

1 **Supplemental information**

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3 **Supplemental Figure 1.** (A) Gating strategy used in flow cytometry to detect TFH and TFR  
4 cells after PCT sensitization. Foxp3 expression was assessed by intracellular staining. (B)  
5 Absolute numbers of *Ii4*-GFP<sup>+</sup> TFH versus TFR cells in mesenteric lymph node (mLN), from  
6 gating in A with *Ii4*-GFP<sup>+</sup> mice and calculation of cell numbers from total mLN cells. n = 5. \*\*\*\* P  
7 < 0.0001 by Students T-test. Data were representative of two independent experiments.

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9 **Supplemental Figure 2.** (A) Gating strategy used in flow cytometry to measure *Ii4*-GFP-  
10 expressing Treg cell populations in 4Get mice, using the same gating approach as in Sup. Fig.  
11 1. CD4<sup>+</sup>Foxp3<sup>+</sup> Treg cells were divided into four groups based on CXCR5 and PD-1 expression.  
12 *Ii4*-GFP<sup>+</sup> cells were measured in these four sub-populations. (B) Percentages of *Ii4*-GFP<sup>+</sup> cells  
13 in pre-TFR (CXCR5<sup>+</sup>PD-1<sup>-</sup>) and TFR (CXCR5<sup>high</sup>PD-1<sup>high</sup>) populations in naive mice, mice with  
14 SRBC immunizations, and mice sensitized with PCT. (C) *Ii4* mRNA expression in TFH cells and  
15 TFR cells after PCT sensitization. Mice were sensitized either with PCT at day 1 and cells  
16 isolated by FACS at day 8 for RNA preparation or PCT at day 1 and day 8, and cells isolated by  
17 FACS at day 12 for RNA preparation for RNA preparation. n =4. (D) Percentages of *Ii4*-GFP<sup>+</sup>  
18 cells in TFR (CXCR5<sup>high</sup>PD-1<sup>high</sup>) cells in different lymphoid organs from naive mice. "other LNs"  
19 is inguinal, popliteal and sublingual LNs. \* P < 0.05, \*\* P < 0.01, \*\*\* P < 0.001, \*\*\*\* P < 0.0001,  
20 ns not significant by ANOVA. (A-B) n = 8. (C-D) n=4.

21

22 **Supplemental Figure 3.** Heatmap of *Ii4* regulating genes, comparing bulk RNAseq of TFH and  
23 TFR cells from day 1 and day 8 PCT sensitized mice.

24

25 **Supplemental Figure 4.** (A) Bone marrow chimera (BMC) design and experimental flow, where  
26 CD45 congenically marked mice could distinguish between the two types of input bone marrow.  
27 Irradiated Rag1-KO mice received equal amount of BM cells from BoyJ (CD45.1) and Bcl6-flox  
28 or Bcl6 cKO mice. Eight weeks later, the mice were primed for peanut allergy with PCT and  
29 donor derived TFH and TFR cells percentages were measured by flow cytometry. (B) The  
30 contribution of each type of donor to the TFH and TFR cell compartments for the two types of  
31 BMC. (C) Percentages of TFH and TFR cells in the two types of BMC. (D) PN-specific IgE  
32 levels in the two types of BMC at day 15 of the PCT response.

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34 **Supplemental Figure 5.** (A-D) WT mice and *Il4*<sup>-/-</sup> mice were sensitized with PCT. The percent  
35 of TFH cells (A), TFR cells (B), GC B cells (C), and IgE<sup>+</sup> GC B cells (D) were analyzed in the  
36 spleen and mLN using flow cytometry. (E) Ab response in mice immunized with OVA plus Alum.  
37 OVA-specific IgE, OVA-specific IgG1, and total IgE in WT, *Il4*<sup>+/-</sup>, and *Il4*<sup>-/-</sup> mice after  
38 immunization with OVA plus Alum. Mice were immunized with OVA plus Alum at day 1 and day  
39 8. Sera were collected at day 15, and Abs were tested using ELISA. n = 8. Data were  
40 representative of two independent experiments. (F-H) TFH, TFR, and GC B cells were  
41 unaffected by anti-IL-4R $\alpha$  blocking or *Il4* gene knockout. Mice were injected with anti-IL-4R $\alpha$   
42 Abs. Mice were also sensitized with PCT. The percent and cell number of TFH cells (F), TFR  
43 cells (G), and GC B cells (H) in the spleen and mLN were analyzed using flow cytometry. n = 7–  
44 9. n = 5. \* P < 0.05, \*\* P < 0.01, \*\*\* P < 0.001, \*\*\*\* P < 0.0001, ns not significant by t-test (A-H).  
45 Data were representative of two or three independent experiments.

