

Supplementary Materials for
Cell type–specific epigenetic priming of gene expression in nucleus accumbens by cocaine

Philipp Mews *et al.*

Corresponding author: Eric J. Nestler, eric.nestler@mssm.edu

Sci. Adv. **10**, eado3514 (2024)
DOI: 10.1126/sciadv.ado3514

This PDF file includes:

Figs. S1 to S11
Tables S1 and S2
References

SUPPLEMENTARY MATERIALS

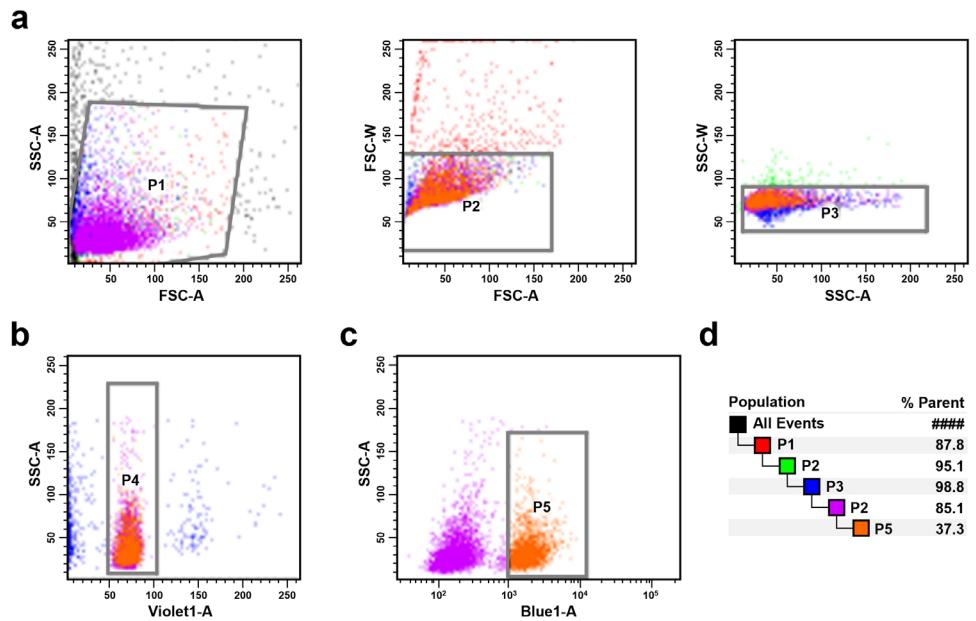


Figure S1. Fluorescence-activated nuclei sorting (FANS) to purify D1 and D2 MSN nuclei from NAc. Representative gating strategy for FANS. **(a)** First three gates on FSC-A vs. SSC-A (P1), FSC-A vs. FSC-W (P2), and SSC-A vs SSC-W (P3) retrieve nuclei as opposed to debris. **(b)** Gating strategy on Violet1-A vs. SSC-A (P4) retrieves single nuclei (DAPI+ events) as opposed to duplets. **(c)** The final gate FSC-A vs. Blue1-A (FITC channel) separates transgenically labeled nuclei (P5, GFP+ events) from wild-type nuclei. **(d)** Summary table of representative hierarchical gating strategy.

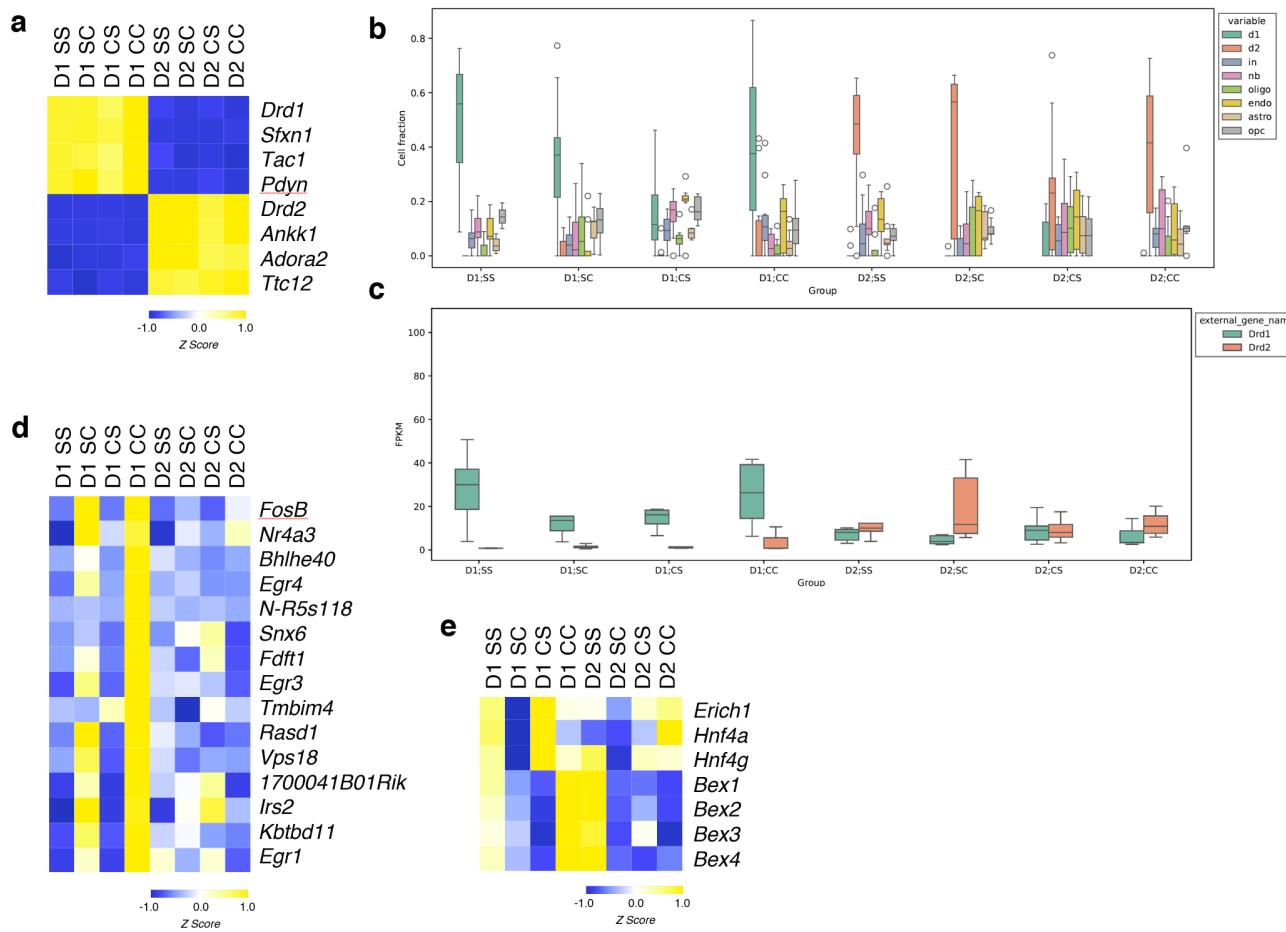


Figure S2. **(a)** Heatmap of mRNA expression (cpm) of well-characterized genes specific to D1 MSNs (*Drd1*, *Sfcn1*, *Tac1*, *Pdyn*) or to D2 MSNs (*Drd2*, *Ankk1*, *Adora2*, *Ttc12*), confirming successful sorting and purification of D1 and D2 MSN nuclei. **(b)** To evaluate the purity of the FACS sorted cells from the perspective of the RNA-seq libraries, the cell type components of the RNA-seq samples were estimated using a deconvolution approach using Unicell. The sorted D1 RNA-seq samples exhibit a high predicted D1 cell type proportion, whereas the sorted D2 RNA-seq samples exhibit a high predicted D2 cell type proportion. **(c)** When examining the *Drd1* and *Drd2* expression of the sorted libraries, the sorted D1 cells show a vastly higher expression of *Drd1* compared to *Drd2*. Conversely, the sorted D2 cells show higher overall expression of *Drd2* compared to *Drd1*. **(d)** mRNA expression heatmap of top genes upregulated in D1 MSNs with cocaine challenge in withdrawal animals (D1 CC), indicating priming of cocaine-responsive genes when compared to acute cocaine (D1 SC). **(e)** mRNA expression heatmap of genes previously shown to be upregulated upon withdrawal from cocaine (*Erich1*, *Hnf4a*, *Hnf4g*) or depressed with cocaine or alcohol addiction in humans (Bex gene family, *Bex1-4*) (62, 63).

