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1723 Supplementary Information

1724 Supplementary Figures

Figure S1 pi5k- cells display cytokinesis and development defects. (a-b) Verification of CRISPR-mediated disruption of PI5K. (a) Genomic PCR sequence analysis confirmed mutations on the PI5K coding genes of PI5K mutant clone. Sequences of WT (Sbjct) and mutation clone (Query) are presented. 2 bp deletions are highlighted in the green box, and induced stop Condon TAA is highlighted with a red box. (b) Representative live-cell time-lapse confocal images (DIC) of CRISPR-mediated pi5k- fan-shaped (left), and pi5k- oscillatory cells (right). (c) Images of DIC channel and nuclear staining by Hoescht (merged) reveal a big increase in number of nuclei in each cell in *pi5k*- cells. (d) Histogram guantification of normalized cell number at different nuclei numbers for (*pi5k*-) PI5K and *pi5k*- after 65h in suspension. (e) WT (top) and *pi5k*⁻ cells (bottom) were plated on development buffer (DB) medium agar for starvation at 6h, 18h, and 24h. WT cells aggregate normally, while pi5k- cells fail to aggregate. (f-h) Representative live-cell time-lapse confocal images of *Dictvostelium pi5k*- oscillatory cells expressing PHcrac-RFP (biosensor for PIP3) (f) or LimE-mCherry (biosensor for F-actin polymerization) (g) or CynA-GFP (biosensor for PI(3,4)P2) (h). Scale bars represent 5 mm.



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Figure S2 pi5k- cells display less Myosin and PI(3,4)P2 activities (a-b) Representative live-cell time-lapse confocal images of Dictyostelium AX2 (WT) cells (left), and pi5k- cells (right) expressing PHcrac-RFP (biosensor for PIP3) (a) or LimE-mCherry (biosensor for F-actin polymerization) (b). Scale bars represent 5 μm. (c-d) Box-and-whisker plots of (c) PHcrac patch length/Cell Perimeter, (d) LimE patch length/Cell Perimeter. nc=22 from at least 3 independent experiments; asterisks indicate significant difference, ****P ≤ 0.0001 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (e-f) Representative live-cell time-lapse confocal images of Dictyostelium AX2 (WT) cells (left), and pi5k- cells (right) expressing Myosin II-GFP (e) or CynA-GFP (biosensor for PI(3,4)P2) (f). Scale bars represent 5 μ m. (g) Box-and-whisker plot of aspect ratio corresponds to Figure 10, 488 nm OFF or 488 nm ON. nc=10 from at least 3 independent experiments: 'ns' indicates non-significant difference, ns denotes P>0.05 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (h-i) Representative membrane kymograph of LimE intensity in AX2(WT) cells (h) and *pi5k*- cells (i) respectively. A linear color map shows that blue has the lowest LimE or Myosin II intensity, whereas yellow has the highest. (i-k) Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 (WT) cells (i), or pi5k- cells (k) expressing Myosin II-GFP and LimE-mCherry (biosensor for F-actin polymerization). Cells show ventral wave propagation in the substrate-attached surface of the cell in (k). Scale bars represent 5 µm.

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Figure S3 Globally recruiting Inp54p in neutrophil and macrophage induced fan-shaped phenotype (a) Time-lapse confocal images of differentiated HL-60 macrophage expressing CRY2PHR-mCherry-Inp54p (magenta; upper panel) and LifeAct-miRFP703 (cyan; lower panel), before or after 488 nm laser was switched on globally. Time in min:sec format. Scale bars represent 5 µm. (b-d) Box-and-whisker plots of (b) cell area, (c) cell migration speed, and (d) aspect ratio correspond to (a), 488 nm OFF or 488 nm ON. n_c=10 from at least 3 independent experiments; **** $P \le 0.0001$, 'ns' indicates non-significant difference, ns denotes P>0.05 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (e) Time-lapse confocal images of differentiated HL-60 macrophage expressing CIBN-CAAX, empty vector CRY2PHR-mCherry-CTRL (magenta; upper panel) and LifeAct-miRFP703 (cyan; lower panel), before or after 488 nm laser was switched on globally. Time in min:sec format. Scale bars represent 5 µm. (f-i) Centroid tracks of differentiated HL-60 neutrophils (f-g) or macrophage (h-i) (n_c=10) showing random motility at 488 nm OFF (f, h), or 488 nm ON (g, i). Each track lasted at least 5 minutes and was reset to the same origin. (j-k) Representative kymograph of cortical LifeAct intensity in Inp54p-expressing neutrophil (j) or macrophage (k) before or after 488 nm laser was turned on. A linear color map shows that blue has the lowest LifeAct intensity, whereas yellow has the highest. Duration of the kymograph is 5 mins. (I-m) color-coded temporal overlay profiles of differentiated HL-60 neutrophil (I) and macrophage (m) expressing CRY2PHR-mCherry-Inp54p. Square brackets indicate the range of recruitment.



