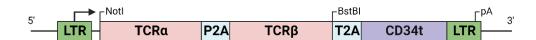
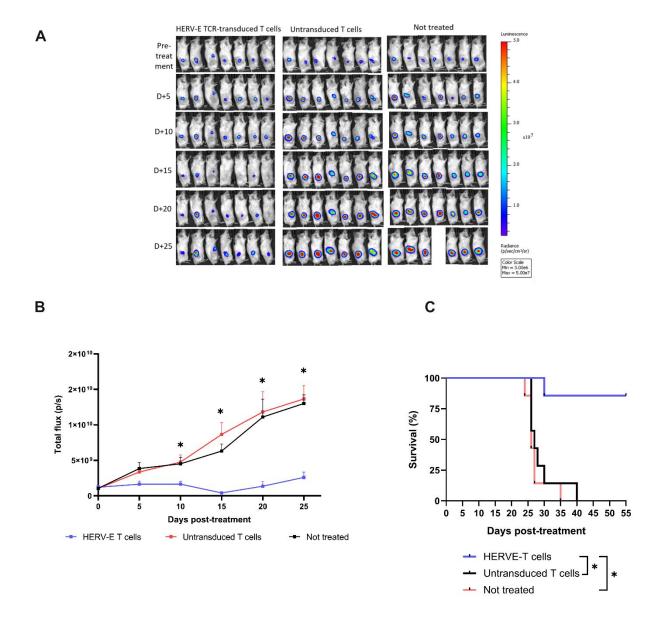
## **Supplementary material:**



**Supplementary item 1. The HERV-E TCR retroviral construct.** SAMEN CMV/SRα retroviral vector encoding HERV-E TCR.



Supplementary item 2. HERV-E T cells mediate regression of ccRCC tumors in vivo in a tumor bearing mouse model. NSG mice with established (10 days) luciferase expressing subcutaneous RCC1 WT tumors were treated with a single intravenous injection of either HERV-E T cells, untransduced T cells from the same donor, or did not receive T cells. Tumor burden was evaluated by serial BLI at the indicated timepoints. (A) Bioluminescence signal shows tumor burden in each treatment group at indicated timepoints after the T cell injection.

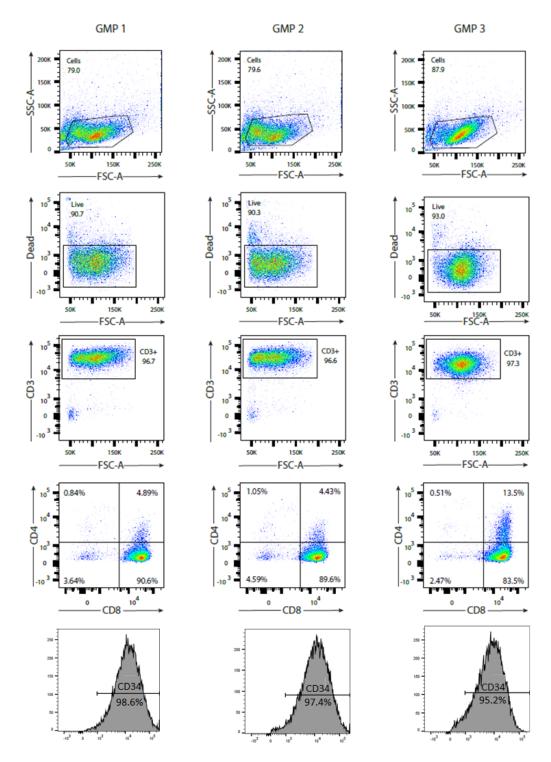
(B) Bioluminescent quantification of ccRCC tumor burden in mice. Error bars represent the SEM

in the group treated with HERV-E T cells (n=7), in the group treated with untransduced T cells (n=7), and in the mice that did not receive T-cells (n=7; Not treated). **(C)** Kaplan-Meyer survival curves. \* p<0.05

Supplementary item 3. Purity and viability of HERV-E TCR transduced T cells manufactured using GMP conditions. HERV-E TCR-transduced T cell products were expanded using GMP conditions at full scale from three healthy donors (GMP 1-3) with cell number, viability, and purity (CD34 expression) assessed at multiple time points after thawing.

		Thawed sample			
		0h	2h	4h	Method
Cell count	GMP 1	1.34 x 10 <sup>8</sup>	1.32 x 10 <sup>8</sup>	1.29 x 10 <sup>8</sup>	
	GMP 2	7.55 x 10 <sup>8</sup>	7.39 x 10 <sup>8</sup>	6.54 x 10 <sup>8</sup>	Cellometer-based
	GMP 3	6.34 x 10 <sup>9</sup>	16.11 x 10 <sup>9</sup>	5.82 x 10 <sup>9</sup>	
Viability (%)*		94.6 ± 2.5	94.3 ± 3.5	92 ± 3	Trypan Blue
CD34+ (%)*		96.5 ± 0.4	n/a	n/a	Flow Cytometry
Tetramer+ (%)*		92 ± 2.2	n/a	n/a	Flow Cytometry

<sup>\*</sup> Data is represented as: Mean ± SD



Supplementary item 4. Scale-up using GMP conditions to manufacture clinical-grade

HERV-E TCR T cells and characteristics of transduced cells. HERV-E T cell final products

were manufactured from three healthy donors (GMP 1-3) using GMP conditions and

characterized by flow cytometry.