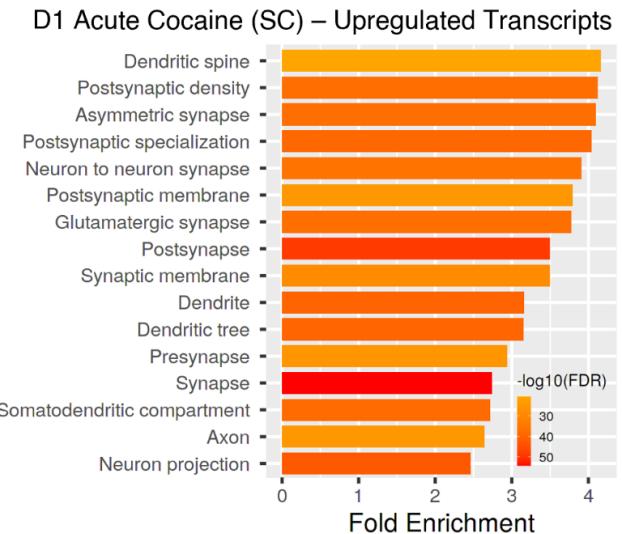


Figure S3. Gene ontology (GO) enrichment of genes that become upregulated with acute cocaine in D1 MSNs using ShinyGO 0.75 (2871 upregulated genes shown in top panel Fig. 1C).

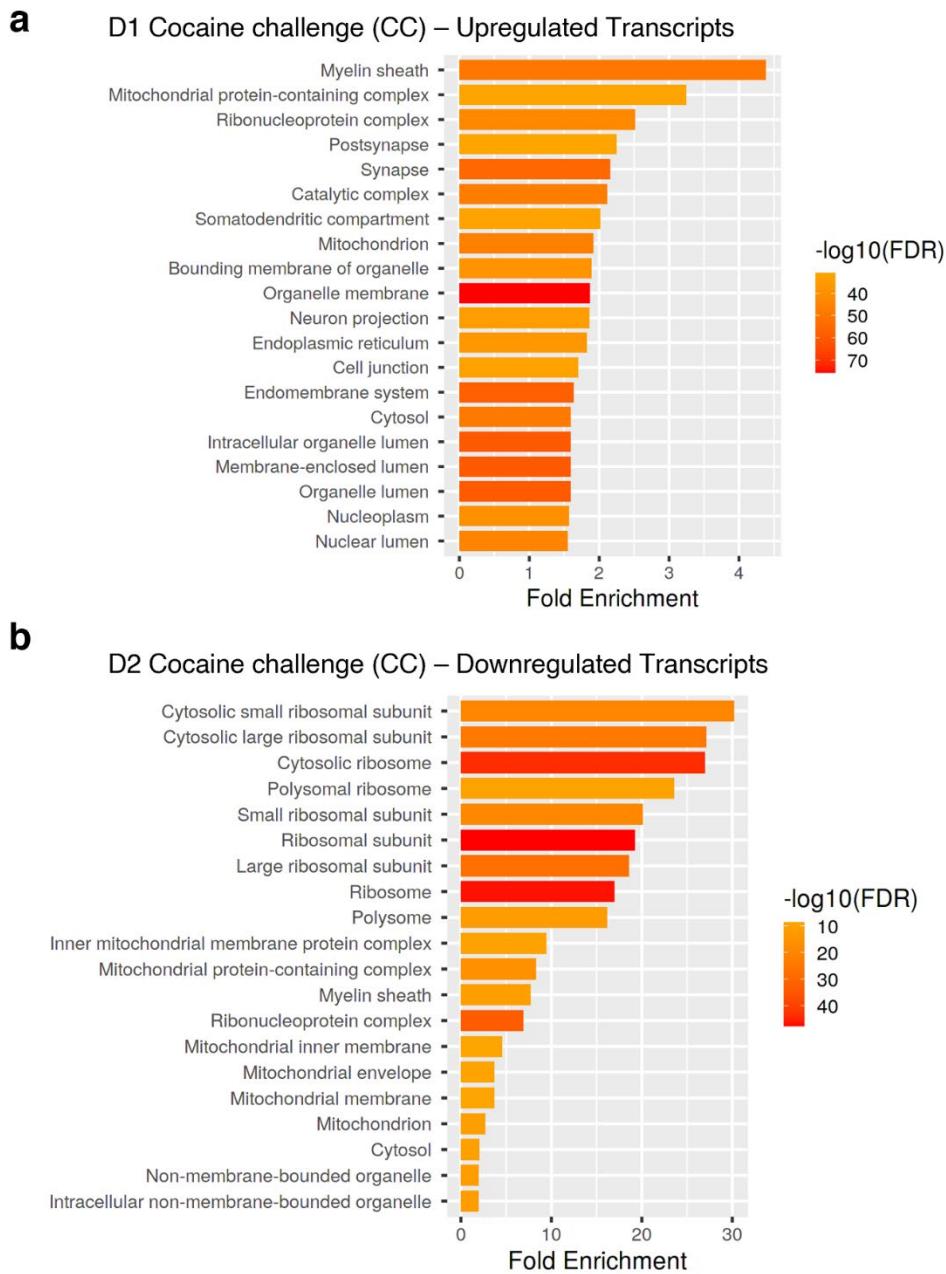


Figure S4. GO term enrichment analysis shows opposite gene regulation in D1 vs. D2 MSNs of NAc with an acute cocaine challenge after prolonged withdrawal from chronic cocaine. **(a)** GO term enrichment of genes that are upregulated in D1 MSNs with cocaine challenge after withdrawal from chronic cocaine using ShinyGO 0.75 (4704 genes shown in lower right panel Fig. 1C). **(b)** GO term enrichment of genes that are downregulated in D2 MSNs with cocaine challenge after withdrawal from chronic cocaine using ShinyGO 0.75 (583 genes shown in lower right panel Fig. 1C).