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Figure S4 PI5K displays dynamic partitioning upon cAMP global stimulation (a) Representative live-cell time-lapse images of Dictyostelium cells coexpressing PI5K-mCherry and RBD-GFP during ventral wave propagation, showing PI5K dynamically localizes to the back-state regions in ventral waves. Line-scan intensity profiles are shown in the bottommost panels. Red line and green line represent PI5K and RBD, respectively. Time in min:sec format. Scale bars represent 5 µm. (b-e) Representative live-cell images of Dictyostelium cells co-expressing PI5K-GFP and PHcrac-RFP (b) or PHPLCδ–GFP and PHcrac-RFP (d) upon global cAMP stimulation, demonstrating that upon transient global activation of cAR1 receptors. PHcrac gets uniformly recruited to membrane whereas PI5K and PHPLCo remained steadily membrane-bound throughout the entire time course of the experiment. PHPLC δ had about 5% response. At time t=53s or 78s, 1 µM (final concentration) cAMP was added. Time series plot of normalized cytosolic intensities of PI5K and PHCrac (c) or PHPLCo and PHcrac (e), showing the kinetics of the response upon global stimulation with cAMP In all these figures, vertical dashed lines are used to indicate the time of stimulation. Mean \pm SEM are shown for n_c=18 cells. (f) Schematic representation of PI5K and the derived truncations. (g) Representative live-cell time-lapse images of Dictyostelium cells expressing PI5K-GFP (301-718aa) during migration showing PI5K (301-718aa) dynamically localizes at the trailing edge in migrating cells. Time in min:sec format. Scale bars represent 5 µm. (h) Time-lapse confocal images of Dictyostelium cells expressing mRFPmars-SspBR73Q-PI5K(316-718aa), before or after 488 nm laser was switched on globally. Time in sec format. Scale bars represent 5 µm. (i) Time-lapse confocal images of Dictyostelium cells expressing mCherry-FRB-PI5K(1-315aa), before or after 5 µM Rapamycin was added. Time in min:sec format. Scale bars represent 5 µm.



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Figure S5 Expressing PI5K induces macropinocytosis defects (a) Representative live-cell 1860 images of *Dictyostelium* cells expressing doxycycline-inducible PI5K with overnight DOX 1861 1862 induction (spiky). Time in sec format. Scale bars represent 5 µm. (b-c) Centroid tracks of cells (n_c=20) showing random motility in cells expressing doxycycline-inducible PI5K without DOX 1863 induction (b) or with overnight DOX induction (c). Each track lasted at least 10 minutes and was 1864 1865 reset to the same origin. (d-f) Box-and-whisker plots cell area (d), mean intensity (e), and aspect 1866 ratio (f). $n_c=20$ from at least 3 independent experiments; asterisks indicate significant difference. ****P ≤ 0.0001 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers 1867 and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (g) 1868 1869 Representative z-stack imaging showing the height of Dictyostelium cells expressing doxycyclineinducible PI5K without DOX induction (left) or with overnight DOX induction (right). Scale bars 1870 1871 represent 5 µm. (h-i) Representative confocal images of Dictyostelium cells without DOX induction (h) or with overnight DOX induction (i). Cells were treated with FITC-dextran (green) for 1872 1873 10mins before imaging. The yellow outline corresponds cell area. Scale bars represent 5 µm. (j) Quantification of macropinocytosis uptake. n_c=58 from at least 3 independent experiments; 1874 asterisks indicate significant difference, ****P ≤ 0.0001 (Mann-Whitney test. Compare ranks). The 1875 1876 median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (k, r) Representative live-cell images of Dictyostelium cells expressing 1877 doxycycline-inducible PI5K with 2h DOX induction (k) or PI5K (K681N, K682N) (r). Time in sec 1878 format. Scale bars represent 5 µm. (I-n) Color-coded temporal overlay profiles of Dictyostelium 1879 cells expressing doxycycline-inducible PI5K with overnight DOX induction (spiky) (I), or 1880 doxycycline-inducible PI5K with 2h DOX induction (m), or PI5K (K681N, K682N) (n). (o) Centroid 1881 tracks of cells (n_c=20) showing random motility in cells expressing doxycycline-inducible PI5K with 1882 2h DOX induction. Each track lasted at least 10 minutes and was reset to the same origin. (p) 1883 Color-coded temporal overlay profiles of differentiated HL-60 neutrophils expressing PIP5K1C 1884 (rounded). (g) Centroid tracks of cells ($n_c=15$) showing random motility in differentiated HL-60 1885 1886 neutrophils expressing PIP5K1B. Each track lasted at least 10 minutes and was reset to the same 1887 origin.

