

**Figure S1. Single and dual lysin kill switches.** (a) TetON single lysin kill switch schematic. Lysin expression is repressed by tetracycline repressor (TetR) and induced by anhydrotetracycline (atc) or doxycycline (doxy) which bind TetR and prevent it from binding DNA.

(b) Impact of atc on growth of BCG-TetON single lysin strains. The paper disc contains 1  $\mu\text{g}$  of atc.

(c) TetOFF dual lysin kill switch schematic. Reverse TetR (Rev TetR) binds to DNA in complex with atc or doxy to repress D29L. TetR represses PipR and in the presence of atc/doxy PipR is expressed and represses L5L.

(d) Impact of atc on growth of BCG-TetOFF-DL. The paper disc contains 1  $\mu\text{g}$  of atc.

**Figure S2. In vitro characterization of BCG kill switch strains.**

(a) Growth of BCG TetON single and dual lysin kill switch strains with and without atc. OD<sub>580</sub> data are means from duplicate cultures.

(b) Western blot analysis of culture filtrates from BCG-TetON-DL and BCG-TetOFF-DL strains grown in the absence of detergent. A whole cell lysate of WT BCG serves as control. Eno and PrcB were enriched in culture filtrate of BCG-TetON-DL after 6 and 9 days of growth in the presence of atc and in culture filtrate of BCG-TetOFF-DL after 6 and 9 days of growth in the absence of atc indicating cell lysis.

(c) Expression of two lysins reduces the fraction of escape mutants compared to expression of single lysins. Cultures were grown in the presence of atc. Symbols represent data from 3-6 individual cultures and means  $\pm$  SD are depicted. Significance was determined by one-way analysis of variance (ANOVA) with Tukey's adjusted p-values shown; \*\*\*\* P < 0.0001.

(d) Growth of BCG TetOFF single and dual lysin kill switch strains with and without atc. Data are means  $\pm$  SD from triplicate cultures. Error bars are too small to be seen.

(e) Expression of two lysins reduces the fraction of escape mutants compared to expression of single lysins. Cultures were grown in the absence of atc. Symbols represent data from 6 individual cultures and means  $\pm$  SD are depicted. Statistical significance was assessed by one-way ANOVA with Tukey's adjusted p-values shown; \*\*\*\* P < 0.0001, \*\*\*, p < 0.001.

**Figure S3. Survival of BCG, BCG-TetOFF-DL and BCG-TetON-DL following intravenous vaccination.**

(a) CFU from lungs and spleens of mice infected with BCG and BCG-TetOFF-DL not receiving doxy.

(b) CFU quantification from lungs and spleens of mice infected with BCG and BCG-TetON-DL treated with doxy. Data are means  $\pm$  SD from 4-5 mice per group and time point.

(c) CFU quantification from lungs and spleens of BCG-TetOFF-DL infected SCID mice treated or not with doxy for the indicated times. Data are means  $\pm$  SD from 4 mice per group and time point.

(a,b,c) Multiple unpaired t-tests run on log<sub>10</sub>-transformed data at each time point with Holm-Šídák adjusted p-values shown. \*\*\*\* p < 0.0001, \*\*\*, p < 0.001, \*\* p < 0.01, \* p < 0.05, # 0.05 < p < 0.10, ns p > 0.10.

(d,e) Lung sections stained with hematoxylin and eosin from SCID mice infected with BCG-TetOFF-DL for 84 days. (d) Mice were treated with doxy from day 1-14. (e) Mice were treated with doxy from day 1- 84. Each section is from an individual mouse and is representative of each lung.

**Figure S4. BCG-TetON-DL and wt BCG provide similar protection against Mtb infection in mice.** Mice were i.v. vaccinated with BCG or BCG-TetON-DL or received PBS.

(a,b) Quantification of effector memory CD4 and CD8 T cells in mouse lungs from mice vaccinated with BCG and BCG-TetON-DL. Symbols are data from individual mice with lines indicating mean  $\pm$  SD. Two-way ANOVA was performed with Tukey's adjusted p-values shown for each time point. \*\*\*\* p < 0.0001, \*\*\*, p < 0.001, \*\* p < 0.01, \* p < 0.05, ns p > 0.10.

(c) Quantification of cytokine producing antigen specific CD4 T cells from mice vaccinated with BCG and BCG-TetON-DL on day 30 post vaccination. Lung cells were restimulated ex vivo with PPD prior to intracellular

cytokine staining. Two-way ANOVA with Tukey's adjusted p-values shown for each cytokine producing population. For most of the cytokine producing cells, there were no statistically significant differences among the treatment groups. \*\*\*\*  $p < 0.0001$ , \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ , ns  $p > 0.10$ .

(d) Bacterial burden in lungs and spleens of vaccinated and PBS treated mice. Mice were infected with Mtb H37Rv by aerosol 90 days post vaccination. Symbols represent data from individual mice with lines indicating mean  $\pm$  SEM. Two-way ANOVA performed on  $\log_{10}$ -transformed data with Tukey's adjusted p-values shown at each time point. \*\*\*\*  $p < 0.0001$ , \*\*\*  $p < 0.001$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ , ns  $p > 0.10$ .