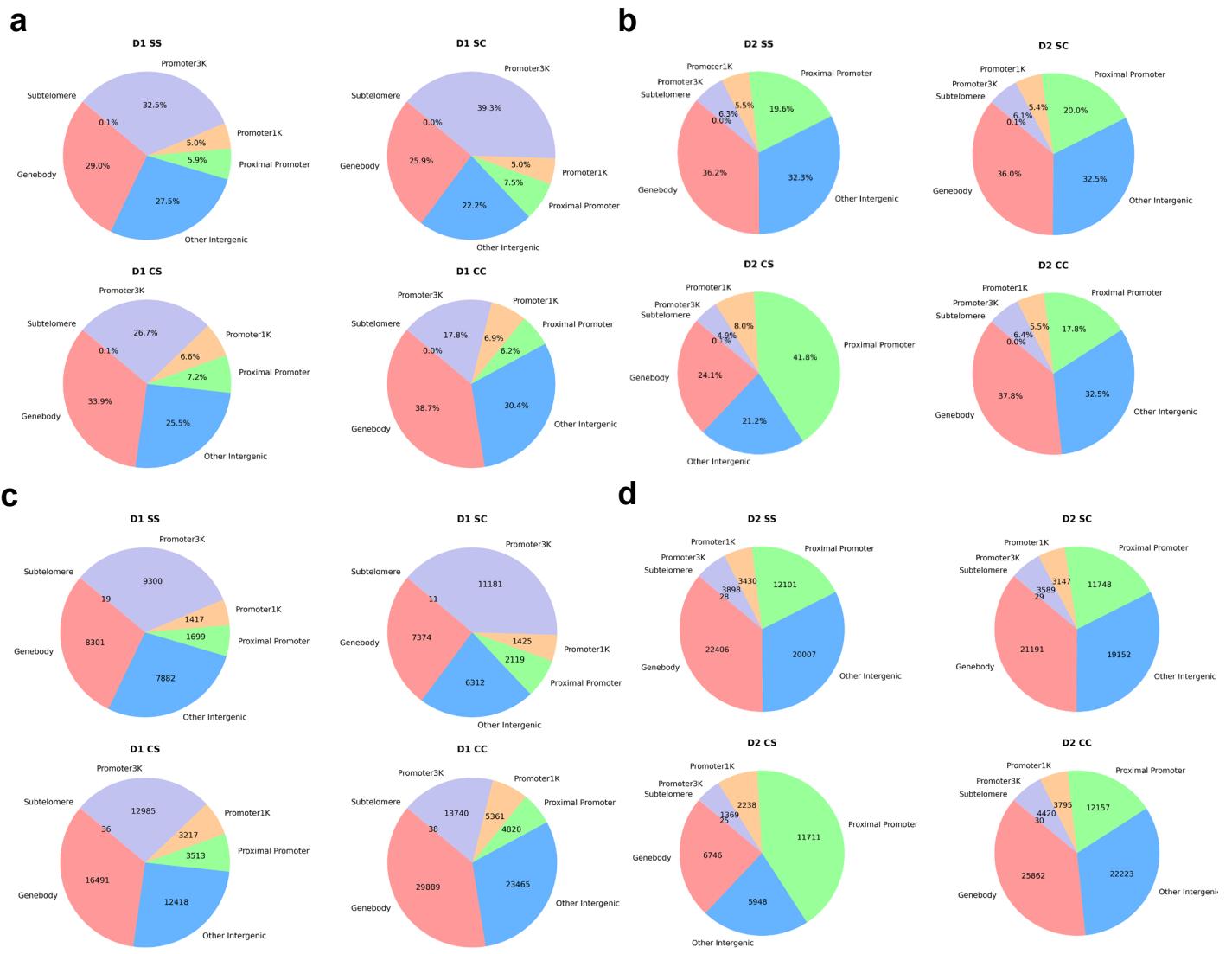


Figure S5. Detailed annotation of ATAC-seq peaks across different conditions as illustrated in pie charts that depict: (a) relative D1 MSN and (b) relative D2 MSN peak distribution across several genomic features, including proximal promoters, longer promoter regions, gene body regions, subtelomeres, and intergenic regions. The respective peak counts across different conditions are shown in (c) for D1 MSNs and (d) for D2 MSNs.

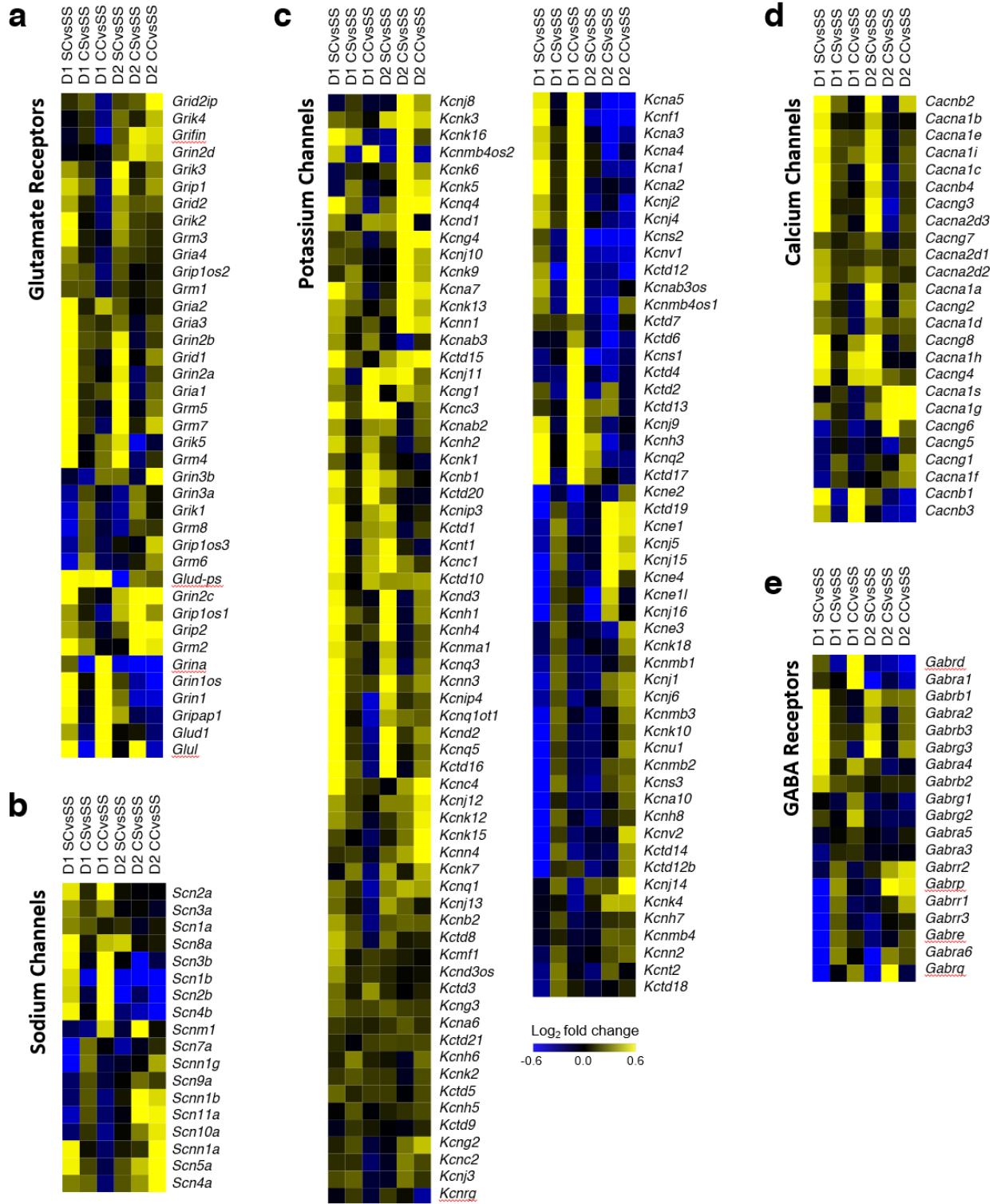


Figure S6. Excitability- and plasticity-related genes exhibit cell-type-specific regulation by cocaine.

Heatmaps show mRNA gene expression of key ionotropic receptor and ion channel subunits upon acute cocaine (SC), prolonged cocaine withdrawal (CS), and cocaine challenge after withdrawal (CC), in both D1 and D2 MSNs of NAc. These plasticity-related transcripts include subunits for (a) glutamate receptors, (b) sodium channels, (c) potassium channels, (d) calcium channels, and (e) GABA receptors.