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Figure S6 Expressing PIP5Ks impairs cell migration and induces cell contraction (a-c) Representative live-cell images of differentiated HL-60 neutrophils expressing PIP5K1C (rounded) (a), PIP5K1C (polarized) (b), or PIP5K1C (polarized) (c). Time in min:sec format. Scale bars represent 5 µm. (d-e) Color-coded temporal overlay profiles of differentiated HL-60 neutrophils expressing PIP5K1B (polarized) (d), or PIP5K1C (polarized) (e). (f) Box-and-whisker plots of cell migration speed, n_c=20 from at least 3 independent experiments; asterisks indicate significant difference, ****P ≤ 0.0001 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (g-h) Centroid tracks of cells (n_c=15) showing random motility in differentiated HL-60 neutrophils WT (g), or expressing PIP5K1C (h). Each track lasted at least 10 minutes and was reset to the same origin. (i-i) Color-coded temporal overlay profiles of differentiated HL-60 neutrophils (WT) (i), or expressing PIP5K1B (j). (k-m) Color-coded temporal overlay profiles of differentiated HL-60 macrophages (WT) (k), or expressing PIP5K1B (I), or expressing PIP5K1C (m). (n-o) Box-and-whisker plots of mean membrane intensity for differentiated HL-60 neutrophils expressing PIP5K1B (n) or PIP5K1C (o). n_c=20 from at least 3 independent experiments; asterisks indicate significant difference, ****P ≤ 0.0001 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (p) Representative live-cell images of differentiated HL-60 macrophages expressing PIP5K1C. Time in min:sec format. Scale bars represent 5 µm. (g) Cartoon illustrating mechanism of PIP5K1C global recruitment on MDA-MB-231 cell membrane with the help of iLiD-SspB optogenetic system.

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Figure S7 Globally recruiting empty vector in MDA-MB-231 cells do not induce cell contraction (a) Time-lapse confocal images of MDA-MB-231 cells expressing crimson-SspB-PIP5K1C-P2A-iLiD-CAAX, before or after 488 nm laser was switched on globally. Time in min:sec format. Scale bars represent 10 µm. Blue arrows indicate where retraction fibers or blebs are formed. Cells are pretreated with 10 ng/ml EGF for 10 mins. (b-d) Color-coded temporal overlay profile corresponds to Figure S7a, 3k, and S7e. (e) Time-lapse confocal images of MDA-MB-231 cells expressing empty vector crimson-SspB-MCS-P2A-iLiD-CAAX, before or after 488 nm laser was switched on globally. Time in min:sec format. Scale bars represent 10 µm. (f-g) Box-and-whisker plots of (f) cell area, (g) aspect ratio correspond to (m-n). $n_c=10$ from at least 3 independent experiments; asterisks indicate significant difference, 'ns' indicates non-significant difference, ns denotes P>0.05 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10).