**Figure S5 Histologic analysis indicates microgranulomas in spleen, liver and lymph nodes 8 weeks post- BCG-TetOFF-DL.** H&E staining of fixed spleen (a), lymph node (b) and liver (c) tissue. (d, e) Lymphocyte composition of lung tissue (d, n=4) and thoracic lymph nodes (e, n=3) from NHP in persistence study at necropsy.

**Figure S6 T cell responses in airways are similar between BCG-TetOFF-DL and WT BCG after vaccination.** Total number of cells, CD4+, CD8+, effector memory CD4+ and CD8+ cells during the vaccination phase with BCG-TetOFF-DL, WT BCG or unvaccinated macaques. BAL samples were obtained pre-vaccination and 4, 12, 20 weeks post vaccination and stimulated with Mtb WCL. Flow cytometry was performed with intracellular staining for effector molecules IFN- $\gamma$ , TNF, IL-17, IL-2, GzmB, GzmK, granulysin or perforin. Multiple unpaired t-tests were used to compare groups at each time point with Holm-Šidák multiple comparison adjusted p-values ( $\# 0.05 < p < 0.10$ ). Median and IQR shown.

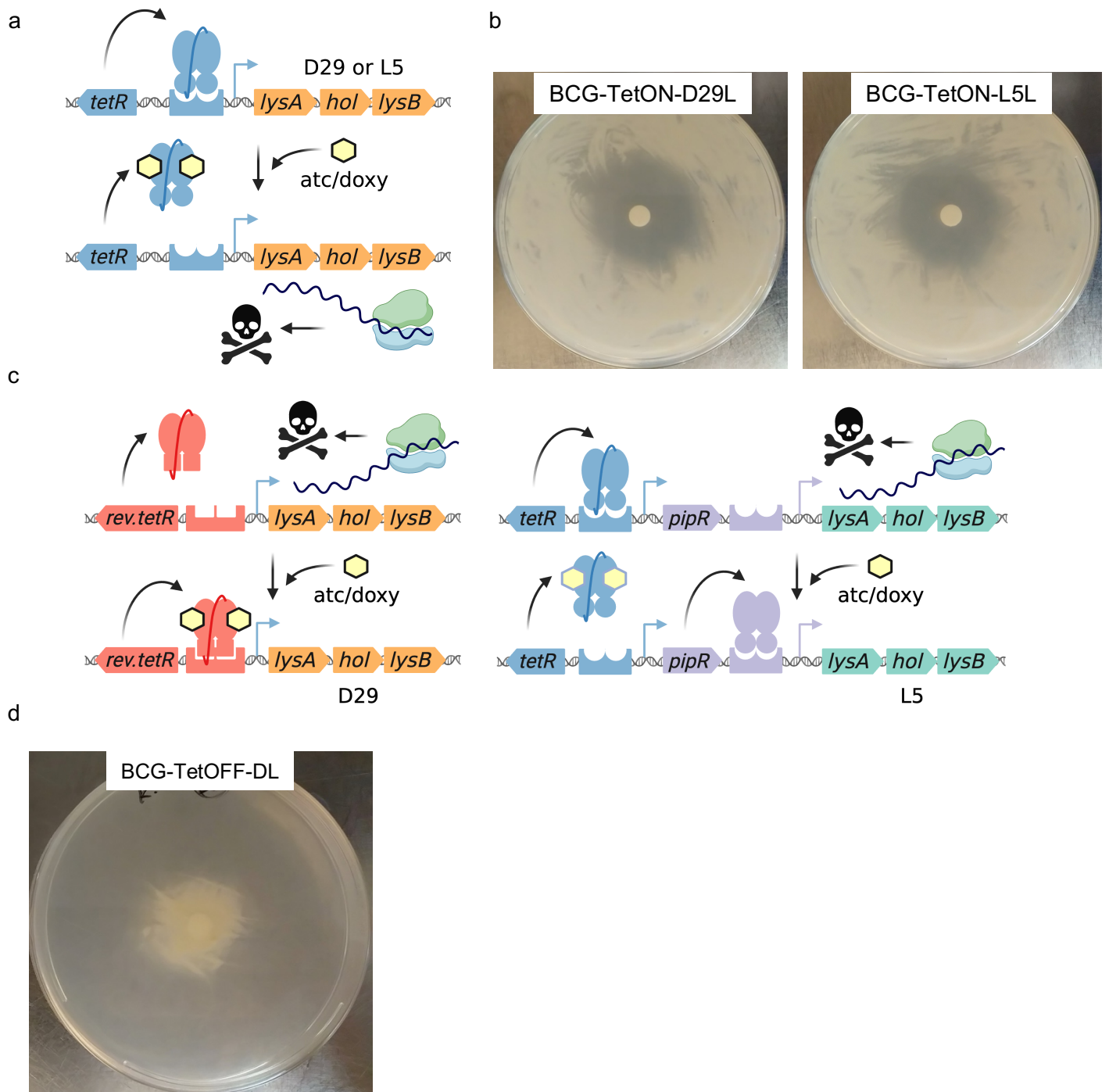
**Figure S7 Individual effector molecules produced by T cells in lungs of vaccinated and challenged NHP.** Total number of CD4+, CD8+, effector memory CD4+ and CD8 $\alpha\beta$ + cells in the lung at necropsy (n=8). Cells producing either cytokines or cytotoxic molecules (IFN- $\gamma$ , TNF, IL-17, IL-2, GzmB, GzmK, granulysin or perforin) were analyzed. Mann-Whitney p-values reported. Median shown.