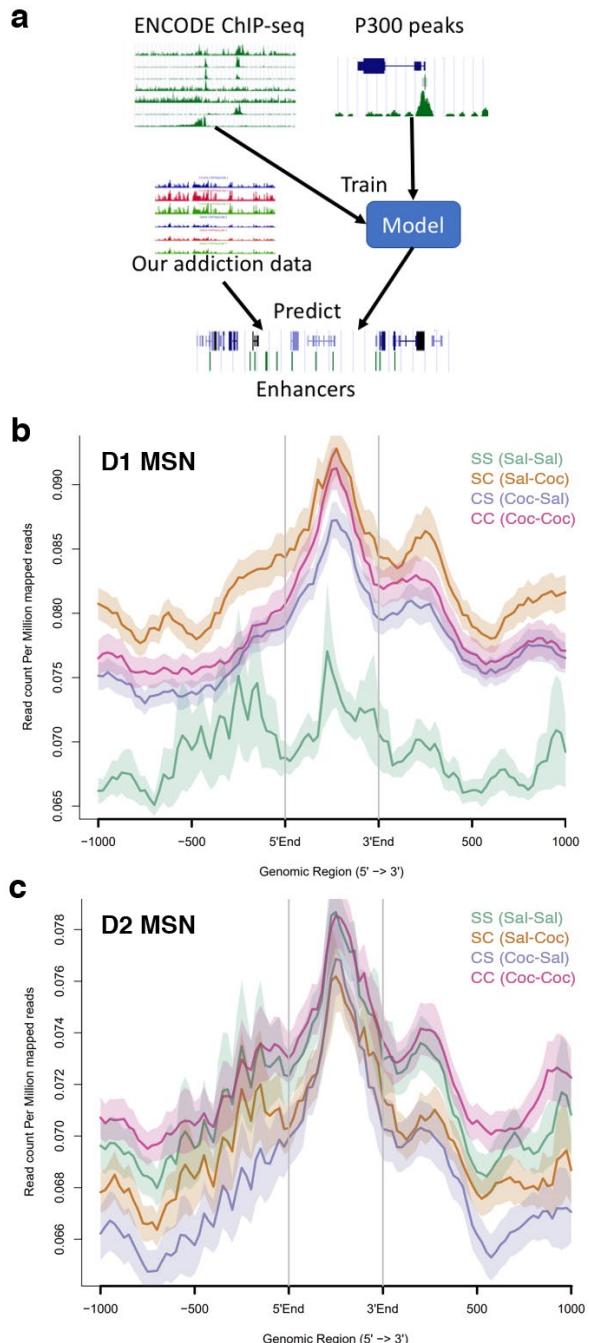


Figure S7. Identification of cocaine-regulated enhancers shows D1-specific accessibility changes. **(a)** Machine learning model, DeepRegFinder (44), identifies putative cocaine-regulated enhancer regions. DeepRegFinder utilizes 1-D convolutional and recurrent neural networks to extract features that represent characteristic chromatin modification patterns of representative enhancers (defined by P300) from ENCODE ChIP-seq data. The trained model was then used to identify enhancers on the whole genome. We used ChIP-seq data from a previous cocaine study (18) to build a model to identify 7,796 cocaine-related enhancers in NAc for this analysis. Accessibility of these 7,796 putative enhancers shown for **(b)** D1 MSNs and **(c)** D2 MSNs in saline control animals (SS), after acute cocaine in drug-naïve mice (SC), after prolonged withdrawal from chronic cocaine (CS), and upon cocaine challenge in withdrawal animals (CC).

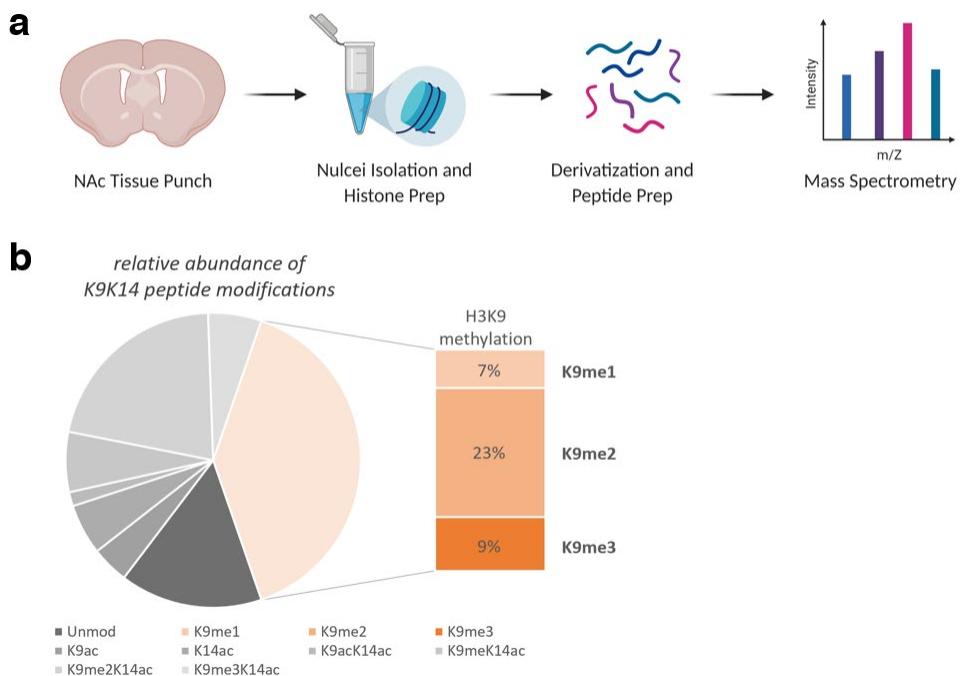


Figure S8. Mass spectrometry reveals changes in chromatin in NAc. **(a)** Experimental outline for histone mass spectrometry starting from NAc tissue punches, which is the starting material for nuclei isolation and histone protein extraction. The prepared histones are first derivatized using propionic anhydride to neutralize charge and block unmodified and monomethylated lysine residues, and are subsequently digested using trypsin, which, under these conditions, cleaves only arginine residues. The generated peptides are then analyzed using online LC-electrospray ionization-tandem mass spectrometry to identify modification sites. **(b)** Relative abundance of H3 lysine 9 (H3K9) and lysine 14 (H3K14) acetylation and methylation (me1/2/3 – mono/di/tri-methylation).

a D1 CC-specific upregulated genes

GO Term	P-value
Regulation of synaptic plasticity	1.43E-12
Anterograde trans-synaptic signaling	2.26E-10
Chemical synaptic transmission	2.26E-10
Trans-synaptic signaling	3.56E-10
Neuron projection morphogenesis	1.51E-08
Neuron projection development	3.11E-08

b D1 CC-specific upregulated genes

Upstream TF Motif	P-value
CREB1	3.62E-05
CTCF	1.53E-03
CEBPB	3.14E-03
EGR1	4.27E-03
NFIC	6.78E-03

Figure S9. **(a)** GO term enrichment analysis of gene sets only induced with cocaine challenge after withdrawal from chronic cocaine, and not with acute cocaine in drug-naïve mice. **(b)** Predicted upstream transcription factors of gene sets only induced with cocaine challenge in withdrawal, and not with acute cocaine in drug-naïve mice.

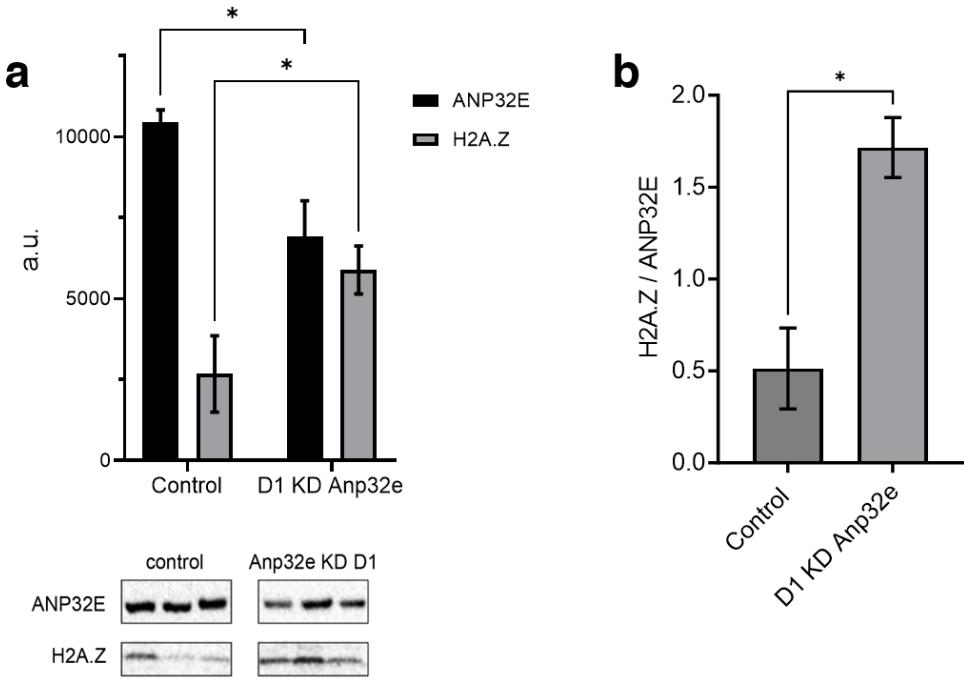


Figure S10. **(a)** Western blot revealed that ANP32E knockdown (KD) in D1 MSNs prevents H2A.Z depletion in the NAc after chronic cocaine exposure (6 days of cocaine following D1 MSN ANP32E KD using Cre-dependent AAV1-siAnp32e delivered to the NAc). Note that all six animals shown in this figure received cocaine treatment (2way ANOVA Interaction $p = 0.0058$; control - Anp32e D1 KD: ANP32E $p = 0.049$, H2A.Z $p = 0.049$, Holm-Šídák's multiple comparisons test). **(b)** Plotted is the relative abundance H2A.Z in control and D1 ANP32E KD animals normalized to ANP32E ($p = 0.012$ unpaired T test). Data are mean \pm s.e.m.

