

Supplementary data**Supplemental Table 1 Number of separated cells from the P-Sp region in each embryo**

	WT	<i>Evi</i> ^{+/+}	<i>Evi</i> ^{-/-}
1	14,168	14,929	12,268
2	20,195	19,199	22,784
3	16,653	16,775	15,949
Average	17,005	16,968	17,000
SD	3,209	2,142	5,336

The P-Sp region from each embryo was isolated at E9.5 and counted the viable separated-cells. The viability is over 90% in each embryo.

Supplemental Table 2 List of gene specific primers for RT-PCR

Gene	Primer sequences (5'-3') (Forward) (Reverse)	Size of products	No. of cycles
Evi1	AATATGAGTCATGCCAACCC CTTGGTGTACTGACATCATC	396	30 or 36
GATA-2	ACACACCACCCGATACCCACCTAT CCTACGCCATGGCAGTCACCATGCT	711	26 or 30
GFP	CTGACCCTGAAGTTCATCTGC GCGGATCTTGAAGTTCACC	381	26
GAPDH	AATCCCATCACCATCTTCCA CCAGGGGTCTTACTCCTTG	700	24

Supplemental Table 3A Oligonucleotide sequences for D1-CONS like DNA binding sites used for EMSA

Name	Position (nt.)	Sequences
D1-CONS		
F		5'-CTAGCGACAAGATAAGATAAGGAGACAAGATAAGGATGACAAGATAAGATAAC-3'
R		5'-TCGAGTTATCTTATCTTGTCACCTTATCTTATCTTGTCCTTATCTTATCTTGTCG-3'
a (3430F)	-6581 to -6554	5'-GAAAGGTCAGGATAGACAGACTGCAAAG-3'
(3430R)		5'-CTTTGCAGTCTGTCTATCCTGACCTTTC-3'
b (3884F)	-6127 to -6108	5'-ACAGTGGTGATGTCCATGTCCCTTTTATTT-3'
(3884R)		5'-AAATAAAAGGGACATGGACATCACCCTGT-3'
c (8543F)	-1468 to -1435	5'-TTAATACAATTGTCAGTAAGTATCCAGACACATT-3'
(8543R)		5'-AATGTGTCTGGATACTTACTGACAATTGTATTAA-3'
d (8721F)	-1290 to -1254	5'-CACTGATTAAGACATACTTATAGATAACCGACTGCCT-3'
(8721R)		5'-AGGCAGTCGGTATCTATAAGTATGTCCTTAATCAGTG-3'

Supplemental Table 3B. Oligonucleotide sequences for D2-CONS like DNA binding sites used for EMSA

Name	Position (nt.)	Sequences
D2-CONS		
D2-CONSF	5'-CTAGC <u>CTCATCTTC</u> GGACTCATCTTCGGTCTCATCTTCC-3'	
D2-CONSR	5'-TCGAG <u>GAAGATGAG</u> ACCGAAGATGAGTCCGAAGATGAGG-3'	
e (3987F)	-6024 to -5999	5'-TGATGCACTG <u>ATCTTC</u> GTGATGAGTC-3'
(3987R)		5'-GACTCATCAC <u>GAAGAT</u> CAGTGCATCA-3'
f (3995F)	-6016 to -5991	5'-TGATCTTCGT <u>GATGAG</u> TCAGACCTGA-3'
(3995R)		5'-TCAGGTCTG <u>ACTCATC</u> ACGAAGATCA-3'
g (5877F)	-4134 to -4109	5'-CCTGCCTGG <u>GATGAG</u> CTAATCCCGC-3'
(5877R)		5'-GCGGGATTAG <u>CTCATCT</u> CCAGGCAGG-3'
h (6180F)	-3831 to -3806	5'-GTACAGCCAT <u>CTCATC</u> CTCACAGCCA-3'
(6180R)		5'-TGGCTGTGAG <u>GATGAG</u> ATGGCTGTAC-3'
i (7023F)	-2988 to -2963	5'-GCTATTCATT <u>ATCTTC</u> GCCGGGAACA-3'
(7023R)		5'-TGTTCCCGG <u>CGAAGAT</u> AATGAATAGC-3'
j (7320F)	-2685 to -2666	5'-CCTTGCTTT <u>ACTCATC</u> TGTTTTCTTT-3'
(7320R)		5'-AAAGAAAAC <u>GATGAG</u> TAAAGCAAGG-3'
k (7657F)	-2354 to -2329	5'-CCTGCCTTCAG <u>GAAGAT</u> CAGATTGAGT-3'
(7657R)		5'-ACTCAATCTG <u>ATCTTC</u> TGAAGGCAGG-3'
l (7995F)	-2016 to -1991	5'-GGCTTATCAG <u>CTCATCT</u> CCCCCTCCC-3'
(7995R)		5'-GGGAGGGGG <u>GATGAG</u> CTGATAAGCC-3'

Supplemental Table 4A List of primers for constructing the reporter plasmid

Name	Nucleotide position (nt)	Sequences (5'-3')
5' <i>SacI</i> gata2US1.6	-1740 to -1717	<u>gagctc</u> GTGTCAACCCAAAGCAATAGCTCCG
5' <i>SacI</i> gata2US3.3	-3326 to -3301	<u>gagctc</u> GCACACTTACTCGCACACTCACATC
3' <i>BglII</i> gata2US	-31 to -53	<u>agatct</u> GAACGCAAGCTCGTCGCGCAGGC
5' <i>SacI</i> gata2US7.0	-7000 to -6977	<u>gagctc</u> GCCTCTGGTCCTACACTAGGCCTT
3' <i>XhoI</i> gata2US7M	-3327 to -3356	<u>ctcgag</u> AAAGTTCACGTGTGCGAGACCCTG
5' <i>XhoI</i> gata2US7U	-3332 to -3303	<u>ctcgag</u> GCACACTTACTCGCACACTCACA
D1MF		TGCACACAGTGGTGATCTCCATGTCCCTTTTATTTACGG
D1MR		CCGTGAAATAAAAAGGGACATGGAGATCACCCTGTGTGCA
D2f-F		TGATGCACTGATCTTCGTTATGAGTCAGACCTGACTGCA
D2f-R		ACTACGTGACTAGAAGCACTACTCAGTCTGGACTGACGT

Sequences with underline are shown to be restriction sites.

Supplemental Table 4B List of PCR primers for chromatin immunoprecipitation

Binding site	Name	Sequences (5'-3')	
GATA-2 (Up)	5ChIPup	CTCACCTGCCCACCATTGTCT	35 cycles
	3ChIPup	GTGGCTAGAGGTACTGCCAGCAAGT	94°C, 56°C and 72°C for 30 sec each
GATA-2 (D1-a)	5ChIP3430	GAGGAGGTTTCGCATAGGACCC	35 cycles
	3ChIP3440	TTGGAAAATGCTCAGTCATGTTAG	94°C, 55°C and 72°C for 30 sec each
GATA-2 (D1-b)	5ChIP38840	CCTCTGCCTCTCTGCCTCTGCCTC	35 cycles
	3ChIP38840	GCA TGTTTTCTAATATCCAGGCCT	94°C, 65°C and 72°C for 30 sec each
GATA-2 (Down)	5ChIPdown	GCTGCAGGGGAGGAGCGGTC	35 cycles
	3ChIPdown	CAGCCAGCCCGCCGCCGAGGTC	94°C, 65°C and 72°C for 30 sec each
b-actin	5ACTB	GCTTCTTGTCACCACCTCAA	35 cycles
	3ACTB	GGGAAGACCCTGTCCTTGTC	94°C, 55°C and 72°C for 30 sec each