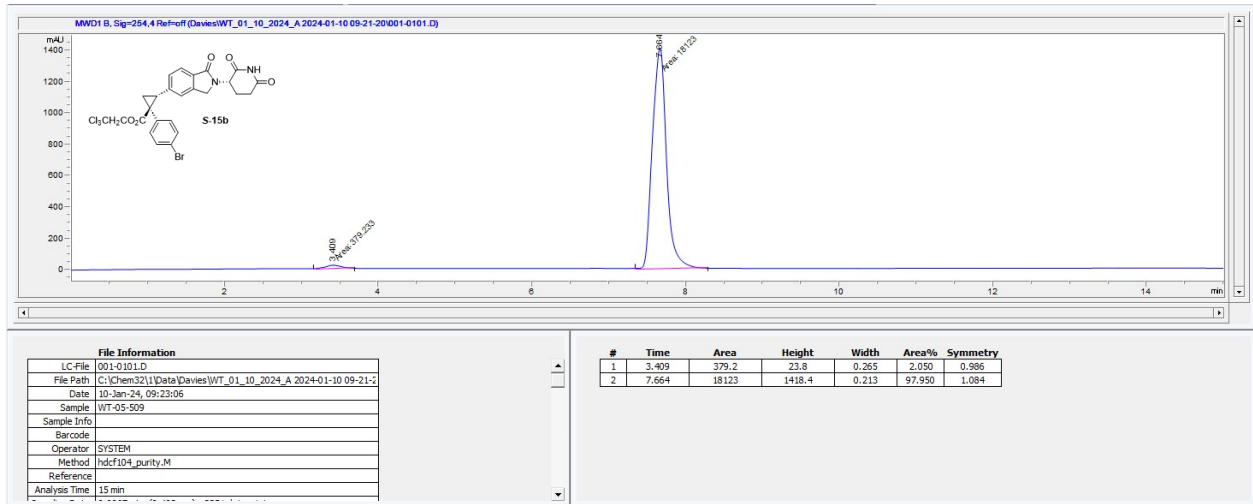
15b



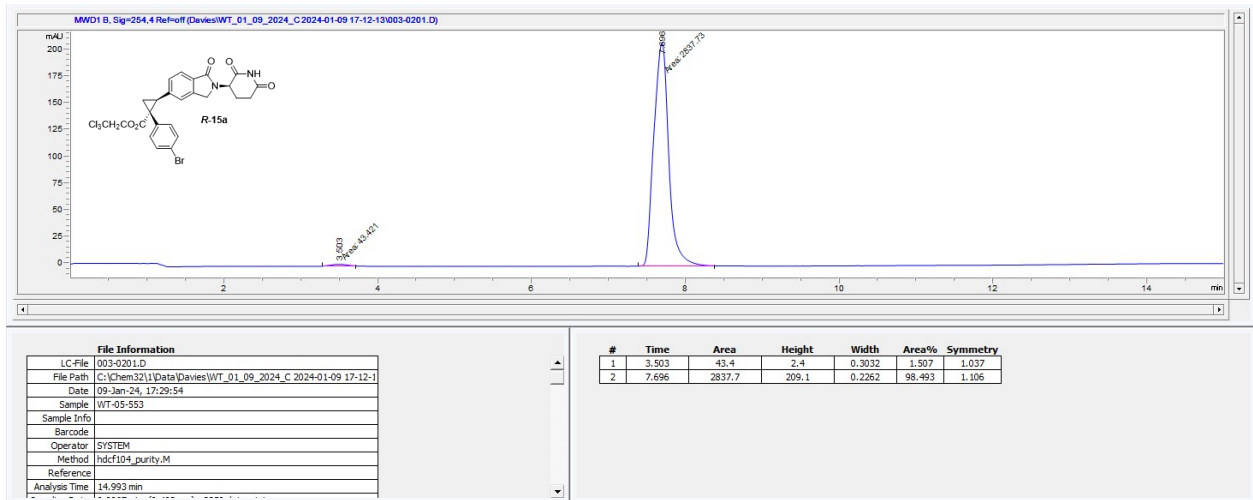
S-15a



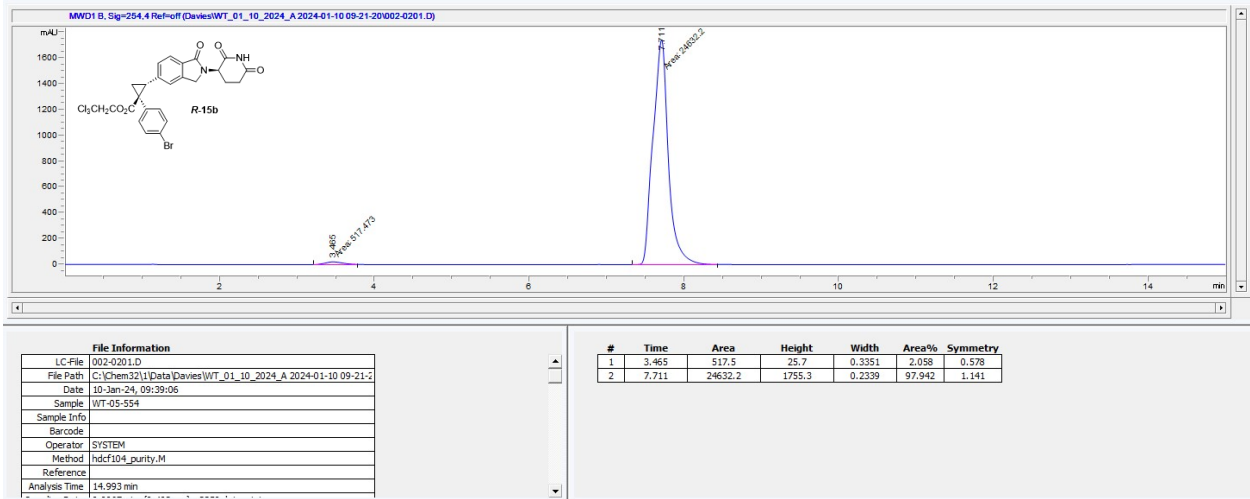
S-15b



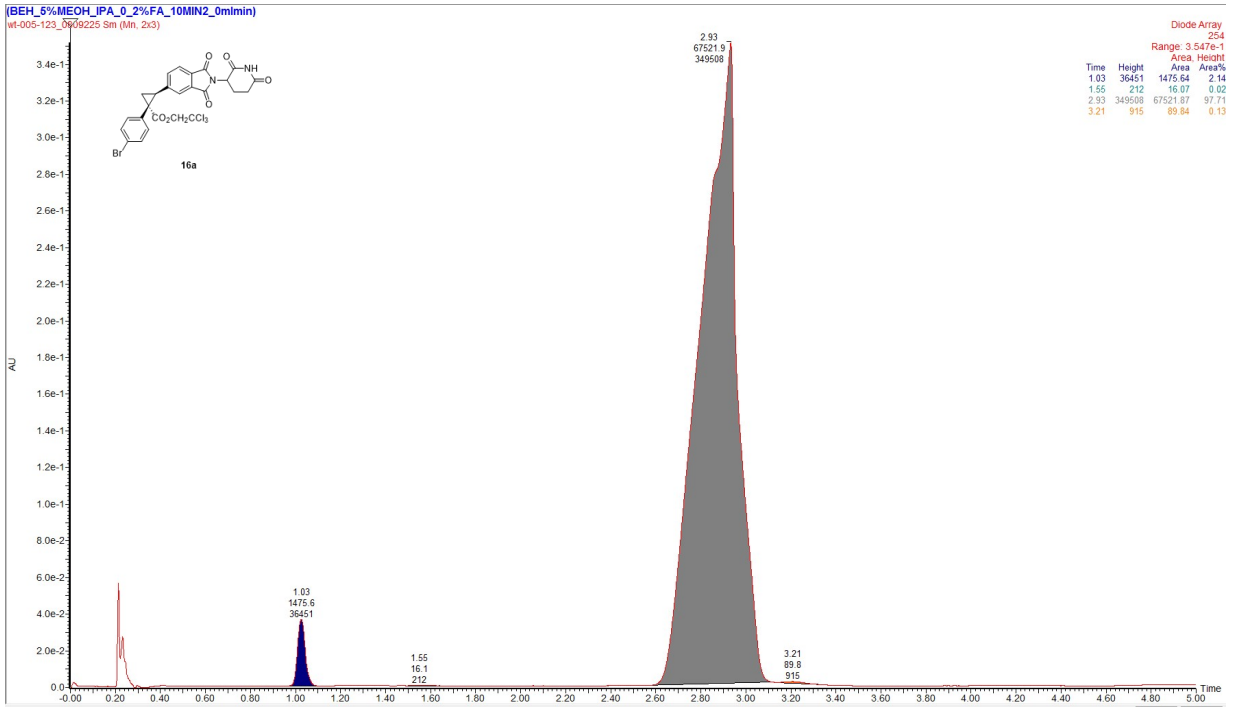
R-15a



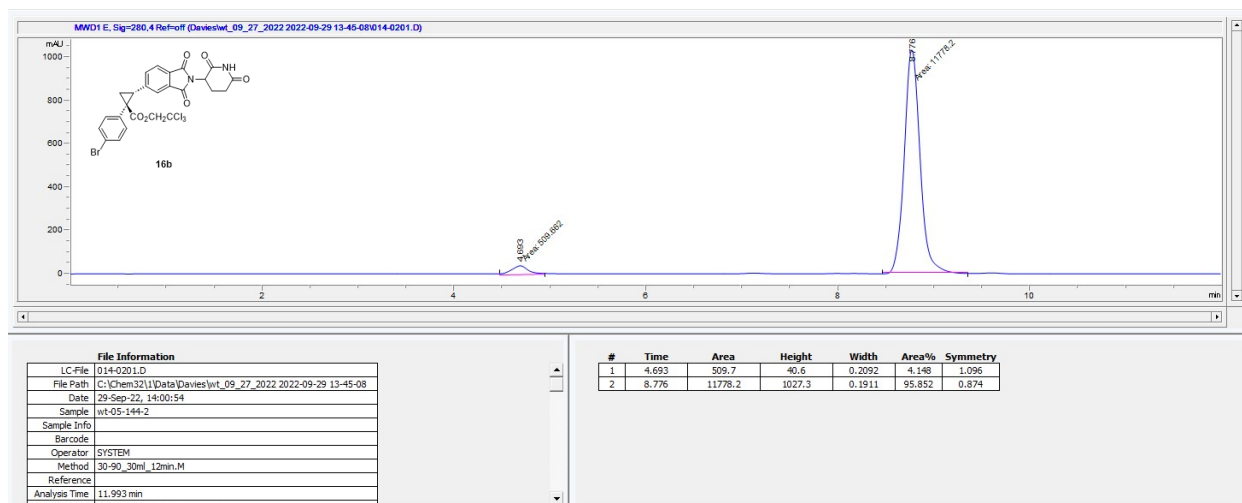
R-15b



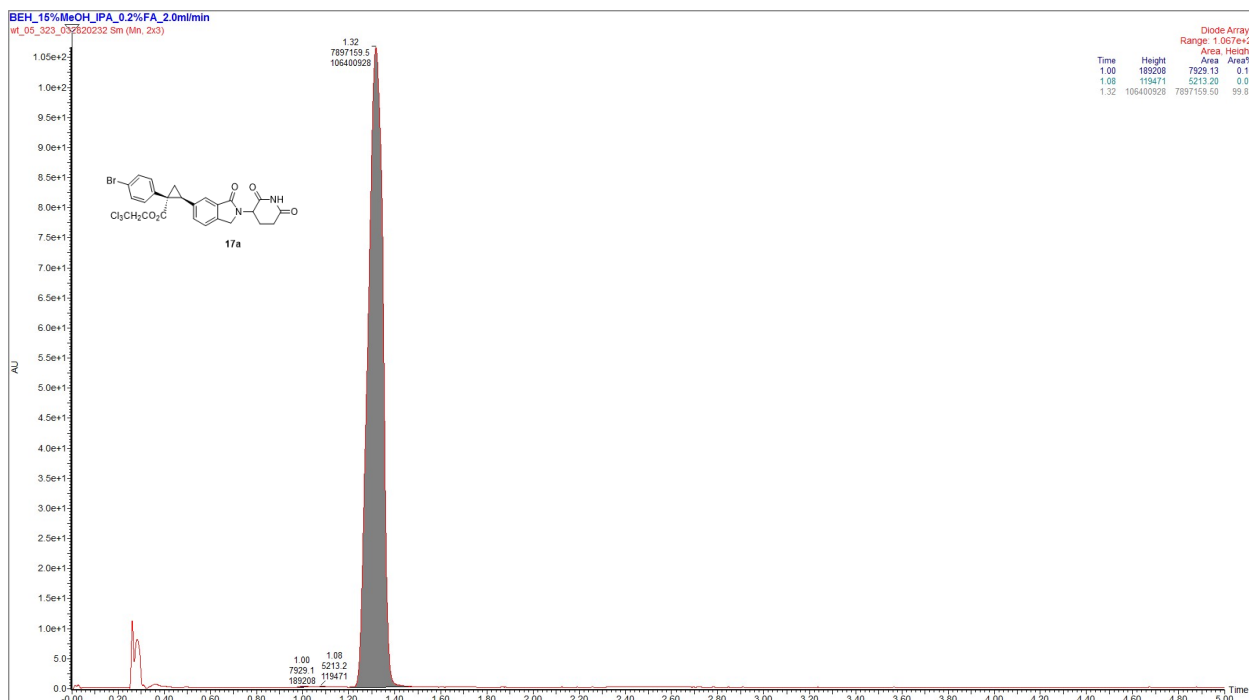
