

Supplemental Figure S1. Factors that affect SKN-1gf driven pathogen apathy. (A) Pathogen response in WT animals across various PA14 cultures: 72 hr (37°C for 24 hours, followed by 48 hours at 25°C), 48 hr (37°C for 24 hours, followed by 24 hours at 25°C), and 24 hr at 37°C. (B-D) Apathy response in WT and skn-1gf worms across developmental stages (L4, Day 1 adulthood, and Day 3 adulthood). Apathy response of (E) WT and (F) skn-1gf animals toward non-virulent PA14 AgacA mutants. (G) WT and skn-1gf animals display similar movement speed towards the first chosen diet. (H) 4hr difference in developmental timing between WT and *skn-1gf* animal does not alter pathogen apathy response in *skn-1gf* animals. (I) Changes in the distance between the two diets does not alter the pathogen apathy response in WT and skn-1gf animals. (J) "Forced" choice assay, where skn-1 of mutant animals reach the PA14 line within 30 min like WT animals but remain on PA14, whereas the WT animals move toward the *E. coli* OP50 option over the 8-hour time course. (K) Images of the apathy response of *skn-1gf* worms at the 1hr and 8hr time points reveals ample food throughout the assay. (L-M) *skn-1gf* animals raised on control RNAi (E. coli HT115) until L4 stage still display apathy toward PA14 in the (L) traditional food choice and (M) forced food choice assays. (N-P) daf-2lf (N), eat-2lf (O), and skn-1lf (P) animals do not display pathogen apathy behavior. Each of the food choice assay comprised of N≥3 (with ≥150 worms per biological replicate per strain/condition) and analyzed via two-way ANOVA test; **(p<0.01) ***(p<0.001) ****(p<0.0001). Individual "p" value numbers for the comparisons within each graph is placed above the respective comparison bars in the graph. Source data for all behavioral responses are provided as a Source Data Table 1.



Supplemental Figure S2. Training and neuronal responses of *skn-1gf*. Chemorepulsion away from (**A**) octanol (OCT) and nonanone (NON) is similar between WT and *skn-1gf* mutants but not (**B**) 1-undecene. Pathogen leaving responses in WT (**C-D**) and *skn-1gf* (**E-F**) worms post-pathogen training (0hr, 2hr, and 4hr exposure). (**G**) Pan-neuronal (*rgef-1p*) degradation of SKN-1wt does not alter pathogen leaving behavior. Panneuronal (*rgef-1p*) expression of SKN-1gf (**H**) isoform a or (**I**) isoform c in the neurons does not elicit an apathy response like *skn-1gf* worms. Each of the chemotaxis assay comprised of N≥3 (with ≥150 worms per biological replicate per strain/condition) and analyzed via one-way ANOVA test data is presented as mean values +/- SEM. Each of the food choice assay comprised of N≥3 (with ≥150 worms per biological replicate per strain/condition) and analyzed via two-way ANOVA test; **(p<0.01) ***(p<0.001) ****(p<0.0001). Individual "p" value numbers for the comparisons within each graph is placed above the respective comparison bars in the graph. Source data for all behavioral responses are provided as a Source Data Table 1.



Supplemental Figure S3. Nuclear accumulation of SKN-1 and intestinal bloating in response to PA14. (A-F) Intestinal accumulation and stabilization of SKN-1gf-GFP and SKN-1wt-GFP upon PA14 exposure in comparison to OP50 (time points: 10 mins, 40 mins, and 60 mins); (G-N) Kinetics of loss of intestinal SKN-1 nuclear localization over 4 hours of post PA14 exposure when transferred to OP50 - the first two intestinal nuclei of each worm are outlined by white-dotted circles and the constitutive expression SKN-1 in the ASI neurons is marked with white arrows; quantification of the (O) intensity and the (P) percent of the population. (Q-T) DIC images of intestinal lumen in (Q, R) WT and (S, T) skn-1gf worms post-exposure to pathogen for 24 hours. skn-1wt-GFP and skn-1gf-GFP on OP50 (Q, S) or PA14 (R, T). (U-V) RNAi of aex-5 or egl-8 in WT animals causes enhanced pathogen leaving even in the absence of PA14, however only eql-8 RNAi was capable of marginally inducing leaving behavior of the skn-1gf mutant. (W) Intestinal-specific (ges-1p) degradation of SKN-1wt does not alter pathogen leaving behavior in SKN-1wt mutants. Intestinal-specific (ges-1p) expression of SKN-1gf isoforms a (X) or c (Y) is not sufficient to elicit the pathogen apathy behavior as observed in skn-1qf worms. For the population studies a total of 60 (N=3; n=20) animals were counted per time point per condition. For quantification studies a minimum of three animals per time point per condition was taken into consideration. Each of the nuclear accumulation time points comprised of N≥3 (with n≥3 per biological replicate per strain/condition) and analyzed via two-way ANOVA test; **(p<0.01) ***(p<0.001) ****(p<0.0001). Each of the food choice assay comprised of N≥3 (with ≥150 worms per biological replicate per strain/condition) and analyzed via two-way ANOVA test; **(p<0.01) ***(p<0.001) ****(p<0.0001). Individual "p" value numbers for the comparisons within each graph is placed above the respective comparison bars in the graph. All experiments were conducted at 25°C. Source data for all nuclear activation responses are provided as a Source Data Table 2. Source data for all behavioral responses are provided as a Source Data Table 1.