**Figure S8 Splenomegaly reduces over time post IV-BCG vaccination.** Spleen size at necropsy for macaques in persistence study and macaques vaccinated and challenged with Mtb. Dashed lines represent normal spleen size range of adult male MCMs.

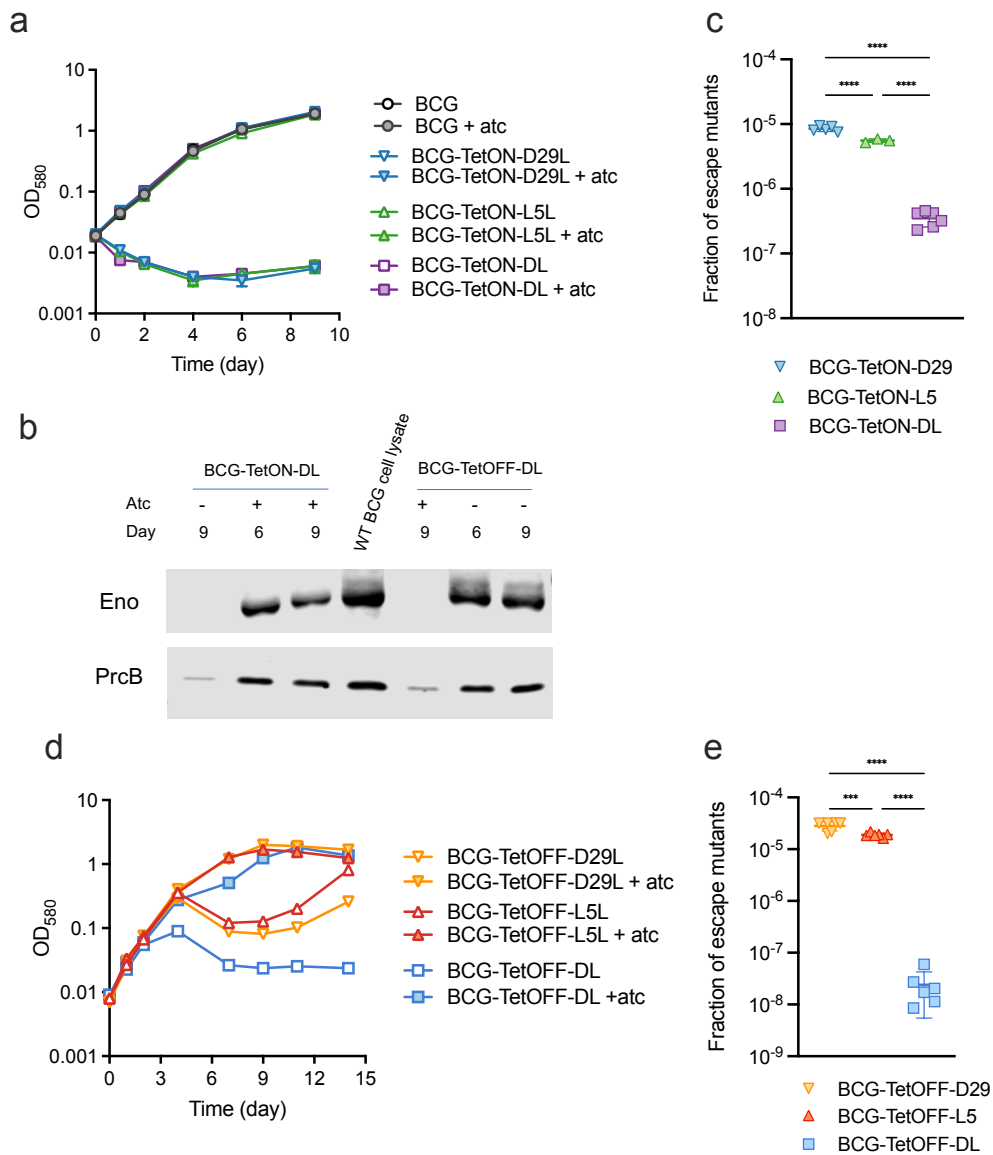
**Figure S9 Gating strategy**

**Table S1: Flow cytometry panel for NHP samples**

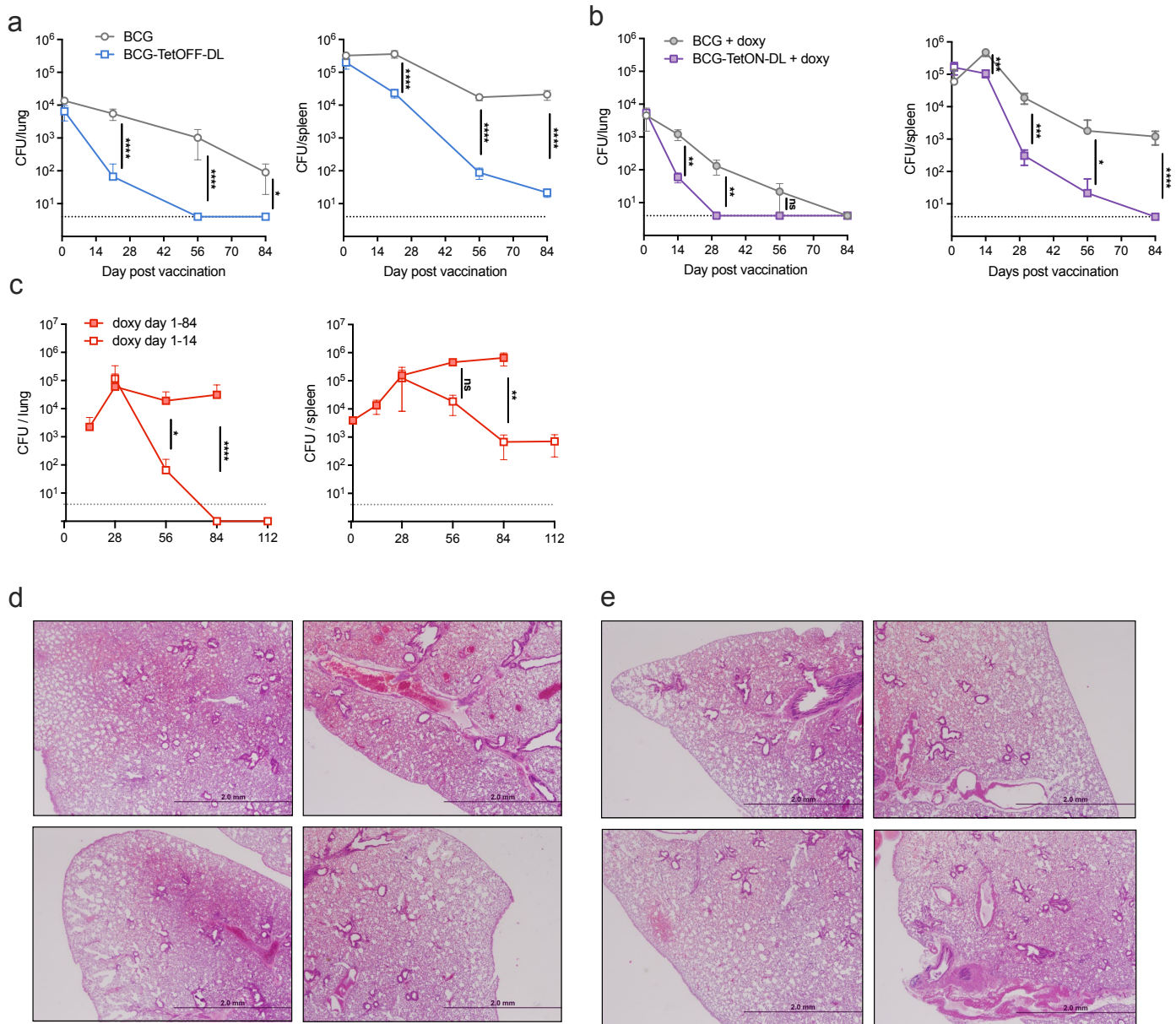
**Table S2 Full details on macaques used in this study**