16a



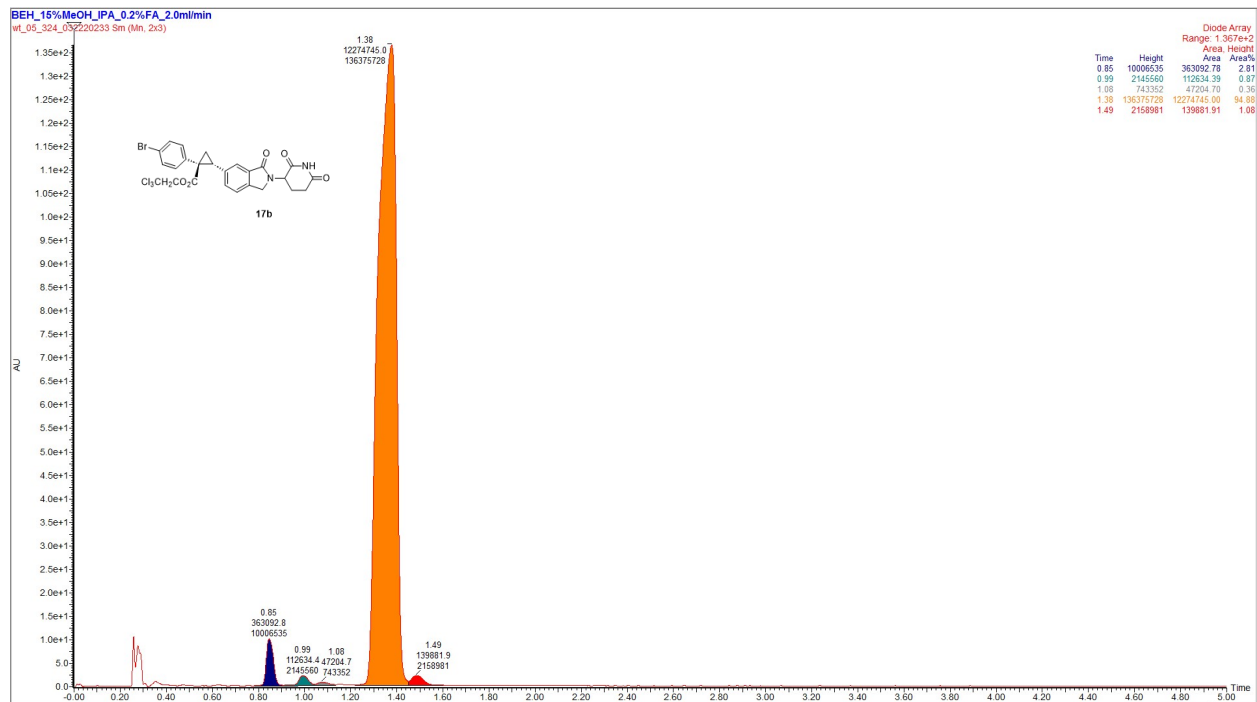
16b



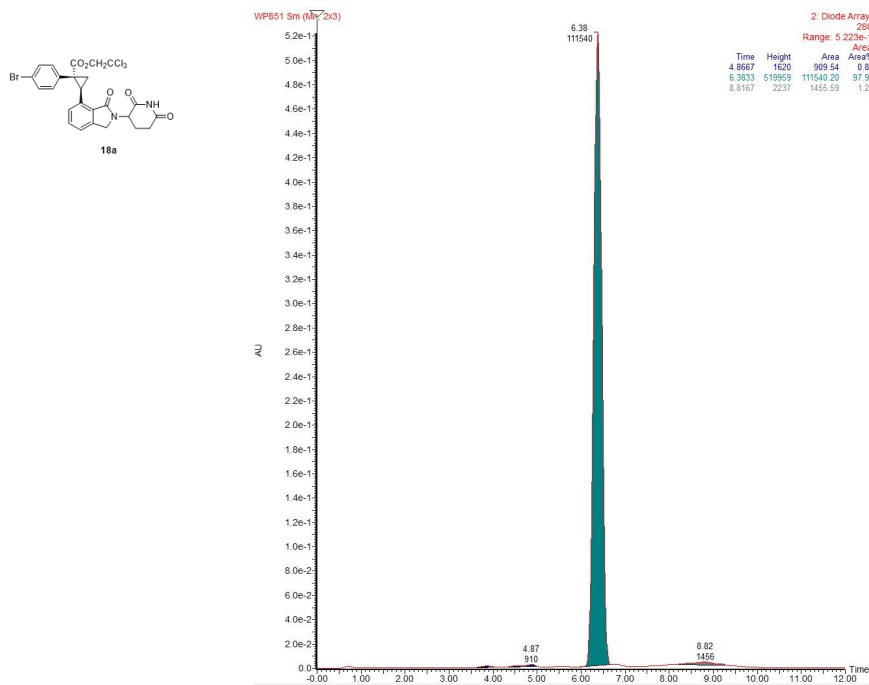
17a



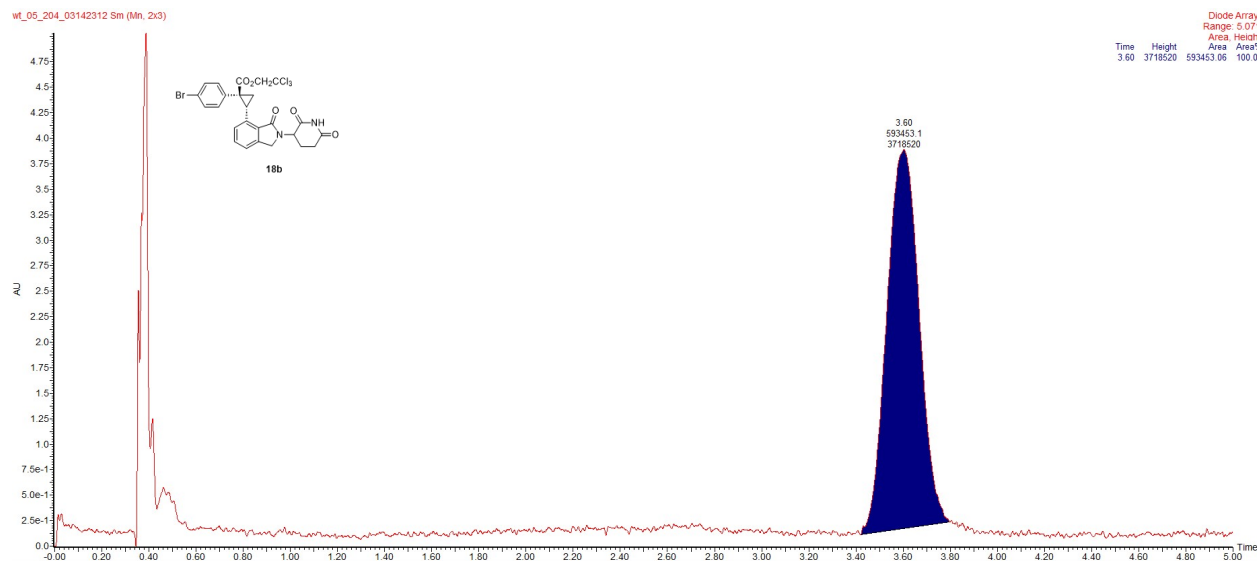
17b



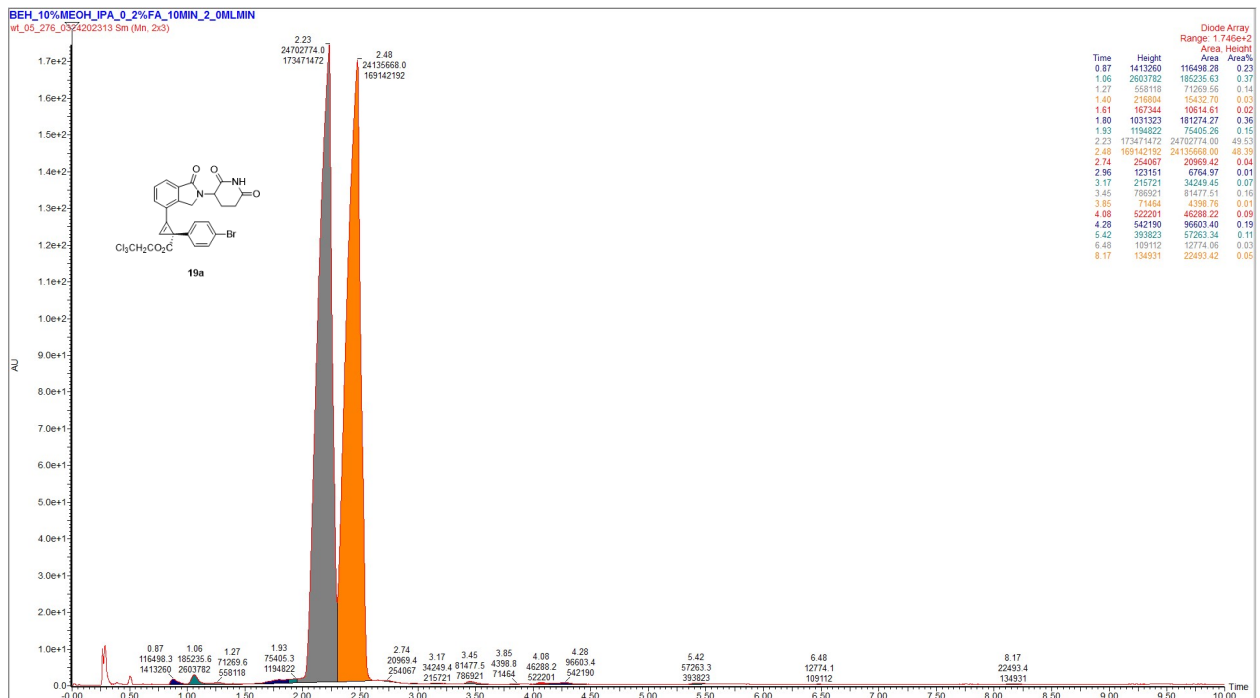
18a



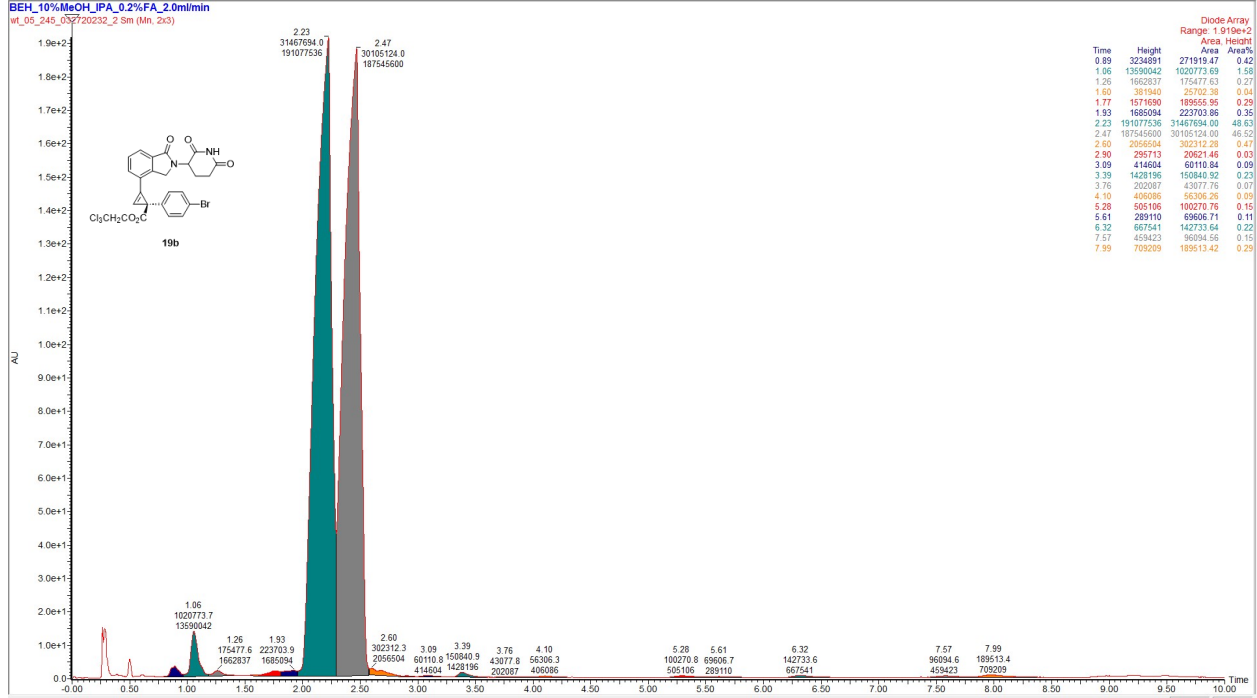
18b