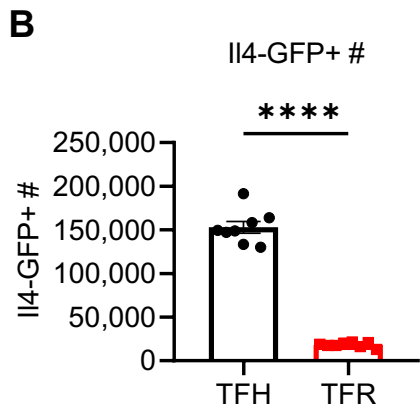
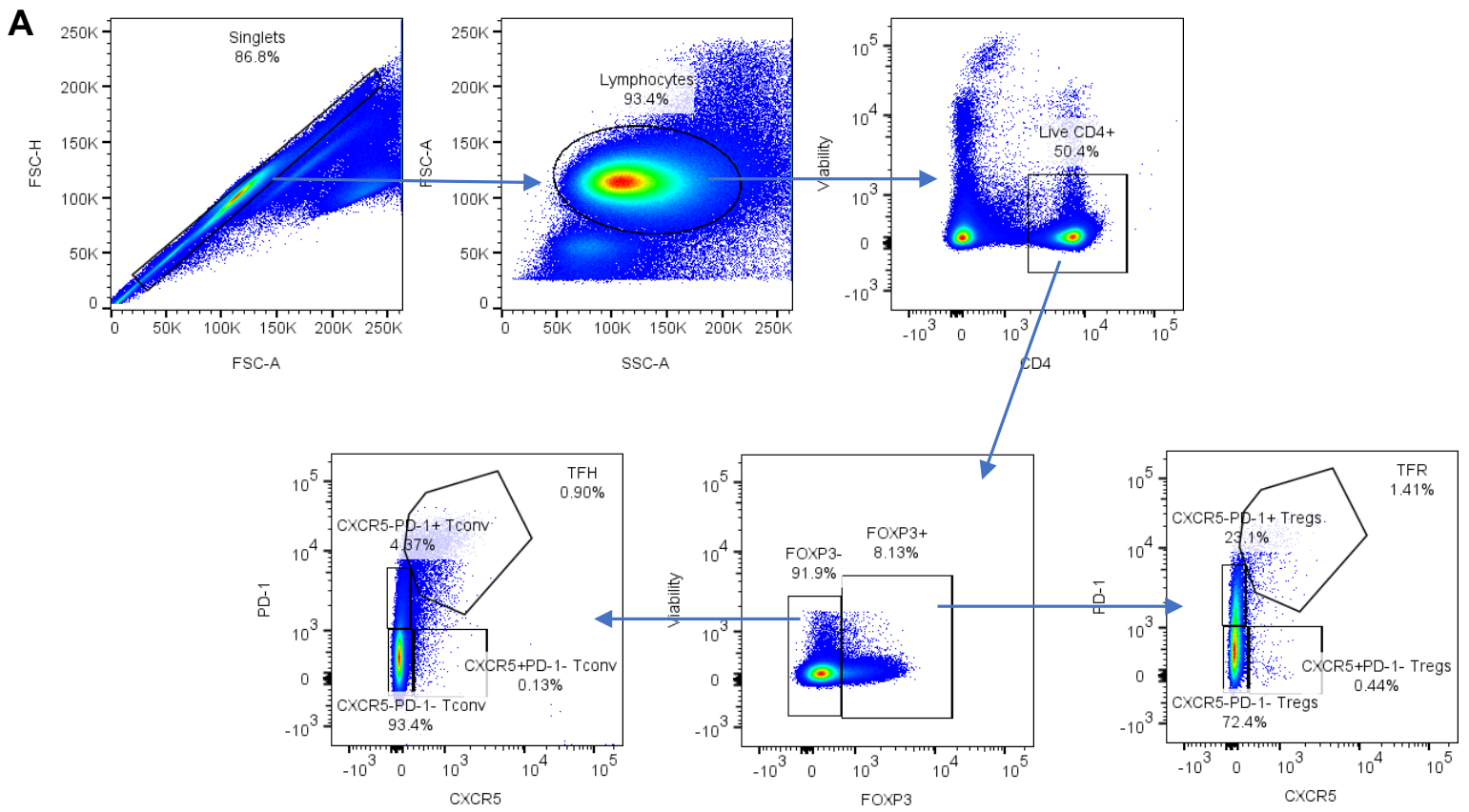
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47 **Supplemental Figure 6.** (A) The expression of IL-4 and IL-10 in TFH and TFR cells after PCT  
48 sensitizations. The percentages of IL-4<sup>-</sup>IL-10<sup>-</sup>, IL-4<sup>+</sup>IL-10<sup>-</sup>, IL-4<sup>-</sup>IL-10<sup>+</sup>, and IL-4<sup>+</sup>IL-10<sup>+</sup> TFH and  
49 TFR cells were measured via flow cytometry after cell fixation, permeabilization and intracellular  
50 cytokine staining. (B) Heatmap of Th2-related genes in TFR cells from HDM allergic airway  
51 primed mice. TFR cells were sorted from the mediastinal LNs of naïve mice and mice after  
52 intranasal HDM challenges. Total RNA was extracted and subjected to bulk RNA-seq and  
53 analysis. The heatmap represents log<sub>2</sub>FPKM values.