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Figure S9 Expressing PI5K inhibits Rac1/Arp2/3 complex/F-actin axis (a) Box-and-whisker plots of LimE patch size (left axis) and LimE patch number (right axis). nc=20 from at least 3 independent experiments; asterisks indicate significant difference, ****P ≤ 0.0001 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (b-c) t-stacks from a cell co-expressing LimE-mCherry and doxycycline-inducible PI5K without DOX induction (b) or with overnight DOX induction (rounded) (c). The white arrow corresponds to the time duration of the t-stack kymograph. (d-f) Representative live-cell time-lapse confocal images of Dictyostelium AX2 co-expressing Pak1-GFP (biosensor for Rac1) and doxycycline-inducible PI5K without DOX induction (d) or with overnight DOX induction (rounded) (e), or with overnight DOX induction (spiky) (f). Time in min:sec format. Scale bars represent 5 µm. (g) Box-and-whisker plots of Pak1 patch size (left axis) and Pak1 patch number (right axis). n_c=20 from at least 3 independent experiments; asterisks indicate significant difference, ****P ≤ 0.0001 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (h-i) Representative live-cell time-lapse confocal images of Dictyostelium AX2 co-expressing ArpC-GFP and doxycycline-inducible PI5K without DOX induction (h) or with overnight DOX induction (i). Time in min:sec format. Scale bars represent 5 µm. (i-k) Representative live-cell time-lapse confocal images of Dictyostelium abnABC- cells co-expressing RBD-GFP and doxycycline-inducible PI5K without DOX induction (i) or with overnight DOX induction (k). Time in min:sec format. Scale bars represent 5 µm.



Figure S10 Expressing less PI5K increases cell polarity but inhibits signal transduction activities (a-b) Representative live-cell time-lapse confocal images of Dictyostelium AX2 coexpressing RBD-GFP (a) or PHcrac-YFP (b) and doxycycline-inducible PI5K with 2h DOX induction. Time in min:sec format. Scale bars represent 5 µm. (c-d) Representative live-cell time-lapse confocal images of Dictvostelium AX2 co-expressing RBD-RFP (c) or LimE-RFP (d) and PI5K (K681N, K682N). Time in min:sec format. Scale bars represent 5 µm. (e-f) Representative membrane kymograph of RBD intensity in *Dictyostelium* AX2 expressing doxycycline-inducible PI5K without DOX induction (e) or with overnight DOX induction (f), even in the absence of actin cytoskeleton. Cells were pre-treated with actin polymerization inhibitor Latrunculin A (final concentration 5µM) and caffeine (final concentration 4mM) for 20min. A linear color map shows that blue has the lowest RBD intensity, whereas yellow has the highest. (g-h) Representative membrane kymograph of PHcrac intensity in *Dictyostelium* AX2 expressing doxycycline-inducible PI5K without DOX induction (g) or with overnight DOX induction (h), even in the absence of actin cytoskeleton. Cells were pre-treated with actin polymerization inhibitor Latrunculin A (final concentration 5µM) and caffeine (final concentration 4mM) for 20min. A linear color map shows that blue has the lowest PHcrac intensity, whereas yellow has the highest.



X-Distance (µm)

X-Distance (µm)

Figure S11 Expressing C2GAPB in *pi5k*- cell increases cell polarity and migration (a) Representative live-cell time-lapse confocal images of Dictyostelium AX2 co-expressing doxycycline-inducible PI5K C2GAPB and doxycycline-inducible PI5K with overnight DOX induction. Time in min:sec format. Scale bars represent 5 µm. (b) Color-coded temporal overlay profile of the cell corresponds to (a). (c) Color-coded temporal overlay profiles of Dictyostelium pi5k- cells expressing doxycycline-inducible C2GAPB without DOX induction (left) or with overnight DOX induction (right). (d) Centroid tracks of cells ($n_c=16$) showing random motility in pi5k- cells expressing doxycycline-inducible C2GAPB without DOX induction (left) or with overnight DOX induction (right). Each track lasted at least 10 minutes and was reset to the same origin. (e) Box-and-whisker plot of cell migration speed corresponds to (c). nc=16 from at least 3 independent experiments; asterisks indicate significant difference, ****P ≤ 0.0001 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10).

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Figure S12 Expressing PIP5Ks in RAW 264.7 cells increases the threshold for PI3K activation (a-h) Representative live-cell time-lapse confocal images of responses of PH-AktmCherry to global simulation C5aR agonist at 0.1 μ M in RAW 264.7 WT cells (a) or cells overexpressing PIP5K1C (b); or at 1 μ M in RAW 264.7 WT cells (c) or cells overexpressing PIP5K1C (d); or at 10 μ M in RAW 264.7 WT cells (e) or cells overexpressing PIP5K1C (f); or at 100 μ M in RAW 264.7 WT cells (g) or cells overexpressing PIP5K1C (h). Time in min:sec format. Scale bars represent 5 μ m.