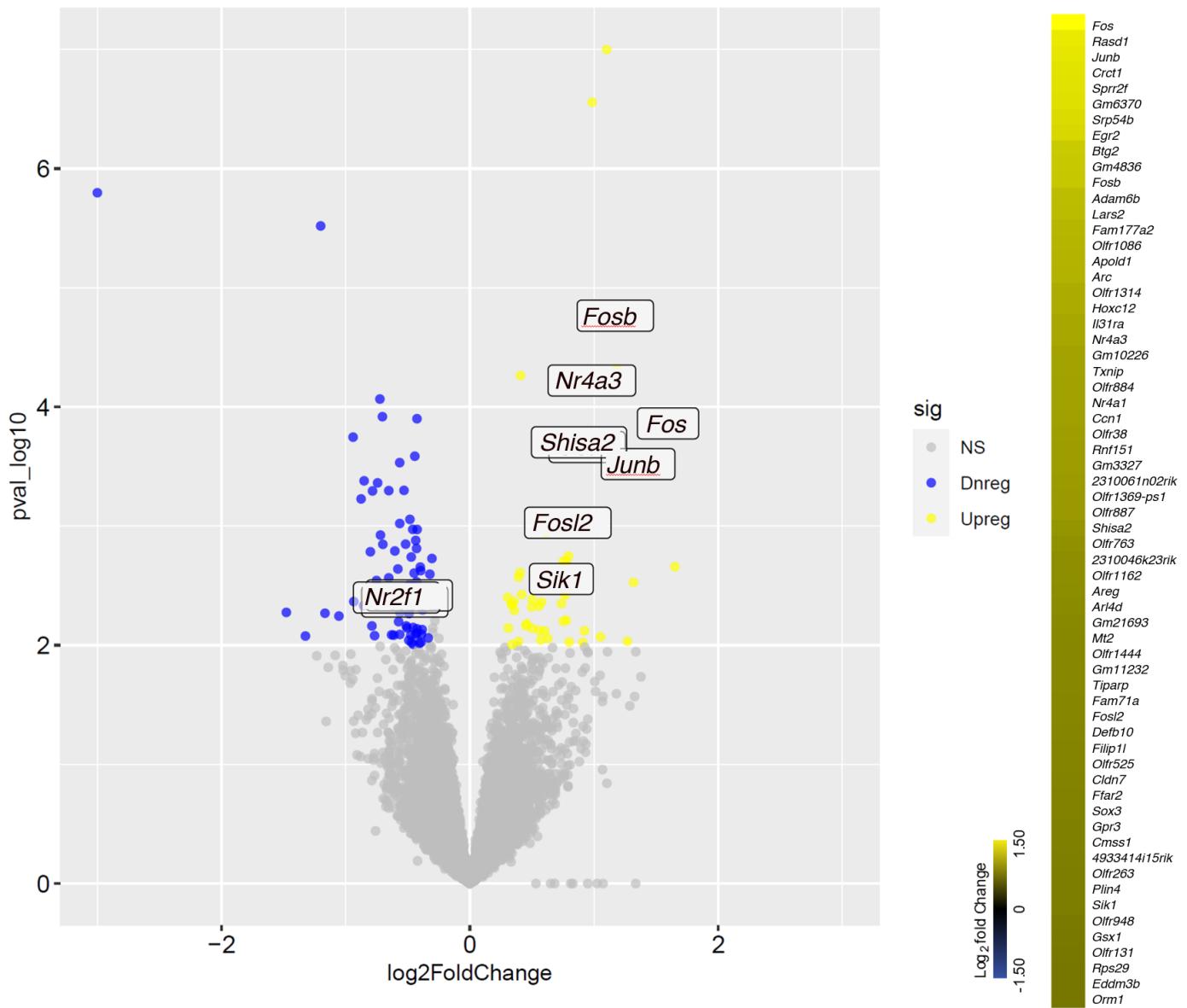


Figure S11. Volcano plot shows protein-coding transcripts that become upregulated (Upreg, yellow) or downregulated (Dnreg, blue) in the NAc upon cocaine challenge injection following prolonged withdrawal from chronic cocaine (CC; whole NAc tissue analyzed for RNAseq). The top genes induced by cocaine challenge encompass well-known immediate early genes (IEGs) such as *Fos*, *Junb*, *FosB*, *Arc*, and *Nr4a*, that have been previously implicated in neural plasticity linked to cocaine action in this brain region.

aD1 cocaine challenge (CC) upregulated transcripts | **Network master regulators**

Network Master Regulator	Type	Predicted Activation	Activation z-score	p-value of overlap
BDNF	growth factor	Activated	5.009	2.12E-20
EGF	growth factor	Activated	8.598	1.85E-19
CAMK1	kinase	Activated	6.591	4.02E-19
RASA2	other	-	-1.584	6.14E-19
EGF	growth factor	Activated	8.391	6.91E-19
CREB1	trxn regulator	-	1.279	1.15E-18
ROR2	kinase	Activated	6.891	3.89E-13

bD1 cocaine challenge (CC) upregulated transcripts | **Upstream chemical regulators**

Upstream Regulator	Molecule Type	Predicted Activation	Activation z-score	p-value of overlap
levodopa	chemical - endogenous	Activated	6.624	6.17E-18
cocaine	chemical drug	Activated	4.539	1.14E-11
haloperidol	chemical drug	Activated	2.645	2.1E-10
kainic acid	chemical toxicant	Activated	3.66	3.25E-09
topotecan	chemical drug	Inhibited	-3.534	9.8E-09

Table S1. Upstream regulators of cocaine-induced gene programs in D1 MSNs of NAc. Network master regulators (**a**) and upstream chemical regulators (**b**) of gene programs that are induced by acute cocaine in D1 MSNs, as revealed by IPA (2871 upregulated genes shown in top panel Fig. 1C).

a

GO Enrichment Genes with increased accessibility in withdrawal (D1 MSNs)		P-value	FDR
regulation of synaptic transmission, glutamatergic		4.06E-07	2.39E-03
calcium ion regulated exocytosis		5.83E-07	1.71E-03
postsynaptic specialization organization		7.70E-07	1.51E-03
postsynaptic density organization		7.70E-07	1.13E-03
postsynaptic density assembly		7.70E-07	9.05E-04
postsynaptic specialization assembly		7.70E-07	7.54E-04
regulation of AMPA receptor activity		7.70E-07	6.46E-04
regulated exocytosis		3.85E-06	2.51E-03
calcium ion transport		5.08E-06	2.98E-03