54

55 **Supplemental Figure 7.** (A) Image shows frozen section of mLN at 7 days after PCT  
56 sensitization after immunofluorescent staining. Red – Bcl6, Blue – CD21/35, Green – Foxp3.  
57 The dark zone of GC stains intensely for Red Bcl6<sup>+</sup> B cells, while the GC light zone will have  
58 blue CD21/CD35<sup>+</sup> FDCs. TFH cells should be Red with a white border and TFR cells should be  
59 red/green/yellow with a white border. Yellow arrows show likely TFH cells. Red and green arrow  
60 shows likely TFR cell. GCs found in the mLN after PCT priming were relatively small when  
61 compared to splenic GCs after i.p. immunization. (B) CD38 expression on Treg cells, TFR cells,  
62 and TFH cells after PCT sensitization, assayed by flow cytometry. \*\* P < 0.01, \*\*\* P < 0.001, ns  
63 not significant by one-way ANOVA. n = 3.

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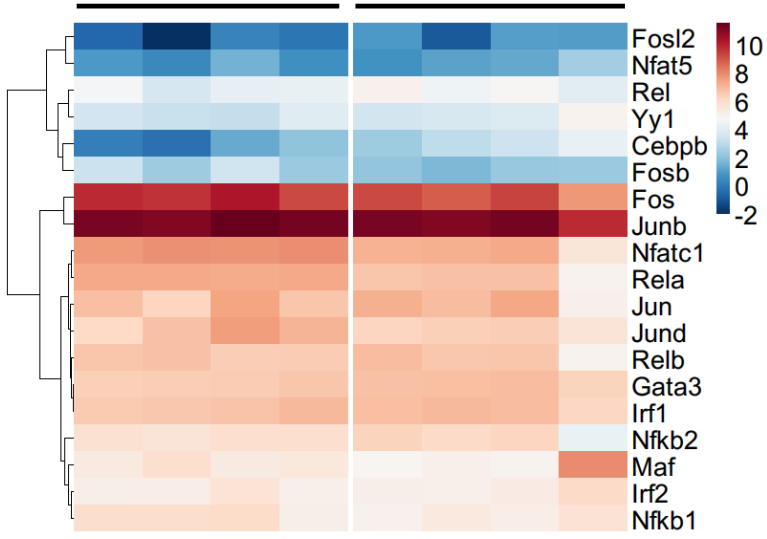