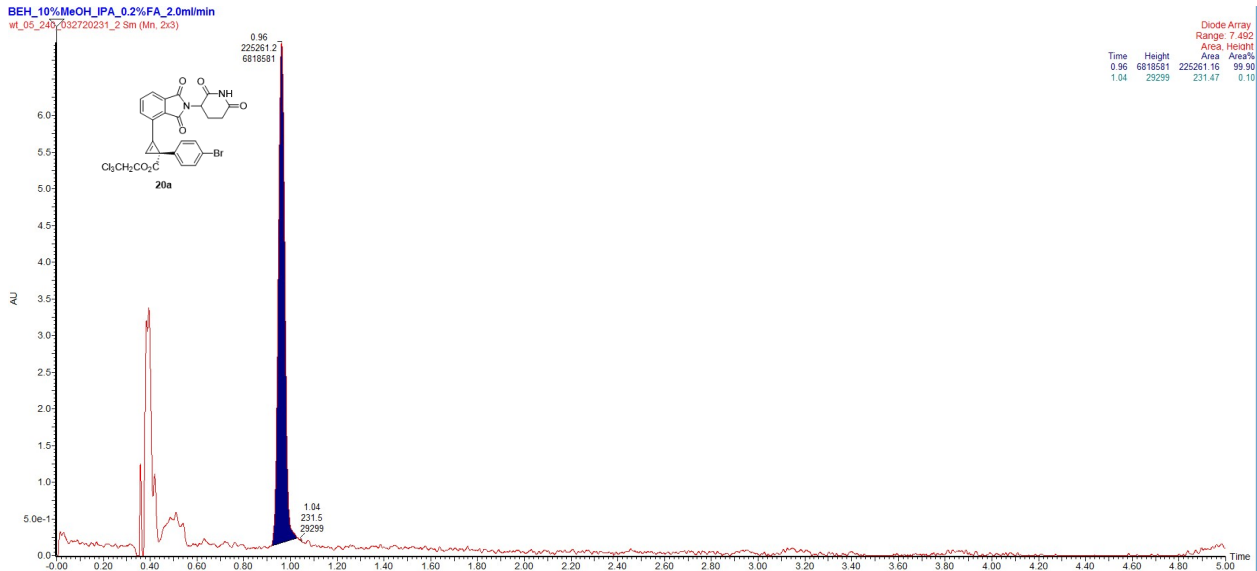
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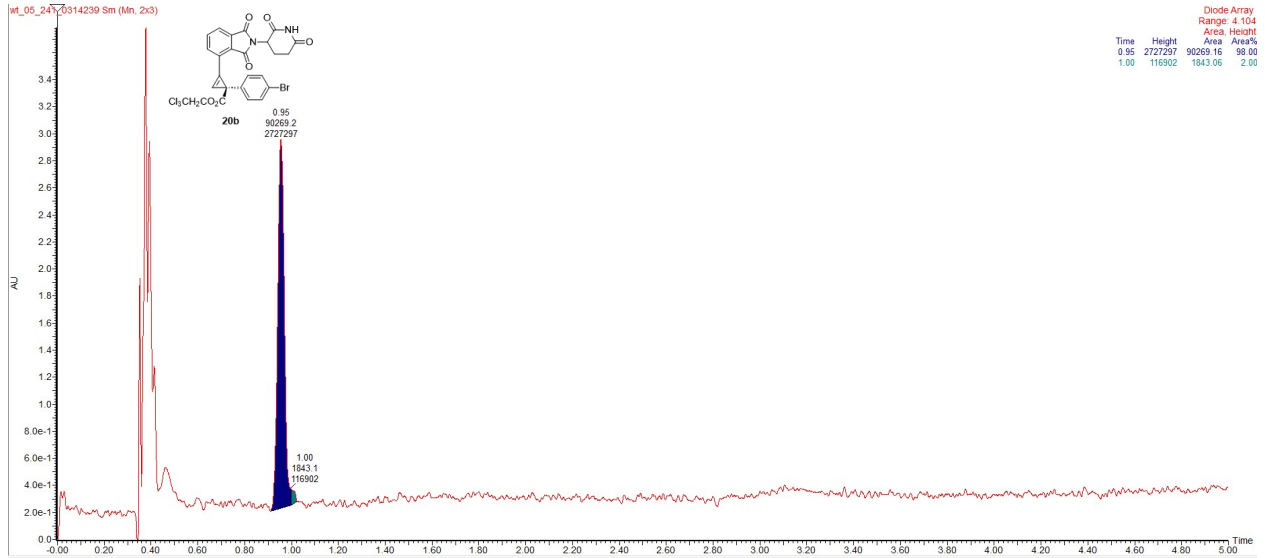
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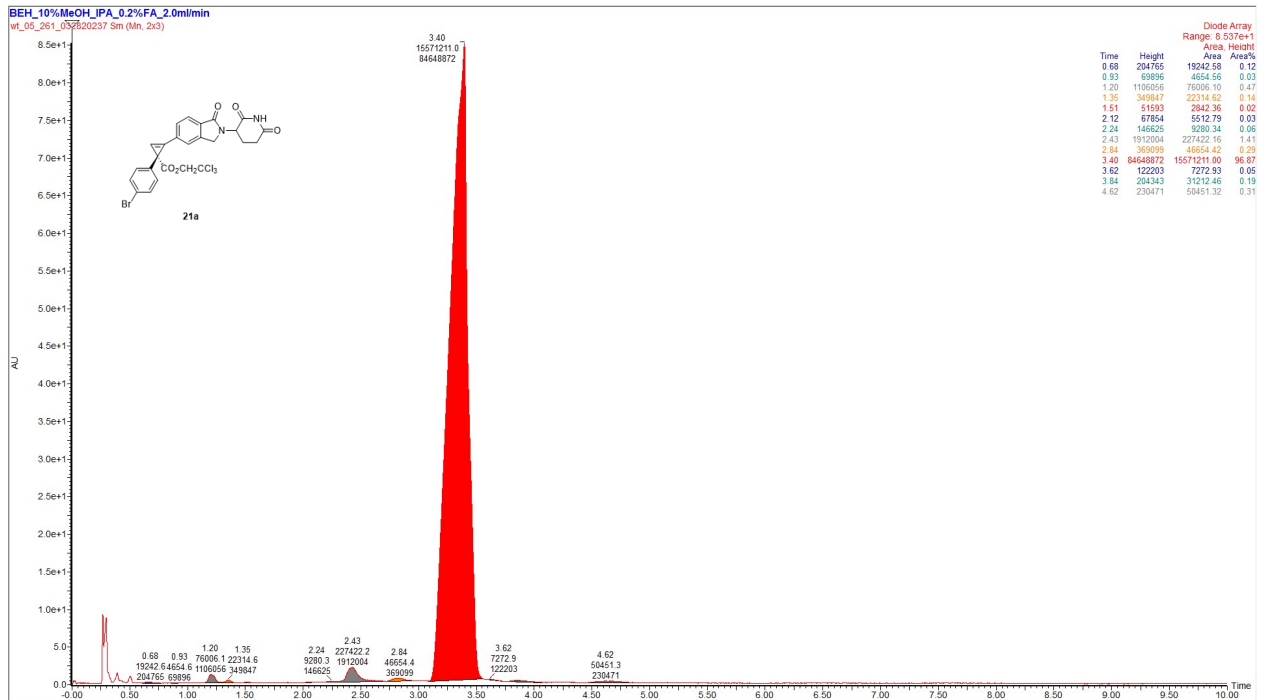
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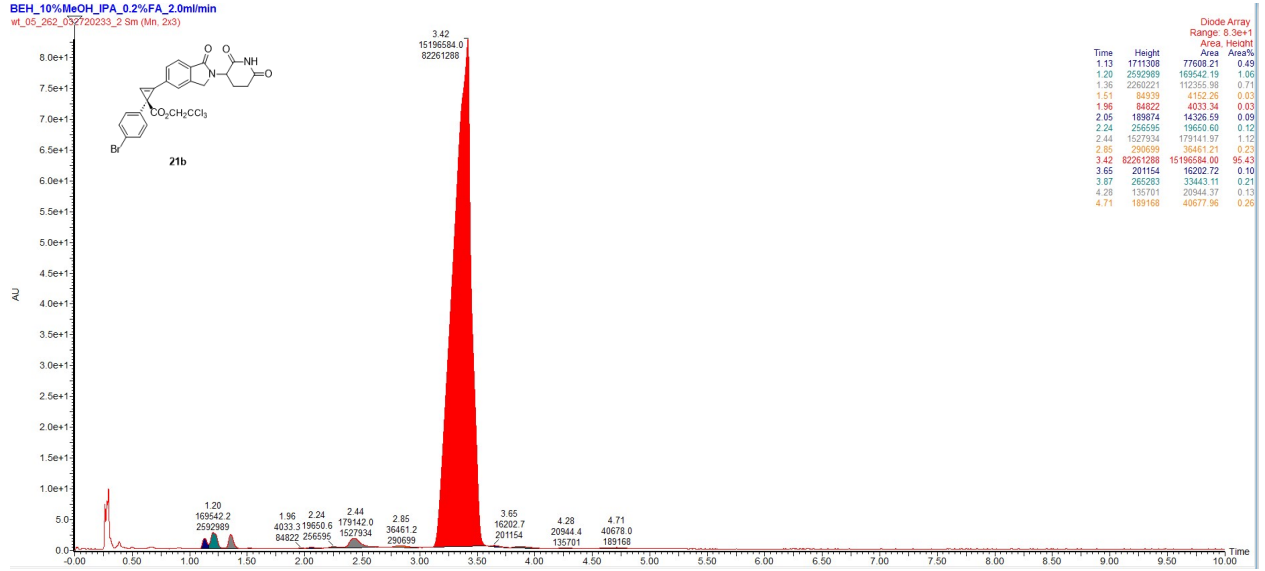