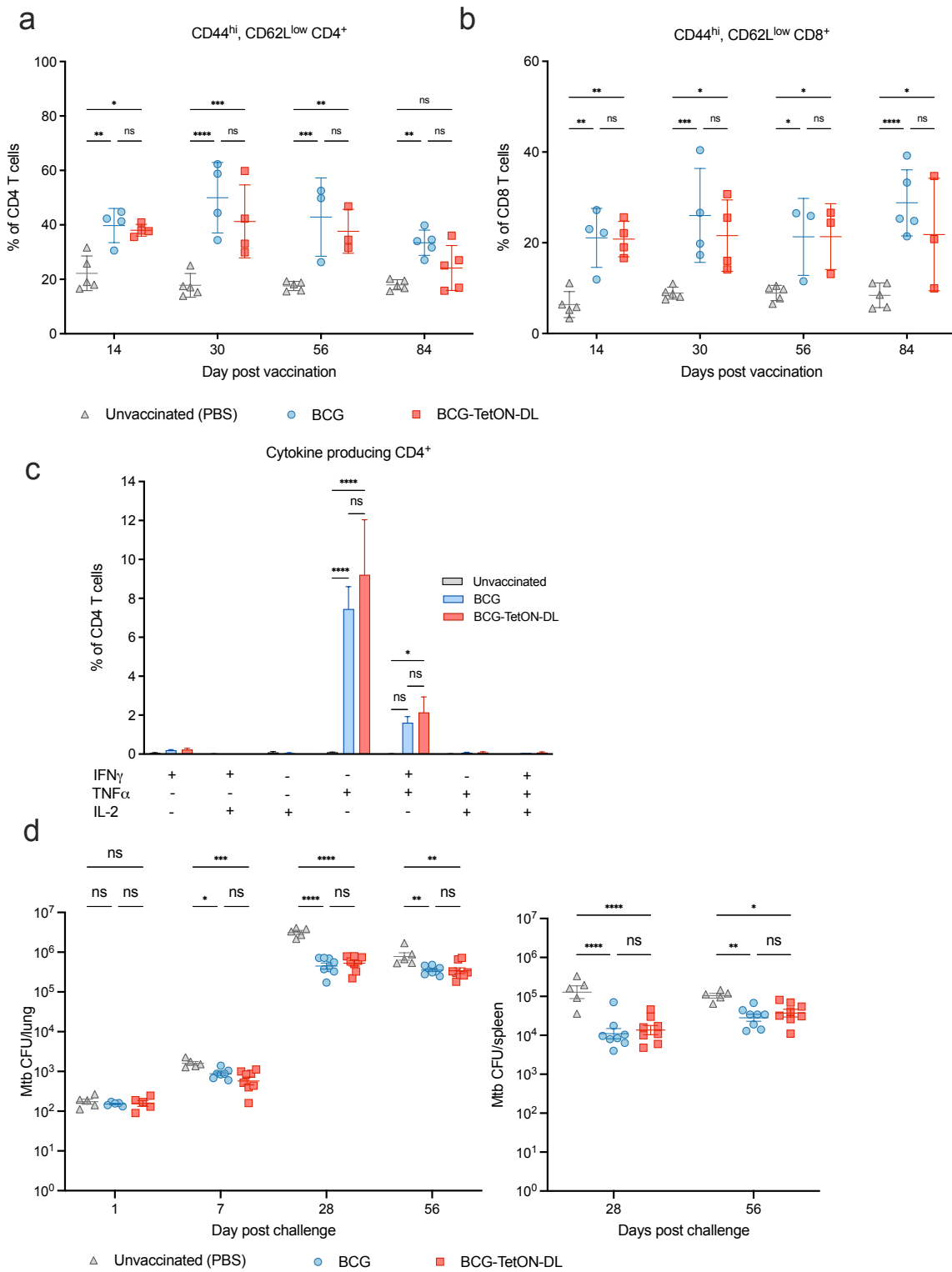
**Figure S1**



**Figure S2**



**Figure S3**