Sup. Fig. 1

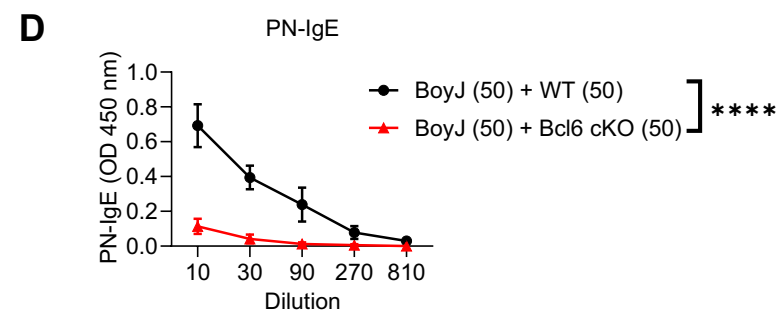
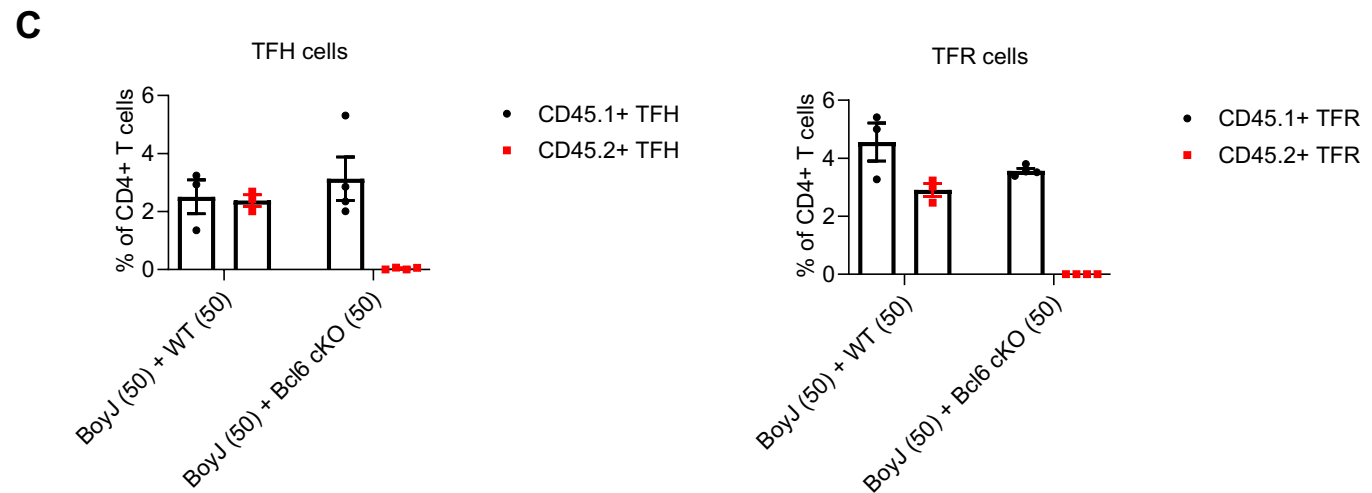