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PI5K -DOX

PI5K +DOX overnight polarized shape PI5K +DOX overnight rounder shape

Figure S13 Expressing PI5K induces chemotaxis defects (a-b) Line-scan intensity profiles correspond to Figure 5d-e. (c) Representative live-cell time-lapse confocal images of 5% (left) or 20% (right) responses of PH-Akt-mCherry to global simulation C5a agonist at 100 µM in RAW 264.7 cells overexpressing PIP5K1C. (d) Representative live-cell time-lapse confocal images of 5% (left) or 20% (right) responses of RBD-GFP to global simulation folic acid (FA) at 100 nM in Dictyostelium AX2 expressing doxycycline-inducible PI5K with overnight DOX induction. (e) Histogram guantification of normalized cell number at different doses of C5a agonist in RAW 264.7 WT cells or cells overexpressing PIP5K1C. Gray columns represent PIP5K1C cells that have < 5% responses. Yellow columns represent PIP5K1C cells that have > 20% responses. Blue columns represent WT cells that have < 5% responses. Orange columns represent WT cells that have > 20% responses. (f) Histogram quantification of normalized cell number at different doses of folic acid (FA) in in Dictyostelium AX2 expressing doxycycline-inducible PI5K without DOX induction (- DOX) or with overnight DOX induction (+ DOX). Green columns represent + DOX cells that have < 5% responses. Cyan columns represent + DOX cells that have > 20% responses. Dark blue columns represent - DOX cells that have < 5% responses. Orange columns represent - DOX cells that have > 20% responses. (g) Color-coded temporal overlay profiles of vegetative Dictyostelium AX2 expressing doxycycline-inducible PI5K without DOX (left), or polarized cells with overnight DOX induction (middle), or rounded cells with overnight DOX induction (right), chemotaxing to 10 μ M folic acid. The green box is where the center of the chemoattractant source.

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Figure S14 Quantifications of directed migration speed with or without PI5K expression (a) Centroid tracks of cells (nc=13) showing chemotaxis motility in cells expressing doxycyclineinducible PI5K without DOX induction (left) or with overnight DOX induction (right). Each track lasted 30 minutes, and the center of the chemoattractant source was reset to origin. (b-c) Scatter dot plots of directed cell migration speed (b) and aspect ratio (c) corresponds to Figure S12e. n_c =18 from at least 2 independent experiments; asterisks indicate significant difference, ****P \leq 0.0001, ***P \leq 0.001 (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10).

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Figure S15 Globally recruiting Mypt169 in neutrophil and macrophage induces cell polarity and protrusive activities (a-b) Line-scan intensity profiles correspond to Figure 6b time point 30: 40 (a) or 35: 47 (b). (c-d) Centroid tracks of cells correspond to Figure 6b (n_c=15), showing random motility before (c) and after (d) 5 µM rapamycin treatment. Each track lasted at least 15 minutes and was reset to the same origin. (e) Representative live-cell time-lapse confocal images of differentiated HL-60 macrophage expressing CIBN-CAAX, CRY2PHR-mCherry-Mypt169 (magenta) and LifeAct-miRFP703 (Cyan), before or after 488 nm laser was switched on globally. Blue arrows indicate where protrusions are formed. Time in min:sec format. Scale bars represent 5 µm. (f) Color-coded temporal overlay profile corresponds to (j). (g) Representative live-cell time-lapse confocal images of RAW 264.7 cells expressing CIBN-CAAX, CRY2PHR-mCherry-Mypt169 (magenta) and PIP5K1C-GFP (green), before or after 488 nm laser was switched on globally. Blue arrows indicate where protrusions are formed. Time in min:sec format. Scale bars represent 5 µm. Cells are pretreated with 10 µM C5a agonist for 10 mins. (h) Color-coded temporal overlay profile corresponds to (I).

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Figure S16 Expressing PI5K induces a shift to cortical actin at the cell rear (a-b) Representative live-cell time-lapse confocal images of Dictyostelium AX2 co-expressing ABD120-GFP (green), LimE-Halo (magenta), and doxycycline-inducible PI5K without DOX induction (I) or with overnight DOX induction (m). Time in sec format. Scale bars represent 5 µm. (c-d) Phalloidin stain of Dictyostelium AX2 doxycycline-inducible PI5K without DOX induction (c) or with overnight DOX induction (d). (e) Box-and-whisker plot of ABD120-LimE ratio corresponds to (a-b). nc=17 from at least 3 independent experiments; asterisks indicate significant difference, **** $P \le 0.0001$ (Mann-Whitney test. Compare ranks). The median is at the center, and whiskers and outliers are graphed according to Tukey's convention (GraphPad Prism 10). (f) Representative live-cell timelapse images of Dictyostelium cells coexpressing LimE-mCherry and doxycycline-inducible PI5K with overnight DOX induction, before and after 100 µM CK666 treatment. Time in min:sec format. Scale bars represent 5 µm.