**Figure S4**

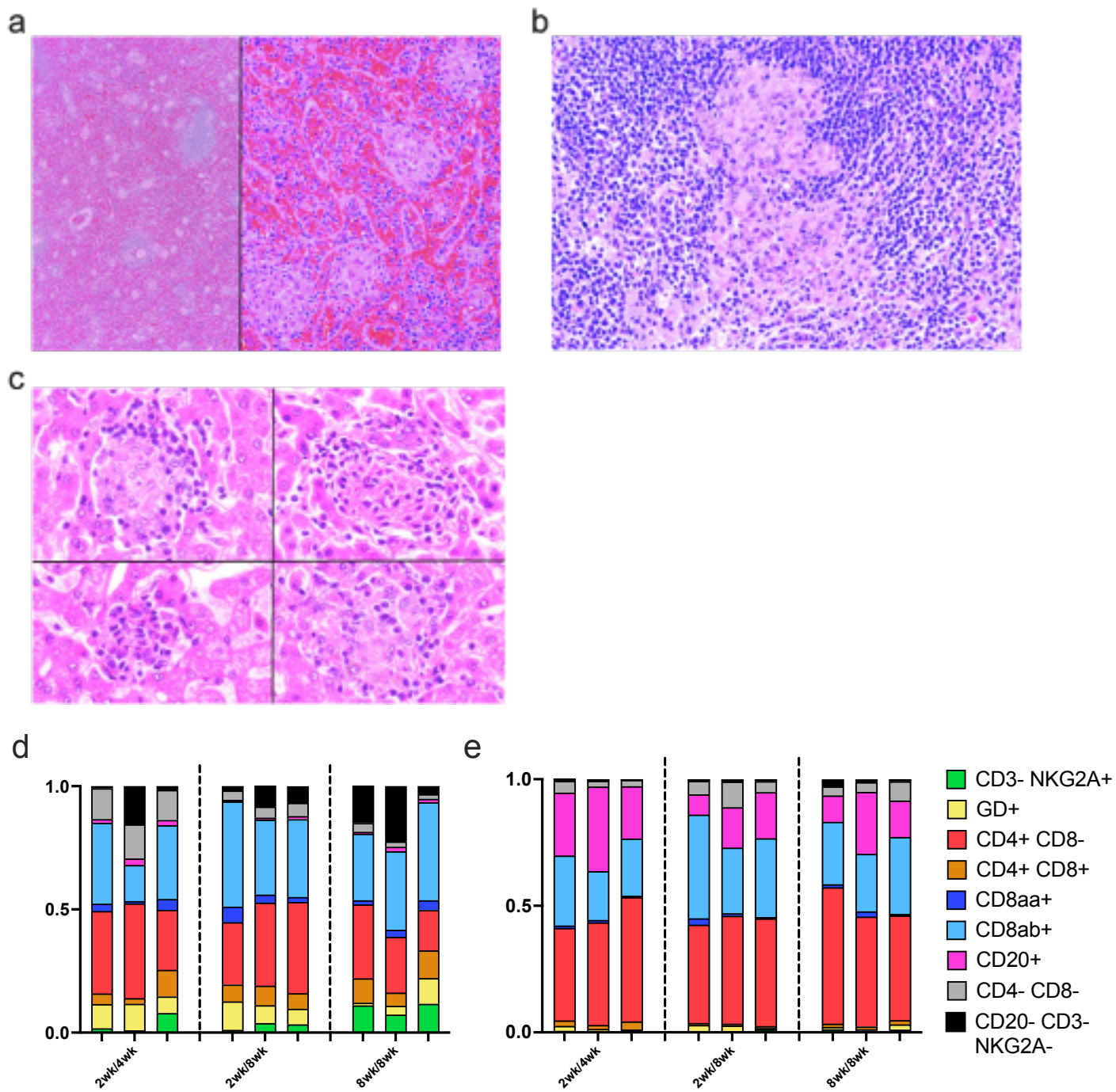
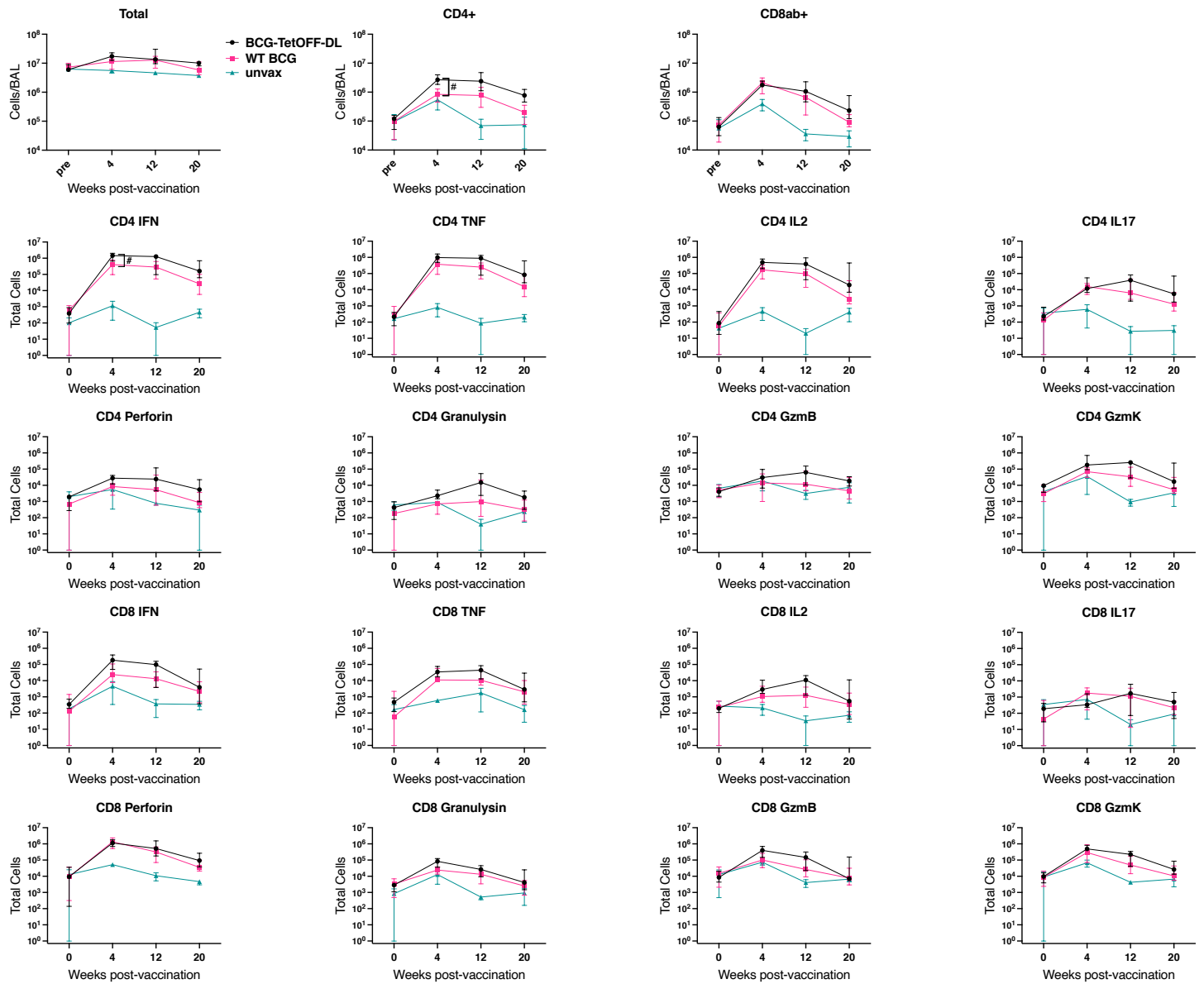