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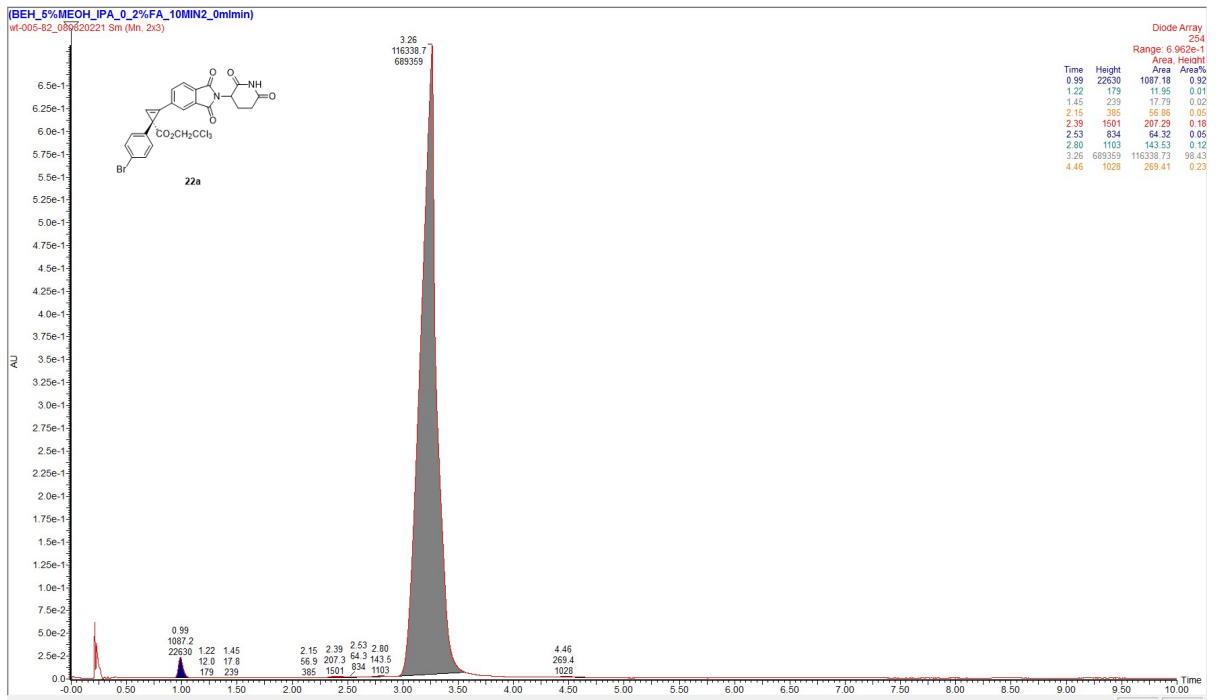
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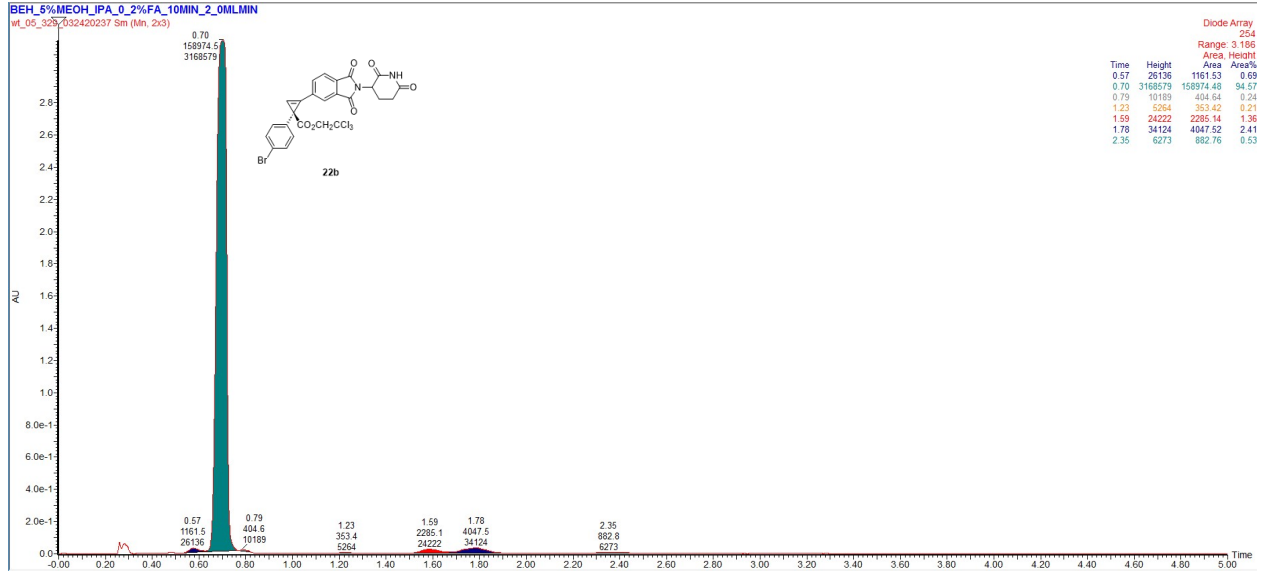
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22a