b

TF name	D1SCvsSS_pvalue	TF name	D1CSvsSS_pvalue	TF name	D1CCvsSS_pvalue
TCFL5	4.63002E-160	ZNF148	5.65836E-172	CTCF	1.49896E-169
KLF15	8.80881E-190	ZNF740	4.34876E-159	EGR2	3.80405E-147
ZBTB14	1.00306E-167	KLF15	3.15907E-162	KLF15	2.23885E-167
NRF1	4.43142E-165	CTCFL	2.74017E-150	ZBTB14	1.31038E-152
SP9	3.86183E-176	MAZ	1.57747E-158	TCFL5	1.07547E-136
HINFP	1.63945E-142	KLF5	3.98330E-172	EGR3	3.70092E-159
TFDP1	7.38029E-158	KLF4	1.72389E-173	NRF1	5.69842E-149
SP3	1.57084E-178	SP9	5.51040E-162	CTCFL	4.06985E-157
HES1	6.06382E-142	KLF2	4.64131E-159	EGR1	8.29278E-170
ZNF148	2.57292E-181	KLF16	1.63022E-165	KLF11	1.42953E-162
SP4	1.33636E-165	EGR1	3.78524E-157	KLF14	1.25149E-153
CTCFL	5.48915E-161	SP3	1.03533E-158	SP3	6.82874E-169
KLF2	3.02082E-170	Plagl1	6.70764E-142	FOSL2::JUN	6.36510E-151
KLF14	1.06983E-162	PLAGL2	2.91769E-120	KLF6	6.44372E-151
KLF3	8.06021E-159	VEZF1	1.24086E-132	KLF3	7.61597E-152
SP1	7.30706E-170	ZIC5	1.76407E-140	FOS::JUND	1.29246E-156
EGR1	4.65410E-164	ZIC3	1.78197E-136	SP4	8.24260E-154
EGR3	1.18135E-154	BHLHE22	6.60473E-139	SP9	1.03807E-160
YY2	2.30903E-138	PLAG1	7.61472E-143	FOS::JUNB	7.95603E-148
KLF5	3.48142E-177	ZBTB14	7.01988E-143	HES1	1.30693E-129
KLF6	1.60169E-160	KLF11	2.86021E-155	SP1	7.33044E-154
Zfx	1.03906E-155	SP8	4.97788E-158	ZIC5	6.19579E-140
ZIC5	1.83773E-147	TFAP2A	2.80698E-131	FOSL2::JUND	5.76016E-147
KLF11	1.03618E-168	TFAP2C	2.07045E-132	FOSB::JUNB	5.57231E-147
KLF16	7.15986E-172	EGR3	3.89172E-143	FOS::JUN	5.99997E-152

Table S2. (a) GO term enrichment analysis of genes that show increased accessibility in D1 MSNs after cocaine withdrawal (CS). (b) TF footprinting analysis using TOBIAS implicates previously known TF dynamics in the response to cocaine exposure, including the AP-1 TF family members FOS, FOSB, JUN, and JUNB.

REFERENCES AND NOTES

1. D. M. Walker, H. M. Cates, E. A. Heller, E. J. Nestler, Regulation of chromatin states by drugs of abuse. *Curr. Opin. Neurobiol.* **30**, 112–121 (2015).
2. M. K. Lobo, E. J. Nestler, The striatal balancing act in drug addiction: Distinct roles of direct and indirect pathway medium spiny neurons. *Front. Neuroanat.* **5**, 41 (2011).
3. A. V Kravitz, L. D. Tye, A. C. Kreitzer, Distinct roles for direct and indirect pathway striatal neurons in reinforcement. *Nat. Neurosci.* **15**, 816–818 (2012).
4. A. F. MacAskill, J. M. Cassel, A. G. Carter, Cocaine exposure reorganizes cell type- and input-specific connectivity in the nucleus accumbens. *Nat. Neurosci.* **17**, 1198–1207 (2014).
5. K.-W. Lee, Y. Kim, A. M. Kim, K. Helmin, A. C. Nairn, P. Greengard, Cocaine-induced dendritic spine formation in D1 and D2 dopamine receptor-containing medium spiny neurons in nucleus accumbens. *Proc. Natl. Acad. Sci. U.S.A.* **103**, 3399–3404 (2006).
6. Z. Luo, N. D. Volkow, N. Heintz, Y. Pan, C. Du, Acute cocaine induces fast activation of d1 receptor and progressive deactivation of d2 receptor striatal neurons: In vivo optical microprobe $[Ca^{2+}]_i$ imaging. *J. Neurosci.* **31**, 13180–13190 (2011).
7. R. J. Smith, M. K. Lobo, S. Spencer, P. W. Kalivas, Cocaine-induced adaptations in D1 and D2 accumbens projection neurons (a dichotomy not necessarily synonymous with direct and indirect pathways). *Curr. Opin. Neurobiol.* **23**, 546–552 (2013).
8. B. A. Grueter, A. J. Robison, R. L. Neve, E. J. Nestler, R. C. Malenka, ΔFosB differentially modulates nucleus accumbens direct and indirect pathway function. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 1923–1928 (2013).
9. L. K. Dobbs, A. R. Kaplan, J. C. Lemos, A. Matsui, M. Rubinstein, V. A. Alvarez, Dopamine regulation of lateral inhibition between striatal neurons gates the stimulant actions of cocaine. *Neuron* **90**, 1100–1113 (2016).

10. E. S. Calipari, R. C. Bagot, I. Purushothaman, T. J. Davidson, J. T. Yorgason, C. J. Peña, D. M. Walker, S. T. Pirpinias, K. G. Guise, C. Ramakrishnan, K. Deisseroth, E. J. Nestler, In vivo imaging identifies temporal signature of D1 and D2 medium spiny neurons in cocaine reward. *Proc. Natl. Acad. Sci. U.S.A.* **113**, 2726–31 (2016).
11. K. E. Savell, J. J. Tuscher, M. E. Zipperly, C. G. Duke, R. A. Phillips, A. J. Bauman, S. Thukral, F. A. Sultan, N. A. Goska, L. Ianov, J. J. Day, A dopamine-induced gene expression signature regulates neuronal function and cocaine response. *Sci. Adv.* **6**, eaba4221 (2020).
12. R. Chandra, T. C. Francis, P. Konkalmatt, A. Amgalan, A. M. Gancarz, D. M. Dietz, M. K. Lobo, Opposing role for Egr3 in nucleus accumbens cell subtypes in cocaine action. *J. Neurosci.* **35**, 7927–7937 (2015).
13. C. A. McClung, E. J. Nestler, Regulation of gene expression and cocaine reward by CREB and ΔFosB. *Nat. Neurosci.* **6**, 1208–1215 (2003).
14. P. Mews, E. S. Calipari, J. Day, M. K. Lobo, T. Bredy, T. Abel, From circuits to chromatin: The emerging role of epigenetics in mental health. *J. Neurosci.* **41**, 873–882 (2021).
15. R. Chen, T. R. Blosser, M. N. Djekidel, J. Hao, A. Bhattacherjee, W. Chen, L. M. Tuesta, X. Zhuang, Y. Zhang, Decoding molecular and cellular heterogeneity of mouse nucleus accumbens. *Nat. Neurosci.* **24**, 1757–1771 (2021).
16. P. Mews, L. Sosnick, A. Gurung, S. Sidoli, E. J. Nestler, Decoding cocaine-induced proteomic adaptations in the mouse nucleus accumbens. *Sci. Signal.* **17**, eadl4738 (2024).
17. I. Maze, H. E. Covington, D. M. Dietz, Q. Laplant, W. Renthal, S. J. Russo, M. Mechanic, E. Mouzon, R. L. Neve, S. J. Haggarty, Y. Ren, S. C. Sampath, Y. L. Hurd, P. Greengard, A. Tarakhovsky, A. Schaefer, E. J. Nestler, Essential role of the histone methyltransferase G9a in cocaine-induced plasticity. *Science* **327**, 213–216 (2010).
18. J. Feng, M. Wilkinson, X. Liu, I. Purushothaman, D. Ferguson, V. Vialou, I. Maze, N. Shao, P. Kennedy, J. Koo, C. Dias, B. Laitman, V. Stockman, Q. LaPlant, M. E. Cahill, E. J. Nestler, L.