Figure S5



**Figure S6**



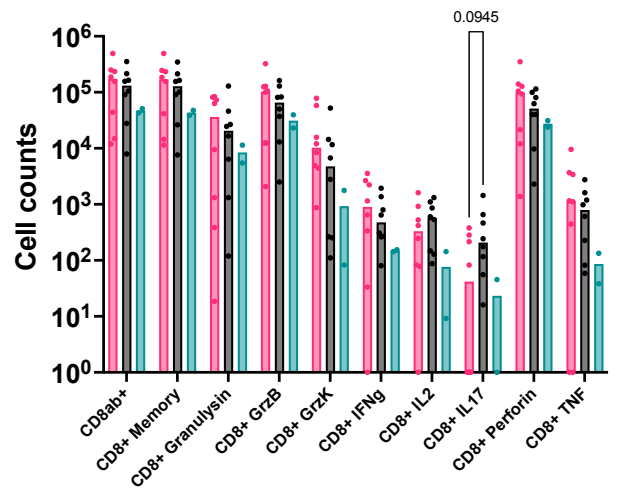
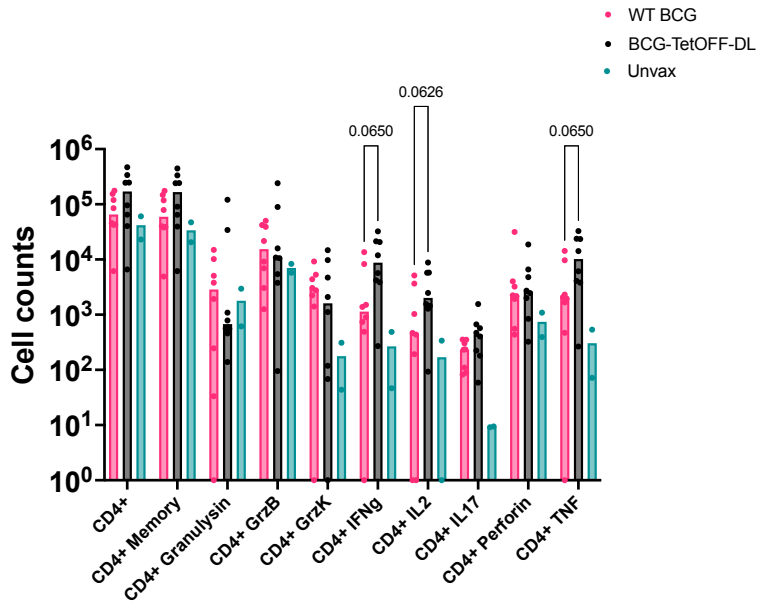