Increased Signal Transduction Network Activity

Figure S17 Schematic illustration showing the effect of PIP5Ks on cell morphology, signal transduction, and cytoskeletal dynamics. Cells display different morphology at different PIP5Ks expression levels as shown on the left of this figure. The front region of the cell at each cell morphology is shown in yellow, while the back region of the cell is shown in green. The blue box on each cell represents the zoom-in region on the right side of the figure. The yellow or green-shaded lipid head groups at each zoomed-in box represent the inner leaflet membrane. The headgroups of the inner leaflet lipid molecules that are enriched in front-state are shown in yellow, while the headgroups that are enriched in back-state are shown in green. Actin or actomyosin structures are shown at the front-state or bac-state of the cell, respectively. The black box on each lipid bilaver represents the zoom-in region of the lipid bilaver and the signaling pathways within this region. In each condition, the lighter color icons represent the depleted signaling components. Green arrow represents the increased change of this signaling molecular, while orange arrow represents the decreased change.

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2305 Supplementary Video Legends

2306 Video S1

2307 Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 (WT) cells and *pi5k*-2308 cells expressing PHPLC δ -YFP (biosensor for PI(4,5)P2), CynA-GFP (biosensor for PI(3,4)P2), 2309 and mhcA-GFP. Top left corner shows time in min:sec format. Scale bar represents 10 µm.

2310 Video S2

2311 Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 (WT) cells and *pi5k*-

2312 cells expressing RBD-GFP (biosensor for activated Ras), PHcrac-RFP (biosensor for PIP3), and 2313 LimE-mCherry (biosensor for actin polymerization). Top left corner shows time in min:sec format.

- 2314 Scale bar represents 10 µm.
- 2315 Video S3

2316 Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 (WT) cells and *pi5k*-

2317 cells expressing mhcA-GFP (green) and LimE-mCherry (magenta). In *pi5k*- cells, ventral wave

activities of mhcA and LimE can be observed. Top left corner shows time in min:sec format. Scale

- 2319 bar represents 10 μ m in *pi5k* cells and 5 μ m in AX2 cells, respectively.
- 2320 Video S4

Time-lapse confocal images of differentiated HL-60 neutrophil and macrophage expressing CIBNCAAX, CRY2PHR-mCherry-Inp54p (magenta) and LifeAct-miRFP703 (cyan), or differentiated
HL60 macrophage expressing untagged CIBN-CAAX, CRY2PHR-mCherry-empty vector
(magenta) and LifeAct-miRFP703 (cyan), before or after 488 nm laser was switched on globally.
Top left corner shows time in min:sec format. To start recruitment (magenta), the laser was
switched on at '03:09', or '03:51', or '03:30', once '488 nm ON' appears at the top of the video.
Cell was not exposed to chemoattractant during the experiment. Scale bar represents 5 µm.

2328 Video S5

Representative live-cell time-lapse images of Dictyostelium cells coexpressing PI5K-GFP and PHcrac–RFP (biosensor for PIP3), differentiated HL-60 neutrophil expressing PIP5K1B, or PIP5K1C and LifeAct (Cyna) during migration showing PI5K dynamically moves away from protrusions in migrating cells. Top left corner shows time in min:sec format. Scale bars represent 5 mm.

2334 Video S6

Representative live-cell time-lapse images of Dictyostelium cells coexpressing PI5K-GFP and
PHcrac-RFP (biosensor for PIP3), PI5K-mCherry and RBD-GFP (biosensor for activated Ras),
PI5K-mCherry and PHPLCδ–GFP (biosensor for PI(4,5)P2), differentiated HL-60 macrophage
expressing PIP5K1B and LifeAct, and Dictyostelium cells coexpressing PI5K-mRFP and RBDGFP (biosensor for activated Ras) upon Latrunculin A treatment, during ventral wave propagation,
showing PI5K dynamically localizes to the back-state regions in ventral waves. Top left corner
shows time in min:sec format. Scale bars represent 5 mm.