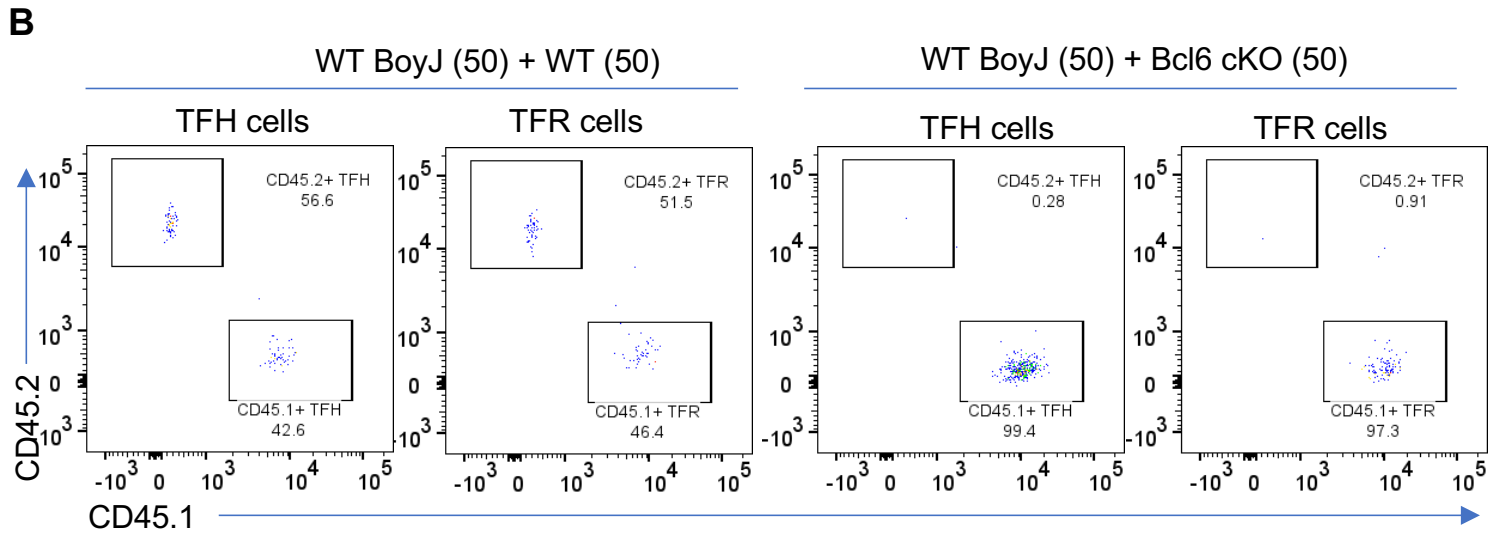
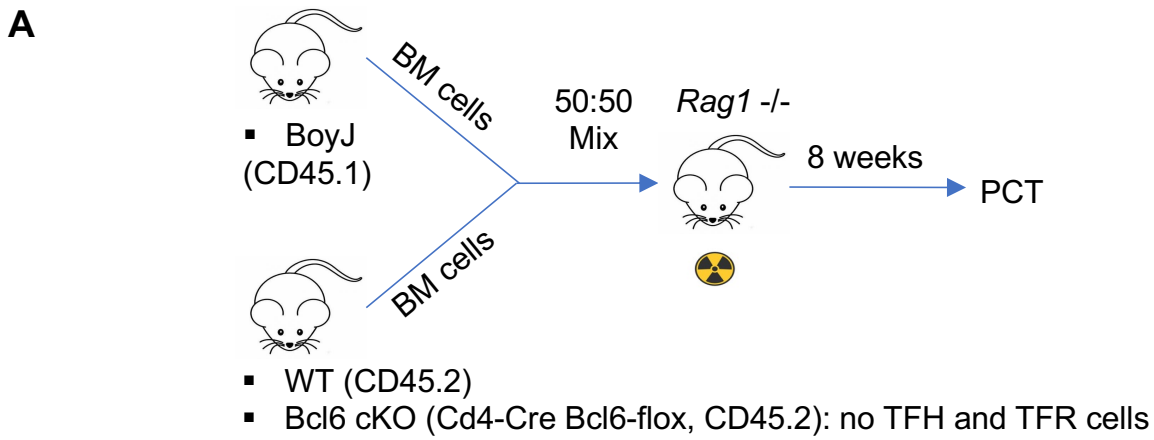