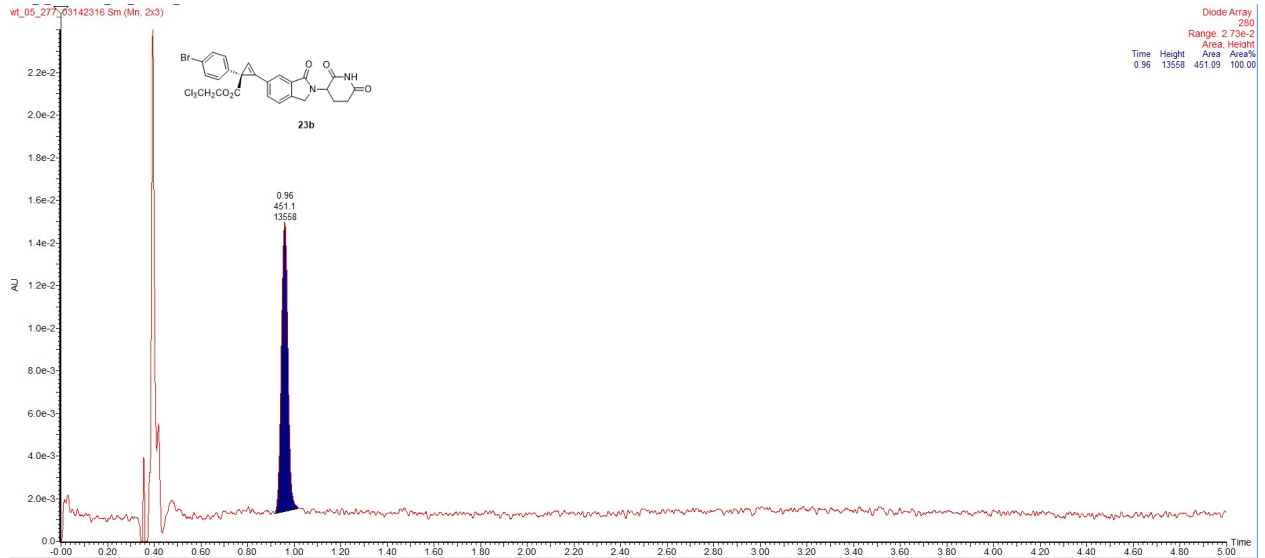
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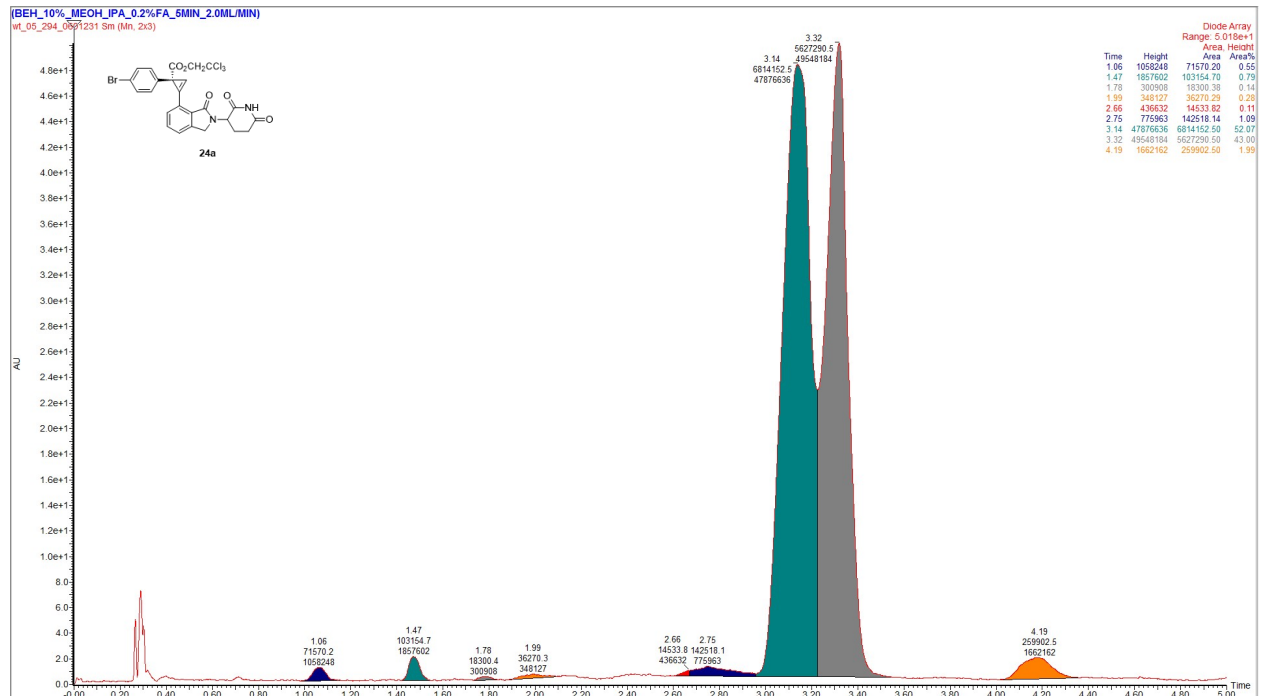
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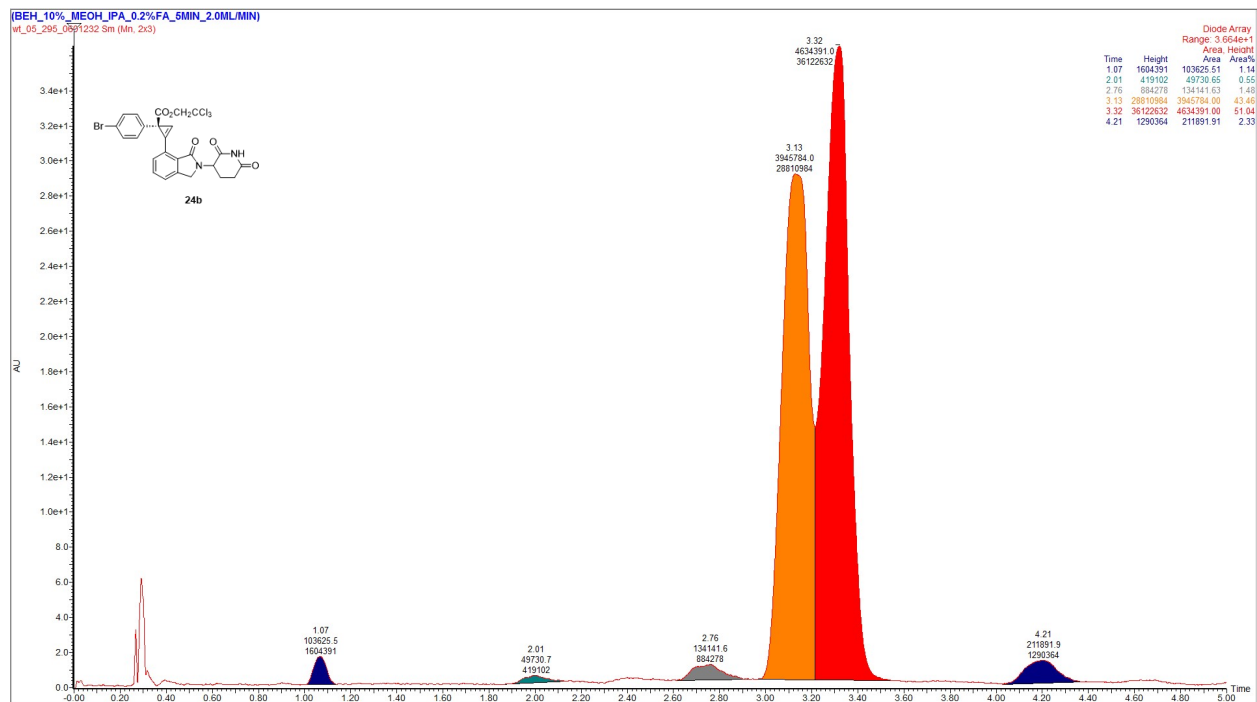
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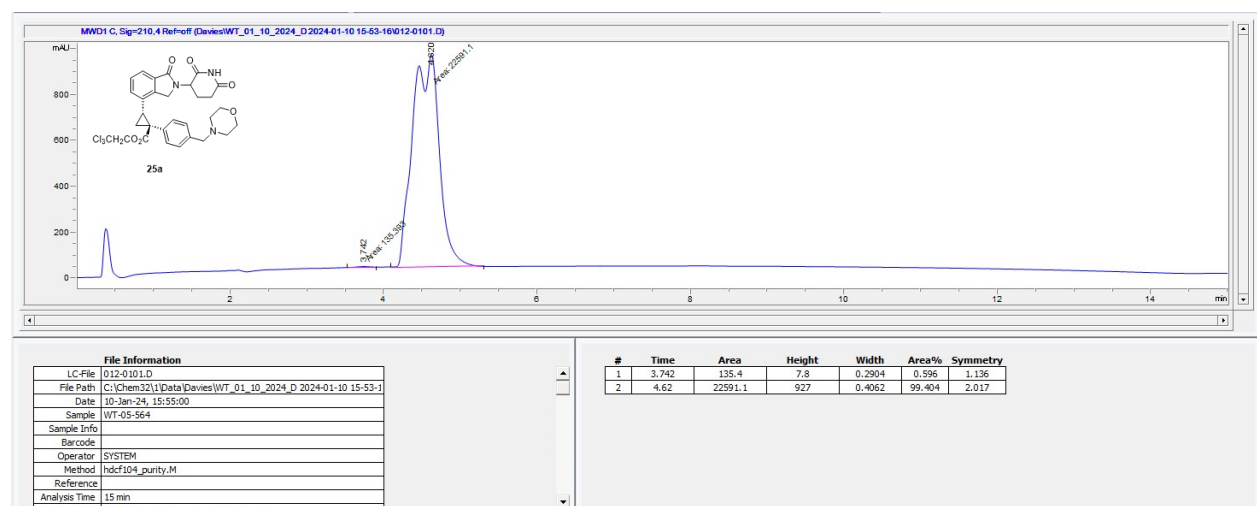
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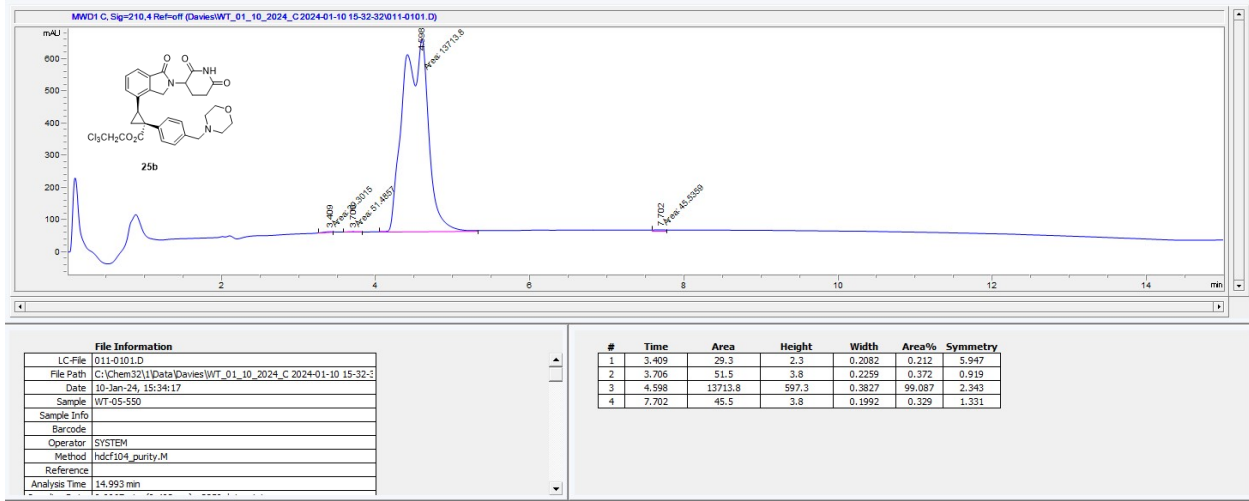