2342 Video S7

- 2343 Representative live-cell images of *Dictyostelium* cells co-expressing PI5K-GFP and PHcrac-RFP, 2344 or PHPLC δ -GFP and PHcrac-RFP upon global cAMP stimulation. Top left corner shows time in 2345 min:sec format. To start global stimulation, cAMP was added at '00:53', or '01:17', once '+ cAMP'
- 2346 appears at the top of the video. Scale bar represents 5 μ m.
- 2347 Video S8

Representative live-cell images of *Dictyostelium* cells expressing doxycycline inducible KikGRPI5K with overnight DOX incubation. Top left corner shows time in min:sec format. Photo
conversion happened at '01:50', '05:30', or '11:34'. Scale bar represents 5 µm.

2351 Video S9

2352 Representative live-cell time-lapse images of Dictyostelium cells expressing PI5K-GFP (301-2353 718aa) during migration showing PI5K (301-718aa) dynamically localizes at the trailing edge in migrating cells, or expressing mRFPmars-SspBR73Q-PI5K(316-718aa), before or after 488 nm 2354 2355 laser was switched on globally, or expressing mCherry-FRB-PI5K(1-315aa), before or after 5 µM 2356 Rapamycin was added. Top left corner shows time in min:sec format. To start recruitment, the 2357 laser was switched on at '01:20', once '488 nm ON' appears at the top of the video, or Rapamycin 2358 was added at '12:51', once '+ Rapamycin' appears at the top of the video. Scale bars represent 2359 5 µm.

2360 Video S10

Representative live-cell images of *Dictyostelium* cells expressing doxycycline-inducible PI5K without DOX induction, or with 2h DOX incubation, or with overnight DOX induction (rounded), or with overnight DOX induction (spiky), or expressing PI5K(K681N, K682N). Top left corner shows time in min:sec format. Scale bars represent 5 mm for first 3 videos and 10 mm for last video.

2365 Video S11

Representative live-cell images of differentiated HL-60 neutrophils (WT) expressing LifeAct as the
 biosensor, or expressing PIP5K1B (rounded), or expressing PIP5K1B (polarized), or expressing
 PIP5K1C (rounded), or expressing PIP5K1C (polarized). Top left corner shows time in min:sec
 format. Scale bars represent 5 µm.

2370 Video S12

Representative live-cell images of differentiated HL-60 macrophage (WT) expressing PH-Akt as
 the biosensor, or expressing PIP5K1B (rounded), or expressing PIP5K1C (rounded). Top left
 corner shows time in min:sec format. Scale bars represent 5 µm.

2374 Video S13

Time-lapse confocal images of MDA-MB-231 cells expressing crimson-SspB-PIP5K1C-P2A-iLiD-CAAX or expressing crimson-SspB-empty vector-P2A-iLiD-CAAX, before or after 488 nm laser was switched on globally. Top left corner shows time in hour:min:sec or min:sec format. To start recruitment, the laser was switched on at '00:12:21', or '09:44', or '04:55', once '488 nm ON' appears at the top of the video. Scale bar represents 10 µm.

2380 Video S14

2381 Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 co-expressing RBD-2382 GFP (biosensor for activated Ras) or PHcrac-YFP (biosensor for PIP3) and doxycycline-inducible 2383 PI5K without DOX induction, or with 2h DOX incubation, or with overnight DOX induction (rounded), or with overnight DOX induction (spiky). Top left corner shows time in min:sec format.
 Scale bars represent 5 µm.

2386 Video S15

Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 co-expressing RBD GFP (biosensor for activated Ras) and doxycycline-inducible PI5K without DOX induction, or with
 overnight DOX induction during ventral wave propagation. Top left corner shows time in min:sec
 format. Scale bars represent 10 µm.

2391 Video S16

Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 co-expressing Pak1 GFP (biosensor for Rac1), or ArpC-GFP, or LimE-mCherry and doxycycline-inducible PI5K
 without DOX induction, or with overnight DOX induction (rounded) or with overnight DOX
 induction (spiky). Top left corner shows time in min:sec format. Scale bars represent 5 μm.

2396 Video S17

Representative live-cell time-lapse confocal images of *Dictyostelium abnABC*- cells co expressing RBD-GFP and doxycycline-inducible PI5K without DOX induction, or with overnight
 DOX induction. Top left corner shows time in min:sec format. Scale bars represent 5 µm.

2400 Video S18

Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 co-expressing RBD RFP, or LimE-RFP and PI5K (K681N, K682N). Top left corner shows time in min:sec format. Scale
 bars represent 5 µm.