*Ii4* regulation

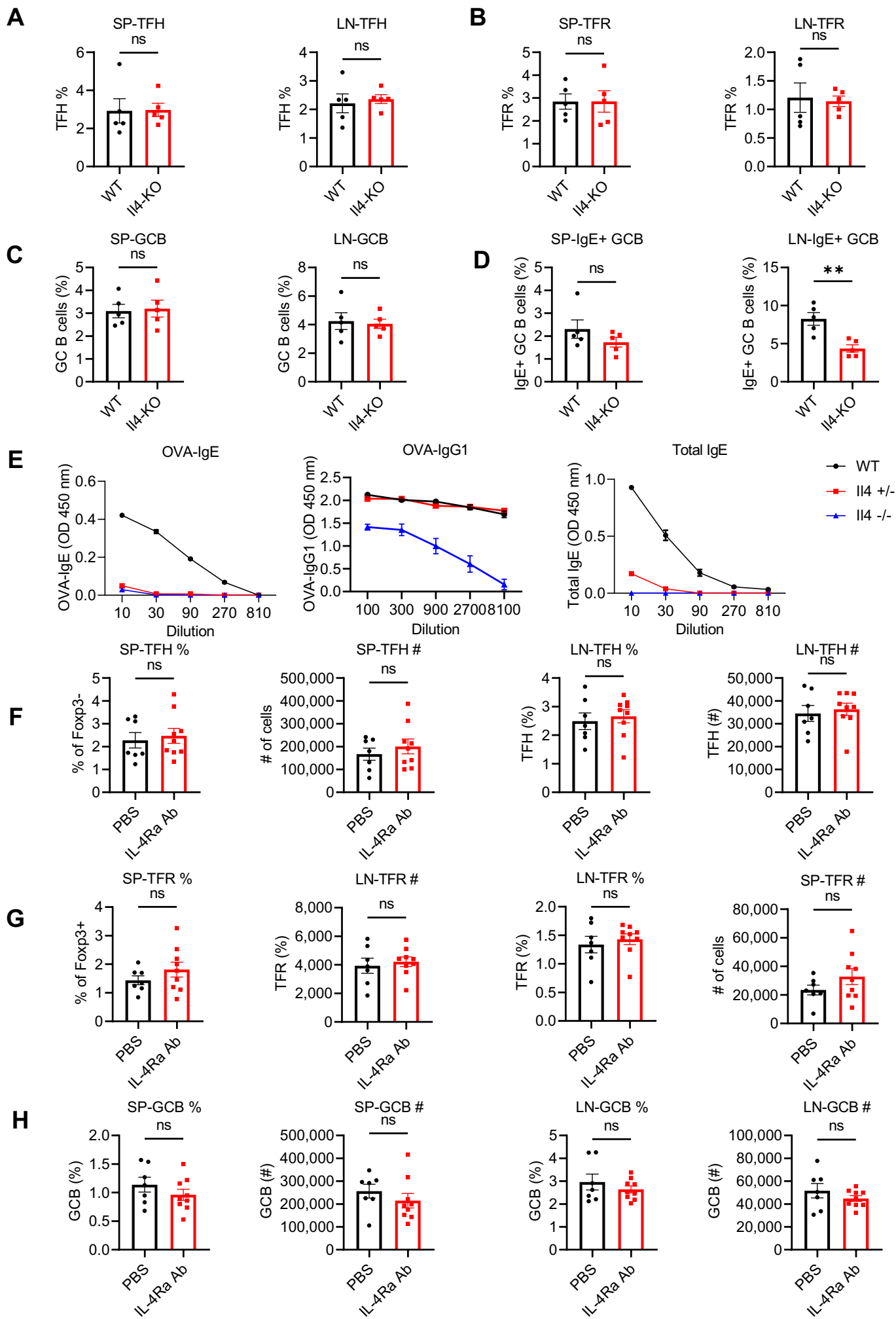
TFH-PCT      TFR-PCT