- Shen, Chronic cocaine-regulated epigenomic changes in mouse nucleus accumbens. *Genome Biol.* **15**, 1–18 (2014).
19. A. J. López, A. R. Johnson, T. J. Euston, R. Wilson, S. O. Nolan, L. J. Brady, K. C. Thibeault, S. J. Kelly, V. Kondev, P. Melugin, M. G. Kutlu, E. Chuang, T. K. T. Lam, D. D. Kiraly, E. S. Calipari, Cocaine self-administration induces sex-dependent protein expression in the nucleus accumbens. *Commun. Biol.* **4**, 883 (2021).
20. G. A. Rogge, M. A. Wood, The role of histone acetylation in cocaine-induced neural plasticity and behavior. *Neuropsychopharmacology* **38**, 94–110 (2013).
21. M. D. Carpenter, Q. Hu, A. M. Bond, S. I. Lombroso, K. S. Czarnecki, C. J. Lim, H. Song, M. E. Wimmer, R. C. Pierce, E. A. Heller, *Nr4a1* suppresses cocaine-induced behavior via epigenetic regulation of homeostatic target genes. *Nat. Commun.* **11**, 504 (2020).
22. M. E. Wimmer, B. Fant, S. E. Swinford-Jackson, A. Testino, D. Van Nest, T. Abel, R. C. Pierce, H3.3 barcoding of nucleus accumbens transcriptional activity identifies novel molecular cascades associated with cocaine self-administration in mice. *J. Neurosci.* **39**, 5247–5254 (2019).
23. I. Maze, J. Feng, M. B. Wilkinson, H. Sun, L. Shen, E. J. Nestler, Cocaine dynamically regulates heterochromatin and repetitive element unsilencing in nucleus accumbens. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 3035–3040 (2011).
24. Z.-J. Wang, J. A. Martin, L. E. Mueller, A. Caccamise, C. T. Werner, R. L. Neve, A. M. Gancarz, J.-X. Li, D. M. Dietz, BRG1 in the nucleus accumbens regulates cocaine-seeking behavior. *Biol. Psychiatry* **80**, 652–660 (2016).
25. D. M. Damez-Werno, H. Sun, K. N. Scobie, N. Shao, J. Rabkin, C. Dias, E. S. Calipari, I. Maze, C. J. Pena, D. M. Walker, M. E. Cahill, R. Chandra, A. Gancarz, E. Mouzon, J. A. Landry, H. Cates, M.-K. Lobo, D. Dietz, C. D. Allis, E. Guccione, G. Turecki, P. Defilippi, R. L. Neve, Y. L. Hurd, L. Shen, E. J. Nestler, Histone arginine methylation in cocaine action in the nucleus accumbens. *Proc. Natl. Acad. Sci. U.S.A.* **113**, 9623–8 (2016).

26. R. M. Bastle, I. S. Maze, Chromatin regulation in complex brain disorders. *Curr. Opin. Behav. Sci.* **25**, 57–65 (2019).
27. P. Mews, E. S. Calipari, Cross-talk between the epigenome and neural circuits in drug addiction. *Prog. Brain Res.* **235**, 19–63 (2017).
28. P. Mews, G. Donahue, A. M. Drake, V. Luczak, T. Abel, S. L. Berger, Acetyl-CoA synthetase regulates histone acetylation and hippocampal memory. *Nature* **546**, 381–386 (2017).
29. S. Sidoli, N. V. Bhanu, K. R. Karch, X. Wang, B. A. Garcia, Complete workflow for analysis of histone post-translational modifications using bottom-up mass spectrometry: From histone extraction to data analysis. *J. Vis. Exp.* **17**, e54112 (2016).
30. I. B. Zovkic, Epigenetics and memory: An expanded role for chromatin dynamics. *Curr. Opin. Neurobiol.* **67**, 58–65 (2021).
31. C. M. Hammond, C. B. Strømme, H. Huang, D. J. Patel, A. Groth, Histone chaperone networks shaping chromatin function. *Nat. Rev. Mol. Cell Biol.* **18**, 141–158 (2017).
32. A. Obri, K. Ouararhni, C. Papin, M. L. Diebold, K. Padmanabhan, M. Marek, I. Stoll, L. Roy, P. T. Reilly, T. W. Mak, S. Dimitrov, C. Romier, A. Hamiche, ANP32E is a histone chaperone that removes H2A.Z from chromatin. *Nature* **505**, 648–653 (2014).
33. D. L. Graham, S. Edwards, R. K. Bachtell, R. J. DiLeone, M. Rios, D. W. Self, Dynamic BDNF activity in nucleus accumbens with cocaine use increases self-administration and relapse. *Nat. Neurosci.* **10**, 1029–1037 (2007).
34. M. K. Lobo, H. E. Covington, D. Chaudhury, A. K. Friedman, H. Sun, D. Damez-Werno, D. M. Dietz, S. Zaman, J. W. Koo, P. J. Kennedy, E. Mouzon, M. Mogri, R. L. Neve, K. Deisseroth, M.-H. Han, E. J. Nestler, Cell type-specific loss of BDNF signaling mimics optogenetic control of cocaine reward. *Science* **330**, 385–90 (2010).
35. G. Schoenbaum, T. A. Stalnaker, Y. Shaham, A role for BDNF in cocaine reward and relapse. *Nat. Neurosci.* **10**, 935–936 (2007).

36. M. G. Murer, R. Moratalla, Striatal signaling in L-DOPA-induced dyskinesia: Common mechanisms with drug abuse and long term memory involving D1 dopamine receptor stimulation. *Front. Neuroanat.* **5**, 51 (2011).
37. H. Kronman, F. Richter, B. Labonté, R. Chandra, S. Zhao, G. Hoffman, M. K. Lobo, E. E. Schadt, E. J. Nestler, Biology and bias in cell type-specific RNAseq of nucleus accumbens medium spiny neurons. *Sci. Rep.* **9**, 8350 (2019).
38. M. K. Lobo, S. Zaman, D. M. Damez-Werno, J. W. Koo, R. C. Bagot, J. A. DiNieri, A. Nugent, E. Finkel, D. Chaudhury, R. Chandra, E. Riberio, J. Rabkin, E. Mouzon, R. Cachope, J. F. Cheer, M.-H. Han, D. M. Dietz, D. W. Self, Y. L. Hurd, V. Vialou, E. J. Nestler, ΔFosB induction in striatal medium spiny neuron subtypes in response to chronic pharmacological, emotional, and optogenetic stimuli. *J. Neurosci.* **33**, 18381–18395 (2013).
39. E. J. Nestler, M. Barrot, D. W. Self, ΔFosB: A sustained molecular switch for addiction. *Proc. Natl. Acad. Sci. U.S.A.* **98**, 11042–11046 (2001).
40. Y. Dong, E. J. Nestler, The neural rejuvenation hypothesis of cocaine addiction. *Trends Pharmacol. Sci.* **35**, 374–383 (2014).
41. D. J. Surmeier, J. Ding, M. Day, Z. Wang, W. Shen, D1 and D2 dopamine-receptor modulation of striatal glutamatergic signaling in striatal medium spiny neurons. *Trends Neurosci.* **30**, 228–235 (2007).
42. K. L. Conrad, K. Y. Tseng, J. L. Uejima, J. M. Reimers, L.-J. Heng, Y. Shaham, M. Marinelli, M. E. Wolf, Formation of accumbens GluR2-lacking AMPA receptors mediates incubation of cocaine craving. *Nature* **454**, 118–121 (2008).
43. D. A. Gallegos, M. Minto, F. Liu, M. F. Hazlett, S. Aryana Yousefzadeh, L. C. Bartelt, A. E. West, Cell-type specific transcriptional adaptations of nucleus accumbens interneurons to amphetamine. *Mol. Psychiatry* **28**, 3414–3428 (2023).
44. A. Ramakrishnan, G. Wangensteen, S. Kim, E. J. Nestler, L. Shen, DeepRegFinder: Deep learning-based regulatory elements finder. *Bioinform. Adv.* **4**, vbae007 (2024).