Figure S7

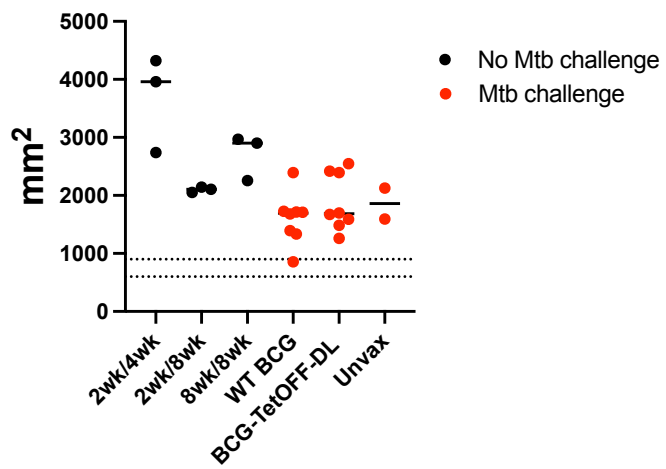
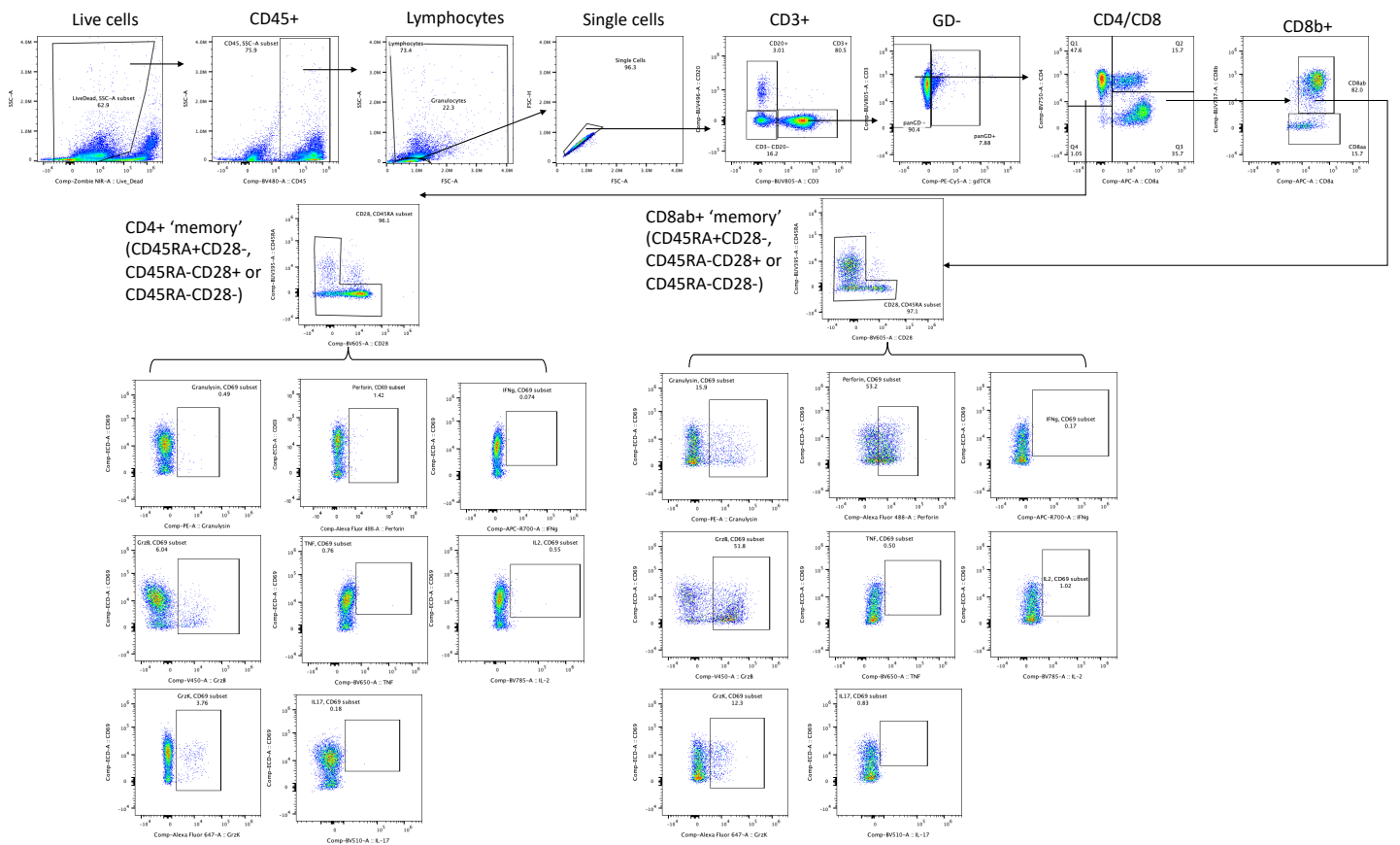


Figure S8



**Figure S9**

	Target	Clone	Fluor	Product #	Vendor
Surface	CD3	SP34-2	APC-Cy7	624072	BD Biosciences
	CD4	L200	BV750	747202	BD Biosciences
	CD8a	DK25	FITC	FCMAB176F	EMD Millipore
	CD8b	2ST8.5H7	BUV737	748324	BD Biosciences
	Pan gd	5A6.E9	PE	MHGD04	Thermo Fischer Scientific
	CD16	3G8	BV570	302036	Biolegend
	CD159a	Z199	PE-Cy7	B10246	Beckman Coulter
	CD45	D058-1283	BV480	566145	BD Biosciences
	CD28	CD28.2	BV605	302968	BD Biosciences
	CD45RA	5H9	BUV395	740315	BD Biosciences
	CD20	2H7	PE-CY5	302308	Biolegend
	CD11b	ICRF44	BUV563	741357	BD Biosciences
	CD11c	3.9	BV421	301628	Biolegend
ICS	CD69	TP1.55.3	ECD	6607110	Beckman Coulter
	Granzyme B	GB11	V450	561151	BD Biosciences
	Granzyme K	G3H69	AF647	566655	BD Biosciences
	IL-17	BL168	BV510	512330	Biolegend
	TNF	MAb11	BV650	502938	Biolegend
	IFNg	B27	APC	506510	Biolegend
	IL-2	MQ1-17H12	BV785	500348	Biolegend
☞	Zombie		NIR	423106	Biolegend

Table S1