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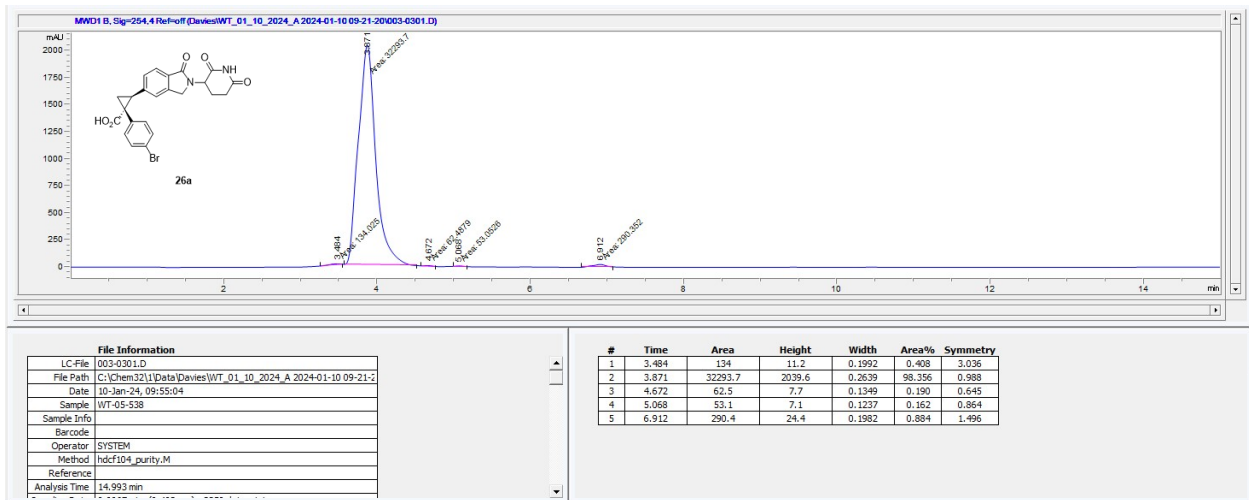
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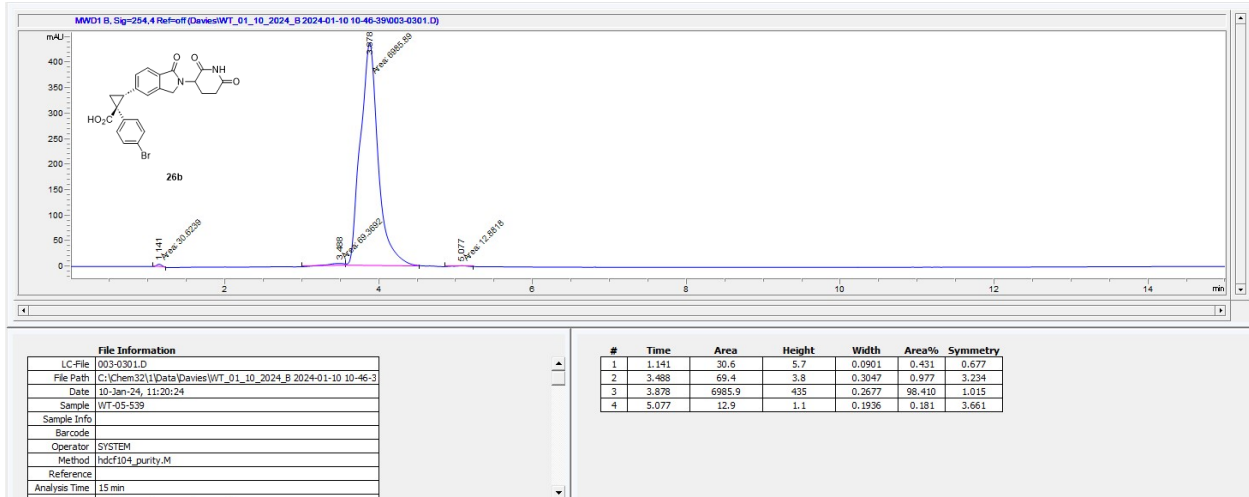
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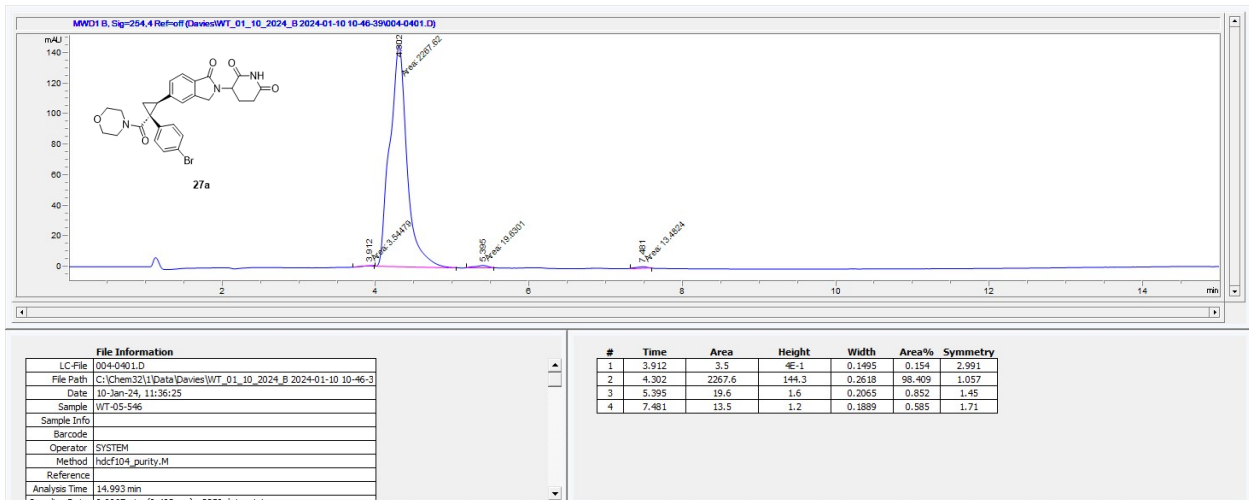
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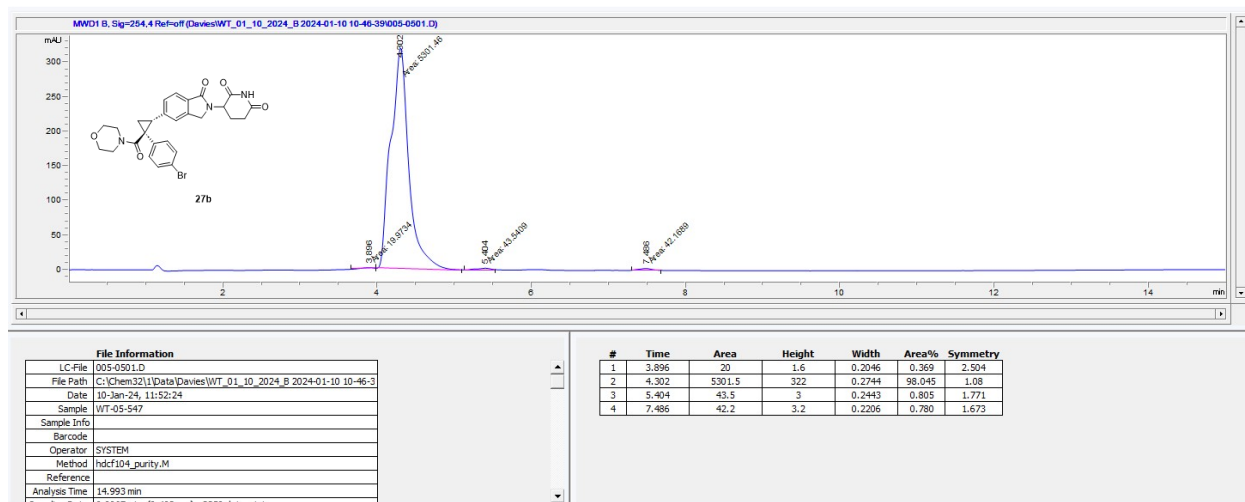
26b