45. D. M. Walker, H. M. Cates, Y.-H. E. Loh, I. Purushothaman, A. Ramakrishnan, K. M. Cahill, C. K. Lardner, A. Godino, H. G. Kronman, J. Rabkin, Z. S. Lorsch, P. Mews, M. A. Doyle, J. Feng, B. Labonté, J. W. Koo, R. C. Bagot, R. W. Logan, M. L. Seney, E. S. Calipari, L. Shen, E. J. Nestler, Cocaine self-administration alters transcriptome-wide responses in the brain's reward circuitry. *Biol. Psychiatry* **84**, 867–880 (2018).
46. H. M. Cates, E. A. Heller, C. K. Lardner, I. Purushothaman, C. J. Peña, D. M. Walker, M. E. Cahill, R. L. Neve, L. Shen, R. C. Bagot, E. J. Nestler, Transcription factor E2F3a in nucleus accumbens affects cocaine action via transcription and alternative splicing. *Biol. Psychiatry* **84**, 167–179 (2018).
47. I. Maze, K.-M. Noh, A. Soshnev, C. D. Allis, Every amino acid matters: Essential contributions of histone variants to mammalian development and disease. *Nat. Rev. Genet.* **15**, 259–271 (2014).
48. G. Stefanelli, A. B. Azam, B. J. Walters, M. A. Brimble, C. P. Gettens, P. Bouchard-Cannon, H.-Y. M. Cheng, A. M. Davidoff, K. Narkaj, J. J. Day, A. J. Kennedy, I. B. Zovkic, Learning and age-related changes in genome-wide H2A.Z binding in the mouse hippocampus. *Cell Rep.* **22**, 1124–1131 (2018).
49. L. Cole, S. Kurscheid, M. Nekrasov, R. Domaschenz, D. L. Vera, J. H. Dennis, D. J. Tremethick, Multiple roles of H2A.Z in regulating promoter chromatin architecture in human cells. *Nat. Commun.* **12**, 2524 (2021).
50. A. J. Robison, E. J. Nestler, Transcriptional and epigenetic mechanisms of addiction. *Nat. Rev. Neurosci.* **12**, 623–637 (2011).
51. R. Chandra, M. K. Lobo, Beyond neuronal activity markers: Select immediate early genes in striatal neuron subtypes functionally mediate psychostimulant addiction. *Front. Behav. Neurosci.* **11**, 112 (2017).
52. E. B. Larson, D. L. Graham, R. R. Arzaga, N. Buzin, J. Webb, T. A. Green, C. E. Bass, R. L. Neve, E. F. Terwilliger, E. J. Nestler, D. W. Self, Overexpression of CREB in the nucleus

- accumbens shell increases cocaine reinforcement in self-administering rats. *J. Neurosci.* **31**, 16447–16457 (2011).
53. C. D. Teague, E. J. Nestler, Key transcription factors mediating cocaine-induced plasticity in the nucleus accumbens. *Mol. Psychiatry* **27**, 687–709 (2022).
54. J. Gräff, L. H. Tsai, Histone acetylation: Molecular mnemonics on the chromatin. *Nat. Rev. Neurosci.* **14**, 97–111 (2013).
55. A. Marco, H. S. Meharena, V. Dileep, R. M. Raju, J. Davila-Velderrain, A. L. Zhang, C. Adaikkan, J. Z. Young, F. Gao, M. Kellis, L.-H. Tsai, Mapping the epigenomic and transcriptomic interplay during memory formation and recall in the hippocampal engram ensemble. *Nat. Neurosci.* **23**, 1606–1617 (2020).
56. J. W. Muschamp, C. L. Nemeth, A. J. Robison, E. J. Nestler, W. A. Carlezon, ΔFosB enhances the rewarding effects of cocaine while reducing the pro-depressive effects of the kappa-opioid receptor agonist U50488. *Biol. Psychiatry* **71**, 44–50 (2012).
57. J. Zhang, L. Zhang, H. Jiao, Q. Zhang, D. Zhang, D. Lou, J. L. Katz, M. Xu, c-Fos facilitates the acquisition and extinction of cocaine-induced persistent changes. *J. Neurosci.* **26**, 13287–13296 (2006).
58. I. B. Zovkic, B. S. Paulukaitis, J. J. Day, D. M. Etikala, J. D. Sweatt, Histone H2A.Z subunit exchange controls consolidation of recent and remote memory. *Nature* **515**, 582–586 (2014).
59. B. D. Giaimo, F. Ferrante, A. Herchenröther, S. B. Hake, T. Borggrefe, The histone variant H2A.Z in gene regulation. *Epigenetics Chromatin* **12**, 37 (2019).
60. K. Narkaj, G. Stefanelli, M. Wahdan, A. B. Azam, F. Ramzan, C. F. D. Steininger, B. J. Walters, I. B. Zovkic, Blocking H2A.Z incorporation via tip60 inhibition promotes systems consolidation of fear memory in mice. *eNeuro* **5**, ENEURO.0378-18.2018 (2018).
61. I. Maze, D. Chaudhury, D. M. Dietz, M. Von Schimmelmann, P. J. Kennedy, M. K. Lobo, S. E. Sillivan, M. L. Miller, R. C. Bagot, H. Sun, G. Turecki, R. L. Neve, Y. L. Hurd, L. Shen, M.-H.

- Han, A. Schaefer, E. J. Nestler, G9a influences neuronal subtype specification in striatum. *Nat. Neurosci.* **17**, 533–539 (2014).
62. D. Kim, J. M. Paggi, C. Park, C. Bennett, S. L. Salzberg, Graph-based genome alignment and genotyping with HISAT2 and HISAT-genotype. *Nat. Biotechnol.* **37**, 907–915 (2019).
63. H. Li, B. Handsaker, A. Wysoker, T. Fennell, J. Ruan, N. Homer, G. Marth, G. Abecasis, R. Durbin, The sequence alignment/map format and SAMtools. *Bioinformatics* **25**, 2078–2079 (2009).
64. Y. Zhang, T. Liu, C. A. Meyer, J. Eeckhoute, D. S. Johnson, B. E. Bernstein, C. Nussbaum, R. M. Myers, M. Brown, W. Li, X. S. Shirley, Model-based analysis of ChIP-Seq (MACS). *Genome Biol.* **9**, R137 (2008).
65. L. Shen, N. Shao, X. Liu, E. Nestler, Ngs.plot: Quick mining and visualization of next-generation sequencing data by integrating genomic databases. *BMC Genomics* **15**, 284 (2014).
66. J. D. Buenrostro, P. G. Giresi, L. C. Zaba, H. Y. Chang, W. J. Greenleaf, Transposition of native chromatin for fast and sensitive epigenomic profiling of open chromatin, DNA-binding proteins and nucleosome position. *Nat. Methods* **10**, 1213–1218 (2013).
67. M. Martin, Cutadapt removes adapter sequences from high-throughput sequencing reads. *EMBnet J.* **17**, 10–12 (2011).
68. L. Shen, N. Y. Shao, X. Liu, I. Maze, J. Feng, E. J. Nestler, diffReps: Detecting differential chromatin modification sites from chip-seq data with biological replicates. *PLOS One* **8**, e65598 (2013).
69. M. Bentsen, P. Goymann, H. Schultheis, K. Klee, A. Petrova, R. Wiegandt, A. Fust, J. Preussner, C. Kuenne, T. Braun, J. Kim, M. Looso, ATAC-seq footprinting unravels kinetics of transcription factor binding during zygotic genome activation. *Nat. Commun.* **11**, 4267 (2020).
70. D. Charytonowicz, R. Brody, R. Sebra, Interpretable and context-free deconvolution of multi-scale whole transcriptomic data with UniCell deconvolve. *Nat. Commun.* **14**, 1350(2023).

71. P. Mews, A. M. Cunningham, J. Scarpa, A. Ramakrishnan, E. M. Hicks, S. Bolnick, S. Garamszegi, L. Shen, D. C. Mash, E. J. Nestler, Convergent abnormalities in striatal gene networks in human cocaine use disorder and mouse cocaine administration models. *Sci. Adv.* **9**, eadd8946 (2023).
72. Z. Zhou, Q. Yuan, D. C. Mash, D. Goldman, Substance-specific and shared transcription and epigenetic changes in the human hippocampus chronically exposed to cocaine and alcohol. *Proc. Natl. Acad. Sci. U.S.A.* **108**, 6626–6631 (2011).