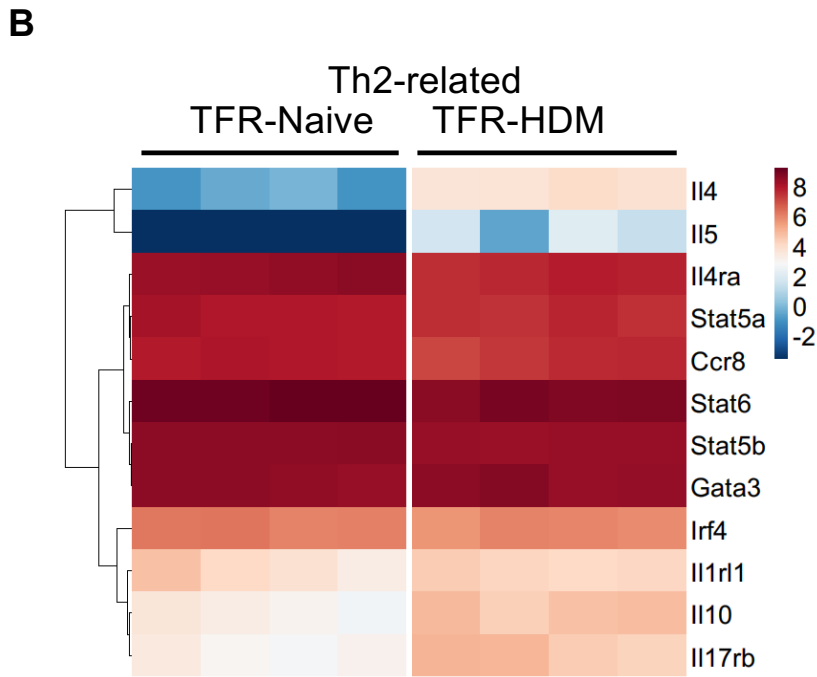
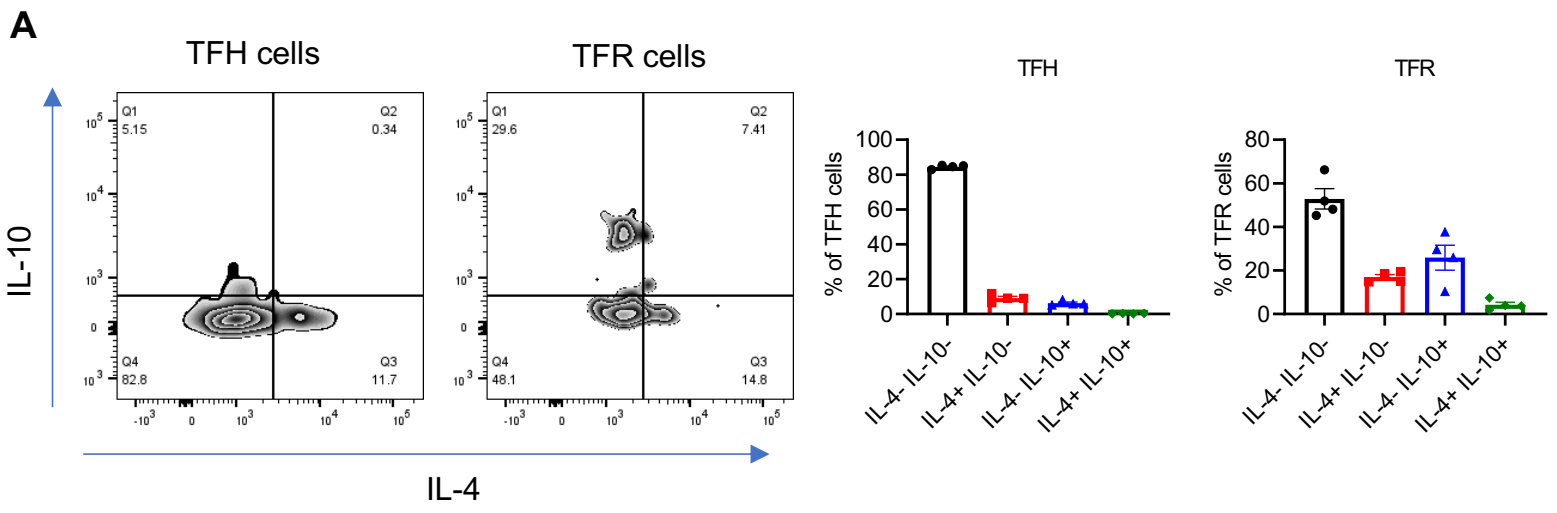
Sup. Fig. 3