27a



27b



Section 6: Assay Protocols

Fluorescence Resonance Energy Transfer-Based NSD@ PWWP Cereblon Binding Assay.

A solution containing 0.5 nM purified 6×His-CRBN_005-DDB1_026 (CRBN a.a. 1–442, DDB1 a.a. 1–1140, generated in-house) and 20 nM Tracer compound (CC0782985) was premixed in FRET-assay buffer (50 mM HEPES pH 7.3, 50 mM NaCl, 0.005% Brij35, 1mg/mL BSA and 0.5mM TCEP). Compounds for testing were spotted into a 1536 well plate (Greiner Cat#782075). CRBN and Tracer solution were incubated in compound wells for 20 minutes. Detection mix containing Anti6xHis Tb Crytate (CisBio Cat#61HI2TLF) was then added to assay wells for a final 0.5x assay concentration of detection antibody (stock is provided at 400x). Plates were incubated for 30 minutes before being read on Pherastar FSX plate reader using TR-FRET module (Excitation 340nm, Emission 615/665nm).

ePL and HiBiT degradation assays

DF15 cells overexpressing ePL tagged GSPT1 (DF15/GSPT1 ePL OE) was used to monitor the degradation of GSPT1 induced by experimental compounds. MDS-L cells overexpressing ePL tagged CK1 α and GSPT1^{G575N} (MDS-L/CK1 α ePL OE_ GSPT1^{G575N} OE) was used to monitor the degradation of CK1 α induced by experimental compounds. NCCIT cells with HiBiT tag knocked in at the c-terminus of *SALL4* (NCCIT/SALL4-HiBiT KI) was used to monitor the degradation of SALL4 induced by experimental compounds. DF15 cells with HiBiT tag knocked in at the n-terminus of *IKZF3* (DF15/HiBiT-IKZF3 KI) was used to monitor the degradation of AIOLOS (IKZF3) induced by experimental compounds.