В

А

Tissue Enrichment Analysis

WT (Up) vs. skn-1gf (Down)

Term	Enrichment	P-value	Term	Enrichment	P-value
germ line	1.6-fold	2.3e-07	amphid socket	2.6-fold	0.0002
reproductive system	1.4-fold	1.6e-05	excratory socket	2.7-fold	0.0024
pharynx	1.4-fold	8.6e-04	phasmid socket	2.4-fold	0.0079

С

Phenotype Enrichment Analysis

WT (Up) vs. skn-1gf (Down)

Term	Enrichment	P-value
eggshell variant	5.4-fold	1.1e-06
shortened lifespan	2.1-fold	0.0013
intestinal cell uptake	4.5-fold	0.0048

D

Gene Ontology (GO) Enrichment Analysis

WT (Up) vs. skn-1gf (Down)

Term	Enrichment	P-value
catabolic process	3.9-fold	3.0e-05
proton transport	6.4-fold	4.8e-05
serine hydrolase	7.5-fold	5.7e-05

WT (Down) vs. skn-1gf (Up)

Term	Enrichment	P-value
amphid socket	2.6-fold	0.0002
excratory socket	2.7-fold	0.0024
phasmid socket	2.4-fold	0.0079

WT (Down) vs. skn-1gf (Up)

Term	Enrichment	P-value
molt	3.8-fold	7.8e-05
ntestinal vacuole	6.6-fold	3.7e-04
pehavior	1.5-fold	0.002

WT (Down) vs. skn-1gf (Up)

Term	Enrichment	P-value
cuticle structure	10-fold	1.1e-13
adherens junction	9-fold	8.3e-05
amino acid biosynthetic function	6.9-fold	2.9e-04

Supplemental Figure S4. Gene set enrichment analysis (GSEA) of context specific transcriptional signature of PA14 exposure. (A) RNAseq of differential transcriptional responses to PA14 exposure in WT and *skn-1gf* animal (WT vs *skn-1gf* up-regulated in blue to the right and down-regulated in red to the left). Gene set enrichment analysis (GSEA) tools in WormBase were used for the RNAseq data in Supplemental Table S3 to identify (B) tissue enrichment, (C) phenotype enrichment, and (D) gene ontology (GO) term enrichment of transcripts that differentially respond to PA14 in WT and *skn-1gf* animals. RNAseq data is available at the NIH (GEO) Gene Expression Omnibus (GSE251677). Source data for all Genotype-specific transcriptional responses are provided as a Source Data Table 3.





Supplemental Figure S5. Pathogen apathy with constitutive SKN-1 is associated exclusively with serotonin. (A) The GO-terms enrichment analysis of serotonin pathway genes that differentially respond to genetic and dietary conditions in WT and *skn-1gf* animal upon exposure to PA14, in comparison to OP50. Supplementation of (B) 5mM 5-HTP, (C) 10-20mM dopamine, (D) or 5-10mM octopamine, or (E) 5-15mM acetylcholine (AcC) does not alleviate the pathogen apathy response in *skn-1gf* animals. (F) Pathogen leaving behavior of *pah-1lf* (*ok687*) is like WT. Quantitative PCR analysis of expression levels of *tph-1* (G) and *pah-1* (H) in WT and *skn-1gf* mutants with and without PA14 exposure. Data is represented as mean values for (N=3; n=3 replicates) (I) Pathogen leaving behavior of *tph-1lf* pah-1lf animals with constitutive SKN-1 activation resulting from *wdr-23* RNAi. Each of the food choice assay comprised of N≥3 (with ≥150 worms per biological replicate per strain/condition) and analyzed via two-way ANOVA test; **(p<0.01) ***(p<0.001) ****(p<0.0001). Individual "p" value numbers for the comparisons within each graph is placed above the respective comparison bars in the graph. Source data for all Genotype-specific transcriptional responses are provided as a Source Data Table 3. Source data for all behavioral responses are provided as a Source Data Table 1.





J





Figure S6. Activation of intestinal SKN-1 in response to PA14 with and without serotonin supplementation. (A) Intestinal accumulation of SKN-1wt-GFP upon PA14 exposure when prior grown on OP50 with and without serotonin supplementation (time point: 10-40 mins). Quantification of the (B) intensity and the (C) percent of the population of nuclear localization over the time course of PA14 exposure. (D) Intestinal accumulation of SKN-1-gf-GFP upon PA14 exposure when prior grown on OP50 with and without serotonin supplementation (time point: 10-40 mins). Quantification of the (E) intensity and the (F) percent of the population with nuclear localization over the time course of PA14 exposure. (G) Measuring the change in intestinal nuclear localization of SKN-1wt-GFP upon removal from PA14 and return to E. coli (time points: 1hr-4hr) with and without serotonin supplementation throughout (prior and post PA14 exposure); Quantification of the (H) intensity and the (I) percent of the population with nuclear localization over the time course. (J) Measuring the change in intestinal nuclear localization of SKN-1gf-GFP upon removal from PA14 and return to E. coli (time points: 1hr-4hr) with and without serotonin supplementation throughout (prior and post PA14 exposure). Quantification of the (K) intensity the (L) percent of the population of nuclear localization over the time course post PA14 exposure. White-dotted circles outline the first two intestinal nuclei of each worm; the constitutive expression in the ASI neurons is marked with white arrows. For the population studies, a total of 60 (N=3; n=20) animals were counted per time point per condition. For quantification studies a minimum of three animals per time point per condition was considered with three biological replicates i.e. N=3 (with n \geq 3 per biological replicate per strain/condition). The data have been analyzed via a two-way ANOVA test: **(p<0.01) ***(p<0.001) ****(p<0.0001). Individual "p" value numbers for the comparisons within each graph is placed above the respective comparison bars in the graph. Source data for all nuclear activation responses are provided as a Source Data Table 2.