2404 Video S19

Representative live-cell time-lapse confocal images of Dictyostelium *pi5k*- cells expressing doxycycline-inducible PI5K C2GAPB without DOX induction, or with overnight DOX induction, or *Dictyostelium* AX2 co-expressing doxycycline-inducible PI5K C2GAPB and doxycycline-inducible PI5K with overnight DOX induction. Top left corner shows time in min:sec format. Scale bars represent 10 µm for first 2 videos, and 5 µm for last video.

2410 Video S20

2411 Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 co-expressing RBD-2412 GFP (biosensor for activated Ras), or PHcrac-YFP (biosensor for PIP3) and doxycycline-inducible 2413 PI5K without DOX induction, or with overnight DOX induction upon Latrunculin A treatment. Top 2414 left corner shows time in min:sec format. Scale bars represent 5 µm.

2415 Video S21

2416 Representative live-cell time-lapse confocal images of responses of PH-Akt-mCherry to global 2417 simulation C5aR agonist in RAW 264.7 WT cells, or cells overexpressing PIP5K1C at at 0.1 μ M-2418 100 μ M. To start global stimulation, C5a agonist was added at '00:53', or '06:09', or '05:45', or 2419 '06:40', or '16:42', or '03:42', or '06:21', or '05:55', or '12:52', once '+ C5a' appears at the top of 2420 the video. Scale bar represents 5 μ m.

2421 Video S22

Representative live-cell time-lapse confocal images of *Dictyostelium Gb*- cells co-expressing
 RBD-GFP and doxycycline-inducible PI5K without DOX induction, or with overnight DOX
 induction. Top left corner shows time in min:sec format. Scale bars represent 5 µm.

2425 Video S23

2426 Representative live-cell time-lapse confocal images of vegetative *Dictyostelium* AX2 expressing

doxycycline-inducible PI5K without DOX, or with overnight DOX induction, chemotaxing to 10 mM

folic acid-filled micropipette. The white box is where the center of the chemoattractant source. Top left corner shows time in min:sec format. Scale bars represent 20 µm.

2430 Video S24

2431 Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 co-expressing CAR1-

2432 FKBP-FKBP, mCherry-FRB-MHCKC, and doxycycline-inducible PI5K with overnight DOX induction before and after 5 μM rapamycin treatment. Top left corner shows time in min:sec format.

To start recruitment, Rapamycin was added at '14:03', once '+ Rapamycin' appears at the top of

- 2435 the video. Scale bars represent 10 μm.
- 2436 Videos S25

(e) Representative live-cell time-lapse images of Dictyostelium cells coexpressing RBD-GFP and
 doxycycline-inducible PI5K with overnight DOX induction during ventral wave propagation, before
 and after 50 µM blebbistatin treatment. Top left corner shows time in min:sec format. Blebbistatin
 was added at '06:33', once '+ Blebbistatin' appears at the top of the video. Scale bars represent
 10 µm.

2442 Videos S26

2443 Representative live-cell time-lapse confocal images of differentiated HL-60 macrophage 2444 expressing untagged CIBN-CAAX, CRY2PHR-mCherry-Mypt169 (magenta), or differentiated HL-2445 60 neutrophil and macrophage expressing untagged CIBN-CAAX, CRY2PHR-mCherry-Mypt169 (magenta), and PIP5K1B-GFP (green), before or after 488 nm laser was switched on globally. 2446 2447 Top left corner shows time in min:sec or hour:min:sec format. To start recruitment (magenta), the 2448 laser was switched on at '03:41', or '00:05:55', or '02:51', once '488 nm ON' appears at the top of 2449 the video. Cell was not exposed to chemoattractant during the experiment. Scale bar represents 2450 10 μ m for the frist video, and 5 μ m for the rest two videos.

2451 Video S27

2452 Representative live-cell time-lapse confocal images of *Dictyostelium* AX2 co-expressing ABD120-

GFP (green), LimE-Halo (magenta), and doxycycline-inducible PI5K without DOX induction, or

- with overnight DOX induction. Top left corner shows time in min:sec format. Scale bars represent
 5 µm.
- 2456 Video S28

2457 Representative live-cell time-lapse images of Dictyostelium cells coexpressing LimE-mCherry 2458 and doxycycline-inducible PI5K with overnight DOX induction, before and after 100 μ M CK666 2459 treatment. Top left corner shows time in min:sec format. CK666 was added at '12:23', once '+ 2460 CK666' appears at the top of the video. Scale bars represent 5 μ m.

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