The cell culture medium recipes, seeding densities and compound incubation time with cells are shown in the table below:

Target	Cell line	Cell culture medium	Seeding density (cells/well)	Compound incubation time (hr)
GSPT1	DF15/GSPT1 ePL OE	RPMI 1640, 10% heat inactivated (HI) FBS, 1mM sodium pyruvate, 25mM Hepes, 0.1% pluronic acid, 1X NEAA	800	20
CK1 α	MDS-L/CK1 α ePL OE_ GSPT1 ^{G575N} OE	RPMI 1640, 20% HI FBS and 50 ng/mL of human recombinant IL3	1,000	4
SALL4	NCCIT/SALL4-HiBiT KI	RPMI 1640 and 10% HI FBS	800	2
IKZF3	DF15/HiBiT-IKZF3 KI	RPMI 1640, 10% HI FBS, 1mM sodium pyruvate, 25mM Hepes, 0.1% pluronic acid, 1X NEAA	800	4

For all degradation assays, compounds were pre-spotted in a 1536 well plate (Corning 3727) starting at 10 μ M with 3-fold serial dilution down 11 points in replicates using an Echo 650 series acoustic liquid handler. 5 μ L/well cells were seeded in the assay ready plate at the density and in the medium as indicated in the table. The final DMSO concentration in the cell culture was 0.25%. The plates were equilibrated at room temperature for 30 minutes before putting into a 37 °C CO₂ incubator. After appropriate incubation period, plates were retrieved from the incubator and set at room temperature to equilibrate for 30 minutes, before adding ePL or HiBiT detection reagent. For ePL detection, the pre-prepared mixture of EA reagent, lysis buffer, and substrate reagent at ratio 1:1:4 from the DiscoverX InCELL Detection Kit (Eurofins 96-0079L) according to the manufacturer's recommendation was added 3 μ L/well into the plate. Plates were incubated in the dark for 1hr at room temperature before reading using a BMG PheraStar luminescence reader. For HiBiT detection, Nano-Glo HiBiT Lytic Reagent (Promega, N3050) prepared according to the manufacturer's recommendation was added 3 μ L/well into the plate. Plates were incubated 30 minutes at room temperature in the dark before reading using a BMG PheraStar luminescence reader.

To determine EC₅₀ values for degradation, a four parameter logistic model (Sigmoidal Dose-Response Model) ($FIT = (A + ((B - A) / (1 + ((C/x)^D))))$) C is the inflection point (EC₅₀), D is the correlation coefficient, A and B are the low and high limits of the fit respectively) was used to determine the compound's EC₅₀ value, which is the half maximum effective concentration. In the degradation assays, the Yconst of each compound was calculated by normalizing the lowest point of the fitted curve to the media only control, which is 0%, and the cells treated with DMSO control, which is 100%. All degradation curves were processed and evaluated using Dotmatics.

Section 7: Biological Data Tables

Table S7.1: Fluorescence Resonance Energy Transfer-Based Cereblon Binding Assay Data

Compound Number	CRBN HTRF IC ₅₀ (μM)
13a	0.66
13b	2.4
14a	>10
14b	7.9
15a	0.39
15b	1.3
16a	5.3
16b	>10
17a	2.1
17b	2.5
18a	>10
18b	>10
19a	0.16
19b	0.35
20a	4.6
20b	35
21a	0.59
21b	0.8
22a	5.7
22b	7.8
23a	0.98
23b	1.7
24a	2.4
24b	2.9

Table S7.2: Neosubstrate Degradation Assay Data*

Compound Number	IKZF3 EC ₅₀ (μM)	IKZF3 Y _{min} (%)	CK1a EC ₅₀ (μM)	CK1a Y _{min} (%)	GSPT1 EC ₅₀ (μM)	GSPT1 Y _{min} (%)	SALL4 EC ₅₀ (μM)	SALL4 Y _{min} (%)
13a	>10 ± 0.00	81 ± 3.3	>10 ± 0.00	87 ± 0.55	>10 ± 0.00	92 ± 7.8	>10 ± 0.00	85 ± 7.3
13b	>10 ± 0.00	97 ± 4.9	>10 ± 0.00	90 ± 7.1	>10 ± 0.00	99 ± 2.2	>10 ± 0.00	97 ± 3.1
14a	>10 ± 0.00	89 ± 3.2	>10 ± 0.00	92 ± 3.8	>10 ± 0.00	91 ± 16	>10 ± 0.00	91 ± 8.0
14b	>10 ± 0.00	95 ± 4.3	>10 ± 0.00	94 ± 5.1	>10 ± 0.00	87 ± 11	>10 ± 0.00	87 ± 13
15a	>10 ± 0.00	100 ± 4.6	>10 ± 0.00	89 ± 5.1	>10 ± 0.00	96 ± 7.8	>10 ± 0.00	99 ± 1.2
15b	>10 ± 0.00	89 ± 5.0	0.22 ± 0.089	47 ± 5.0	>10 ± 0.00	83 ± 7.0	0.16 ± 0.05	67 ± 5.4
16a	>10 ± 0.00	94 ± 8.3	>10 ± 0.00	94 ± 9.9	>10 ± 0.00	94 ± 11	>10 ± 0.00	92 ± 8.2
16b	>10 ± 0.00	97 ± 3.6	>10 ± 0.00	85 ± 4.1	>10 ± 0.00	86 ± 8.2	0.41 ± 0.16	60 ± 6.1
17a	>10 ± 0.00	95 ± 7.6	>10 ± 0.00	98 ± 12	>10 ± 0.00	95 ± 5.4	>10 ± 0.00	97 ± 2.2
17b	>10 ± 0.00	89 ± 6.5	>10 ± 0.00	96 ± 12	>10 ± 0.00	93 ± 5.8	>10 ± 0.00	96 ± 3.1
18a	>10 ± 0.00	91 ± 0.6	>10 ± 0.00	89 ± 1.7	>10 ± 0.00	87 ± 6.5	>10 ± 0.00	93 ± 3.8

18b	>10 ± 0.00	96 ± 5.7	>10 ± 0.00	91 ± 7.1	>10 ± 0.00	83 ± 15	>10 ± 0.00	96 ± 5.4
19a	0.012 ± 0.010	7.9 ± 1.8	0.17 ± 0.18	64 ± 4.3	0.17 ± 0.13	64 ± 8.7	0.019 ± 0.010	23 ± 4.5
19b	0.11 ± 0.05	69 ± 3.3	1.5 ± 1.1	60 ± 7.4	0.55 ± 0.47	66 ± 5.5	0.17 ± 0.06	35 ± 5.9
20a	0.90 ± 0.18	40 ± 2.2	1.5 ± 0.94	56 ± 14	4.3 ± 1.9	12 ± 13	0.56 ± 0.27	34 ± 7.4
20b	3.7 ± 0.62	10 ± 8.2	2.1 ± 1.3	54 ± 8.9	5.1 ± 1.4	17 ± 13	2.8 ± 1.6	57 ± 7.7
21a	>10 ± 0.00	95 ± 7.6	>10 ± 0.00	89 ± 9.3	>10 ± 0.00	81 ± 17	>10 ± 0.00	89 ± 4.2
21b	4.8 ± 2.6	79 ± 6.4	0.25 ± 0.37	57 ± 8.0	3.3 ± 3.0	7.7 ± 4.7	0.075 ± 0.050	68 ± 2.1
22a	1.3 ± 2.1	73 ± 6.2	1.2 ± 1.2	60 ± 26	1.1 ± 0.69	2.7 ± 0.7	3.6 ± 0.74	67 ± 7.8
22b	0.31 ± 0.11	71 ± 9.7	1.6 ± 0.71	55 ± 39	1.4 ± 0.88	1.8 ± 0.86	2.4 ± 1.7	58 ± 3.2
23a	>10 ± 0.00	94 ± 2.3	>10 ± 0.00	93 ± 10	>10 ± 0.00	82 ± 7.1	>10 ± 0.00	95 ± 6.5
23b	>10 ± 0.00	81 ± 4.6	>10 ± 0.00	95 ± 6.0	>10 ± 0.00	88 ± 10	>10 ± 0.00	91 ± 10
24a	>10 ± 0.00	97 ± 0.58	>10 ± 0.00	96 ± 12	>10 ± 0.00	85 ± 24	>10 ± 0.00	95 ± 5.0
24b	>10 ± 0.00	93 ± 2.5	>10 ± 0.00	93 ± 6.8	>10 ± 0.00	80 ± 4.3	>10 ± 0.00	97 ± 5.2

* N ≥ 3 for all data

Section 8: References

1. Qin, C.; Davies, H. M. L., Role of Sterically Demanding Chiral Dirhodium Catalysts in Site-Selective C–H Functionalization of Activated Primary C–H Bonds. *Journal of the American Chemical Society* **2014**, *136*, 9792-9796.
2. Garlets, Z. J.; Boni, Y. T.; Sharland, J. C.; Kirby, R. P.; Fu, J.; Bacsa, J.; Davies, H. M. L., Design, Synthesis, and Evaluation of Extended C₄-Symmetric Dirhodium Tetracarboxylate Catalysts. *ACS Catalysis* **2022**, *12*, 10841-10848.
3. Tracy, W. F.; Davies, G. H. M.; Grant, L. N.; Ganley, J. M.; Moreno, J.; Cherney, E. C.; Davies, H. M. L., Anhydrous and Stereoretentive Fluoride-Enhanced Suzuki–Miyaura Coupling of Immunomodulatory Imide Drug Derivatives. *The Journal of Organic Chemistry* **2024**.
4. Berlin, M.; Dong, H.; Sherman, D.; Snyder, L.; Wang, J.; Zhang, W. Bifunctional Molecules Containing an E3 Ubiquitin Ligase Binding Moiety Linked to a BCL6 Targeting Moiety. US 2022323547A1 2021.
5. Tortoreto, C.; Rackl, D.; Davies, H. M. L., Metal-Free C–H Functionalization of Alkanes by Aryldiazoacetates. *Organic Letters* **2017**, *19*, 770-773.
6. Tilden, J. A. R.; Lubben, A. T.; Reeksting, S. B.; Kociok-Köhn, G.; Frost, C. G., Pd(II)-Mediated C–H Activation for Cysteine Bioconjugation. *Chemistry – A European Journal* **2022**, *28*, e202104385.