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Supplemental Figure 1, in regard to Main Figure 2. **Impact of age, CMV infection and influenza vaccination on the healthy immune landscape. A.** Normalized protein expression (NPX) of inflammatory markers TNF, IL-6, IL-1b and IL-11 over time in young (teal) and older (bronze) adult plasma. **B**. Volcano plot of the age-related expression differences in circulating plasma proteome at baseline (Flu Vax Year 2 Day 0). **C**. Spearman correlation between age-related protein expression difference at Year 1 and Year 2. **D**. The number of down-regulated or up-regulated differential expressed genes (DEGs) from DEseq2 analysis of immune cell subsets from young (n=40) and older (n=44) adults at 'Flu Vax year 2 day 0' using DEseq2 analysis. **E**. Spearman correlation between the number of age-related DEGs in each immune cell subsets in 'Flu Vax year 1 day 0 and 'Flu Vax year 2 day 0'. **F**. The number of DEGs in immune cell subsets ("Flu Vax", Day 0 and Day 7 post-flu vaccination) and No Vax time series ("No Vax", Day 0 vs Day 7 after no vaccination) to that of age. DEGs were defined as log2fc >0.1 and padj<0.05 for all comparisons.

Where:

А

Age Composite Metric_i =
$$\sum_{j=1}^{n} \frac{E_{ij} - \mu_j}{\sigma_j}$$

(Up or Down)

• E_i is the mean pseudo-bulked expression for gene j in sample i
• n is the total number of genes in the set, with n
$$\ge 20$$
.
• μ_{i} is the mean expression of gene j across all samples.
• σ_{i} is the standard deviation of expression of gene j across all samples.
• The Age Composite Metric is calculated from genes that are differently regulated in older
adults comparing with young adults. The 'Up' metric is based on up-regulated genes, and
the 'Down' metric is based on down-regulated genes.

· Age Composite Metric is the composite score for sample i.



Supplemental Figure 2, in regard to Main Figure 2. Maintained, age-related transcriptional signatures in healthy immune cell subsets. A. Equation for calculating the composite age score in each immune cell subset with more than 20 DEGs between young and older adults. B. RNA age metric (upregulated and downregulated genes) in 8 subsets (all subsets with >20 DEGs) comparing young and older adults on year 1 day 0 samples. Each dot is from a single donor. P-value was determined using the Wilcoxon rank-sum test. C. RNA age metric (upregulated genes) in select subsets over time in young and older adults. Each donors' samples are connected with a thin line. D. RNA age metric (downregulated genes) in select

subsets over time in young and older adults. Each donors' samples are connected with a thin line.

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Supplemental Figure 3, in regard to Main Figure 4. Transcriptional landscape of healthy immune cell subsets altered by CMV and age. A. Bubble plot comparison of the change in frequency (using centered log-ratio (CLR) transformation) and number of DEGs between CMV+ (n=136) and CMV- (n=97) adults in our follow-up cohort. Bubble size shows a combined metric of change defined as -log10(p.adj from CLR freq comparison) x DEG_Counts. B. Select subset frequencies in PBMCs shown over time. Teal dots are young adults. Bronze dots are older adults. Regression line shown. C. RNA age metric (upregulated genes) in select T cell subsets split by age and CMV infection status. P-values were calculated using Wilcoxon rank sum test. D-E. The number of DEGs (log2fc >0.1 and p.adj<0.05) in immune cell subsets comparing CMV+ and CMV- individuals, in young and older adults separately from our longitudinal cohort at D. 'Flu Vax year 1 day 0' and E. 'Flu Vax year 2 day 0'.



Supplemental Figure 4, in regard to Main Figure 5. **Age-associated responses of non-naive B cell subsets to flu vaccination. A**. Correlation plots of RNA-quantified (y-axis) and flow cytometry-quantified (x-axis) level 3 B cell population (individual plots) frequencies of total live PBMC. Data shown for 6 young and 6 older adult subjects in flu year 1 that were represented in both scRNA-seq and cytometry analyses. P-value and r values of the Pearson correlation are displayed. B-D. Sample level enrichment analysis (SLEA) scores for the top pathways in the Hallmark database at each timepoint for **B**. plasma cells, **C**. core memory B cells and **D**. CD27+ effector B cells in young and older adults.

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Supplemental Figure 5, in regard to Main Figure 6. **Age-related transcriptional alterations in CM CD4 T cells independent from circulating cytokine signatures. A.** The CLR transformed frequency of ICOS+ CD38+ Tfh cells determined by spectral flow cytometry, at day 0 and day 7 post-influenza vaccination, comparing responses in young and older adults. P-values were calculated using Wilcoxon's signed-rank test (paired) for the comparison between Day 0 and Day 7, and using the Wilcoxon rank-sum test for all other comparisons. B. Tfh activity score was

determined by NMF projection in CM CD4 T cells in young and older adults. **C.** Expression of leading-edge genes in Tfh activity score in young and older adults. **D.** CXCR5 expression in CM CD4 T cells across age in our follow-up cohort. **E.** Receptor-ligand interaction prediction between CM CD4 T cells and core memory B cells in young (n=40) and older (n=44) adults from a single time point (Flu Year 2 Day 0). **F.** CD40LG expression in CM CD4 T cells in young and older adults. **G.** CD40 expression in core memory B cells in young and older adults. **H.** TEAseq UMAP of T cells based on RNA module, with CM CD4 T cells highlighted. **I.** IRF4 and STAT6 transcription factor (TF) activity based on Chromvar analysis of scATACseq data in CM CD4 T cells from children (n=8) and older adults (n=8). P-value was determined by Wilcoxon rank-sum or Wilcoxon signed-rank test, as appropriate, unless otherwise indicated in legend. **J.** Chromatin accessibility tracks of the *IL4R* gene region in CM CD4 T cell subsets from TEA-seq data, showing normalized read coverage. **K.** IL-4 normalized protein expression (NPX) in young (teal) and older (bronze) adult plasma. **L.** Normalized protein expression (NPX) of select Th1-, Th2- and Th17-related serum proteins in our follow-up cohort, with donors ordered by age.