Sup.  
Fig. 5

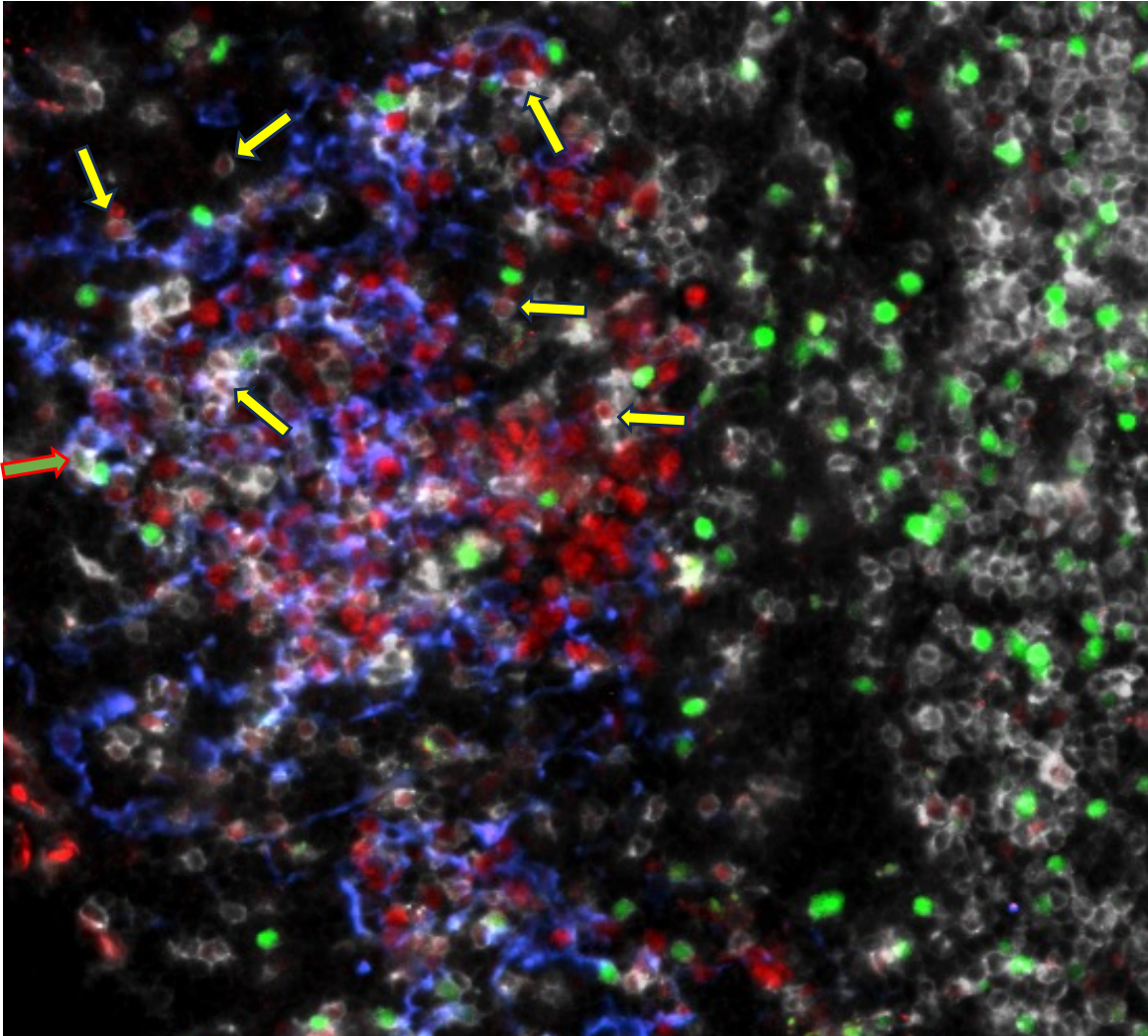




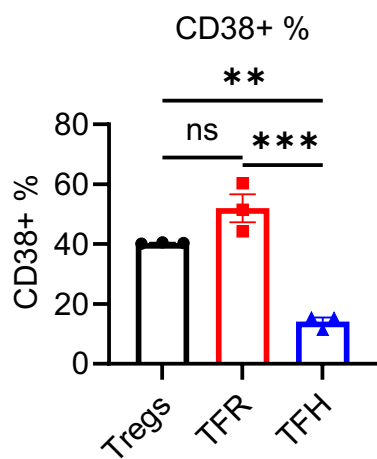


Sup. Fig. 6

A



B



Sup. Fig. 7